

Table I (Continued)

Compd	Formula	Mp, °C (recrystn solvent)	Yield, %	Antileukemic activity							
				P388				L1210			
				Dose, mg/kg	Sur- vival	Wt diff	T/C, %	Dose, mg/kg	Sur- vival	Wt diff	T/C, %
9	C ₂₂ H ₂₃ NO ₅	140–142	83	160	6/6	–1.6	59				
				80	6/6	–1.0	90				
				40	6/6	+2.0	100				
				20	6/6	+0.5	154				
				10	6/6	–0.2	173				
10a	C ₂₁ H ₂₄ INO ₄	158–160 dec (MeOH–Et ₂ O)	83	5	6/6	–0.5	127	400	1/6		
								200	4/6	+0.5	95
								100	6/6	+0.8	98
								50	12/12	–0.7	106
								25	6/6	+0.5	102
10b	C ₂₂ H ₂₆ BrNO ₄	165–167 dec (EtOH–Et ₂ O)	89	200	0/6			12.5	6/6	–0.0	104
				100	5/6	–0.5	104	200	2/6		
				50	6/6	+0.7	90	100	6/6	–0.2	98
								50	5/6	–0.4	115
								25	6/6	–0.4	94
11	C ₂₄ H ₂₇ NO ₅ S	277–279 dec (MeOH)	68					200	0/6		
								100	10/12	–4.1	116
								50	6/6	–5.6	107
								25	5/6	–4.6	104

^aFor general screening procedure and data interpretation, cf. R. I. Geran, N. H. Greenberg, M. M. MacDonald, A. M. Schumacher, and B. J. Abbott, *Cancer Chemother. Rep.*, **3** (2), 1 (1972); Instruction Booklet 14, "Screening Data Summary Interpretation," Drug Research and Development, Chemotherapy, National Cancer Institute, Bethesda, Md., 1972. ^b1–2 cures in groups of six to ten mice were occasionally observed at doses of 25–200 mg/kg. ^c1–3 cures against B16 melanocarcinoma in groups of ten mice were noted at doses of 12.5–200 mg/kg. ^dThe corresponding chloride salt was prepared in 85% yield, mp 292–294° dec. ^eCell culture (KB): ED₅₀ at 1.0 × 10⁻¹ μg/ml. Activity confirmed. ^fOne cure each in one group of six mice was observed at doses of 12.5 and 8.33 mg/kg. ^gThe corresponding chloride salt was prepared in 85% yield, mp 252–254° dec. ^hTwo cures in one group of six mice were observed at dose of 100 mg/kg. ⁱThe corresponding chloride salt was prepared in 85% yield, mp 235–237° dec. ^jCell culture (KB): ED₅₀ at 2.2 × 10⁻¹ μg/ml. Activity confirmed. ^kThe corresponding nitrate salt was prepared in 80% yield, mp 303–305° dec.

ine and CH₃I in a manner similar to that described for 10b. *Anal.* (C₂₁H₂₄INO₄) C, H, N.

5,6-Dihydro-8-methyl-2,3,10,11-tetramethoxydibenzo[a,g]-quinolizinium acetosulfate (11), mp 277–279° dec, was prepared from dihydropapaverine (5, R₁, R₂, R₃, R₄ = OCH₃) in a manner similar to that used for the preparation of coralyne. Uv absorption of this compound is similar to that of dehydro-α-coralydine.¹¹ *Anal.* (C₂₄H₂₇NO₅S) C, H, N.

Acknowledgment. This investigation was supported by Contract N01-CM-33743 with Drug Research and Development, Division of Cancer Treatment, National Cancer Institute of National Institutes of Health. The authors thank Dr. Harry B. Wood, Jr., for his interest and encouragement. Thanks are also due to Mr. John R. Gravatt, Mrs. Margaret L. Rounds, and Mr. George W. Vaughn for performing instrumental measurements.

