

**cis-3a,4,5,9b-Tetrahydrobenzo[e]isoindoline Hydrochloride (6a).** The above salt, 5 g in 100 ml of EtOH, was hydrogenated with 0.2 g of PtO<sub>2</sub> and H<sub>2</sub> at 4.2 kg/cm<sup>2</sup> until uptake ceased. The catalyst and solvent were removed and the product was crystallized several times from EtOH: yield 4.2 g; mp 228–230°. *Anal.* (C<sub>12</sub>H<sub>15</sub>N·HCl) C, H, N.

**cis-2-Methyl-3a,4,5,9b-tetrahydrobenzo[e]isoindoline (6b).** **Method A.** The base from 3 g of the 6a salt above, 14.5 ml of 90% HCOOH, and 9.5 ml of 37% HCHO were heated under reflux in an oil bath for 4 hr. The mixture was cooled, 10 ml of 10% HCl was added, and it was distilled to dryness. Excess dilute NaOH was added to the residue and the product was extracted with ether, dried over K<sub>2</sub>CO<sub>3</sub>, and distilled: bp 130–132° (4 mm). *Anal.* (C<sub>13</sub>H<sub>17</sub>N) C, H, N.

**6b hydrochloride** was crystallized from MeCN: mp 175–177°. *Anal.* (C<sub>13</sub>H<sub>18</sub>NCl) C, H, N.

**Method B.** *cis*-1,2-Dihydronaphthalene-1,2-dicarboxylic anhydride,<sup>4</sup> 15 g in 200 ml of EtOAc, was hydrogenated with 2 g of 5% Pd/C and H<sub>2</sub> at 4.2 kg/cm<sup>2</sup>. This gave 11 g of the *cis*-1,2,3,4-tetrahydro anhydride,<sup>5</sup> mp 61–63°. Anhydrous MeNH<sub>2</sub> was passed through a solution of 10 g of this anhydride in 200 ml of xylene while the temperature was raised slowly to 100°. The mixture was refluxed with removal of H<sub>2</sub>O for 2 hr and evaporated *in vacuo* and the residue was crystallized from ether, yielding 7 g of *cis*-N-methyl-1,2,3,4-tetrahydronaphthalene-1,2-dicarboximide, mp 79–80°. *Anal.* (C<sub>13</sub>H<sub>13</sub>NO<sub>2</sub>) C, H, N. This material, refluxed for 12 hr with 4 g of LiAlH<sub>4</sub> in 500 ml of ether and worked up as described for 7, gave 4.2 g of amine identical with that obtained by method A (ir and nmr spectra, melting point, and mixture melting point of the hydrochlorides).

**cis-2-Acetyl-3a,4,5,9b-tetrahydrobenzo[e]isoindoline (6c).** The base 6a with (CH<sub>3</sub>CO)<sub>2</sub>O in the usual manner gave crystals, mp 76–78°, from ether. *Anal.* (C<sub>14</sub>H<sub>17</sub>NO) C, H, N.

**cis-2-Ethyl-3a,4,5,9b-tetrahydrobenzo[e]isoindoline Hydrochloride (6d).** The reduction of 5 g of 6c with 2 g of LiAlH<sub>4</sub> in 200 ml of refluxing ether for 7 hr gave the amine which with dry HCl in ether yielded 3.8 g of this salt, mp 179–180° from MeCN. *Anal.* (C<sub>14</sub>H<sub>19</sub>N·HCl) C, H, N.

**3-Cyclopropylmethyl-2,3,4,5-tetrahydro-1H-3-benzazepine (11).** A solution of 17.3 g (0.1 mol) of 7 in 500 ml of benzene was stirred and 5.3 g (0.05 mol) of cyclopropylcarbonyl chloride in 50 ml of benzene was added. After 0.5 hr the salt was filtered and the benzene was washed with dilute HCl, dilute NaOH, and H<sub>2</sub>O. Distillation of the solvent left a viscous residue of the amide which was reduced to 11 as described for 6d.

