

## Biological Effects of Modified Colchicines. 2. Evaluation of Catecholic Colchicines, Colchifolines, Colchicide, and Novel *N*-Acyl- and *N*-Aroyldeacetylcolchicines

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A series of natural and synthetic colchicine derivatives was examined for their potency in the lymphocytic leukemia P388 screen in mice, for their toxicity in mice, and for their binding to microtubule protein. The natural alkaloids cornigerine and colchifoline and several N,O-substituted analogues of colchifoline were found to be as potent and as toxic as colchicine in the P388 screen with good affinity for tubulin. The 1,2-(methylenedioxy)-substituted isomer of cornigerine was considerably less potent *in vivo* than could have been anticipated from the *in vitro* tubulin binding data. Several *N*-acyl and *N*-aroyl derivatives prepared from deacetylcolchicine showed high potency in the *in vitro* and *in vivo* screens. Colchicide was found to be highly potent *in vivo*, and *N*-carbethoxydeacetylcolchicine, a synthetic analogue of colchicine with a *N*-carbethoxy instead of an *N*-acetyl function, showed interesting biological properties.

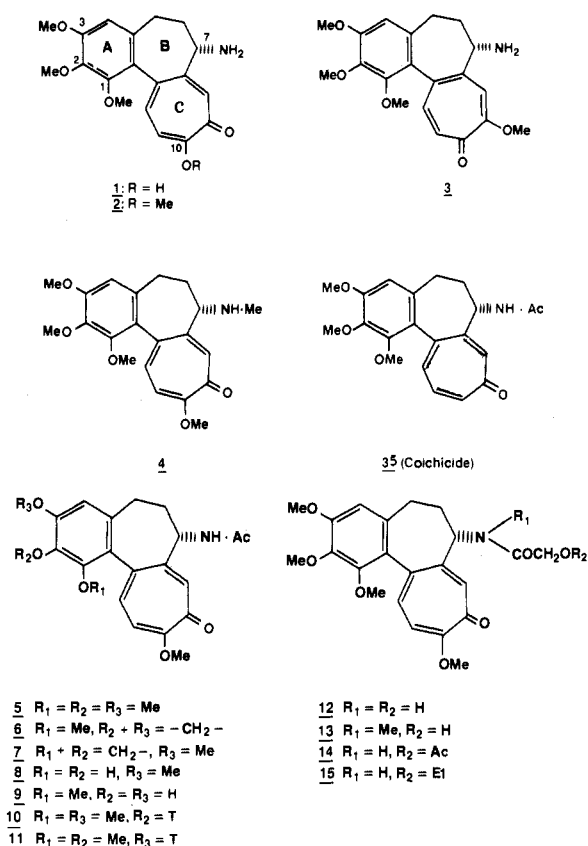
We reported in our first paper that tropolonic colchicines which showed weak binding to microtubule protein were essentially inactive in the lymphocytic leukemia P388 assay and that compounds showing low toxicity in mice always displayed significantly lower potency in the P388 screen.<sup>1</sup> Evaluation of the results obtained from the P388 assay, tubulin binding assay, and toxicity data appeared, then, to be a reasonable method for ascertaining the biological effects of colchicine derivatives. It was hoped that by appropriate modification of the colchicine structure a potent but much less toxic substitute for colchicine could ultimately be found.

Selective O-demethylation at C<sub>1</sub> of colchicine gave 1-demethylcolchicine, which was found to be practically inactive *in vitro* and *in vivo*.<sup>1</sup> O-Demethylation at C<sub>3</sub> of colchicine, on the other hand, afforded 3-demethylcolchicine, which was found remarkably active *in vivo*, although its O-glucoside, colchicoside, is practically inactive.<sup>1</sup> The substitution in the aromatic ring A has thus proven to be important both for tubulin binding and antimetabolic effect<sup>2,3</sup> and this conclusion has been supported by the additional compounds examined herein.

Modification of the *N*-acetamido side chain also significantly influenced antimetabolic activity. Introduction of electron-withdrawing groups at the acetamido terminal increased potency,<sup>4,5</sup> whereas a change from an *N*-acyl to a *N*-aroyl group seemed to cause a decrease in potency.<sup>5,6</sup> In order to verify this point, which seemed very critical for our study, a series of novel *N*-acyldeacetylcolchicines was needed, and deacetylcolchicine (1), its methyl ethers 2 and 3 (obtained by O-methylation of 1) and demecolcine (4) were utilized for their preparation.

A third, interesting, group of compounds is represented by colchicide (35), a compound prepared by Rapoport et al. from thicolchicine<sup>7</sup> and reported to be biologically active. Since the C<sub>10</sub>-OMe group of colchicine was long considered to be essential for antimetabolic activity and gave rise to the development of thicolchicine and colchicineamide, we considered a reevaluation of colchicide worthwhile.

