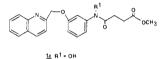
# Leukotriene $D_4$ Antagonists and 5-Lipoxygenase Inhibitors. Synthesis of Benzoheterocyclic [(Methoxyphenyl)amino]oxoalkanoic Acid Esters

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A series of novel benzoheterocyclic [(methoxyphenyl)amino]oxoalkanoic acid esters has been prepared. These compounds were tested as inhibitors of rat polymorphonuclear leukocyte 5-lipoxgenase (LO) in vitro and as inhibitors of leukotriene  $D_4$  (LTD<sub>4</sub>) and ovalbumin (OA) induced bronchospasm in the guinea pig (GP) in vivo. In general, inhibitory activity against 5-LO, LTD4, and OA was broadest for benzthiazole-containing analogues (benzthiazole > benzimidazole >> benzoxazole, benzofuran). The most potent 5-LO inhibitor, 4-[[3-(2-benzthiazolylmethoxy)phenyl]hydroxyamino]-4-oxobutanoic acid methyl ester (7), had an IC<sub>50</sub> of 0.36  $\mu$ M. Compound 7, however, was inactive vs. OA. The most potent compound in vivo, 4-[[3-[(1-methyl-2-benzimidazolyl)methoxy]phenyl]amino]-4-oxobutanoic acid methyl ester 4, inhibited both LTD4- and OA-induced bronchospasm by 83% and 60%, respectively, at 50 mg/kg intraduodenally. Compound 4 was studied in the Ames assay employing five strains of bacteria (TA1535, TA1537, TA1538, TA98, and TA100) with and without S-9 rat liver enzyme metabolic activation, and there was no significant number of reversions noted.

We have recently shown that 4-[[3-(2-quinolinylmethoxy)phenyl]hydroxyamino]-4-oxobutanoic acid methyl ester (1a) is a potent inhibitor of rat polymorphonuclear leukocyte (PMN) 5-lipoxygenase (5-LO) in vitro and is a potent inhibitor of both leukotriene  $D_4$  (LTD<sub>4</sub>) and ovalbumin (OA) induced bronchospasm in the guinea pig (GP).<sup>1,2</sup> This paper covers a portion of our continuing investigation of this novel structural class and is concerned with the synthesis and biological evaluation of heterocyclic groups other than quinoline and/or phenyl side chains other than aminooxoalkanoic acid esters.



As pointed out by us, compound 1a is not being further developed as a drug candidate because of its mutagenic activity in the Ames<sup>3</sup> assay. Certain aryl hydroxamic acids are known to form sulfate or acetate esters in vivo, which then heterolytically cleave to give arylacylnitrenium ions as the metabolic ultimate electrophilic mutagens.<sup>4</sup> Since 1a contains an aryl hydroxamic acid, we thought this portion of the molecule was suspect. Alternatively, the (quinolinylmethoxy)phenyl system of 1a may be acting as an intercalating agent.<sup>5</sup>

Therefore, our goals were to discover new nonmutagenic analogues of compound 1 that retain potent  $LTD_4$  antagonist activity in vivo. Past experience in this series has shown that a 2-oxymethyl nitrogen heterocycle, meta substitution, and a terminal alkyl carboxylate are elements needed for  $LTD_4$  antagonist activity. We chose to sub-

stitute a benzthiazole because it is a bioisostere of quinoline.<sup>6</sup> Benzimidazole and benzoxazole were employed because they extend the 2-oxymethyl nitrogen heterocyclic series and benzofuran would examine the effect of a 2oxymethyl oxygen heterocycle. With respect to phenyl substitution, we prepared some 2-oxopyrrolidine-4carboxylates because this moiety is found in previously reported LT antagonists.7

### Chemistry

The synthetic pathways for the preparation of compounds 2-14 listed in Table I are shown in Scheme I. Reaction of 2-aminophenol, N-methyl-2-phenylenediamine, or 2-aminophenol with methyl chloroacetimidate<sup>8</sup> gave 2-chloromethyl heterocycles 16. Treatment of 16 with 3-nitrophenol in acetone with cesium carbonate<sup>9</sup> at reflux provided the bis-aryl system 17. Reduction of 17 with iron and ethanolic HCl followed by treatment with 3-carbomethoxypropionyl chloride gave compounds 2-4. The reagents used in the preparation of the remaining target compounds 5-14 are noted in Scheme I and details of the reactions are provided in the Experimental Section.

The preparation of target compounds 18-24 listed in Table II started with 3-aminophenol (Scheme II). Fusion of 3-aminophenol with itaconic acid followed by acidcatalyzed esterification provided intermediate 25. Alkylation of 25 with the appropriate chloromethyl benzoheterocycle 16 gave compounds 18-21. Condensation of 3-aminophenol with 3-carbomethoxypropionyl chloride provided intermediate 26. Reaction of 26 with the appropriate benzoheterocycle 16 in acetone with cesium carbonate at reflux effected both alkylation and cyclization to give target compounds 22-24.

Physical data for key intermediates in Schemes I and II are summarized in Table IV.

### **Biological Results and Discussion**

The results obtained for compounds 2-14 and 18-24 as inhibitors of rat PMN 5-LO and LTD<sub>4</sub>- and OA-induced bronchospasm in the GP are listed in Tables I and II. As expected on the basis of bioisosteric equivalence, benzthiazole 2 was equiactive both in vitro and in vivo with quinoline 1b. Unexpectedly, benzimidazole 4 was

Musser, J. H.; Kubrak, D. M.; Chang, J.; Lewis, A. J. J. Med. (1)Chem. 1986, 29, 1429.