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## References

- (1) L. A. Walter, U. S. Patent 2,520,264 (1950); *Chem. Abstr.*, 45, 675 (1951); B. V. Shetty, U. S. Patent 3,719,669 (1973); B. Pecherer, R. Sunbury, and A. Brossi, *J. Heterocycl. Chem.*, 9, 617 (1972); American Home Products Corporation, Netherlands Patent 6,906,604 (1969).
- (2) W. von E. Doering and M. J. Goldstein, *Tetrahedron*, 5, 53 (1953).
- (3) S. Irwin, "Clinical Pharmacological Techniques," J. H. Nodine and P. S. Sigler, Ed., Yearbook Medical Publishers, Chicago, Ill., 1964, Chapter 4.
- (4) K. Alder and K. Triebeneck, *Chem. Ber.*, 87, 237 (1954).
- (5) T. Lyssy, *J. Org. Chem.*, 27, 5 (1962).

## Correlation of Psychotomimetic Activity of Phenethylamines and Amphetamines with 1-Octanol–Water Partition Coefficients

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In an attempt to relate the hallucinogenic potencies in man of some biologically important amphetamines and phenethylamines, the 1-octanol–water partition coefficients for 11 amphetamines were determined. Using these values and published Hansch  $\pi$  constants, the log *P* for 17 additional amines was estimated. It was found that lipophilicity, as measured by the log of the partition coefficient, may be a significant determinant of the level of hallucinogenic potency. The study also suggests that an ideal log *P* value for psychotomimetic activity in man may be from 2.89 to 3.72.

The persisting major problem of research with psychotomimetic agents is the relationship of activity to physicochemical and structural properties of the active drugs. For the psychotomimetic amines, correlations have been attempted between activity and (1) the energy of the highest occupied molecular orbital;<sup>1,2</sup> (2) ultraviolet absorption maxima and molar absorptivity;<sup>3</sup> (3) degree of fluorescence;<sup>4</sup> and (4) stability of molecular complexes with dinitrobenzene.<sup>5</sup> Several correlations have been suggested between activity and conformation<sup>6–8</sup> and between activity and ability to stimulate various physiological receptors.<sup>9,10</sup> Numerous other studies have focused on relating metabolism, substitution patterns, and other chemical or metabolic factors to psychotomimetic activity.

The relationship between lipophilicity and activity has not been established. While drug action ultimately may be related to chemical or electronic factors, distribution and transport to the receptor may also be important in

assigning relative contributions to the various components of drug action *in vivo*. We decided to study 1-octanol–water partition coefficients of psychotomimetic amines and examine whether a relationship exists between this parameter and activity.

**Method.** Partition coefficients were determined in the 1-octanol–water system according to published procedures.<sup>11</sup> The aqueous phase was buffered to pH 7.4 using phosphate buffer and the partitioning was done at room temperature. Under these conditions the amines are partially ionized. The partition coefficients are reported as that of the neutral species with correction for ionization being made according to Albert.<sup>12</sup> The p*K*<sub>a</sub> of 2,5-dimethoxyamphetamine, which is reported to be 9.60,<sup>13</sup> was used in the correction. This is identical with the p*K*<sub>a</sub> reported for mescaline.<sup>14</sup> The additive nature of log *P*<sup>15</sup> was used to estimate this parameter for those agents for which it was not available experimentally.