**Chemistry.** The first group of colchicine derivatives evaluated included the catechols 8 and 9, prepared earlier



in connection with our synthesis of cornigerine,<sup>8</sup> and the two *N*-phenyltetrazolyl ethers 10 and 11, obtained from

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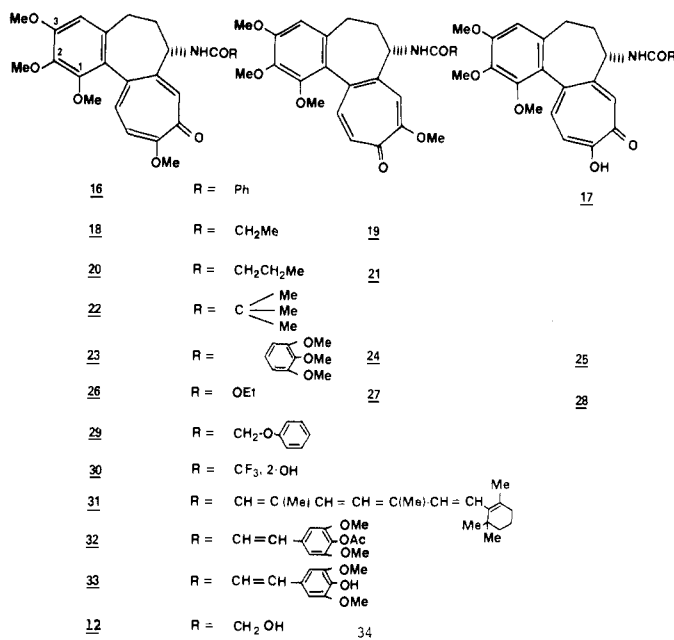
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their phenolic precursors<sup>1</sup> by O-alkylation with *N*-phenyltetrazolyl chloride. Both heterocyclic ethers were amorphous materials, but their chemical integrity was assured by MS and TLC analysis.

A second group included colchifoline (12), a minor contaminant of commercial colchicine,<sup>9</sup> and its derivatives 13–15. The *N*-methyl derivative 13 was prepared by reacting the now readily available demecolcine (4)<sup>10</sup> with (trifluoroacetyl)glycolyl chloride, a method analogous to the synthesis of colchifoline from deacetylcolchicine.<sup>11</sup>

Other compounds of this group are represented by novel *N*-acyldeacetylcolchicines, *N*-aroyldeacetylcolchicines, and the ethyl carbamate 26. The analogues 18, 20, 23, and 26



were obtained from a mixture of deacetylcolchicine and deacetylcolchicine<sup>12</sup> by *N*-acylation and separation of the isomers 19, 21, 24, and 27 by chromatography over silica gel (method A). The two analogues 18 and 20 have been reported elsewhere,<sup>13,14</sup> but no experimental details were given. Mild acid hydrolysis of mixtures of 23 and 24, and 26 with 27, afforded the colchicine analogues 25 and 28, which could be recycled. The *N*-benzoyldeacetylcolchicine (16), evaluated earlier as a mixture of the two tautomeric ethers,<sup>9</sup> has now been prepared as the natural isomer, which on mild acid hydrolysis gave 17. The analogues 16, 20, 22 and 29 were prepared from deacetylcolchicine, obtained from *N*-(trifluoroacetyl)deacetylcolchicine<sup>10</sup> and kept in the form of their *d*-tartrate salts<sup>12</sup> (method B).

The assignment of the normal and iso structures was based on <sup>1</sup>H NMR analysis<sup>15</sup> and the MS-fragmentation

pattern, the iso compounds show more intense fragmentation peaks than the natural isomers,<sup>9</sup> and the well established behavior of the two isomers during chromatography, with the normal isomer moving faster on silica gel with CHCl<sub>3</sub>/MeOH/NH<sub>4</sub>OH (90:9:1) than the iso isomer. Furthermore, the optical rotation of the iso forms were consistently more negative than the natural isomers in chloroform solution.

The partial ether cleavage of colchicine with concentrated sulfuric acid has now been applied to *N*-(trifluoroacetyl)deacetylcolchicine,<sup>10</sup> affording the 2-demethyl congener 30, an interesting compound for further exploration of the 2-demethyl analogues of colchicine. The phenol 30 showed signals for C<sub>1</sub>-OMe at δ 3.72, for C<sub>3</sub>-OMe at 3.86, and for C<sub>10</sub>-OMe at 4.00.

Colchicide (35), which lacks the C<sub>10</sub>-OMe group of colchicine, was prepared from thiocolchicine by the reported procedure<sup>7</sup> and obtained in 46% yield, the byproduct being hexahydrodemethoxycolchicine. Colchicide, purified by chromatography and obtained as a glassy solid, showed one spot on TLC but had a considerably lower rotation than that reported in the literature.<sup>7</sup> This might be due to the presence of solvents originating from the chromatography. The hydrochloride salt prepared for biological evaluation showed an unusually high optical rotation, [α]<sub>D</sub><sup>25</sup> -456° (CHCl<sub>3</sub>).

The *N*-sinapinoyl compound 33, prepared via its *O*-acetate 32, and the *N*-retinoyl compound 31 were both prepared as potential UV-affinity labels, and for this reason they were only tested *in vitro*. Their UV absorption, however, can not be distinguished from that of colchicine and thus 33 and 31 were not useful as affinity labels, although 33 showed the expected bathochromic shift in alkaline solution.

