# Synthesis and Biological Activity of Cocaine Analogues. 2.  $6H$ -[2]Benzopyrano[4,3-c]pyridin-6-ones

Edward S. Lazer, Gilbert J. Hite, Karl A. Nieforth,\*

*Section of Medicinal Chemistry and Pharmacognosy, School of Pharmacy, The University of Connecticut, Storrs, Connecticut 06268* 

## and Eugene S. Stratford

*School of Pharmacy, University of West Virginia, Morgantown, West Virginia 26506. Received September 5, 1978* 

l,2,3,4-Tetrahydro-2-methyl-6/Y-[2]benzopyrano[4,3-c]pyridin-6-one (20) and *cis-* and *trans-l,*2,3,4,4a,lOb-hexahydro-2-methyl-6H-[2]benzopyrano[4,3-c]pyridin-6-one (3a and 3b) were synthesized. The design of 3b was based on the proposal that the active conformation of cocaine is one in which the phenyl and amino groups are arranged in a manner that will superimpose upon a  $\beta$ -phenethylamine in a trans-staggered conformation. The compounds were compared with cocaine and tropacocaine for their ability to inhibit uptake of [3H]norepinephrine by rat brain synaptosomal preparations. The test compounds  $(IC_{50} = 3.2 \times 10^{-4} M, 20; 6.5 \times 10^{-4} M, 3a;$  and  $3.2 \times 10^{-4} M, 3b;$ <br>respectively) were considerably weaker than cocaine  $(IC_{50} = 5.8 \times 10^{-7} M)$  and tropacocaine  $(IC_{50} = 5.6$ dopamine (3%) and serotonin (0%) uptake to a much lesser extent, if at all, at this concentration.

Structural modification of cocaine has been undertaken to obtain compounds that interact at the same biological sites as cocaine but with an altered or reduced spectrum of activity.<sup>1</sup> Such an approach might ultimately lead to a new antidepressant or a compound with the ability to reverse or block cocaine activity at the receptor level. It may also provide a compound which is more selective than cocaine as an inhibitor of biogenic amine uptake. Fluoxetine, a selective inhibitor of serotonin reuptake,<sup>2</sup> and nisoxetine, a selective inhibitor of norepinephrine reup $take,$ <sup>3</sup> have generated clinical interest.

In this paper, we describe the synthesis and biogenic amine uptake inhibition activity of compounds based upon what has been proposed as an active conformation of  $cocaine$ ,  $1<sup>4</sup>$  In this conformation, the phenyl ring and



nitrogen are arranged in a manner that will superimpose upon a trans-staggered phenethylamine. Removal of the ethylene bridge on cocaine and formation of a bond between the phenyl and piperidine rings provided the novel 6H-[2]benzopyrano[4,3-c]pyridin-6-one system represented by 2 and 3. Compound 3 is a rigid analogue of tropacocaine (cocaine without the carbomethoxy group). Tropacocaine has been shown to possess weak cocaine-like activity.<sup>5</sup> The synthesis and activity of 3a (cis), 3b (trans), and unsaturated analogue 20 are described.

Chemistry. A direct five-step route to 2 which failed in the fourth step is illustrated in Scheme I. The magnesium iodide salt of indene (4) was prepared by reaction of 4 with ethylmagnesium iodide. This was allowed to react with  $\beta$ -chloropropionaldehyde to give 5. Ozonolysis, followed by oxidative workup, gave the diacid, which lactonized in situ to give 6. Esterification of 6 was achieved with diazomethane. At this point, the goal was to convert 7 to the amine 8 and cyclize to the piperidine ring via a Mannich reaction. However, rather than displace the halide, methylamine abstracted the benzylic proton of 7

Scheme I



and the benzoate ester eliminated to give 9. Attempts to aminate 5 or 6 to circumvent this problem were unsuccessful and the route had to be abandoned.

The synthesis of 3 is illustrated in Scheme II. This route was based on the synthesis of l-methyl-3-phenyl-4-piperidone from ethyl  $\alpha$ -phenylacrylate by Patchett and Giarrusso.<sup>8</sup> Homophthalic acid (10) was prepared by oxidation of 4 according to an established procedure.<sup>9</sup> Esterification of 10 was carried out according to the procedure of Sheehan and O'Neill.<sup>10</sup> Carboxylation with lithium diisopropylamide and carbon dioxide according to a general procedure for esters<sup>11</sup> provides 12. Formation of Mannich base 13 with formaldehyde and diethylamine, followed by in situ HOAc/NaOAc decarboxylation and elimination, gave 14. The conversion of 11 to 14 basically followed a procedure by Carlson et  $al<sup>12</sup>$  for synthesis of  $\alpha$ -methylene lactones from lactones.