References

- (1) K. Y. Zee-Cheng and C. C. Cheng, *J. Pharm. Sci.*, **61**, 969 (1972).
- (2) K. Y. Zee-Cheng and C. C. Cheng, *J. Pharm. Sci.*, **59**, 1630 (1970).
- (3) N. T. LeQuang Thuan and J. Gardent, *C. R. Acad. Sci., Ser. C*, **267**, 1340 (1968).
- (4) W. E. McEwan and R. I. Cobb, *Chem. Rev.*, **55**, 511 (1955).
- (5) B. C. Uff and J. R. Kershaw, *J. Chem. Soc. C*, **666** (1969).
- (6) W. Schneider and E. Nitze, *Ber.*, **56**, 1036 (1923).
- (7) K. Y. Zee-Cheng and C. C. Cheng, *J. Pharm. Sci.*, **62**, 1572 (1973).
- (8) J. Axelrod and R. Tomchick, *J. Biol. Chem.*, **233**, 702 (1958).
- (9) W. F. Herblin, *Anal. Biochem.*, **51**, 19 (1973).
- (10) B. C. Pal, *J. Sci. Ind. Res., Sect. A*, **17**, 270 (1958).
- (11) V. Preininger, L. Hruban, V. Šimánek, and F. Šantavý, *Collect. Czech. Chem. Commun.*, **35**, 124 (1970).

9,11-Seco Steroids. An Attempt to Separate Biological Activities via Ring Cleavage

Leland J. Chinn,* John H. Dygos, Stanley E. Mares, Richard L. Aspinall, and Robert E. Ranney

Departments of Chemical, Biological, and Metabolic Research, Searle Laboratories, Division of G. D. Searle & Co., Chicago, Illinois 60680. Received September 27, 1973

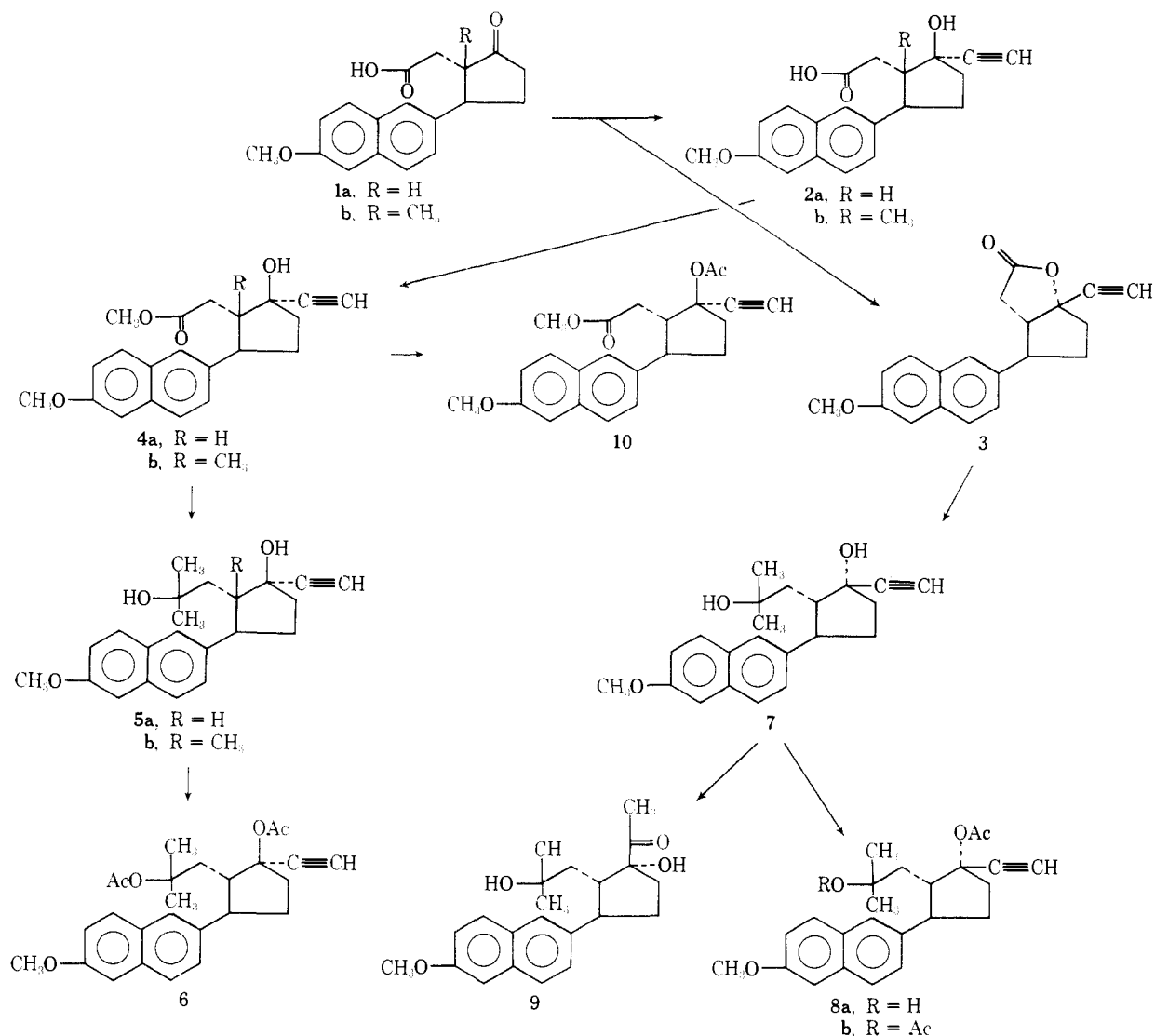
A series of seco steroids with rings A and B aromatic was synthesized in order to determine the effect which scission of the 9,11 bond has on biological activities. One compound, **5a**, was found to have antiestrogen, antifertility, and antiinflammatory properties, with an estrogenic activity approximately 0.003% that of estrone.