**Structure-Activity Relationships.** It can be seen from Table I that a correlation exists between the *in vivo* potency of these compounds in the lymphocytic leukemia P388 screen of the National Cancer Institute (NCI) (see Experimental Section) and their toxicity in that screen. With few exceptions, molecular modifications that produced compounds with potencies equal to or less than colchicine resulted in correspondingly altered toxicities. Variations in structure that resulted in compounds having potencies similar to that of colchicine are embodied in the colchifolines 12 and 13 and the derivative 15 (Table I). The *N*-benzoyl analogue 16, when tested earlier as a mixture of the natural and iso isomer,<sup>6</sup> was not found to be very potent *in vivo*. However, the pure isomer 16 is potent, almost comparable to colchicine in the P388 assay, as is the *N*-carbethoxy analogue 26. In contrast, the 3,4,5-trimethoxybenzoyl analogue 23 is considerably less potent. *N*-Substituted compounds, which show binding to microtubule protein similar to colchicine, are the *N*-propionyl, *N*-butyryl, *N*-phenoxyacetyl, *N*-trifluoroacetyl, and the *N*-benzoyl derivatives.

The 10-demethoxycolchicine (35, colchicide) has surprising activity *in vitro*, since it has been generally accepted that the C<sub>10</sub>-methoxy group of colchicine is essential for activity and is the place of interaction with biopolymers.

The colchifoline acetate 14, which was only available in minute quantities, showed good affinity for microtubule protein. The isocolchicines 19 and 34 showed poor *in vitro* activity and were not investigated *in vivo*. The colchicines 17, 25, and 28, which belong to an inactive group of colchicine, were also not further evaluated. The phenyl-

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Table I. Potency in P388 Lymphocytic Leukemia Screen, Tubulin Binding, and Toxicity of Colchicine and Derivatives

no.	compound	potency, <sup>a</sup> μmol/kg	tubulin binding, <sup>b</sup> %	toxicity	
				I <sup>c</sup>	II <sup>d</sup>
5	colchicine	0.4 <sup>e</sup>	90	2.5	3.0 (2.5-4.0)
6	cornigerine	<10 <sup>f</sup>	59	39.1	9.7 (7.0-13.1)
7	1,2-(methylenedioxy)-1,2-demethylcolchicine	inactive	48	104	>167 <sup>g</sup>
8	1,2-didemethylcolchicine	inactive	0	>320	>450 <sup>h</sup>
9	2,3-didemethylcolchicine	20 <sup>e</sup>	18	54	>450 <sup>i</sup>
10	2- <i>O</i> -( <i>N</i> -phenyltetrazolyl)-2-demethylcolchicine		52		
11	3- <i>O</i> -( <i>N</i> -phenyltetrazolyl)-3-demethylcolchicine		27		
12	colchifoline	0.7 <sup>e</sup>	97	3.0	3.9 (2.9-5.1)
13	<i>N</i> -methylcolchifoline	0.6 <sup>e</sup>	99	2.9	14.5 (11.2-18.9)
14	<i>O</i> -acetylcolchifoline		86		8.5 (6.6-11.2)
15	<i>O</i> -ethylcolchifoline	0.4	74	2.8	9.0 (6.5-12.6)
16	<i>N</i> -benzoyldeacetylcolchicine	1.0 <sup>e</sup>	83	4.3	60.5 (43.2-85.0)
18	<i>N</i> -propionyldeacetylcolchicine		98		8.2 (4.8-14.0)
19	<i>N</i> -propionyldeacetylcolchicine		10		
20	<i>N</i> -butyryldeacetylcolchicine	0.7 <sup>e</sup>	98	5.8	5.8 (4.0-8.9)
22	<i>N</i> -pivaloyldeacetylcolchicine		65		9.3 (6.8-12.9)
23	<i>N</i> -(3,4,5-trimethoxybenzoyl)deacetylcolchicine	14.0 <sup>e</sup>	46	72.6	>180 <sup>j</sup>
26	<i>N</i> -carbethoxydeacetylcolchicine	0.7 <sup>e</sup>	86	11.7, 1.5 <sup>k</sup>	114 (88.3-147.1)
29	<i>N</i> -(phenoxyacetyl)deacetylcolchicine	1.3 <sup>e</sup>	81	5.1	61.3 (45.0-83.5)
30	2-demethyl- <i>N</i> -(trifluoroacetyl)deacetylcolchicine	3.8 <sup>e</sup>	96	23.5	32.5 (20.0-52.9)
31	retinoyldeacetylcolchicine		41		
32	<i>N</i> -(acetoxysinapinoyl)deacetylcolchicine		69		
33	<i>N</i> -inapinoyldeacetylcolchicine		69		
34	isocolchifoline		0		
35	colchicide	9.2 <sup>e</sup>	87	54.2	168.9 (119.5-238.8)

<sup>a</sup> Obtained by graphical estimation of the potency, which would give a T/C = 140. <sup>b</sup> Percentage by which binding is reduced by the presence of the inhibitor at 25 μM with [<sup>3</sup>H]colchicine at 2.5 μM. Values are the average of triplicate assays. <sup>c</sup> Toxicity in the P388 NCI screen. Defined as that dose, in micromoles per kilogram, that caused an average weight loss > 3 g in at least one out of the set of mice or the death of one or more of the mice in the set. Six mice were used at each of the dose levels. Deaths were noted on 5th day of a series of nine injections in 9 days (1 per day) of the compound. <sup>d</sup> Toxicity found after a single intramuscular injection, in micromoles per kilogram. The total number of deaths were counted after 7 days at various dose levels, and the LD<sub>50</sub> was determined by probit analysis. Parenthesized numbers represent the 95% confidence interval. <sup>e</sup> Confirmed by a second P388 assay. <sup>f</sup> Estimated from two nontoxic dose levels of 19.6 and 9.8 μmol/kg, which gave T/C values of 161 and 155, respectively. <sup>g</sup> Twelve percent of mice were dead at 167 μmol/kg. <sup>h</sup> Ten percent of mice were dead at 450 μmol/kg. <sup>i</sup> No deaths occurred at highest dose level tested (450 μmol/kg). <sup>j</sup> Ten percent of mice were dead at 180 μmol/kg. <sup>k</sup> A second P388 assay showed markedly different toxicity. This compound is being further evaluated.

tetrazolyl ethers 10 and 11 and analogues of colchicine with unsaturated N-CO side chains were prepared as potential affinity labels and only tested in vitro.