Conjugate addition of ethyl  $\beta$ -(methylamino)propionate to 14 gave 15. Dieckmann condensation was carried out



with sodium hydride. Theoretically, the condensation could have given 16 and 17, as well as 18 and 19. Reduction of 17 would have provided 2; however, only 18 and 19 were detected as reaction products. The failure of 16 or 17 to form may be due to their relative instability compared to 18 and 19. Under the reaction conditions, any of 16 that might have initially formed could have been converted back to starting material and ultimately to 18 or 19. Hydrolysis and decarboxylation to 20 were carried out in refluxing 10% hydrochloric acid. Catalytic reduction with platinum oxide gave 3a and 3b in a ratio of 13:1.

16 17

CH<sub>3</sub>

 $CH<sub>3</sub>$ 

**Biological Results and Discussion.** Compounds 3a and 3b, as well as their unsaturated precursor 20, were compared to cocaine and tropacocaine for their ability to inhibit the reuptake of [<sup>3</sup>H]norepinephrine by rat-brain synaptosomal preparations. The results are shown in Figure 1.

The compounds may be separated into three groups on the basis of their potency. Cocaine was the most potent compound with an  $IC_{50}$  of approximately 5.75  $\times$  10<sup>-7</sup> M. The  $IC_{50}$  of tropacocaine had an intermediate value of approximately  $5.6 \times 10^{-6}$  M. The three other compounds formed a group of weak inhibitors. Compounds 3b and 20 were equipotent with  $IC_{50}$  values of approximately 3.2  $\times$  10<sup>-4</sup> M. Compound 3a was somewhat weaker, with an IC<sub>50</sub> of about 6.5  $\times$  10<sup>-4</sup> M.

Compounds 3a and 3b were also evaluated in a separate study<sup>13</sup> using a similar test. Both compounds were evaluated at concentrations of  $1 \times 10^{-5}$  M. Compound 3b inhibited reuptake of norepinephrine by 36%, while 3a showed no inhibition at all. At  $1 \times 10^{-5}$  M (Figure 1), 3b inhibited reuptake by about 11%. Compound 3a (Figure 1) was not tested at this concentration, but if the line obtained for  $3a$  is extrapolated to  $1 \times 10^{-5}$  M a value of 0% is not unreasonable. Some variations between the

values shown in Figure 1 and those obtained by Ferris and Maxwell<sup>13</sup> may be expected, since slightly different procedures were used. Also, as indicated by the bars in Figure 1, variation occurred to some extent for each point on the curve.

The design of 3 was based on what has been proposed to be the active conformation of cocaine. The trans-fused ring system 3b is superimposable on the proposed active conformation represented by 1, whereas the cis-fused system 3a is not. Therefore, it was expected that 3b would be more potent than 3a. Although the results in Figure 1 show that the  $IC_{50}$  of 3b is slightly less than half that of 3a, the low potency of both compounds and small difference in activity is not adequate to conclude that 3b is an accurate representation of the active conformation of cocaine. Therefore, these results do not lend any support to the active conformation of cocaine proposed by support to the active componation of cocame proposed by<br>Maxwell et al.<sup>4</sup> The unsaturated compound 20 was found to be about equipotent with 3b. Assuming that both compounds act at a cocaine site, this suggests that the enol double bond has no observable effect on potency.

In an examination of uptake inhibitors,  $\mathrm{\tilde{K}oe^{14}}$  proposed that the active conformation of most of these compounds is represented by an extended phenylbutylamine. Accordingly, he proposed that the active conformation of cocaine was that in which the benzoate ester was extended, rather than folded back as in 1. Koe's proposal did not rule out the possibility that uptake inhibitors can act in a phenethylamine-type orientation. He postulated alternate binding sites for aromatic rings to permit the uptake site to accommodate either a phenethylamine or a phenylbutylamine.

