Bisnorpenicillin V

the two hydroxyl groups for the estrogenic potency of diethylstilbestrol.^{20,21} Replacement of one of the hydroxyl groups in diethylstilbestrol by an amino group had been shown to reduce the estrogenic activity 25-fold, and larger substitutions (methoxy or bromine) caused much greater reductions in activity.¹⁰

Electrophilic Reactivity. The reaction of esters of Narylacethydroxamic acids with methionine-methyl- ${}^{3}H$ to yield sulfonium derivatives which decompose to benzenesoluble [3H]methylmercaptoarylamides has served as a convenient assay for the electrophilic reactivity of these compounds.¹⁴ Under our assay conditions 0.65% of the ³H in methionine-methyl- ${}^{3}H$ was converted to a benzenehexane-soluble form on incubation with N-acetoxy-4'-hydroxy-7,7'-diethyl-N-4-trans-stilbenylacetamide. Under the same conditions reactions of 0.58 and 5.3% were obtained with N-acetoxy-N-4-trans-stilbenylacetamide or N-acetoxy-N-2-fluorenylacetamide, respectively. From the similar activity of 11 and N-acetoxy-N-4-stilbenylacetamide toward methionine, 11 might be expected to be reactive toward certain nucleophilic sites in cellular macromolecules. Carcinogenicity tests on N-hydroxy- and N-acetoxy-4'-hydroxy-7,7'-diethyl-N-4-trans-stilbenylacetamide are in progress.

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Total Synthesis of Bisnorpenicillin V

Jos Hoogmartens, Paul J. Claes, and Hubert Vanderhaeghe*

Rega and Pharmaceutical Institute, University of Leuven, B-3000 Leuven, Belgium. Received November 13, 1973

Bisnorpenicillin V, an analog of penicillin V in which the two methyl groups are replaced by hydrogen atoms, was prepared by total synthesis. The reaction scheme was essentially that described by Sheehan, *et al.*, for the synthesis of penicillins. The antibacterial activity of bisnorpenicillin was lower than that observed for the parent penicillin V. Replacement of the two C-2 methyl groups of penicillin has no or little effect on its sensitivity to β -lactamase.

Several modifications at the C-2 position of penicillins have been reported recently.¹⁻⁴ These modifications concern the introduction of an acetoxy group or a halogen into one of the C-2 methyl groups. The present report deals with the synthesis of bisnorpenicillin V (1a), a penicillin V analog in which the two C-2 methyl groups are replaced by hydrogen atoms.[†] From earlier work⁶ in the field of total synthesis of penicillins, one would expect bisnorpenicillins to be biologically active. Antibacterial activity was found for the reaction mixture of pL-cysteine with 2-benzyl-4-methoxymethylene-5(4)-oxazolone, a condensation which yields penicillin in very low yields.

For the preparation of pure 1a the two reaction schemes reported by Sheehan, *et al.*, 7,8 for the synthesis of penicillin V (1b) were followed. Both reaction sequences, out-

lined in reaction schemes A and B, start with *tert*-butyl (3carboxy-5-thiazolidine)phthalimidoacetate (2a).[‡] In order to obtain a biologically active penicillin, it is necessary to use the isomer of 2a with the configuration of natural penicilloate. This isomer can be obtained by condensation of *tert*-butyl phthalimidomalonaldehyde⁹ (3) with D-cysteine in aqueous ethanol containing sodium acetate. The reaction proceeds in a similar way as reported for the condensation of 3 with D-penicillamine⁷ and afforded a mixture of two diastereoisomers of 2a in a ratio of about 5:1. In analogy to the synthesis of 2b,⁷ the natural configuration was assigned to the minor component, which will be designated as α isomer (the major component as γ isomer). The α and γ isomers can be differentiated by tlc (silica gel, solvent system I). In both cases the α isomer showed

‡To avoid confusion the penam numbering will be used in this paper for the thiazolidine nucleus of compounds related to penicillin.

[†]According to the nomenclature, proposed by Sheehan, *et al.*,⁵ compound la should be called 6-phenoxyacetamidopenam-3-carboxylic acid.