Table I. Structure and Activity Data for Psychotomimetic Amines

	2	3	4	5	6	R	Log P	Log act.		[Δ log act.]
								Obsd	Calcd	
1		OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>		H	1.18 <sup>a</sup>	0.00	-0.15	0.15
2		OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	1.48 <sup>b</sup>	0.34	0.39	0.05
3	OCH <sub>3</sub>		OCH <sub>3</sub>			CH <sub>3</sub>	1.75 <sup>b</sup>	0.70	0.81	0.11
4			OCH <sub>3</sub>			CH <sub>3</sub>	1.77 <sup>b</sup>	0.70	0.83	0.13
5	OCH <sub>3</sub>			OCH <sub>3</sub>		CH <sub>3</sub>	1.88 <sup>b</sup>	0.90	0.98	0.08
6	OCH <sub>3</sub>		OCH <sub>3</sub>		OCH <sub>3</sub>	CH <sub>3</sub>	1.57 <sup>b</sup>	1.00	0.54	0.46
7	OCH <sub>3</sub>		OCH <sub>3</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	1.74 <sup>b</sup>	1.23	0.79	0.44
8	OCH <sub>3</sub>		CH <sub>3</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	2.08 <sup>b</sup>	1.90	1.21	0.69
9	OCH <sub>3</sub>		<i>n</i> -C <sub>3</sub> H <sub>7</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	3.31 <sup>c</sup>	1.90	1.76	0.14
10	OCH <sub>3</sub>		<i>n</i> -C <sub>4</sub> H <sub>9</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	3.81 <sup>d</sup>	1.56	1.55	0.01
11	OCH <sub>3</sub>		C <sub>2</sub> H <sub>5</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	2.81 <sup>b</sup>	2.00	1.72	0.28
12	OCH <sub>3</sub>		Br	OCH <sub>3</sub>		CH <sub>3</sub>	2.58 <sup>b</sup>	2.60	1.62	0.98
13		-OCH <sub>2</sub> O-				CH <sub>3</sub>	1.68 <sup>e</sup>	0.48	0.71	0.23
14		OCH <sub>3</sub>	-OCH <sub>2</sub> O-			CH <sub>3</sub>	1.80 <sup>f</sup>	0.43	0.87	0.44
15	OCH <sub>3</sub>		-OCH <sub>2</sub> O-			CH <sub>3</sub>	2.42 <sup>g</sup>	1.08	1.52	0.44
16	OCH <sub>3</sub>	-OCH <sub>2</sub> O-				CH <sub>3</sub>	2.04 <sup>h</sup>	1.00	1.17	0.17
17	-OCH <sub>2</sub> O-		OCH <sub>3</sub>			CH <sub>3</sub>	1.72 <sup>i</sup>	0.48	0.76	0.28
18	OCH <sub>3</sub>	-OCH <sub>2</sub> O-		OCH <sub>3</sub>		CH <sub>3</sub>	2.16 <sup>j</sup>	1.08	1.29	0.21
19	OCH <sub>3</sub>	OCH <sub>3</sub>	-OCH <sub>2</sub> O-			CH <sub>3</sub>	2.54 <sup>k</sup>	0.70	1.59	0.89
20	OCH <sub>3</sub>		OCH <sub>3</sub>	OCH <sub>3</sub>		H	1.44 <sup>l</sup>	0.00	0.32	0.32
21		-OCH <sub>2</sub> O-				H	1.38 <sup>m</sup>	0.00	0.22	0.22
22	OCH <sub>3</sub>	OCH <sub>3</sub>		OCH <sub>3</sub>		CH <sub>3</sub>	1.61 <sup>n</sup>	0.60	0.60	0.00
23	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	1.48 <sup>o</sup>	0.78	0.39	0.39
24	OCH <sub>3</sub>	OCH <sub>3</sub>			OCH <sub>3</sub>	CH <sub>3</sub>	1.73 <sup>p</sup>	1.11	0.78	0.33
25	OCH <sub>3</sub>		OC <sub>2</sub> H <sub>5</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	2.24 <sup>q</sup>	1.18	1.37	0.19
26	OCH <sub>3</sub>		C <sub>5</sub> H <sub>11</sub>	OCH <sub>3</sub>		CH <sub>3</sub>	4.31 <sup>r</sup>	1.00	1.09	0.09
27		OCH <sub>3</sub>	OCH <sub>3</sub>			CH <sub>3</sub>	1.00 <sup>b</sup>	<0.00	-0.52 <sup>s</sup>	0.52
28	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>			CH <sub>3</sub>	1.36 <sup>b</sup>	0.30	0.18 <sup>s</sup>	0.12

<sup>a</sup>Log P = log P<sub>3,4,5-trimethoxyamphetamine</sub> - π<sub>branched methyl</sub> = 1.48 - 0.30 = 1.18.

