N-(2-Cyanoethyl) Derivatives of Meperidine, Ketobemidone, and a Potent 6,7-Benzomorphan

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The N-(2-cyanoethyl)- 9α -ethyl-5-methyl-6,7-benzomorphan (1c) is a more potent antinociceptive and has stronger receptor binding affinity than its N-methyl analogue 1b. The N-(2-cyanoethyl)-4-phenylpiperidine compounds 2b and 3b were almost inactive compared to their N-methyl congeners 2a and 3a, respectively. It appears that the pharmacological effect of the N-(2-cyanoethyl) moiety is dependent on the opioid on which it is substituted.

Recently it was reported that replacement of the *N*methyl substituent of (-)-3-hydroxy-*N*-methylmorphinan (levorphanol) and metazocine by 2-cyanoethyl resulted in a marked increase in antinociceptive potency without a corresponding increase in opiate receptor affinity and with a considerable decrease in acute toxicity.² Furthermore, the physical dependence capacity of the resultant compounds was nil. We wish to report our findings on a similar chemical alteration of three strong analgesics, meperidine (**2a**), ketobemidone (**3a**), and 2,5-dimethyl-9 α -ethyl-2'-



hydroxy-6,7-benzomorphan (1b), along with the carbamido compound 1d.

Chemistry. The cyanoethyl derivatives 1c, 2b, and 3b were prepared by alkylation of their respective nor bases with acrylonitrile. The carbamido analogue 1d was similarly prepared by alkylation with acrylamide.

Biological Results and Discussion. Compounds 1c, 1d, 2b, and 3b were evaluated for analgesic activity in the hot-plate assay^{2a,b} and for receptor affinity as determined by the capacity to displace bound, radiolabeled di-hydromorphine from rat brain homogenates (Table I).^{2c} Additionally 1c and 1d were evaluated in the tail-flick,⁴ phenylquinone writhing (PPQ),⁵ and tail-flick antagonism assays.⁴ Also, 1c, 1d, 2b, and 3b were assayed by single-dose suppression tests (SDS) in morphine-dependent monkeys,⁶ and 3b was evaluated in the Nilsen assay.⁷

The N-(2-cyanoethyl) benzomorphan analogue 1c was six times more potent than its N-methyl parent 1b,⁸ though the increase was not as large as that observed for the 9α -methyl (metazocines) congeners. There was a corresponding increase in receptor affinity; however, both 1b and 1c exhibit lower binding affinities than might be predicted from their hot-plate ED₅₀ values when compared to morphine. This type of discrepancy may be rationalized on the basis of transport, such as for the case of meperidine

Table I. Pharmacology of N-(2-Cyanoethyl) Compounds

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ED _{so} ^a	EC ₅₀ ^b	PDC ^c
$1.3 (0.9-2.0)^d$	15	high ^d
$0.21 (0.15 - 0.30)^{e}$	4	intermed
$19.3 (13.3 - 27.9)^{f}$	9	none
$4.7 (4.2-5.4)^{g}$	700^{h}	high
inact to 100	8000	none
0.8(0.7-0.9)	2	high
i	1500	low
1.2(0.9-1.3)	3	high
1.18(1.05 - 1.33)	9	high
	$\frac{\text{ED}_{50}{}^{a}}{1.3 (0.9-2.0)^{d}} \\ 0.21 (0.15-0.30)^{e} \\ 19.3 (13.3-27.9)^{f} \\ 4.7 (4.2-5.4)^{e} \\ \text{inact to 100} \\ 0.8 (0.7-0.9) \\ i \\ 1.2 (0.9-1.3) \\ 1.18 (1.05-1.33)^{j} \\ \end{array}$	$\begin{array}{c c} & ED_{s0}{}^{a} & EC_{s0}{}^{b} \\ \hline ED_{s0}{}^{a} & EC_{s0}{}^{b} \\ \hline 1.3 & (0.9-2.0)^{d} & 15 \\ 0.21 & (0.15-0.30)^{e} & 4 \\ 19.3 & (13.3-27.9)^{f} & 9 \\ 4.7 & (4.2-5.4)^{g} & 700^{h} \\ \text{inact to } 100 & 8000 \\ 0.8 & (0.7-0.9) & 2 \\ i & 1500 \\ 1.2 & (0.9-1.3) & 3 \\ 1.18 & (1.05-1.33)^{j} & 9 \end{array}$