Table I. Minimum Inhibitory Concentrations^a $(\mu g/ml)$ of 1a and 1b

Microorganism	1a ^b	1 b ^b
Staphylococcus aureus ATCC 6538P	0.3	0.025
Staphylococcus aureus (resistant strain)	>100	>100
Micrococcus flavius ATCC 10240	1.5	0.17
Sarcina lutea ATCC 9341	1.5	0.08
Bacillus subtilis NCTC 8236	0.35	0.17
Streptococcus faecalis NICB 8192	1.56	3.12
Escherichia coli 9661	>100	>100

^aAssayed by the agar-streak method on nutrient agar plates. ^bAs potassium salt.

the highest $R_{\rm f}$ value. In both series, the α isomer was the most dextrorotatory product. Analogy between the cysteine and penicillamine series was also observed in the nmr spectra of the benzyl esters of α and γ isomers of 2a and **2b.** In both series the H-3 signals of the γ isomers were shifted upfield (0.23 ppm) with respect to the α isomers. From an nmr study¹⁰ of various carboxythiazolidines[‡] derived from penicillamine it can be concluded that the α series have a 5(R) and the γ series a 5(S) configuration.§

Higher yields of α -2a and also of α -2b were obtained when the condensation of 3 with cysteine or penicillamine was carried out in pyridine. Reaction temperatures ranging from 40 to 110° were investigated. Optimal temperatures were 55° for α -2a and 80° for α -2b. Almost pure α isomer crystallized upon evaporation of the pyridine solution. The γ isomers, which are more soluble in pyridine, were isometized into a mixture of α and γ isomets by heating in pyridine. Thus, the total yield was 89.6% for α -2a and 67.5% for α -2b. It should be noted that the isomerization of α -2b in pyridine has been reported by Sheehan, et al.⁷

Initially, the reaction given in scheme A, which proceeded in four steps, was investigated. It appeared that the intermediate α -4a and especially the intermediates α -5a and α -6a were not stable (in contrast with the analogs in the penicillamine series) and could not be isolated in a pure state. Similar difficulties were reported with thiazane analog.¹¹ The yields observed for phenoxyacetylation of α -4a and for removal of the *tert*-butyl group of α -5a were extremely low. Therefore, this method was abandoned and the reaction sequence shown in scheme B was investigated. Thus, the isomer of tert-butyl (3-benzyloxycarbonyl-5-thiazolidino)phthalimidoacetate (α -7a), obtained in a 76% yield by esterification of α -2a with benzyl bromide in dimethylformamide, was subjected to hydrazinolysis as described for the parent α -7b.⁸ Reaction of α -8a with anhydrous hydrogen chloride (for 4 hr at 0° instead of 27 hr as reported for α -8b⁸) afforded the hydrochloride of α -9a as an amorphous powder which decomposed at room temperature. When α -9a was N-tritylated in aqueous 2-propanol or aqueous tetrahydrofuran,⁸ extensive degradation of the tritylated compound α -10a occurred. Therefore, the hydrochloride of α -9a was tritylated by reaction at -18° with trityl chloride in CH₂Cl₂ in the presence of N-ethyldiisopropylamine. The crude reaction mixture, containing α -10a, was freed from the amine and treated with diisopropylcarbodiimide (DICI) in nitromethane, yielding, after column chromatography over silica gel, crystalline benzyl 6-tritylaminobisnorpenicillanate (11a). The overall yield for the conversion of α -8a to 11a was 14.5% # It was found that nitromethane was a better

These authors concluded that the stereochemistry at C-5 affects thechemical shift of the C-3 proton. The latter appears at the highest field when C-3 and C-5 protons possess a cis configuration.

=When the conversion of α -8b to 11b was carried out, under the reaction conditions described in the present report, the overall yield which has been reported to be 15%8 was improved to 52%.

solvent for the cyclization of α -10a than aqueous dioxane, which was reported for the β -lactam closure of α -11b.