Molecular modification has been employed as a means of enhancing specific biological activities while reducing or eliminating less desirable effects. Application of this process has led to the successful development of potent

antiinflammatory steroids,¹ anabolic agents with reduced androgenic properties,² orally active progestins,³ and long acting estrogens.⁴

Of the four classes of steroid hormones (corticosteroids,

Scheme I



androgens, progestins, and estrogens), the estrogens are structurally the least specific. A variety of nonsteroidal substances show feminizing activity.⁵

The naturally occurring steroid estrogens have also been reported to possess hypocholesterolemic, antiinflammatory, and calcium-retaining effects.⁶ These effects are useful in the treatment of certain diseases. However, the potent feminizing property of the estrogens often militates against their use as therapeutic agents.

Suppression of the feminizing effect of the estrogens without concomitant loss of other biological activities would be highly desirable. As an extension of work which had previously been reported,⁷ we prepared a series of 9,11-seco steroids in order to determine whether rupture of the 9,11 bond would produce the desired result.

It was our hope that ring cleavage ("entropy effect"⁸) would alter the substrate-receptor interaction in the uterus⁹ as to provide a favorable separation of activities.

Each of the compounds studied contained a *gem*-dimethyl function at C-11 (steroid numbering). The purpose of inserting two methyl groups into that position was to prevent the molecule from adopting the nearly planar conformation of the natural estrogens, thereby reducing the possibility of the compound being estrogenic.

The starting substance, *trans*-2-(6-methoxy-2-naphthyl)-5-oxocyclopentaneacetic acid (1a), was treated with ethynylmagnesium bromide to afford a mixture of *trans-anti*-

2-(6-methoxy-2-naphthyl)-5-ethynyl-5-hydroxycyclopentaneacetic acid (2a)[†] and *trans-syn*-2-(6-methoxy-2-naphthyl)-5-ethynyl-5-hydroxycyclopentaneacetic acid lactone (3) in a ratio of 2:3. In contrast, the reaction of ethynylmagnesium bromide with the homologous 1-methylketo acid 1b proceeded stereoselectively to give the *trans-anti* acid 2b as the sole product.¹⁰ Esterification of 2a followed by treatment with methylmagnesium bromide yielded *trans-anti*-2-(2-hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol (5a). In a similar manner, 2b afforded 5b.¹⁰ When 5a was heated at 95° with acetic anhydride and pyridine, the diacetate 6 was obtained.