<sup>b</sup>Experimental value, this report.

<sup>c</sup>Log P = log P<sub>2,5-dimethoxy-4-ethylamphetamine</sub> + π<sub>methyl</sub> = 2.81 ± 0.50 = 3.31.

<sup>d</sup>Log P = log P<sub>2,5-dimethoxy-4-ethylamphetamine</sub> + 2π<sub>methyl</sub> = 2.81 + 2(0.50) = 3.81.

<sup>e</sup>Log P = log P<sub>amphetamine</sub> + π<sub>methylenedioxy</sub> = 1.73 - 0.05 = 1.68.

<sup>f</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> + π<sub>3-methoxy</sub> = 1.68 + 0.12 = 1.80.

<sup>g</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> + π<sub>2-methoxy</sub> = 1.68 + 0.74 = 2.42.

<sup>h</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> + π<sub>2-methoxy'</sub> = 1.68 + 0.36 = 2.04.

<sup>i</sup>Log P = log P<sub>4-methoxyamphetamine</sub> + π<sub>methylenedioxy</sub> = 1.77 + 0.05 = 1.72.

<sup>j</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> + π<sub>2-methoxy'</sub> + π<sub>3-methoxy</sub> = 1.68 + 0.12 = 2.15.

<sup>k</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> + π<sub>2-methoxy</sub> + π<sub>3-methoxy</sub> = 1.68 + 0.74 + 0.12 = 2.54.

<sup>l</sup>Log P = log P<sub>2,4,5-trimethoxyamphetamine</sub> - π<sub>branched methyl</sub> = 1.74 - 0.30 = 1.44.

<sup>m</sup>Log P = log P<sub>3,4-methylenedioxyamphetamine</sub> - π<sub>branched methyl</sub> = 1.68 - 0.30 = 1.38.

<sup>n</sup>Log P = log P<sub>2,3-dimethoxyamphetamine</sub> + π<sub>3-methoxy</sub> = 1.49 + 0.12 = 1.61.

<sup>o</sup>Log P = log P<sub>2,3,4-trimethoxyamphetamine</sub> + π<sub>3-methoxy</sub> = 1.36 + 0.12 = 1.48.

<sup>p</sup>Log P = log P<sub>2,4,6-trimethoxyamphetamine</sub> - π<sub>4-methoxy</sub> + π<sub>3-methoxy</sub> = 1.57 - (-0.04) + 0.12 = 1.73.

<sup>q</sup>Log P = log P<sub>2,4,5-trimethoxyamphetamine</sub> + π<sub>methyl</sub> = 1.74 + 0.50 = 2.24.

<sup>r</sup>Log P = log P<sub>2,5-dimethoxy-4-ethylamphetamine</sub> + 3π<sub>methyl</sub> = 2.81 + 1.50 = 4.31.

<sup>s</sup>A prediction of the activity based on the calculated log P was made, but the compound was not included in the regression analysis.

Note: π constants were determined for crowded and uncrowded 2-methoxys as follows: π<sub>2-methoxy</sub> (uncrowded) = log P<sub>2,4,5-trimethoxyamphetamine</sub> - log P<sub>3,4-dimethoxyamphetamine</sub>; π<sub>2-methoxy'</sub> (crowded) = log P<sub>2,3,4-trimethoxyamphetamine</sub> - log P<sub>3,4-dimethoxyamphetamine</sub>.

The hallucinogenic data are from the compilation of Shulgin, *et al.*<sup>16</sup> The activity values of the 4-propyl, 4-butyl, and 4-pentyl derivatives of 2,5-dimethoxyphenylisopropylamine were obtained by personal communication from A. T. Shulgin. The potencies are reported in Mescale Units, M. U., as defined by Shulgin.<sup>16</sup> The data are given in Table I.