Other workers have attempted to modify the structure of cocaine in an attempt to obtain a stimulant or antidepressant. 3 $\beta$ -Phenyltropan-2-ols<sup>15</sup> and 3 $\beta$ -phenyltropane-2-carboxylate esters<sup>16</sup> have shown cocaine-like



**Figure 1.** Relative potencies of cocaine  $(\bullet)$ , tropacocaine  $(\triangle)$ , 20 (C), **3b** (O), and **3a** (O) as inhibitors of norepinephrine uptake by rat-brain synaptosomal preparations: abscissa, -log [drug]; ordinate, percent inhibition. The points on the curve represent the average percent inhibition of two determinations (except for cocaine, which had six determinations) at each concentration. The bars extend out to the two values that were used to determine each point.

Table I. Inhibition of Uptake of Biogenic Amines by  $3b^a$ 

biogenic amine	$%$ inhibn at $10^{-5}$ M
norepinephrine serotonin	36
dopamine	

<sup>a</sup> Data obtained by Ferris and Maxwell<sup>13</sup> according to the procedure in ref 19.

activity in various tests. In both of these systems, removal of the ethylene bridge from the tropane ring resulted in loss of activity. If this represents a general rule at the cocaine receptor, lack of an ethylene bridge might contribute to the low activity of **3a, 3b,** and 20.

The data in Table I suggest that at  $1 \times 10^{-5}$  M  $3b$  exhibits selectivity in inhibition of biogenic amine uptake. At this concentration, **3b** inhibited norepinephrine uptake by 36% while having virtually no effect on serotonin or dopamine uptake. If appropriate substitutions on **3b** or perhaps 20 could increase potency while retaining selectivity, a useful compound might result.

# **Experimental Section**

**Chemistry.** Melting points were determined on a Thomas-Hoover unimelt apparatus and are uncorrected. Elemental analyses were performed by Microlytics, Inc., South Deerfield, Mass., and are within  $\pm 0.4\%$  of the theoretical values. All compounds gave IR and proton NMR spectra compatible with their assigned structures. NMR spectra were obtained on a Hitachi Perkin-Elmer R-24 high resolution NMR spectrometer, using Me<sub>4</sub>Si as an internal standard with  $CDCl<sub>3</sub>$  or external with D20. IR spectra were obtained on a Beckman Acculab 3. Mass spectra were obtained with an A.E.I. Scientific Inc. mass spectrometer MS902. TLC were run on Baker-flex silica gel 1B-F sheets,  $7.52 \times 2.5$  cm. Column chromatography was run on silica gel 60, particle size 0.063-0.200 *nm* (70-230 mesh ASTM; EM Reagents). References to removing or evaporating solvent under reduced pressure refer to evaporations on a rotary flash evaporator. Centrifugations at lOOOOg were carried out on a Sorvall superspeed RC2-B automatic refrigerated centrifuge. Centrifugation at 23000g was carried out on a Beckman L2-65B ultracentrifuge.

**l-(l-Hydroxy-3-chloropropyl)-lif-indene** (5). EtMgl was prepared from 116.7 g of EtI (0.748 mol) dissolved in 120 mL of anhydrous  $Et_2O$  and 17.8 g (0.74 mol) of magnesium turnings. The vigorous reaction was slowed by the rapid addition of 160 mL of anhydrous  $Et<sub>2</sub>O$ . After the addition of EtI was complete, the reaction mixture was warmed on a hot water bath for an additional 1 h, then cooled to room temperature, and a solution of 86 g of indene (0.74 mol) in 220 mL of toluene was added rapidly. The condenser was removed and the reaction heated at 100 °C with stirring for 10 h. The brown reaction mixture was then cooled in an ice bath, causing indenemagnesium iodide to precipitate. As much brown supernatant as possible was decanted, and the precipitated salt was dissolved in anhydrous  $Et<sub>2</sub>O$ . The solvent was removed under reduced pressure, leaving a pea-green paste which was dissolved in 350 mL of anhydrous  $Et_2O$ . An ethereal solution of  $\beta$ -chloropropionaldehyde<sup>17</sup> prepared earlier was added. The reaction temperature was kept at 15  $\rm{^{\circ}C}$  or below by cooling in ice. The flask was packed in ice and stirred overnight. The reaction mixture was then poured into 350 mL of ice-water saturated with NH<sub>4</sub>Cl and extracted with Et<sub>2</sub>O (12  $\times$  300 mL). The combined  $Et_2O$  extracts were washed with saturated NH<sub>4</sub>Cl solution (2  $\times$  400 mL) and saturated NaCl solution (2  $\times$  400 mL), dried  $(Na<sub>2</sub>SO<sub>4</sub>)$ , filtered, and evaporated under reduced pressure to give 116 g of a red-brown oil. After seeding and cooling in a freezer, 43.3 g of dark-brown crystals was obtained. The seed was obtained by column chromatography purification of an earlier reaction [silica gel, petroleum ether-Et<sub>2</sub>O (7:3), which eluted some impurity, followed by petroleum ether- $Et<sub>2</sub>O(1:1)$ , which eluted the product which readily solidified]. The product was recrystallized from hexane four times to give 26.8 g (0.128 mol, 17.4%) of light-yellow product, mp 73.5-76.5 °C. After several days in a desiccator, the product turned light brown. Therefore, it was sublimed to give 16.2 g of stable, white, analytically pure product, mp 75-77 °C. Anal.  $(C_{12}H_{13}C_{10})$  C, H.