Conversion of the lactone 3 to *trans-syn*-2-(2-hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol (7) was accomplished directly with methylmagnesium bromide. Compound 7 was converted into a mixture of the monoacetate 8a and the diacetate 8b by warming at 60° with acetic anhydride and pyridine.

Hydration of the triple bond of the *trans-syn*-ethynylcyclopentanol 7 furnished *trans-syn*-1-acetyl-2-(2-hydroxy-2-methylpropyl)-3-(6-methoxy-2-naphthyl)cyclopentanol (9) (Scheme I).

The estrogenicity of the *seco* steroids was determined in

[†]In this paper the configuration is designated on the basis of the relative spatial orientation of the following three groups attached to the cyclopentane ring and in this order: methoxynaphthyl ring, hydroxyalkyl or the acetic acid moiety, and hydroxyl group. Unless specified otherwise the compounds are racemic.

immature (22–25 days old) mice. A total of 1 mg of each compound was administered subcutaneously in corn oil over a period of 3 days. Increase in uterine weight in the 72-hr interval was employed as the index for activity. A compound inactive at this dose has less than 0.01% the activity of estrone.¹¹

All of the compounds were tested for estrogenic activity. Only two compounds, namely, 6 and 9, showed activity at the 1-mg dose level. In view of their estrogenic response, the ability of these two compounds to antagonize estrone-stimulated uterine growth was also studied. Only 9 showed estrogen antagonism when given subcutaneously in oil at a total dose of 1 mg in 3 days simultaneously with a total dose of 0.3 μ g of estrone. Estrogen-stimulated uterine growth can be antagonized by progesterone;¹² the antiestrogen activity of 9 corresponds to 5–10% that of progesterone.

While 6 and 9 possess side chains which are characteristic of the progestins, neither compound showed progestational or antiprogestational activity in immature female rabbits primed with 5 μ g of 17 β -estradiol daily for 6 days.¹³

Although the diacetate 6 was inactive as an estrogen antagonist, the corresponding diol 5a was found to be active. In a control group of ten mice, each treated only with 0.3 μ g of estrone, the average uterine weight was 74.2 mg. Subcutaneous administration of a total of 1 mg of 5a, as described above, reduced the average uterine weight to 52.9 mg of the estrone-treated animals (ten in the group). To establish whether this effect might possibly be due to 5a being an agonist, the estrogenicity of 5a was determined at 2 and 4 mg, as well as at the 1-mg dose level. The diol 5a was found to be active at 4 mg but not at 1 or 2 mg. Surprisingly, the optically active, homologous diol 5b,¹⁰ which possesses the same stereochemistry as the natural estrogens, was inactive as an estrogen or as an antiestrogen at 1 mg.

The diol 5a displayed antifertility activity in reducing the number of rats with normal-size fetuses 15 days after mating when these rats were given 4 mg/day of the compound daily for 7 days following mating.¹⁴ No other compounds in this series showed this activity.

When given 4 mg/day of 5a subcutaneously, only one out of five rats became pregnant. At the same dose level but administered orally, 5a prevented pregnancy in three out of five rats. In the untreated group of animals, 80% or more were pregnant. The ED₅₀ values for 5a are 3.1 (subcutaneously) and 3.3 mg (orally). The corresponding subcutaneous and oral values for estrone are 3 and 360 μ g, respectively.

5a showed neither progestational or antiprogestational activity. However, it did exhibit antiinflammatory activity. It was found to be active in the carrageenin-induced foot edema rat assay¹⁵ at 40 mg/kg and in the mycobacterium butyricum-induced polyarthritic, hypersensitive rat test¹⁶ at 25 mg/kg.