The reaction sequence used for the conversion of 11a into 1a was different from that reported⁸ for the synthesis of penicillin V (scheme B). The benzyl ester 11a was detritylated (84%) (p-toluenesulfonic acid in acetone¹²). N-Phenoxyacetylation of 12a (phenoxyacetyl chloride in CH₂Cl₂ in the presence of triethylamine) afforded (87%) 13a, which was debenzylated (Pd/C) and neutralized with KOH to yield 1a (71.5%). Spectral data of compounds 1a, 11a, 12a, and 13a were consistent with the proposed structures; ir spectra showed the presence of a β -lactam carbonyl group (1769, 1785, 1782, and 1796 cm⁻¹, respectively); the coupling constants (4-4.5 Hz) between the C-5 and C-6 protons observed in the spectra of 1a, 11a, and 13a were in agreement with a cis relationship between these protons.

series a R = H; series b R = Me; Ft = phthalimidoFtCHCHO



3

reaction sequence A

2

5

7

$$2 \xrightarrow{N,H_{t}} 4 \xrightarrow{PhOCH_{1}COC} 5 \xrightarrow{H^{+}} 6 \xrightarrow{DCCI} 1$$
reaction sequence B
$$2 \xrightarrow{B_{2}B_{1} \text{ or }} 7 \xrightarrow{N,H_{t}} 8 \xrightarrow{H^{-}} 9 \xrightarrow{Ph,CCI} 10 \xrightarrow{DICI} 11$$

$$11 \xrightarrow{Pd C} 14 \xrightarrow{H^{-}} 15 \xrightarrow{PhoCH_{2}COCI} 1$$
modification used in the present work

$$11 \xrightarrow{H^*} 12 \xrightarrow{PhOCH_COCI} 13 \xrightarrow{Pd \cdot C} 1$$

Biological Activity. The in vitro activity of bisnorpenicillin V was compared to that of the parent penicillin V. The data given in Table I indicate that replacement of the C-2 methyl groups by hydrogen atoms results in a marked decrease of the activity against penicillin-sensitive organisms. No activity was observed for 1a against a penicillin-resistant Staphylococcus aureus strain and against Escherichia coli.

It has been pointed out that the structure of penicillin is very similar to that of D-ala-D-ala fragment, which is cleaved by a penicillin-sensitive transpeptidase during the biosynthesis of peptidoglycan of bacterial cell wall.¹³ From an inspection of models one could conclude that the influence of the two C-2 methyl groups of penicillin would be negligible. These results indicate that their importance cannot be neglected.

It should be noted that the replacement of the two C-2 methyl groups of penicillin by hydrogen atoms has little or no effect on its sensitivity to β -lactamase. The rate of hydrolysis of **1a** by this enzyme is almost the same as that observed for **1b**.

Experimental Section

Melting points were determined with a Büchi-Tottoli apparatus and are uncorrected. Solvents were evaporated under reduced pressure below 30° unless otherwise stated. Tlc was performed on silica gel F-254 plates (Merck) using the following solvent systems: I, C_6H_6 -EtOAc-HCOOH (20:10:0.25); H, C_6H_6 -Me₂CO III, n-BuOAc-n-BuOH-H₂O-MeOH-HOAc (95:5):(80:15:24:5:40); IV, C₆H₆; V, C₆H₆-Me₂CO (70:30). Spots were located by uv illumination and exposure to iodine vapor. Column chromatography was performed over silica gel (Merck, 0.05-0.2 mm). Petroleum ether refers to the fraction of boiling range 40-60°. Ir spectra were run on a Perkin-Elmer 257 spectrometer. Mass spectra were recorded on a A.E.I. MS12 apparatus and nmr spectra on a Varian A-60 or XL-100 spectrometer with tetramethylsilane (TMS) or sodium 2,2-dimethyl-2-silapentane-5-sulfonate (DSSA) as internal standard. The β -lactamase used was a penicillinase (Penase) obtained from Leo Pharmaceutical Products. Ballerup, Denmark.