A comparison between the 1,2-didemethyl compound 8 and its methylene ether 7 with the corresponding isomers 9 and 6 of the 2,3-didemethyl series in the P388 assay pointed again to the importance of the substitution at C-1 observed in the series of monophenols prepared earlier.<sup>1</sup> The C<sub>1</sub>-OMe group in 6 is out of the plane of the aromatic ring, and its hydrogen bonding interaction is now being studied.<sup>16</sup> The very toxic compounds showed, with the exception of 16, 26, 29, a reasonable correlation between the toxicities determined through single intramuscular injection in mice (toxicity II in Table I) and those found during the multiple-dose studies at NCI (toxicity I in Table I; see Experimental Section). A considerable difference was also found between the toxicities determined after multiple and single injection with compounds 9 and 35. The compounds examined were almost always found to be more toxic in the multiple-dose study used in the NCI P388 assay than in the single-injection study. It is possible that the multiple doses used in the P388 assay at NCI tend to increase the toxic effect of the drug due to bioaccumulation, compared with the lower toxicity generally observed from a single dose where bioaccumulation would not be a problem. It can be observed from Table I that all the compounds found highly active in the P388 system did bind well to rat brain tubulin, confirming again that

this assay is a valuable prescreen for colchicine-related compounds. Both catecholic colchicines 8 and 9, the retinoyl derivative 31, and the *N*-phenyltetrazolyl ethers 10 and 11 showed relatively poor binding and, thus, were not tested in vivo. Nevertheless, it is noteworthy that the *N*-acyl residue can be chemically altered considerably before the tubulin binding disappears.

Colchicide, which lacks the 10-methoxy groups of colchicine, and two compounds with altered *N*-substituents, *N*-benzoyldeacetylcolchicine and *N*-carbethoxydeacetylcolchicine, all have somewhat different molecular structure than colchicine and are among the most interesting candidates discovered in this study and deserve further attention. *N*-Butyryl- and *N*-propionyldeacetylcolchicine were also found to be extremely potent, as noted earlier.<sup>13,14</sup>

### Experimental Section

Melting points are uncorrected and were taken in a Büchi-Tottoli capillary apparatus and Fisher-Johns apparatus. Alumina Stratocrom ALF 254 (5 × 10 cm) and silica gel 60 F 254 (5 × 10 cm) were used for TLC, and silica gel 60 (0.04-0.06 mm) was used for column chromatography. The solvent system used for TLC, if not otherwise noted, was CHCl<sub>3</sub>/MeOH/NH<sub>4</sub>OH, 90:9:1. The analytical samples were dried in vacuo (10 mm) over P<sub>2</sub>O<sub>5</sub> at 90-100 °C for 24 h. Microanalyses were performed in the Microanalytical Section of Istituto Superiore di Sanita (Rome) and the Laboratory of Chemistry, NIADDK, NIH, Bethesda, MD. Optical rotations were measured with a Perkin-Elmer Model 141 and 241 MC polarimeter in CHCl<sub>3</sub> and concentrations specified. UV spectra were measured in EtOH (otherwise stated) with a Hewlett Packard 8450 A spectrophotometer. Absorption maxima (λ<sub>max</sub>) are reported in nanometers, followed by log ε values in

parentheses. NMR spectra were obtained with a Varian 220 spectrometer with  $(\text{CH}_3)_4\text{Si}$  as the internal standard. Electron-ionization mass spectra (EIMS) were obtained with a LKB 2091 mass spectrometer: equipped with a Digital PDP 11 data processing system, samples applied by direct inlet, and probe usually heated from 25 to 250 °C, 70 eV, source temperature 250 °C, acceleration potential 3500 V. Chemical-ionization mass spectra (CIMS) were determined by using a Finnigan 1015D spectrometer with a Model 6000 data collection system. Relative intensities are given in parentheses.

**2-O-(*N*-Phenyltetrazolyl)-2-demethylcolchicine (10).** A mixture of 2-demethylcolchicine, (600 mg, 1.55 mmol), 5-chloro-1-phenyl-1*H*-tetrazole (330 mg, 1.83 mmol), finely ground  $\text{K}_2\text{CO}_3$  (330 mg, 2.39 mmol), and DMF (15 mL) was stirred for 48 h until the starting material disappeared according to TLC. After the addition of 50 mL of water, the material was extracted with  $\text{CH}_2\text{Cl}_2$  ( $4 \times 50$  mL), and the combined organic extracts were washed with brine, dried ( $\text{Na}_2\text{SO}_4$ ), and evaporated to dryness to give 1 g of a slightly yellowish residue. The product behaves on TLC as a single entity. The material needed for the biological experiments was obtained by flash column chromatography over silica gel and eluted with  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  (30:1) to afford a residue, which was triturated with cyclohexane to afford a pure compound (550 mg, 67%): CIMS,  $m/e$  529 ( $M^+ + 1$ ); UV  $\lambda_{\text{max}}$  233 nm ( $\log \epsilon$  2.83), 345 (2.58).