With the exception of the acetoxy methyl ester 10, which was obtained by acetylating 4a, none of the compounds in this series was found to be active in reducing the plasma cholesterol level of male rats made hypercholesterolemic and hypothyroid with propylthiouracil¹⁷ when the test compound was administered at 10 mg/kg daily for 10 days.

In the eight animals employed as control, the average plasma cholesterol concentration was 86.7 mg %. The value for the eight rats treated orally with compound 10 was 78.7 mg %.

Although the present study cannot be regarded as definitive, the results appear sufficiently interesting to us

that we have undertaken further studies on the seco steroids.

Experimental Section†

Melting points were determined on a Fisher-Johns melting block and are corrected. Nmr spectra were taken on a Varian A-60 instrument in deuteriochloroform (unless specified otherwise) with tetramethylsilane as the standard.

(\pm)-*trans-anti-2-(6-Methoxy-2-naphthyl)-5-ethynyl-5-hydroxycyclopentaneacetic Acid* (2a) and (\pm)-*trans-syn-2-(6-Methoxy-2-naphthyl)-5-ethynyl-5-hydroxycyclopentaneacetic Acid Lactone* (3). To 820 ml of redistilled THF saturated with acetylene was added with stirring at 20° 330 ml of 3 M EtMgBr in THF over a period of 75 min. Acetylene was continuously passed into the mixture. The reaction mixture was cooled in an ice bath and while acetylene was continually being passed into the mixture, a solution of 16.4 g of *trans-2-(6-methoxy-2-naphthyl)-5-oxocyclopentaneacetic acid* (1a)⁷ in 190 ml of THF was added portionwise over a period of 50 min. The reaction mixture was stirred in the ice bath for an additional 2 hr and then allowed to come to room temperature over the course of 15 hr with stirring. The reaction mixture was concentrated under reduced pressure. The residue was acidified with 300 ml of 7 N H₂SO₄, and the resultant mixture was extracted with ether. The ether extract in turn was extracted with successive portions of H₂O, 5% NaHCO₃, and H₂O again. The ether solution was dried (Na₂SO₄) and distilled to dryness under reduced pressure to afford ca. 9.7 g of the crude lactone 3. The crude product was chromatographed on 1500 g of SiO₂. The column was eluted with C₆H₆-EtOAc. Elution with 1% EtOAc in C₆H₆, followed by crystallization from pentane, gave 6.8 g of 3, mp 78–80°. Another crystallization from pentane raised the melting point to 82–84°; nmr (Hz) multiplet 425.5–468, 233, 164.5. *Anal.* (C₂₀H₁₈O₃) C, H.

The combined bicarbonate extracts were acidified with 6 N HCl. The acidified mixture was extracted with ether. The ether extract was washed with H₂O, dried (Na₂SO₄), and distilled to dryness under reduced pressure to afford ca. 6.8 g of the crude acid 2a. The crude acid was chromatographed on 980 g of SiO₂. The column was eluted with C₆H₆-EtOAc. Elution with 5% EtOAc in C₆H₆, followed by crystallization from chloroform-pentane, furnished 5.3 g of the acid 2a: mp 132–134°; nmr (Hz) multiplet 423–474, 233.5, 158.

(\pm)-*Methyl trans-anti-2-(6-Methoxy-2-naphthyl)-5-ethynyl-5-hydroxycyclopentaneacetate* (4a). To a solution of 889 mg of the acid 2a in 15 ml of ether, cooled in an ice bath, was added 20 ml of an ethereal solution of diazomethane (prepared from 3.5 g of *N*-nitrosomethylurea, 40 ml of ether, 7 g of KOH, and 7 ml of H₂O). The reaction mixture was allowed to stand at 5° for 3 hr. Then it was evaporated to dryness to afford a viscous yellow oil. The viscous oil was crystallized from ether-pentane to afford 565 mg of the ester 4a: mp 102.5–105°; nmr (Hz) multiplet 427–468, 235, 214, 158.5.