tert-Butyl (3-Carboxy-5-thiazolidino)phthalimidoacetate α Isomer (α -2a). A slurry of D-cysteine hydrochloride (23.625 g, 150 mmol) and tert-butyl phthalimidomalonaldehydate (43.35 g, 150 mmol) in 250 ml of anhydrous pyridine was heated for 22 hr at 55° under N_2 atmosphere. The solution was cooled and concentrated until a crystalline precipitate was observed. After storage for 24 hr at room temperature the crystals were collected, washed (Et₂O and H₂O), and dried, yielding 47.45 g (80.7%) of α -2a: mp 124-127° dec (after recrystallization from MeOH-H₂O); $[\alpha]^{25}$ D +67° (c 0.5, dioxane); mass spectrum M⁺ 406; ir (KBr) 3280 (NH), 1780, 1715 (imide), 1740, 1155 (ester), 1715, 1380 (COOH), 715 cm⁻¹ (phenyl); tlc (system I) $R_{\rm f}$ 0.21. The residue obtained on evaporation of the pyridine and Et₂O filtrates (which contained the γ isomer) was converted into the α isomer by heating (55°) in pyridine for 22 hr. This afforded another 5.26 g (8.9%) of α -2a.

 γ Isomer (γ -2a). To a solution of tert-butyl phthalimidomalonaldehydate (11.5 g, 40 mmol) in EtOH (80 ml) was added p-cysteine hydrochloride (6.3 g, 40 mmol) dissolved in 80 ml of H₂O containing NaOAc (4.95 g). After storage for 12 hr at room temperature under N₂ atmosphere, crystals of γ -2a (10.3 g, 65.7%) were collected and recrystallized from MeOH: mp 161–163° dec; [α]²⁵D +6° (c 0.5, dioxane); mass spectrum M⁺ 406; ir (KBr) 3260 (NH), 1785, 1720 (imide), 1745, 1160 (ester), 1720, 1380 (COOH), 715 cm⁻¹ (phenyl); tlc (system I) R_f 0.12. Addition of H₂O to the filtrate of the first fraction yielded 2.26 g (14.5%) of a mixture of the α and γ isomer which can be separated by fractional crystallization from methanol.

tert-Butyl (3-Carboxy-2,2-dimethyl-5-thiazolidine)phthalimidoacetate α Isomer (α -2b). D-Penicillamine (18.55 g, 100 mmol) and tert-butyl phthalimidomalonaldehyde (28.9 g, 100 mmol) in 500 ml of pyridine were heated for 22 hr at 80° under N₂ atmosphere. The reaction mixture was worked up as described for α -2a, yielding 28.87 g (67.5%) of α -2b: $[\alpha]^{25}$ D +11° (c 1, dioxane); tlc (system I) $R_{\rm f}$ 0.34.

 γ Isomer (γ -2b). This compound was prepared according to Sheehan, et al.:⁷ [α]²⁵D -13° (c 1, dioxane); tlc (system I) R_f 0.23. Other physical constants of both isomers were in agreement with the reported values.⁷

tert-Butyl (3-Carbobenzyloxy-5-thiazolidino)phthalimidoacetate α Isomer (α -7a). Freshly distilled NEt₃ (3.5 ml, 25 mmol) and freshly distilled benzyl bromide (4.5 ml, 38 mmol) were added to a solution of α -2a (7.84 g, 20 mmol) in 100 ml of dimethylformamide. The mixture was stirred for 26 hr at room temperature and poured into a mixture of 600 ml of ice-water, 100 ml of C₆H₆, and 4 ml of concentrated HCl. The C₆H₆ layer was separated and the aqueous layer extracted with C₆H₆. The combined organic layer was successively washed with NaHCO₃ (5%) and H₂O, dried (Na₂SO₄), and evaporated to a yellow oil which was crystallized from 50 ml of ether yielding 5.25 g (59.4%) of α -7a: mp 115–117°. Addition of petroleum ether to the ether filtrate afforded a second fraction (2.26 g, 16.7%). Recrystallization from Et₂O-petroleum ether increased the melting point to 117–118°: $[\alpha]^{25}_{D}$ +45° (c 1, dioxane); mass spectrum M⁺ 482; ir (KBr) 3365 (NH), 1780, 1715 (imide), 1730 (ester), 745, 710, 698 cm⁻¹ (phenyl); nmr (CDCl₃) δ 1.48 (s, *tert*-butyl), 3.06 (AB part of ABX pattern, $J_{AX} = 6$ Hz, $J_{HX} = 9$ Hz, $J_{AB} = 10$ Hz, H-2), 4.06 (br, H-3), 4.80 (d, J = 10 Hz, FtCH<), 5.18 (s, OCH₂C₆H₅), 5.38 (d, J = 10 Hz, H-5), 7.33 (s, C₆H₅), 7.6–8.0 ppm (m, C₆H₄) (after addition of D₂O, the signal at 4.06 ppm showed the typical four lines for the X part with $J_{AX+BX} = 15$ Hz); tlc (system II) R_f 0.65.