Regression equations were generated using a nonweighted, multivariable, linear regression program and the IBM 370/155 computer of the Research Resources Laboratory of the University of Illinois at the Medical Center.

## Results and Discussion

Attempts to relate the hallucinogenic potencies in humans of the psychotomimetic amines resulted in eq 1. The

$$\log \text{act.} = -3.17 (\pm 1.61) +$$

$$3.15 (\pm 1.33) \log P - 0.50 (\pm 0.25) \log P^2 \quad (1)$$

$$n = 26; r = 0.79; s = 0.41;$$

$$\log P_0 = 3.14 (2.89-3.72)$$

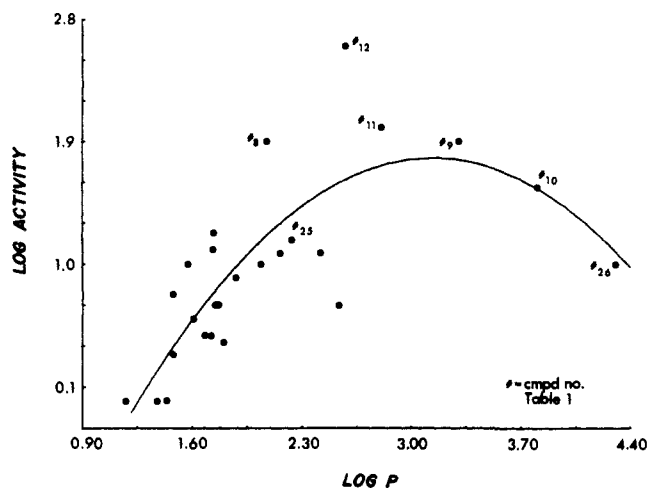


Figure 1.

terms in eq 1 and the statistical parameters are defined as follows:  $P$  is the 1-octanol-water partition coefficient,  $n$  is the number of agents in the data set,  $r$  is the correlation coefficient, and  $s$  is the standard deviation of the regression. The optimum lipophilicity,  $\log P_0$ , is obtained by finding the partial derivative,  $\delta(\log \text{act.})/\delta(\log P)$ , equating it with zero, and solving for  $\log P$ . The equation is significant of the 99.5% level of confidence as determined from the  $F$  statistic:  $F_{2,23} = 19.436$ ,  $F_{2,23}: \alpha 0.005 = 6.730$ . Figure 1 illustrates eq 1.

The correlation coefficient for our analysis is not as high as one would hope from studies of this nature. Because of variability in the biological data, which is accurate to only  $\pm 25\%$ ,<sup>16</sup> one can at best expect to account for 75% of the variation. However, our equation accounts for 62% ( $R^2 = 0.62$ ) of the variation, leaving about 10% for approximations introduced in applying the model and other factors that may effect activity. The best previous correlation using the quantum mechanically derived parameter, HOMO, accounted for 56% of the variance using only 13 of the hallucinogenic drugs. Therefore, our results indicate that lipophilicity is important in determining hallucinogenic potency. Due to the high degree of steric crowding in several of the compounds, there is a degree of uncertainty regarding the applicability of additivity in each case. If anything, however, we believe that inclusion of the calculated  $\log P$  values gives a regression which tends to be less highly correlated than would be the case for the actual experimentally determined values. For example, regression equations using only the 11 experimental values give approximations for the portion of the curve  $0.9 < \log P \leq 2.81$ . Both the linear and parabolic regressions have  $R^2 = 0.83$ . This value is somewhat better than the overall correlation when the calculated  $\log P$  values are included.

The optimum lipophilicity is of interest since it predicts that psychotomimetic amines more or less lipophilic than those with  $\log P_0 = 3.14$  should have less than optimum activity. To our knowledge an optimum  $\log P$  has not previously been observed in the analysis of biological data obtained on human subjects. Our finding indicates that this relative affinity for lipoidal biophases is necessary or maximum passive permeation of the CNS. This could be an important physicochemical parameter to consider in the design of active CNS agents.