(\pm)-*trans-anti-2-(2-Hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol* (5a). To 30 ml of 1 M MeMgBr in diethyl ether, stirred and heated under reflux, was added over a period of 5 min a solution of 443 mg of the ester 4a in 30 ml of ether. The reaction mixture was stirred and heated under reflux for 7 hr. The cooled reaction mixture was treated with a saturated solution of NH₄Cl. The ether phase was separated, washed successively with H₂O and saturated NaCl, dried (Na₂SO₄), and distilled to dryness under reduced pressure. The residual viscous yellow oil was crystallized from ether-pentane to afford 396 mg of a crystalline product, mp 164.5–167.5°. Two more crystallizations from ether-pentane gave 318 mg of 5a: mp 167.5–168°; nmr [Hz, (CD₃)₂SO] multiplet 424–473, 234, 207, 201 (one of the two signals belongs to H₂O), 64, 56. *Anal.* (C₂₂H₂₆O₃) C, H.

(\pm)-*trans-anti-2-(2-Hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol Diacetate* (6). A solution of 880 mg of the diol 5a, 10 ml of C₅H₅N, and 10 ml of Ac₂O was maintained at 95° for 36 hr in an N₂ atmosphere. The cooled reaction mixture was poured into ice H₂O. The resultant mixture was extracted with ether. The ether extract was washed with H₂O, dried (Na₂SO₄), and evaporated to a small volume. The residue was diluted with hexane whereupon the product 6 crystallized out of the mixture: yield 887 mg; mp 119.5–121.5°. Another crystallization from ether-hexane raised the melting point to 122–123°; nmr (Hz) multiplet 426–467.5, 233.5, 165, 124, 95.5, 79, 77. *Anal.* (C₂₆H₃₀O₅) C, H.

(\pm)-*trans-syn-2-(2-Hydroxy-2-methylpropyl)-1-ethynyl-3-(6-*

methoxy-2-naphthyl)cyclopentanol (7). To 210 ml of 1 M MeMgBr in diethyl ether, stirred and heated under reflux, was added over a period of 25 min a solution of 2.93 g of the lactone 3 in 200 ml of ether. The reaction mixture was stirred and heated under reflux for 3.5 hr. The cooled reaction mixture was treated with a dilute solution of NH₄Cl. The ether phase was separated, washed successively with H₂O and saturated NaCl, dried (Na₂SO₄), and concentrated to a small volume by distillation under reduced pressure. The residue, which contained a crystalline product, was allowed to stand at 0° for 1 hr. The solid was collected by filtration: yield 1.78 g; mp 178–180°. From the mother liquor, an additional 0.67 g of 7, mp 173–184.5°, was obtained. Crystallization of 7, mp 178–180°, from acetone–hexane raised the melting point to 179.5–182.5°; nmr [Hz, (CD₃)₂SO] multiplet 422–471.5, 232.5, 199.5, 60.5, 49. *Anal.* (C₂₂H₂₆O₃) C, H.

(±)-*trans-syn*-2-(2-Hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol 1-Acetate (8a) and (±)-*trans-syn*-2-(2-Hydroxy-2-methylpropyl)-1-ethynyl-3-(6-methoxy-2-naphthyl)cyclopentanol Diacetate (8b). A mixture of 412 mg of the diol 7, 5 ml of C₆H₅N, and 5 ml of Ac₂O was maintained at 60° in an N₂ atmosphere for 36 hr. The cooled reaction mixture was poured into ice H₂O and rubbed. The resultant gum was collected by filtration, washed with H₂O, and dried. Then it was crystallized from ether–pentane to afford 328 mg of a crystalline product, mp 119.5–130°.

A 249-mg sample of this product was chromatographed on 8 g of SiO₂. The column was eluted with C₆H₆–EtOAc. Elution with 10% EtOAc in C₆H₆ gave initially a solid product which was crystallized from ether–pentane to afford 68 mg of the diacetate 8b, mp 135.5–137°. Further crystallization from ether–pentane raised the melting point to 137–138.5°; nmr (Hz) multiplet 424.5–466, 234.5, 158.5, 127, 93, 76. *Anal.* (C₂₈H₃₀O₅) C, H.