**traus-3,4-Dihydro-l-oxo-3-(2-chloroethyl)-lH-2-benzopyran-4-carboxylic Acid (6).** A solution of sublimed 5 (16.2 g, 77.6 mmol) and 385 mL of anhydrous EtOH was cooled with stirring in an ice bath. Ozone was bubbled into the reaction via a Welsbach T-23 ozonator for 3 h and 10 min. The EtOH was removed under reduced pressure to give 23.9 g of a sticky foamy oil, which was dissolved in 108 mL of glacial HOAc and 163 mL of 15%  $H_2O_2$ . The mixture was warmed, with stirring, for 17 h in an oil bath set at 55 °C. The solvent was removed under reduced pressure, and the residue was dissolved in 200 mL of Et^O and carefully extracted (violent reaction) with a 10% solution of NaHSO<sub>3</sub> (7  $\times$  25 mL) to destroy the excess H<sub>2</sub>O<sub>2</sub>. The ether phase was then extracted with 10%  $\mathrm{Na_{2}CO_{3}}$  (4  $\times$  25 mL). The combined base extracts were washed with  $Et_2O$  (2  $\times$  30 mL) and then made acidic with 3 N HC1. The cloudy aqueous phase was extracted with  $Et_2O$  (4  $\times$  50 mL). The combined  $Et_2O$  extracts were dried (saturated NaCl solution extract; then  $Na<sub>2</sub>SO<sub>4</sub>$ ) and filtered, and the solvent was removed under reduced pressure to give 11 g of yellow oil. The product was crystallized from a small amount of  $CHCl<sub>3</sub>$  with a few drops of hexane. Two crops gave 5.2 g. The residue from the mother liquors was purified again by the base-acid extractions to give another 0.64 g of product, for a total of 5.84 g (23.3 mmol, 30%). An analytical sample, mp 134-135.5 °C, was obtained by recrystallization from chloroform: NMR (CDCI<sub>3</sub>) decoupling at  $\delta$  2.0 shows  $J_{\text{CH-CH}} = 6$  Hz. On this basis, the trans configuration was assigned. Anal.  $(C_{12}H_{11}ClO_4)$  C, H.

**Methyl traus-3,4-Dihydro-l-oxo-3-(2-chloroethyl)-lH-2 benzopyran-4-carboxylate** (7). An ethereal solution of diazomethane prepared from 8.56 g (40 mmol) of Diazald® ( $p$ tolysulfonylmethylnitrosamide, Aldrich) was added to a solution of 6 (650 mg, 2.55 mmol) dissolved in 50 mL of  $Et_2O$ . The reaction mixture was allowed to sit in a hood overnight. The faint yellow reaction mixture was then filtered, and the solvent was removed under reduced pressure to give a yellow oil. The product was crystallized from acetone-hexane to give 300 mg (1.116 mmol, 44%), mp 97-99 °C. An analytical sample, mp 99.5-101 °C, was obtained by recrystallization from acetone-hexane. Anal. (C13H13C104) C, **H.** 

**Methylammonium o-[l-(Methoxycarbonyl)-4-chlorobut-1-enyl]benzoate (9).** CH<sub>3</sub>NH<sub>2</sub> gas was bubbled into a solution of 222 mg (0.826 mmol) of 7 in 14 mL of anhydrous  $Et_2O$ for 15 min. The mixture was cooled and filtered to give 194 mg  $(0.647 \text{ mmol}, 78\%)$  of analytically pure product: mp 121-122 °C. Anal. (C14H18C1N04) C, **H,** N.