**3-O-(*N*-Phenyltetrazolyl)-3-demethylcolchicine (11)** was prepared in a manner similar to 10, from 3-demethylcolchicine, and was obtained as a yellowish powder, which was pure on TLC: CIMS,  $m/e$  529 ( $M^+ + 1$ ); UV  $\lambda_{\text{max}}$  231 nm ( $\log \epsilon$  4.05), 339 (3.87).

***N*-(Hydroxyacetyl)demecolcine (13).** To a solution of demecolcine (4; 0.8 g, 2.24 mmol) in 20 mL of  $\text{CH}_2\text{Cl}_2$  and 0.7 mL of pyridine, cooled in an ice bath, was carefully added, under stirring and anhydrous conditions, 0.43 mL (0.64 g, 3.36 mmol) of (trifluoroacetyl)glycolyl chloride. The mixture was stirred for 1 h (TLC showed the absence of 4 and the appearance of a new spot running slightly slower than demecolcine). The organic layer was washed with  $\text{H}_2\text{O}$  ( $2 \times 5$  mL), dried ( $\text{Na}_2\text{SO}_4$ ), and concentrated to leave a residue, which was chromatographed through a column of silica gel and eluted with  $\text{CHCl}_3/\text{MeOH}/\text{NH}_4\text{OH}$  (97.0:2.5:0.5) to give pure compound 13 (540 mg): mp 208–209 °C;  $[\alpha]_{\text{D}}^{22}$   $-223^\circ$  ( $c$  1.06,  $\text{CHCl}_3$ ). Anal. ( $\text{C}_{23}\text{H}_{27}\text{NO}_7$ ) C, H, N.

***O*-Acetylcolchifoline (14).** To a solution of colchifoline (12; 200 mg, 0.48 mmol) dissolved in 20 mL of dry benzene were added acetic anhydride (1 mL) and pyridine (1 mL), and the mixture was refluxed for 1 h until the TLC, on silica gel, showed the absence of colchifoline in the reaction mixture. The solvent was removed in vacuo, and the residue was diluted with ice- $\text{H}_2\text{O}$  and ammonia and extracted with  $\text{CH}_2\text{Cl}_2$ . The organic phase was dried ( $\text{Na}_2\text{SO}_4$ ) and evaporated to give a frothy yellow solid (180 mg, 82%): mp 120–124 °C; EIMS,  $m/e$  457 ( $M^+$ ).

***O*-Ethylcolchifoline (15).** To a solution of deacetylcolchicine (400 mg, 1.12 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) and (2.26 mL) pyridine (0.4 mL) was added freshly distilled ethoxyacetic anhydride at ice-bath temperature under stirring. After the solution was left standing at room temperature overnight, the solvent and the pyridine were evaporated, and the residue was taken into  $\text{CHCl}_3$ , washed with brine, dried over  $\text{Na}_2\text{SO}_4$ , and passed through a column of 20 g of silica gel. Elution with  $\text{CHCl}_3/\text{MeOH}/\text{NH}_4\text{OH}$  (80:20:2) afforded an amorphous product (640 mg) showing two spots on TLC originating from a small contamination with deacetylcolchicine and the isomeric ether: EIMS,  $m/e$  443 ( $M^+$ ).

**Preparation of *N*-Acyldeacetylcolchicine Derivatives. Method A. From a Mixture of Deacetylcolchicine and Deacetylisocholecolchicine.** The procedure adopted is as follows for compounds 18 and 19.

***N*-Propionyldeacetylcolchicine (18) and *N*-Propionyldeacetylisocholecolchicine (19).** To a crude mixture of 2 and 3, prepared from deacetylcolchicine<sup>12</sup> (2 g, 5.60 mmol) dissolved in dry benzene (50 mL) and pyridine (3 mL), was added propionic anhydride (3 mL) in one portion, and the solution was heated under stirring at 55–60 °C for 4 h [TLC:  $\text{Al}_2\text{O}_3$ ,  $\text{CHCl}_3/\text{MeOH}$  (97.5:2.5)]. The solvent was removed in vacuo, and the residue, diluted with ice- $\text{H}_2\text{O}$  and ammonia, was extracted with  $\text{CH}_2\text{Cl}_2$ . The organic layer was dried ( $\text{Na}_2\text{SO}_4$ ) and evaporated to afford an oil. The oily residue was chromatographed on a column (2.5  $\times$  80 cm) of silica gel packed with  $\text{CHCl}_3$  and eluted with

$\text{CHCl}_3/\text{MeOH}/\text{NH}_4\text{OH}$  (97.0:2.5:0.5). The more readily eluted *N*-propionyldeacetylcolchicine (18) was crystallized from EtOAc/ $\text{Et}_2\text{O}$  (300 mg, 13% from deacetylcolchicine): mp 155–157 °C (with gas evolution, shrinks at 139 °C);  $[\alpha]_{\text{D}}^{21}$   $-129^\circ$  ( $c$  1.11,  $\text{CHCl}_3$ ); EIMS,  $m/e$  413 ( $M^+$ ). Anal. ( $\text{C}_{23}\text{H}_{27}\text{NO}_6$ ) C, H, N.