Continued elution of the column with 10% EtOAc in C₆H₆ gave a second solid product which was crystallized from ether–pentane to afford 45 mg of 8a, mp 137–138.5°. Admixed with 8b, it melted at 124–137.5°; nmr (Hz) multiplet 425–468, 234.5, 164, 127.5, 61, 53. *Anal.* (C₂₄H₂₈O₄) C, H.

When a mixture of 1.8 g of the diol 7, 30 ml of C₆H₅N, and 30 ml of Ac₂O was heated under reflux in an N₂ atmosphere for 6.5 hr and worked up in the usual manner, 1.36 g of the diacetate 8b was obtained: mp 134–138°.

(±)-*trans-syn*-1-Acetyl-2-(2-hydroxy-2-methylpropyl)-3-(6-methoxy-2-naphthyl)cyclopentanol (9). A mixture of 285 mg of the ethynyl diol 7, 70 mg of HgO, 6 ml of MeOH, and 0.4 ml of BF₃–Et₂O was stirred at room temperature for 2 hr. Afterward a 2-ml solution of aqueous–methanolic HCl (prepared from 2 ml of 6 N HCl, 13 ml of H₂O, and 7 ml of MeOH) was added to the reaction mixture. Stirring at room temperature was continued for an additional 1.5 hr. The reaction mixture was then diluted with 30 ml of 2% NaHCO₃. The resultant mixture was allowed to stand at 0° for 1 hr. The gum which formed was collected by filtration, washed with H₂O, and dried. Crystallization from ether–pentane afforded 145 mg of 9, mp 114.5–117°. Another crystallization from ether–pentane raised the melting point to 116–117.5°; λ^{KBr} 2.82, 2.96, 5.87, 6.13, 6.23 μ. *Anal.* (C₂₂H₂₈O₄) C, H.

(±)-Methyl *trans-anti*-2-(6-Methoxy-2-naphthyl)-5-ethynyl-5-acetoxycyclopentaneacetate (10). A solution of 1.5 g of the hydroxy ester 4a, 12 ml of C₆H₅N, and 12 ml of Ac₂O was maintained at 60° for 66 hr in an N₂ atmosphere, after which time it was poured into ice H₂O. The resultant solid was collected by filtration, washed with water, and dried: yield 1.52 g; mp 143.5–145.5°. Crystallization from ether–pentane afforded 1.32 g of 10: mp 148.5–149°; λ^{KBr} 3.03, 4.72, 5.73, 6.12, 6.22, 8.07 μ. *Anal.* (C₂₃H₂₄O₅) C, H.

Acknowledgment. We gratefully acknowledge the technical assistance of Messrs. Gatis Plume and Mark Wenger and Miss Mary Ann Oram in the preparation of some of the seco steroids and of Mr. Wally Pautsch and Mrs. Esther Muir and their staffs in the biological evaluation of the compounds.