**2-(MethoxycarbonyI)-2-[2-(methoxycarbonyl)phenyl] acetic Acid (12).** A solution of diisopropylamine (25.3 g, 253 mmol) in 250 mL of dry THF under an  $N_2$  atmosphere was cooled to between -10 and -15 °C in a dry ice-acetone bath, and 162 mL of  $1.6$  N *n*-butyllithium was added. After stirring the solution for 15 min, 45 g (216 mmol) of 11 dissolved in 170 mL of dry THF was added over a 30-min period at -30 to -15 °C. After the addition was complete, the  $N_2$  inlet was removed and  $CO_2$  was vigorously bubbled through the reaction mixture for 20 min. The mixture was maintained at 2 °C or less and slowly acidified with 10% HCl. The reaction mixture was then extracted with  $CHCl<sub>3</sub>$  $(1 \times 375 \text{ mL}, 3 \times 100 \text{ mL})$ . The combined CHCl<sub>3</sub> extracts were extracted with 10%  $\text{Na}_2\text{CO}_3$  (2 × 200 mL, 4 × 100 mL). The combined aqueous extracts were cooled below 2 °C and were made acidic by the careful addition of, initially,  $6 \text{ N HCl}$  and then  $10\%$ HC1 as the pH approached neutrality. The product was extracted into  $CHCl<sub>3</sub>$  (5 × 200 mL). The combined  $CHCl<sub>3</sub>$  extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), and the solvent removed under reduced pressure to give 38 g (150 mmol, 69%) of 12 as a light-yellow solid. Recrystallization from  $Et_2O$  gave 30.8 g (122 mmol) of white crystals, the yield now 56.5%. An analytical sample, mp 117-118 °C, was obtained by a second recrystallization from  $Et<sub>2</sub>O$ . Anal.  $(C_{12}H_{12}O_6)$  C, H.

**Methyl 2-[2-(Methoxycarbonyl)phenyl]acrylate** (14). A solution of 100 mL of diethylamine and 190 mL of 35-40% formaldehyde was cooled in an ice bath and 51.5 g (204 mmol) of 12 was added. The reaction mixture was stirred at room temperature for 15 min and then heated on a steam bath for 30 min. The reaction was removed from the steam bath, and 19.1 g of NaOAc and 190 mL of glacial HOAc were added carefully. The reaction was heated for an additional 15 min and cooled, and the contents of the flask were extracted with  $Et_2O$  (1 × 800 mL,  $1 \times 200$  mL). The combined Et<sub>2</sub>O extracts were washed with 10% HCl ( $3 \times 200$  mL), H<sub>2</sub>O ( $2 \times 200$  mL), saturated NaHCO<sub>3</sub> solution  $(2 \times 200 \text{ mL})$ , and saturated NaCl solution  $(2 \times 200 \text{ mL})$ . The  $Et<sub>2</sub>O$  was then dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and evaporated to give 40.8 g (185 mmol, 91%) of 17 as a colorless oil. An analytical sample was obtained by distillation: bp 90-93 °C (0.06 mm). Anal.  $(C_{12}H_{12}O_4)$  C, H.

Methyl  $\alpha$ -[[N-[2-(Ethoxycarbonyl)ethyl]-N-methyl**amino]methyl]-o-(methoxycarbonyl)phenylacetate (15).**  Ethyl  $\beta$ -(methylamino)propionate<sup>18</sup> (13.8 g, 105 mmol) and 21 g (95.4 mmol) of 14 were stirred overnight at room temperature. The reaction mixture was then dissolved in 150 mL of cold 2 N HCl and extracted with 100 mL of  $Et_2O$ . The  $Et_2O$  was back extracted with 70 mL of 2 N HC1. The combined acid extracts were made basic with 20% NaOH, and the product was extracted with  $Et_2O$  (6 × 100 mL). The ether solution was dried  $(Na_2SO_4)$ and evaporated to give 18 as an oil. An attempt was made to distill the product (160-168 "C, 0.15 mm); however, examination of the distillate showed it was not 15 but probably the retro-Michael product 14. Since the product apparently decomposed upon distillation, it was purified by filtering through silica gel, eluting with benzene-ether (1:1). This separated the product from any remaining ethyl $\beta$ -(methylamino)propionate, which remained in the silica gel. The product was an oil, 19.3 g (55 mmol, 55%). No sample was submitted for elemental analysis. The NMR spectrum was consistent for the structure of 15. An attempt was made to make a hydrochloride salt, but it proved to be gummy and hygroscopic: NMR (CDC13) *8* 7.2-8.0 (4 H, m, aromatic), 4.9  $(1 H, m, benzylic proton), 4.1 (2 H, m, -CH<sub>2</sub>OCO<sup>-</sup>), 3.9 (3 H, s,$ phenyl COOCH3), 3.65 (3 H, s. C-COOCH3), 2.3 (3 H. s, N-CH3), 1.2 (3 H, t,  $CH_3CH_2OCO-$ ).