 γ Isomer. This compound was obtained in a similar way as described for the α isomer: mp 99.5–100.5°; $[\alpha]^{25}{}_{\rm D}$ –1° (c 1, dioxane); mass spectrum M⁺ 482; ir (KBr) 3280 (NH), 1780, 1720 (imide), 1750, 1195 (ester), 745, 730, 698 cm⁻¹ (phenyl); nmr (CDCl₃) δ 1.48 (s, tert-butyl), 3.08 (AB part of ABX pattern, $J_{\rm AX}$ = 6 Hz, $J_{\rm HX}$ = 10 Hz, $J_{\rm AB}$ = 10 Hz, H-2), 3.83 (br, H-3), 5.08 (d, J = 8.5 Hz, FtCH<), 5.21 (s, OCH₂C₆H₅), 5.24 (d, J = 8.5 Hz, H-5), 7.35 (s, C₆H₅), 7.6–8.0 ppm (m, C₆H₄) (after addition of D₂O, the signal at 3.83 ppm showed the typical four lines of the X part with $J_{\rm AX+BX}$ = 16 Hz); tlc (system II) $R_{\rm f}$ 0.60.

tert-Butyl (3-Carbobenzyloxy-2,2-dimethyl-5-thiazolidine)phthalimidoacetate (α - and γ -7b). Both α - and γ -2b were benzylated with PhCHN₂ as described by Sheehan, et al..¹⁴ nmr (CDCl₃) δ 1.12 (s, CH₃), 1.47 (s, tert-butyl), 1.62 (s, CH₃), 3.28 (br, NH), 3.82 (br, H-3), 4.9 (d, J = 10.5 Hz, FtCH<), 5.13 and 5.18 (AB pattern, CH₂Ph), 5.28 (d, J = 10.5 Hz, FtCH<), 5.13 (s, C₆H₅), 7.6-8.0 ppm (m, C₆H₄) for α -7b and 1.14 (s, CH₃), 1.46 (s, tert-butyl), 1.6 (s, CH₃), 3.36 (d, J = 12 Hz, NH), 3.59 (d, J =12 Hz, H-3), 5.02 (d, J = 8 Hz, FtCH<), 5.14 and 5.18 (AB pattern, CH₂Ph), 5.34 (d, J = 8 Hz, H-5), 7.34 (s. C₆H₅), 7.6-8.0 ppm (m, C₆H₄) for γ -7b (after addition of D₂O, the H-3 signals gave a sharp singlet in both spectra, H-5 signals were also sharper). Other physical constants were in agreement with the reported values.

tert-Butyl (3-Carbobenzyloxy-5-thiazolidino)aminoacetate Hydrochloride (α -8a). Removal of the phthalyl group of α -7a was performed as described for the parent α -7b.⁸ The product obtained on treatment of α -7a (9.24 g, 20 mmol) with hydrazine and decomposition of the resulting phthalhydrazide complex with HCl was crystallized from MeOH-Et₂O yielding 6.98 g (76.3%) of crystalline α -8a: mp 141-142.5° dec; $[\alpha]^{25}$ D +84° (c 1, MeOH); ir (KBr) 3300 (NH), 2850 (NH₃⁺), 1740, 1205, 1720, 1190 (ester). 755, 700 cm⁻¹ (phenyl); tlc (system III) R_f 0.72.