The second eluted component, *N*-propionyldeacetylisocholecolchicine (19), crystallized from EtOAc- $\text{Et}_2\text{O}$  (450 mg, 19% from deacetylcolchicine): mp 205–207 °C;  $[\alpha]_{\text{D}}^{22}$   $-257^\circ$  ( $c$  0.92,  $\text{CHCl}_3$ ); EIMS,  $m/e$  413 ( $M^+$ ). Anal. ( $\text{C}_{23}\text{H}_{27}\text{NO}_6$ ) C, H, N.

Other derivatives listed below were similarly prepared by method A. The yield of the individual isomers based on deacetylcolchicine varied between 10 and 20%.

***N*-Butyryldeacetylcolchicine (20) and *N*-Butyryldeacetylisocholecolchicine (21) (with Butyryl Chloride).** Compound 20 was obtained as an amorphous yellow powder after trituration with  $\text{Et}_2\text{O}$ : mp 129–133 °C;  $[\alpha]_{\text{D}}^{21}$   $-157^\circ$  ( $c$  1.08,  $\text{CHCl}_3$ ); EIMS,  $m/e$  427 ( $M^+$ ). Anal. ( $\text{C}_{24}\text{H}_{29}\text{NO}_6 \cdot 1/2\text{H}_2\text{O}$ ) C, H, N. Compound 21: light yellow solid; mp 127–131 °C (shrinks at 110 °C);  $[\alpha]_{\text{D}}^{26}$   $-254^\circ$  ( $c$  1.05,  $\text{CHCl}_3$ ); EIMS,  $m/e$  427 ( $M^+$ ). Anal. ( $\text{C}_{24}\text{H}_{29}\text{NO}_6$ ) C, H, N.

***N*-(3,4,5-Trimethoxybenzoyl)deacetylcolchicine (23) and *N*-(3,4,5-Trimethoxybenzoyl)deacetylisocholecolchicine (24) (with 3,4,5-Trimethoxybenzoyl Chloride).** Compound 23: amorphous yellow powder; mp 166–169 °C (shrinks before melting);  $[\alpha]_{\text{D}}^{20}$   $-16^\circ$  ( $c$  0.97,  $\text{CHCl}_3$ ); EIMS,  $m/e$  551 ( $M^+$ ). Anal. ( $\text{C}_{30}\text{H}_{33}\text{NO}_9 \cdot 1/2\text{H}_2\text{O}$ ) C, H, N. Compound 24: crystallized from EtOAc-MeOH; mp 274–275 °C;  $[\alpha]_{\text{D}}^{20}$   $-92^\circ$  ( $c$  0.95,  $\text{CHCl}_3$ ); EIMS,  $m/e$  551 ( $M^+$ ). Anal. ( $\text{C}_{30}\text{H}_{33}\text{NO}_9$ ) C, H, N.

***N*-Carbethoxydeacetylcolchicine (26) and *N*-Carbethoxydeacetylisocholecolchicine (27) (with Ethyl Chloroformate and Triethylamine).** Compound 26: crystallized from EtOAc-MeOH; mp 209–211 °C;  $[\alpha]_{\text{D}}^{20}$   $-173^\circ$  ( $c$  1.0,  $\text{CHCl}_3$ ); EIMS,  $m/e$  429 ( $M^+$ ). Anal. ( $\text{C}_{23}\text{H}_{27}\text{NO}_7$ ) C, H, N. Compound 27: crystallized from EtOAc; mp 202–203 °C;  $[\alpha]_{\text{D}}^{20}$   $-249^\circ$  ( $c$  0.96,  $\text{CHCl}_3$ ); EIMS,  $m/e$  429 ( $M^+$ ). Anal. ( $\text{C}_{23}\text{H}_{27}\text{NO}_7$ ) C, H, N.

**Method B. From Deacetylcolchicine. *N*-Pivaloyldeacetylcolchicine (22).** A solution of *N*-deacetylcolchicine (500 mg, 1.40 mmol) in dry benzene (30 mL), pyridine (1 mL), and trimethylacetic anhydride (1 mL) was stirred at 55–60 °C. Acylation was complete after 7–8 h (TLC). The mixture was concentrated in vacuo, and the residue was diluted with  $\text{H}_2\text{O}$  and ammonia and extracted with  $\text{CH}_2\text{Cl}_2$ . The organic layer was dried ( $\text{Na}_2\text{SO}_4$ ) and evaporated to give a yellow solid, which was purified through column chromatography [ $\text{Al}_2\text{O}_3$  activity II, with  $\text{CHCl}_3/\text{MeOH}$  (99.5:0.5) as eluent] to give *N*-pivaloyldeacetylcolchicine (22) as an amorphous powder (350 mg, 57%): mp 139–143 °C (with gas evolution);  $[\alpha]_{\text{D}}^{21}$   $-168^\circ$  ( $c$  0.94,  $\text{CHCl}_3$ ); EIMS,  $m/e$  441 ( $M^+$ ). Anal. ( $\text{C}_{25}\text{H}_{31}\text{NO}_6$ ) C, H, N.

Other derivatives prepared by method B are listed below. The yield of the desired compounds varied between 50 and 70% based on deacetylcolchicine.

***N*-Benzoyldeacetylcolchicine (16):** triturated with  $\text{Et}_2\text{O}$  to afford a pure solid compound; mp 260 °C;  $[\alpha]_{\text{D}}^{26}$   $-47^\circ$  ( $c$  0.96,  $\text{CHCl}_3$ ); EIMS,  $m/e$  461 ( $M^+$ ); CIMS,  $m/e$  462 ( $M^+ + 1$ ). Anal. ( $\text{C}_{27}\text{H}_{27}\text{NO}_6$ ) C, H, N.