The studies of Kang and Green<sup>2</sup> indicate that some of the variation in hallucinogenic activity of substituted amphetamines may be due to the electronic nature of the substituents. While our data do not directly support this, the unexplained variance from our analysis does suggest that other factors affect the level of potency. For the substituents in the series electronic effects are difficult to parameterize. For those amphetamines for which potency could be determined with some degree of certainty, a  $\log P$  of approximately 1.40 is required for activity. The exception to this is mescaline, which indicates that an electronic or other effect may be present. The low to marginal activity of 3,4-dimethoxyamphetamine is interesting since its  $\log P$  is determined to be 1.00. This is near that of mescaline, which is active. The additional *m*-methoxy group in mescaline must be responsible for the increased potency. The influence of the methoxy somehow overcomes the effect of lipophilicity even in the absence of a secondary  $\alpha$ -methyl group as in the amphetamines.

Amphetamine, with  $\log P$  estimated to be 1.71, is reported to be inactive as an hallucinogen<sup>2</sup> (except with chronic usage where it precipitates a toxic psychosis), while 2,4,5-trimethoxyamphetamine with  $\log P$  determined to be 1.74 has significant activity. Since the lipophilicities of these two amphetamines are similar, this would imply that the methoxy substitution contributes in addition to a lipophilic effect, an electronic or chemical effect. However, within the series of hallucinogenic compounds examined, the change in activity due to structural variation appears to result mainly from changes in hydrophobicity. Activity drops off for highly lipophilic methoxy-substituted amphetamines ( $\log P > 4.00$ ) as can be noted from the data for the 2,5-dimethoxy-4-pentyl analog. This is consistent with the proposal of Hansch<sup>17</sup> regarding the role of lipophilicity and drug action.

## References

- (1) S. H. Snyder and C. R. Merrill, *Proc. Nat. Acad. Sci. U. S.*, **54**, 258 (1965).
- (2) S. Kang and J. P. Green, *Nature (London)*, **226**, 645 (1970).
- (3) K. Bailey and D. Verner, *J. Pharm. Sci.*, **61**, 480 (1972).
- (4) F. Antun, J. R. Smythies, J. Benington, R. D. Morin, C. F. Barknecht, and D. E. Nichols, *Experientia*, **27**, 62 (1971).
- (5) M. T. Sung and J. A. Parker, *Proc. Nat. Acad. Sci. U. S.*, **69**, 1346 (1972).
- (6) C. Chothia and P. Pauling, *Proc. Nat. Acad. Sci. U. S.*, **63**, 1063 (1969).
- (7) S. Kang, C. L. Johnson, and J. P. Green, *Mol. Pharmacol.*, **9**, 640 (1973).
- (8) R. W. Baker, C. Chothia, P. Pauling, and H. P. Weber, *Mol. Pharmacol.*, **9**, 23 (1973).
- (9) H. C. Cheng, J. P. Long, D. E. Nichols, and C. F. Barknecht, *J. Pharmacol. Exp. Ther.*, **188**, 114 (1974).
- (10) D. C. Dyer, D. E. Nichols, D. B. Rusterholz, and C. F. Barknecht, *Life Sci.*, **13**, 885 (1973).
- (11) T. Fujita, J. Iwasa, and C. Hansch, *J. Amer. Chem. Soc.*, **86**, 5175 (1964).
- (12) A. Albert, "Selective Toxicity," Wiley, New York, N.Y., 1960.
- (13) E. B. Liffler, H. M. Spencer, and A. Burger, *J. Amer. Chem. Soc.*, **73**, 2611 (1951).
- (14) K. J. Taska and J. C. Schooler, *Arch. Int. Pharmacodyn. Ther.*, **202**, 66 (1973).
- (15) A. Leo, C. Hansch, and D. Elkins, *Chem. Rev.*, **71**, 525 (1971).
- (16) A. T. Shulgin, T. Sargent, and C. Naranjo, *Nature (London)*, **221**, 537 (1969).
- (17) C. Hansch, *Accounts Chem. Res.*, **2**, 232 (1969).