**Dieckmann Condensation of 15 with NaH. Isolation of Ethyl l,2,3,4-Tetrahydro-2-methyI-6-oxo-6H-[2]benzopyrano[4,3-c]pyridine-4-carboxylate (19).** Benzene was distilled from a solution of 13.2 g (37.6 mmol) of 15 in 250 mL

of anhydrous benzene until the volume was reduced to 200 mL to remove any traces of water. NaH-mineral oil dispersion (50% sodium hydride, 3.94 g of dispersion, 1.97 g of NaH, 82.4 mmol) was added to a dry flask, the mineral oil was removed by rinsing with hexane, and then 80 mL of anhydrous benzene was added. The solution of 15 was then poured into the reaction flask. The mixture was refluxed, and gradually began to darken. After 6.75 h, the reaction mixture was cooled in an ice bath and poured into 100 mL of ice-cold 2 N HC1. After shaking the mixture, the aqueous phase was separated, and the benzene was extracted with more 2 N HCl  $(4 \times 25 \text{ mL})$ . The combined acid washes were extracted with  $Et<sub>2</sub>O$  (2  $\times$  50 mL) and then neutralized with 10% NaOH. As soon as the aqueous solution remained cloudy, it was extracted with 100 mL of  $Et<sub>2</sub>O$ . The process was repeated twice, and then the aqueous phase was extracted four times with 100 mL of  $Et_2O$ . The combined extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and evaporated under reduced pressure to give 7.9 g of a light-yellow amorphous solid. This material was then used directly in the next reaction without further purification.

In an earlier run of the same reaction, the product was characterized. TLC of the crude product (silica gel, benzene-Et<sub>2</sub>O, 1:1) showed four spots between  $R_f$  0.23 and 0.48. There was also a small amount of material with a higher  $R_f$  and some at the base line. Crude product (1.6 g) was passed through a silica gel column (36 g), eluting with benzene- $Et_2O$ , 1:1. Only the compound with the lowest  $R_f$  among the group of four with close  $R_f$  values was isolated in pure form. Spectral evidence and elemental analysis were consistent with the proposed structure 19 ( $R = CH_3$ ). An analytical sample, mp 122-123.5 °C, was obtained by four recrystallizations from methanol. The fact that this structure had a methyl ester at the 4 position indicated that ester exchange occurred. Mechanistic considerations and NMR spectra of the crude products suggested structures 18 and 19 ( $R = C<sub>2</sub>H<sub>5</sub>$ ) for the other compounds in the group of four with  $R_f$  values between 0.23 and 0.48. None of the spectra were consistent for structures 16 or 17: NMR (CDC13) *8* 7.25-8.45 (4 H, m, aromatic), 3.75 (3 H, s, methyl ester), 3.65 (2 H, br s, N-CH<sub>2</sub>C=C), 2.65-3.5 (3 H, m, N-CH2CH), 2.5 (3 H, s, N-CH3); IR (KBr) broad band centering on  $1720 \text{ cm}^{-1}$ , ester and enol lactone carbonyl; MS calcd for M<sup>+</sup>. 273.0996; found, 273.1001. Anal.  $(C_{15}H_{15}NO_4)$  C, H, N.

**l,2,3,4-Tetrahydro-2-methyl-6#-[2]benzopyrano[4,3-c] pyridin-6-one (20).** The crude product from the previous reaction  $(7.9 \text{ g})$  was dissolved in 120 mL of 10% HCl and refluxed for 3.5 h. The hot reaction mixture was filtered, and the filtrate was cooled to give 4.56 g (18.1 mmol) of product as the HC1 salt, an overall 48% yield based on 18. Recrystallization from water gave an analytical sample, which decomposed at 285-287 °C. Anal.  $(C_{13}H_{14}CINO_2)$  C, H, N.

The free base was prepared by dissolving 4.46 g (17.7 mmol) of the HCl salt in 20 mL of warm  $H<sub>2</sub>O$  and cautiously adding 50 mL of saturated  $\text{Na}_2\text{CO}_3$  solution. After cooling and filtration of the mixture, 3.6 g (16.7 mmol, 94%) of free base, mp 111.5-113 °C, was obtained. An analytical sample, mp 112-113 °C, was obtained by recrystallization from hexane. Anal.  $(C_{13}H_{13}NO_2)$ C, H, N.