***N*-Butyryldeacetylcolchicine (20):** mp 129–133 °C; all the data (IR and MS) were identical with the material obtained earlier by method A.

***N*-(Phenoxyacetyl)deacetylcolchicine (29):** crystallized from 2-propanol- $\text{Et}_2\text{O}$ ; mp 235 °C;  $[\alpha]_{\text{D}}^{26}$   $-91^\circ$  ( $c$  1.11,  $\text{CHCl}_3$ ); EIMS,  $m/e$  491 ( $M^+$ ); CIMS,  $m/e$  492 ( $M^+ + 1$ ). Anal. ( $\text{C}_{28}\text{H}_{29}\text{NO}_7$ ) C, H, N.

**Acetate of *N*-sinapoyldeacetylcolchicine (32) (with acetoxysinapoyl chloride<sup>17</sup>):** obtained on trituration with  $\text{Et}_2\text{O}$  (2 mL) to afford pure solid; mp 168 °C;  $[\alpha]_{\text{D}}^{26}$   $+96^\circ$  ( $c$  0.60,  $\text{CHCl}_3$ ); UV  $\lambda_{\text{max}}$  231 nm ( $\log \epsilon$  2.08), 296 (4.07), 344 (3.93); EIMS,  $m/e$  605 ( $M^+$ ); CIMS,  $m/e$  606 ( $M^+ + 1$ ). Anal. ( $\text{C}_{33}\text{H}_{35}\text{NO}_{10}$ ) C, H, N.

***N*-Sinapoyldeacetylcolchicine (33).** To a solution of 32 (100 mg, 0.16 mmol) in acetone (5 mL) was added dropwise concentrated ammonia solution (4.5 mL), and the reaction mixture was stirred at room temperature for 2 h, until TLC showed the absence of starting material in the reaction mixture. Acetone was

removed at high vacuum at room temperature to leave the aqueous layer. It was diluted with water (10 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (4  $\times$  5 mL). The combined organic layer was washed with 10% aqueous  $\text{NaHCO}_3$  solution (3  $\times$  10 mL), dried ( $\text{Na}_2\text{SO}_4$ ), and concentrated at 40 °C under reduced pressure to leave a solid residue, which was taken into  $\text{Et}_2\text{O}$  (2 mL) and triturated to afford a pure (33; 85 mg, 94%): mp 160 °C;  $[\alpha]^{26}_{\text{D}} +98^\circ$  (c 0.40,  $\text{CHCl}_3$ ); UV  $\lambda_{\text{max}}$  242 nm (log  $\epsilon$  3.44), 334 (3.40); UV ( $\text{EtOH} + \text{NaOH}$ )  $\lambda_{\text{max}}$  247 nm (log  $\epsilon$  3.78), 376 (3.79); EIMS,  $m/e$  563 ( $\text{M}^+$ ). Anal. ( $\text{C}_{31}\text{H}_{33}\text{NO}_5$ ) C, H, N.

**N-Retinoyldeacetylcolchicine (31).** A solution of deacetylcolchicine (2; 100 mg, 0.28 mmol), retinoic acid (100 mg, 0.33 mmol), dicyclohexylcarbodiimide (DCC; 100 mg, 0.48 mmol), and 4-pyrrolidinopyridine (10 mg, catalytic amounts) in dry  $\text{CH}_2\text{Cl}_2$  (3 mL) was stirred overnight at room temperature. The reaction mixture was filtered, and the filtrate was washed with  $\text{H}_2\text{O}$  (3  $\times$  3 mL), 10% aqueous  $\text{NaHCO}_3$  solution (3  $\times$  3 mL), 0.1 N  $\text{HCl}$  (3  $\times$  3 mL), and  $\text{H}_2\text{O}$  (3  $\times$  3 mL), dried ( $\text{Na}_2\text{SO}_4$ ), and evaporated to leave a residue, which was taken into  $\text{Et}_2\text{O}$ . The solid was filtered off, and the filtrate was concentrated to afford an oily residue, which was crystallized from a mixture of  $\text{Et}_2\text{O}$ -petroleum ether to afford a yellow crystalline solid (116 mg, 65%): mp 149 °C;  $[\alpha]^{21}_{\text{D}} +16^\circ$  (c 0.65,  $\text{CHCl}_3$ ); UV  $\lambda_{\text{max}}$  243 nm (log  $\epsilon$  3.81), 350 (4.15); EIMS,  $m/e$  639 ( $\text{M}^+$ ); CIMS,  $m/e$  640 ( $\text{M}^+$ ). Anal. ( $\text{C}_{40}\text{H}_{49}\text{NO}_6$ ) C, H, N.

**Hydrolysis of N-Acyldeacetylcolchicine Derivatives.** The general procedure followed is illustrated with the preparation of 25.