Benzyl 6-Tritylaminobisnorpenicillanate (11a). A stream of anhydrous HCl was passed for 6 min through a cooled (0°) and stirred suspension of α -8a (3.495 g, 9 mmol) in CH₃NO₂ (60 ml). After a few minutes the solid material dissolved. The solution was stored for 4 hr at 0°, the major part of the HCl was removed under reduced pressure at 10°, and anhydrous Et₂O (120 ml) was added to the stirred solution. After 45 min the precipitate of α -9a was filtered off, washed (Et₂O), and dried (3 hr) over P₂O₅ and KOH. Ph₃CCl (7.56 g, 27 mmol) was added to the precipitate followed by a solution of N-ethyldiisopropylamine (9.36 g, 72 mmol) in CH₂Cl₂ (120 ml) and the reaction mixture was cooled (Dry Ice) immediately. After storage for 20 hr at -13° (until this stage, contact with humidity was avoided as much as possible) the solution was diluted with $CHCl_3$, poured into ice-water (150 ml). and adjusted to pH 6 with dilute H₃PO₄. The organic layer containing α -10a was washed (H₂O), dried (Na₂SO₄) for 0.5 hr, and evaporated at low temperature to an oil which was taken up in CH₃NO₂ (60 ml) containing diisopropylcarbodiimide (2.925 g, 22.5 mmol). After storage for 19 hr at room temperature the solvent was evaporated. Chromatography of the residue over silica gel (110 g) using benzene as eluent yielded after crystallization from C₆H₆-CHCl₃-Et₂O 680 mg (14%) of 11a: mp 196-197.5° dec; $[\alpha]^{25}_{D}$ +117° (c 1, CHCl₃); mass spectrum M⁺ 520; ir (KBr) 3290 (NH), 1785 (β -lactam), 1748, 1210 (ester), 740, 704, 695 cm⁻¹ (phenyl); nmr (CDCl₃) δ 3.22 (d, J = 5 Hz, H-2), 4.14 (d, J = 4Hz, H-5), 4.52 (d, J = 4 Hz, H-6), 4.90 (t, J = 5 Hz, H-3), 5.08 (s, $CH_2C_6H_5$), 7.0–7.7 ppm (m, C_6H_5); tlc (system IV) R_1 0.29.

Benzyl 6-Aminobisnorpenicillanate *p*-Toluenesulfonate (12a). A solution of *p*-toluenesulfonic acid monohydrate (760 mg, 4 mmol) in anhydrous Me₂CO (15 ml) was added to a suspension of 11a (2.08 g, 4 mmol) in Me₂CO (15 ml). The reaction was stirred vigorously at room temperature for 2.5 hr. CH₂Cl₂ (4 ml) and Me₂CO (20 ml) were added, and the precipitate was filtered off, washed (Et₂O), dried, and recrystallized from MeOH-Et₂O-Me₂CO yielding 12a (1.47 g, 81.5%): mp 169-170° dec: $[\alpha]^{25}$ D

+118° (c 1, MeOH); ir (KBr) 2950, 1540 (NH₃+), 1782 (β -lactam), 1740, 1190 (ester), 810, 745, 695 cm⁻¹ (phenyl); tlc (system V) R_f 0.47.

Benzyl Phenoxyacetamidobisnorpenicillanate (13a). A suspension of 12a (1.35 g, 3 mmol) in CH₂Cl₂ (120 ml) was neutralized with 1 equiv of Et_3N in CH_2Cl_2 and cooled (0°). Solutions of Et₃N (304 mg, 3.3 mmol) and phenoxyacetyl chloride (560 mg, 3.3 mmol) in CH₂Cl₂ (15 ml for each) were added gradually (in 1 hr) to the cooled solution of 12a. After storage for 2 hr at 0° the reaction mixture was extracted successively with 0.05 N HCl, NaHCO₃ (5%), and H₂O. The organic layer was dried (Na₂SO₄) and evaporated and the residue was crystallized from Et2O-petroleum ether, yielding 13a (1.075 g, 87%): mp 113–114°; $[\alpha]^{25}$ D +145° (c 0.5, CHCl₃); mass spectrum M+ 412; ir (KBr) 3370, 1685, 1520 (amide), 1796 (β-lactam), 1725, 1205 (ester), 745, 690 cm⁻¹ (phenyl); nmr (CDCl₃) δ 3.45 (d, J = 5 Hz, H-2), 4.54 (s, OCH_2CO), 5.03 (5, J = 5 Hz, H-3), 5.22 (s, $OCH_2C_6H_5$), 5.38 (d, J = 4.5 Hz, H-5), 5.72 (dd, J = 4.5 and 9 Hz, H-6), 6.7-7.6 ppm (m, C_6H_5) ; tlc (system II) R_f 0.26.