^a Hot-plate assay in mg/kg. Parentheses indicate 95% SE limits on probit analysis, subcutaneous injection in mice.^{3a,b} ^b Binding constant from rat-brain homogenates in nM.^{3c} ^c Physical dependence capacity in rhesus monkeys from SDS (single-dose suppression).⁶ ^d Reference 8a, 6.4 (3.1-12.8) in tail-flick assay and 0.2 (0.008-0.7) in PPQ. ^e 0.8 (0.4-1.5) in the tail-flick assay and 0.09 (0.04-0.2) in the PPQ. ^f Inactive in the tail-flick assay and PPQ. ^g Reference 9. ^h W. A. Klee, N.I.H., personal communication. ⁱ Erratic dose-response curve in the hot plate assay; ED₅₀ > 50 in the Nilsen assay. ^j Reference 7.

(2a), or possibly metabolism. Such discrepancies might also be expected if, perhaps, the receptor is somewhat distorted in its in vitro environment. A potential metabolite of 1c, the carbamido analogue 1d, was only weakly active as an analgesic. However, the affinity of 1d for the binding site is equal to that of (\pm) -metazocine, reflecting perhaps a decreased ability to cross the blood-brain barrier. Compounds 1c and 1d did not show antagonist activity in the tail-flick assay, an interesting observation since short chains on the nitrogen in analgesics having tricyclic or larger ring systems usually induce antagonist activity. In SDS tests, 1d did not substitute for morphine at the doses tested (to 16 mg/kg), and 1c partially substituted (briefly) for morphine at 1.2 mg/kg, while 1b substituted completely^{8a} for morphine.

The N-(cyanoethyl)-substituted 4-phenylpiperidines **2b** and **3b** showed a complete loss of analgesic activity for **2b**, and only minor activity remained for **3b**, which gave an erratic dose-response curve in the hot plate assay. These results were reflected in their binding constants. In SDS tests, **2b** was inactive (to 20 mg/kg) and **3b** showed a partial suppression of abstinence signs at 20 mg/kg that was complete at 40 mg/kg. The low activity observed with **2b** and **3b** is somewhat surprising in view of the substantial analgesic activity of the N-propyl and N-butyl analogues of **2a**⁹ and **3a**.¹⁰ Thus, short to moderate length chains on the nitrogen atom of **2a** and **3a** continue to show intriguing and unexplained variations in opiate effects.

In conclusion, our results further substantiate the finding of a recent report that the effect of substituting the N-(2-cyanoethyl) moiety in strong analgesics is dependent on the parent opiate employed.² It appears that the utility of this group as an analgesic pharmacophore may be limited to tricyclic and larger ring systems.

Experimental Section

All melting points were obtained on a Thomas-Hoover Unimelt, capillary, melting point apparatus and are uncorrected. IR spectral data were obtained on a Perkin-Elmer 257 and NMR spectral data were obtained on a Perkin-Elmer R-24. Spectra were obtained on all compounds and these are compatible with the structural assignments. Microanalyses obtained as indicated by the symbols of the elements are within $\pm 0.4\%$ of the theoretical values.

2-(2-Cyanoethyl)-9 α -ethyl-2'-hydroxy-5-methyl-6,7benzomorphan (1c). Compound 1a (1 g, 4.3 mmol),⁸ acrylonitrile (0.3 g, 5.6 mmol), absolute EtOH (10 mL), and Et₃N (5 drops) were stirred overnight at room temperature (23–25 °C), after which TLC showed no 1a. Evaporation to dryness at 20–26 mm and cooling gave a solid which crystallized from EtOAc (5 mL)-ligroin (30–60 °C, 3–4 mL) in prisms: yield 0.96 g, 90%; mp 95-96 °C (gas evolution). Anal. (C₁₈H₂₄N₂O) C, H, N.