**N-(Trimethoxybenzoyl)deacetylcolchicine (25).** A mixture of 23 and 24 (600 mg) dissolved in 25 mL of  $\text{MeOH}$  and 25 mL of 0.5 N  $\text{HCl}$  was heated under stirring (oil bath, 100–110 °C) until TLC indicated that almost all of the starting material had disappeared (ca. 10 h). It was kept overnight in a refrigerator, and the precipitate was collected and then washed thoroughly with cold  $\text{H}_2\text{O}$ , and the solid was crystallized with  $\text{MeOH-H}_2\text{O}$  to afford 25 (460 mg, 78%): mp 147 °C (shrinks at 137 °C);  $[\alpha]^{20}_{\text{D}} -92^\circ$  (c 0.95,  $\text{CHCl}_3$ ); EIMS,  $m/e$  537 ( $\text{M}^+$ ). Anal. ( $\text{C}_{29}\text{H}_{31}\text{NO}_6$ ) C, H, N.

Other *N*-acyldeacetylcolchicine derivatives were hydrolyzed in the same way, and the colchicine derivatives were obtained in 60 to 80% yield and characterized as follows.

**N-Benzoyldeacetylcolchicine (17):** crystallized from  $\text{MeOH}$ ; mp 250 °C;  $[\alpha]^{26}_{\text{D}} -149^\circ$  (c 0.20,  $\text{CHCl}_3$ ); EIMS,  $m/e$  447 ( $\text{M}^+$ ). Anal. ( $\text{C}_{26}\text{H}_{25}\text{NO}_6$ ) C, H, N.

**N-Carboethoxydeacetylcolchicine (28):** mp 172–173 °C;  $[\alpha]^{20}_{\text{D}} -240^\circ$  (c 1.0,  $\text{CHCl}_3$ ); EIMS,  $m/e$  415 ( $\text{M}^+$ ). Anal. ( $\text{C}_{22}\text{H}_{25}\text{NO}_7$ ) C, H, N.

**2-Demethyl-N-(trifluoroacetyl)deacetylcolchicine (30).** A solution of *N*-(trifluoroacetyl)deacetylcolchicine (1.35 g, 3.07 mmol) in concentrated  $\text{H}_2\text{SO}_4$  (5 mL) was stirred for 3 h at 60–70 °C bath temperature. The reaction mixture was cooled, ice was added, and the pH was adjusted to 11.0 with 2 N  $\text{NaOH}$  and extracted several times with  $\text{CH}_2\text{Cl}_2$ /2-propanol (1:1) to afford an organic residue. This material was dissolved in  $\text{CH}_2\text{Cl}_2$ , the insoluble material was filtered off, and the residue obtained on

evaporation of the filtrate was passed through a column of silica gel, with  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  (95:5) as eluent. The solid obtained was crystallized from a mixture of  $\text{CH}_2\text{Cl}_2$ /hexane to afford yellow crystals (325 mg, 24%): mp 270 °C (softens at 170 °C); EIMS,  $m/e$  439 ( $\text{M}^+$ ). Anal. ( $\text{C}_{21}\text{H}_{20}\text{NF}_3\text{O}_5$ ) C, H, N.

**Colchicide (35).** To a solution of thiocholchicine (430 mg, 1.03 mmol) in dry acetone (80 mL), under a nitrogen atmosphere, was added 5 g of wet Raney nickel catalyst (washed with 3  $\times$  10 mL of acetone), and the mixture was stirred at room temperature. The reaction was monitored by TLC [silica gel,  $\text{CHCl}_3/\text{MeOH}$  (95:5)]. After 2 h, the second portion of catalyst (5 g) was added, and stirring was continued for 15 h. TLC showed a main spot corresponding to colchicide ( $R_f$  0.30) and minor spots corresponding to thiocholchicine ( $R_f$  0.42) and hexahydrodemethoxycolchicine ( $R_f$  0.13).<sup>7</sup> The catalyst was filtered over a pad of Celite, and the red solution was evaporated. The residue was chromatographed on a column (40  $\times$  2 cm) of silica gel and eluted with  $\text{CHCl}_3/\text{MeOH}$  (97.5:2.5) to afford a glassy solid (176 mg, 46%):  $[\alpha]^{24}_{\text{D}} -166^\circ$  (c 0.53,  $\text{CHCl}_3$ ) [lit.<sup>7</sup>  $[\alpha]^{27}_{\text{D}} -196^\circ$  (c 0.47,  $\text{CHCl}_3$ )]. Anal. ( $\text{C}_{21}\text{H}_{23}\text{O}_5\text{N} \cdot 1/2\text{H}_2\text{O}$ ) C, H, N. The hydrochloride salt was crystallized from  $\text{EtOAc-MeOH}$ : mp 124–126 °C (lit.<sup>7</sup> 119–121 °C);  $[\alpha]^{18}_{\text{D}} -456^\circ$  (c 0.50,  $\text{CHCl}_3$ ); EIMS,  $m/e$  369 ( $\text{M}^+$ );  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  2.00 (s, 3 H, COMe), 2.2–2.6 (m, 4 H, 2  $\text{CH}_2$ ), 3.69 (s, 3 H,  $\text{C}_1$  OMe), 3.92 and 3.90 (2 s, each 3 H,  $\text{C}_2$  OMe and  $\text{C}_3$  OMe), 4.68 (m, 1 H, C, H), 6.58 (s, 1 H,  $\text{C}_4$  H) 7.40 (s, 1 H,  $\text{C}_8$  H), 7.20–7.60 (m, 3 H,  $\text{C}_{10}$  H,  $\text{C}_{11}$  H,  $\text{C}_{12}$  H), 7.66 (d, NH, exchanges on addition of  $\text{D}_2\text{O}$ ).

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