Bisnorpenicillin V Potassium Salt (1a). A solution of 13a (412 mg, 1 mmol) in EtOAc (30 ml) was hydrogenated over Pd/C (10%) (412 mg) for 5 hr at room temperature and at a pressure of 3 kg/cm^2 . The catalyst was filtered off and washed with EtOAc. The combined filtrates were concentrated to 50 ml and H_2O (50 ml) was added. The cooled mixture was adjusted to pH 6.3 with KOH (0.2 N). Freeze-drying of the aqueous layer yielded the potassium salt of 1a (258 mg, 71.5%) which was crystallized from H₂O-Me₂CO: mp 175° dec; $[\alpha]^{25}$ D +185° (c 1, H₂O); ir (KBr) 3350 (NH), 1680, 1525 (amide), 1769 (β-lactam), 1610, 1405 (COO⁻), 690, 750 cm⁻¹ (phenyl); nmr (D₂O, DSSA) δ 3.40 (AB part of ABX pattern, $J_{AX} = 4$ Hz, $J_{BX} = 6$ Hz, $J_{AB} = 11.5$ Hz, H-2), 4.53 (s, OCH₂C₆H₅), 4.88 (X part of ABX pattern, J_{AX+BX} = 10 Hz, H-3), 5.37 (d, J = 4 Hz, H-5), 5.48 (d, J = 4 Hz, H-6), 6.7-7.5 ppm (m, C₆H₅); tlc (system III) $R_{\rm f}$ 0.66. Anal. (C14H13N2O5SK) C, H, N.

Determination of the Sensitivity to β -Lactamase. The rate of hydrolysis of 1a and 1b was determined at 30° and at pH 7 on 4-ml samples containing 12.5 μ mol of penicillin and 13 units** of β -lactamase using the method described by Zyk.¹⁶ Under these conditions the rate of hydrolysis was 17 μ mol/hr for 1a and 19.5 μ mol/hr for 1b.

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**One unit of β -lactamase is defined as the amount of enzyme that hydrolyzes 1 μmol of benzylpenicillin per hour at 30° and at pH 7.15

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New Streptozotocin Analogs with Improved Antileukemic Activity[†]

Allan N. Fujiwara, Edward M. Acton,* and David W. Henry

Life Sciences Division, Stanford Research Institute, Menlo Park, California 94025. Received October 17, 1973

The 3-methyl-3-nitrosoureido derivatives of the following amino sugars were prepared as analogs of streptozotocin with the anomeric carbon protected, by nitrosating the methylureas in water with N₂O₃: 3-amino-3-deoxy-1,2-O-isopropylidene- α -D-ribofuranose, methyl 3-amino-3-deoxy- β -D-xylopyranoside, methyl 3-amino-3-deoxy- α -D-altropy-ranoside, methyl 3-amino-3-deoxy- α -D-glucopyranoside, methyl 6-amino-6-deoxy- α -D-glucopyranoside, and methyl 3-amino-2,3,6-trideoxy- α -L-lyxo-hexopyranoside. Tests against murine leukemia L1210 show that the anticancer activity of streptozotocin not only was retained but was enhanced in most of these derivatives.

Streptozotocin (1) is an antibiotic with antileukemic and diabetogenic activity in animals.¹ In human patients it has been used with success for the treatment of malignant insulinoma² and is being tested against the whole range of common tumor types.³ Kidney damage is the most common and most severe of various toxic side effects, so that analogs and derivatives of 1 are of interest for either enhanced anticancer activity or reduced toxicity. Recently observed separation of antileukemic, diabetogenic, and antibacterial activities in a series of new streptozotocin isomers and analogs^{1.4} emphasized the promise of further structural variations. Analogs blocked at the anomeric carbon of the sugar moiety are of interest for their greater ease and convenience in preparation, relative to free reducing sugars. The methyl glycosides^{4.5} of 1 have been studied *in vitro:* the β anomer was just as active as 1, but the α anomer was twice as active.⁴ The 6-

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