2-(2-Carbamidoethyl)-9 α -ethyl-2'-hydroxy-5-methyl-6,7benzomorphan (1d). A mixture of 1.0 g (4.33 mmol) of 1a,⁸ 0.41 g (5.76 mmol) of acrylamide, 5 drops of Et₃N, and 10 mL of absolute EtOH was stirred at room temperature (4 days) and monitored by TLC [EtOAc-MeOH-concentrated NH₄OH (16:3:1) and CHCl₃-MeOH-concentrated NH₄OH (74:24:2), silica gel, iodoplatinate reagent]. The reaction mixture was then filtered and the filtrate concentrated (in vacuo) to a brownish oil. Et₂O was added, and the resulting crystals from overnight standing were recrystallized from EtOAc/Me₂CO (3:1): yield 1.07 g, 90.7% (based on covered starting material); mp 171.5–172.5 °C. Anal. (C₁₈H₂₆N₂O₂) C, H, N.

N-(2-Cyanoethyl)normeperidine (2b). A solution of normeperidine (norpethidine,¹¹ 3.00 g, 10.7 mmol), acrylonitrile (0.81 mL, 12.25 mmol), and anhydrous K₂CO₃ in absolute EtOH (50 mL) was stirred for 6 h at room temperature. The solution was filtered and evaporated in vacuo to yield a brown oil. The oil was dissolved in Et₂O (50 mL), washed with saturated NaCl solution, dried (MgSO₄), and evaporated in vacuo to yield an oil from which the hydrobromide salt was made. Recrystallization twice from Me₂CO afforded 0.88 g (29%), mp 207–208 °C. Anal. (C₁₇-H₂₃N₂O₂Br) C, H, N.

N-(2-Cyanoethyl)norketobemidone (3b). A solution of norketobemidone¹⁰ (3.00 g, 12.9 mmol) and acrylonitrile (1.26 mL, 19.3 mmol) in absolute EtOH (50 mL) was stirred for 2 h at room temperature. The solvent was evaporated in vacuo, yielding a brown tar from which the hydrobromide salt was made. Recrystallization twice from absolute EtOH afforded 3.45 g (73%), Acknowledgment. This work was supported in part by grants from the National Institute of Drug Abuse (DA-00490) and from Hoffmann-La Roche, Inc. Thanks are also due Dr. A. E. Jacobson, National Institutes of Health, for hot-plate data; Dr. W. A. Klee, National Institute of Mental Health, for binding constants; and Dr. E. L. May, Medical College of Virginia, for helpful discussions. The encouragement by Dr. L. S. Harris (Chairman, Pharmacology Department, Medical College of Virginia) is gratefully acknowledged by I.M.U.

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Active-Site Studies of Neurohypophyseal Hormones: Synthesis and Pharmacological Properties of $[5-(N^4, N^4-Dimethylasparagine)]$ oxytocin¹

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Synthesis and biological properties of $[5 - (N^4, N^4 - \text{dimethylasparagine})]$ oxytocin are reported. In this analogue, the hydrogens of the primary carboxamide moiety in the side chain of the asparagine residue in position 5 of the posterior pituitary hormone oxytocin have been replaced by two methyl groups. The protected nonapeptide intermediate was prepared by a stepwise procedure using solution techniques. The analogue possesses 4.60 ± 0.03 units/mg (mean \pm SEM) uterotonic activity on the isolated rat uterine horn and 9.14 ± 0.03 units/mg of avian vasodepressor activity. Moreover, it displays an identical intrinsic activity in the in vitro rat uterotonic assay as oxytocin, when tested in the presence of either 0.5 mM Ca²⁺ (standard assay conditions) or at reduced levels of Ca²⁺ (0.3, 0.15, and 0.05 mM). This result is significant in view of the proposed biologically active model of oxytocin, in which the side chain of the 5 position residue was assigned to contain an "active element" responsible for the intrinsic activity of the hormone when bound to the uterine receptor.

The 5-position asparaginyl residue plays a key role in the preferred solution conformation of $xytocin^2$ and in the proposed "biologically active" model of the hormone (Figure 1) at the uterine receptor.³⁻⁵ Both in Me₂SO and aqueous medium⁶ the peptide NH of the asparagine residue helps to stabilize the β turn involving the sequence