*cis-* **and £rans-l,2,3,4,4a,10b-Hexahydro-2-methyl-6H-** [2]benzopyrano[4,3-c]pyridin-6-one (3a and 3b).  $PtO<sub>2</sub>$  (150) mg), 23 (2 g, 9.29 mmol), and 50 mL of glacial HOAc were shaken under a hydrogen atmosphere that ranged from 58 to 20 psi for 21 h. TLC of the product (free base on silica gel, eluted with  $CHCl<sub>3</sub>$  with 5% MeOH) showed that there was some starting material left. An additional 0.5 g (2.32 mmol) of 23 and 150 mg of  $PtO<sub>2</sub>$  catalyst were added to the reaction mixture and shaken under 60 psi of hydrogen. After 18.5 h, the pressure had decreased to 48 psi and the reaction was stopped. The reaction mixture was filtered and the catalyst rinsed with a small amount of glacial HOAc. The HOAc was subsequently evaporated under reduced pressure, providing a residual oil which was dissolved in  $H_2O$  and made basic with  $10\%$   $Na<sub>2</sub>CO<sub>3</sub>$ . The product was extracted into CHCl<sub>3</sub> (3  $\times$  25 mL). The combined CHCl<sub>3</sub> extracts were washed with saturated NaCl solution  $(2 \times 20 \text{ mL})$ , dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and evaporated to give 2.3 g of oil which according to TLC showed no starting material: *R,* (CHC13, 5% MeOH) 23, 0.52; 3b, 0.47; 3a, 0.34. The product was purified by passing through a 65-g silica gel column. The fractions containing pure **3b** were combined and gave 56 mg (0.258 mmol, 2.2%). The fractions containing pure

3a were combined to give 456 mg. The fractions containing a mixture were combined and concentrated. The resulting oil was passed through a second column to give an additional 272 mg of 3a for a total of 728 mg (3.35 mmol, 29%). Analytical samples of 3b, mp 101-101.5 °C, and 3a, mp 83-84 °C, were prepared by recrystallizations from hexane. Anal.  $(C_{13}H_{15}NO_3)$  C, H, N. The hydrochloride salts of 3a, 279-280 °C dec, and 3b, mp 268-270 °C, were also prepared and recrystallized from water. Anal. (C13H16C1N02) C, **H,** N.

**Biological Test Methods.** Eight 60-day-old Charles River CD. albino male rats weighing between 300 and 400 g were sacrificed, and their brains were immediately dissected on ice. The midbrains, containing the hypothalamus, hippocampus, and striatum were separated and homogenized in 0.32 M sucrose (10% w/v) using eight rapid up and down strokes with a size B Thomas Teflon pestle tissue homogenizer spinning at approximately 800 rpm. The homogenate was centrifuged at 3000g for 10 min, and the supernatant was separated and centrifuged at 10000g for 10 min. The pellet was resuspended in 0.32 M sucrose and layered on sucrose gradients consisting of 1.2 and 0.8 M sucrose. The gradients were centrifuged at 23000g for 70 min. The white synaptosomal layer, between the 0.8 and 1.2 M sucrose, was pipetted off and diluted with an equal volume of distilled water. The resulting suspension was diluted with an equal volume of 0.32 M sucrose and centrifuged at lOOOOg for 20 min, and the resulting pellet was resuspended in Kreb's Ringer phosphate buffer containing 1.6 mg of pargyline hydrochloride and 237 mg of glucose per 100 mL.