References

- (1) (a) L. Mantica, R. Ciceri, J. P. Cassagne, and E. Mascietti-Coriandoli, *Arzneim.-Forsch.*, **20**, 109 (1970); (b) G. E. Arth, J. Fried, D. B. R. Johnston, D. R. Hoff, L. H. Sarett, R. H. Silber, H. C. Stoerk, and C. A. Winter, *J. Amer. Chem. Soc.*, **80**, 3161 (1958); (c) J. Fried, A. Borman, W. B. Kessler, P. Grabowich, and E. F. Sabo, *ibid.*, **80**, 2338 (1958).
- (2) (a) R. M. Scribner, R. I. Dorfman, and W. H. Rooks II, *J. Med. Chem.*, **13**, 952 (1970); (b) A. Boris, R. H. Stevenson, and T. Trmal, *Steroids*, **15**, 61 (1970).
- (3) (a) E. J. Bailey, H. Fazakerley, M. E. Hill, C. E. Newall, G. H. Phillipps, L. Stephenson, and A. Tulley, *J. Chem. Soc. D*, 106 (1970); (b) J. S. Baran, H. D. Lennon, S. E. Mares, and E. F. Nutting, *Experientia*, **26**, 762 (1970).
- (4) (a) U. K. Banik, C. Revesz, and F. Herr, *Steroids*, **16**, 289 (1970); (b) J. A. Epstein, *Int. J. Fert.*, **12**, 181 (1967); (c) T. Giannina, B. G. Steinetz, and A. Meli, *ibid.*, **12**, 142 (1967).
- (5) (a) D. Lednicer, J. C. Babcock, S. C. Lyster, J. C. Stucki, and G. W. Duncan, *Chem. Ind. (London)*, 2098 (1961); (b) F. W. Schuele, *Science*, **103**, 221 (1946); (c) E. C. Dodds, L. Golberg, N. Lawson, and R. Robinson, *Nature (London)*, **141**, 247 (1938); (d) R. J. Boscott, "The Ovary," S. Zuckerman, Ed., Vol. 2, Academic Press, New York, N. Y., 1962, p 1.
- (6) (a) R. W. Robinson, N. Higano, W. D. Cohen, R. C. Sniffen, and J. W. Sherer, *Circulation*, **14**, 365 (1965); (b) A. S. Spangler, H. N. Antoniadis, S. L. Sofman, and T. M. Inderbitzin, *J. Clin. Endocrinol. Metab.*, **29**, 650 (1969); (c) M. N. Mueller and A. Kappas, *Proc. Soc. Exp. Biol. Med.*, **117**, 845 (1964); (d) P. H. Penneman and S. Wallach, *Arch. Int. Med.*, **100**, 715 (1957).
- (7) L. J. Chinn, E. A. Brown, R. A. Mikulec, and R. B. Garland, *J. Org. Chem.*, **27**, 1733 (1962).
- (8) L. H. Sarett, A. A. Patchett, and S. L. Steelman, "Drug Research," E. Jucker, Ed., Vol. 5, Birkhauser Verlag, Basel, Switzerland, 1963, p 11.
- (9) E. V. Jensen, H. L. Jacobson, J. W. Flesher, N. N. Saha, G. N. Gupta, S. Smith, V. Colucci, D. Shiplacoff, H. G. Neumann, E. R. DeSombre, and P. W. Jungblut, "Steroid Dynamics," G. Pincus, T. Nakao, J. Tait, Ed., Academic Press, New York, N. Y., 1966, p 133.
- (10) J. H. Dygos and L. J. Chinn, *J. Org. Chem.*, **38**, 4319 (1973).
- (11) (a) B. L. Rubin, A. S. Dorfman, L. Black, and R. I. Dorfman, *Endocrinology*, **49**, 429 (1951); (b) R. A. Edgren, *Proc. Soc. Exp. Biol. Med.*, **92**, 569 (1956).
- (12) R. A. Edgren, R. C. Jones, and D. L. Peterson, *Fert. Steril.*, **18**, 238 (1967).
- (13) (a) C. W. Emmens, "Hormone Assay," C. W. Emmens, Ed., Academic Press, New York, N. Y., 1950, p 422; (b) C. Clauberg, *Zentralbl. Gynaekol.*, **54**, 2757 (1930).
- (14) R. A. Edgren, D. Peterson, M. A. Johnson, and G. C. Shipley, *Fert. Steril.*, **12**, 172 (1961).
- (15) C. A. Winter, E. A. Risley, and G. N. Nus, *Proc. Soc. Exp. Biol. Med.*, **111**, 544 (1962).
- (16) R. L. Aspinall and P. S. Cammarata, *Nature (London)*, **224**, 1320 (1969).
- (17) R. E. Counsell, P. D. Klimstra, R. E. Ranney, and D. L. Cook, *J. Med. Pharm. Chem.*, **5**, 720 (1962).