The incubation tubes contained 20  $\mu$ L of test drug (except three control tubes), 20  $\mu$ L of 4.5% NaCl solution, 50  $\mu$ L of 10<sup>-5</sup> M norepinephrine, 10  $\mu$ Ci of a 1  $\mu$ Ci/10  $\mu$ L solution of [<sup>3</sup>H]norepinephrine (except two control tubes), and 500  $\mu$ L of synaptosomal suspension. The tubes were incubated at 30 °C for 5 min. Immediately after incubation, 10  $\mu$ L of the  $[3H]$ norepinephrine was added to the set of two control tubes. Approximately 3 mL of the pargyline-glucose buffer solution was added to each tube as it was removed from the incubation bath to dilute the suspension and, therefore, to quench uptake of biogenic amine by the synaptosomes. The tubes were kept in ice and after 2 min were centrifuged at lOOOOg for 15 min. The supernatant was discarded and the sides of the tubes were carefully cleaned with Q-tips® without touching the pellet. Two glass beads and 200  $\mu$ L of 50% EtOH were added to each tube. The tubes were then vortexed for 30 s, and 100  $\mu$ L of the final suspension was added to 10 mL of Bray's solution in a scintillation vial. The radioactivity was counted on a Packard Tri-Carb liquid scintillation spectrometer Model 3380. The set of two control tubes represented the amount of [<sup>3</sup>H]norepinephrine taken up by the synaptosomes after the incubation period was over. This value (cpm) was subtracted from the average cpm obtained in each of the other sets of vials. Percent inhibition was then calculated for each sample as (control cpm  $-$  sample cpm)/control cpm. The cpm were not converted to dpm, since counting efficiency varied only  $\pm 0.5\%$  (35-36%) between samples.

Uptake studies on 3a and 3b were also conducted by Dr. R. M. Ferris and Dr. R. A. Maxwell<sup>13</sup> at Burroughs Wellcome Co. in Research Triangle Park, North Carolina. The procedure was similar to the one just described but contained some variations.<sup>19</sup>

**Acknowledgment.** The authors express their gratitude to Drs. Robert A. Maxwell and R. M. Ferris of Burroughs Wellcome Co. for biological testing of the compounds. We also thank Dr. Paul Y. Sze and Dr. Naranjan D. Aggarwal for their valuable contributions to the biological testing at the University of Connecticut.

## **References and Notes**

- (1) For the first paper in this series, see E. S. Lazer, N. D. Aggarwal, G. J. Hite, K. A. Nieforth, R. T. Kelleher, R. D. Spealman, C. R. Schuster, and W. Wolverton, *J. Pharm. Sci.,* 67, 1656 (1978).
- (2) D. T. Wong, F. P. Bymaster, J. S. Hornig, and B. B. Molloy, *J. Pharmacol. Exp. Ther.,* **151,** 339 (1966).
- (3) L. Lemberger, S. Terman, H. Rowe, and R. Billings, *Br. J. Clin. Pharmacol.,* 3, 215 (1976).
- (4) R. A. Maxwell, E. Chaplin, S. Batmanglidj Eckhardt, J. R. Soares, and G. Hite, *J. Pharmacol. Exp. Ther.,* **173,** 158 (1970).
- (5) S. B. Ross and A. L. Renyi, *Acta Pharmacol. Toxicol.,* 36, 56 (1975).
- (6) E. J. Corey, L. S. Melvin, Jr., and M. F. Haslanger, *Tetrahedron Lett.,* 3117 (1975).
- (7) M. Elliot, N. F. Jones, and B. C. Peerson, *J. Sci. Food Agric,*  18, 325 (1967).
- (8) A. A. Patchett and F. F. Giarrusso, *J. Med. Chem.,* 4, 393 (1961).
- (9) O. Grummitt, R. Egan and A. Buck, "Organic Syntheses", Collect. Vol. 3, Wiley, New York, 1955, p 449.
- (10) J. C. Sheehan and R. C. O'Neill, *J. Am. Chem. Soc,* 72, 4614 (1950).
- (11) S. Reiffers, H. Wynberg, and J. Strating, *Tetrahedron Lett.,*  3001 (1971).
- (12) R. G. Carlson, R. L. Coffin, W. W. Cox, and R. S. Givens, *Chem. Commun.,* 501 (1973).
- (13) R. M. Ferris and R. A. Maxwell, personal communication.
- (14) B. K. Koe, *J. Pharmacol. Exp. Ther.,* **199,** 649 (1976).
- (15) S. J. Daum, M. D. Aceto, and R. L. Clarke, *J. Med. Chem.,*  16, 667 (1973).
- (16) R. L. Clarke, S. J. Daum, A. J. Gambino, M. D. Aceto, J. Pearl, M. Levitt, W. R. Cuminskey, and E. F. Bogado, *J. Med. Chem.,* 16, 1260 (1973).
- (17) T. L. Jacobs, S. Winstein, G. B. Linden, and D. Seymour, *J. Org. Chem.,* 11, 223 (1946).
- (18) R. W. Holley and A. D. Holley, *J. Am. Chem. Soc,* 71, 2124 (1949).
- (19) R. M. Ferris, F. L. M. Tang, and R. A. Maxwell, *J. Pharmacol. Exp. Ther.,* 181, 407 (1972).