Compound 1a was recently retested as an antagonist of LTD<sub>4</sub>-induced contractions of isolated GP tracheal spiral strips and was shown to be a competitive inhibitor with a  $pk_B$  value of 6.60, which is different than the  $\mathrm{p}k_\mathrm{B}$  value of 5.33 reported in ref 1. The reason for the difference is that L-cysteine was added to inhibit conversion of LTD<sub>4</sub> to LTE<sub>4</sub>. Furthermore, 1a does not inhibit  $LTC_4$  in the presence of glutathione and it does not inhibit histamine dose-response curves.

<sup>(3)</sup> Ames, B. N.; McCann, J.; Yamasaki, E. Mutat. Res. 1975, 31,

<sup>(4)</sup> Gassman, P. G.; Granrud, J. E. J. Am. Chem. Soc. 1984, 106, 1498 and references cited therein.

Epler, J. L.; Winton, W.; Ho, T.; Larimer, F. W.; Rao, T. K.; (5)Hardigree, A. A. Mutat. Res. 1977, 39, 285.

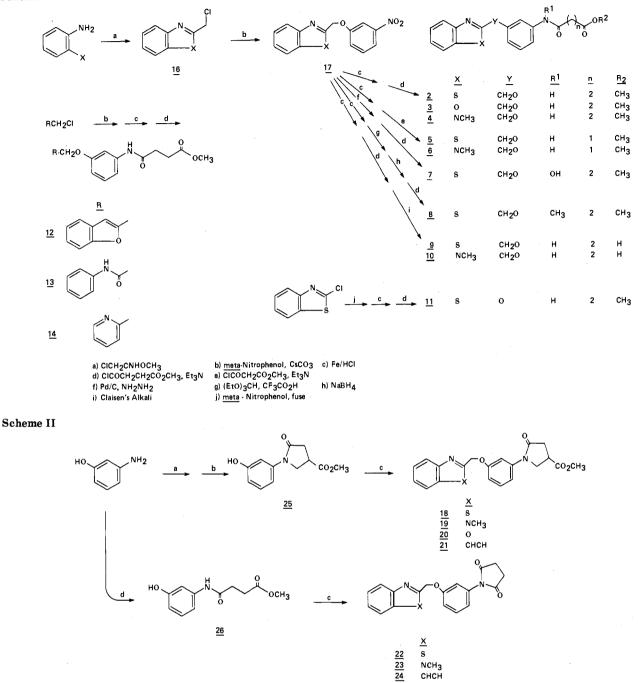
<sup>(6)</sup> Thornber, C. W. Chem. Soc. Rev. 1979, 18, 564.

Kadin, S. B. U. S. Patent 4 296 120. (7)

Roger, R.; Neilson, A. G. Chem. Rev. 1961, 61, 179. Zaugg, H. E. J. Org. Chem. 1976, 41, 3419. (8)

<sup>(9)</sup> 

### Scheme I



a) Itaconic acid, fuse b) MeOH, pTSA c) Cs<sub>2</sub>CO<sub>3</sub>, <u>16</u> d) CICOCH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>CH<sub>3</sub>, Et<sub>3</sub>N

equiactive with 1b, whereas benzoxazole 3 was essentially inactive in all three assays. Furthermore, the benzthiazole malonate 5 was equiactive with 1b and 2, whereas the benzimidazole malonate 6 was only active vs.  $LTD_4$ -induced bronchospasm.

In general, further modification of the benzoheterocyclic system lowered potency or eliminated activity. The pyrido analogue of 1b was prepared to determine the importance of the fused benzyl ring; pyrido analogue 14 was completely inactive. Results obtained for benzofuran 12 and acetanilide 13 indicated that a 2-oxymethyl oxygen heterocycle and an oxyacetanilide, respectively, could not be substituted for the 2-oxymethylquinoline and retain activity.

Several modifications made in the quinoline series<sup>1</sup> were also made in this study for comparison purposes. In most cases the results were consistent with our earlier findings. For example, carboxylic acids (9, 10) were less potent than the corresponding esters (2, 4) and removing the ether link (11) or methylating the amide (8) abolished activity in vivo. However, benzthiazole hydroxamic acid 7 relative to quinoline hydroxamic acid 1a was more potent in vitro, less potent vs. LTD<sub>4</sub>-induced bronchospasm, and inactive vs. OA-induced bronchospasm.

Certain compounds with a pyrrolidinone ring instead of the succinate group on the phenyl retained activity. Like the above series, methyl 2-oxopyrrolidine-4-carboxylates containing benzthiazole (18) and quinoline (21) were active as inhibitors of 5-LO and  $LTD_4$ -induced bronchospasm and benzoxazole (20) was inactive. In contrast, the benzimidazole 2-oxopyrrolidine-4-carboxylate 19 was not significantly active. It is interesting that compound 8 was inactive. Possibly the methyl group presents some steric

402

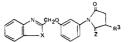
Table I. Inhibition of Rat PMN 5-LO and GP Bronchospasm

# $\begin{array}{c} \begin{array}{c} & & & \\ & &$

												vitro IN:	in v	vivo (50	mg/kg id	)
									synth			$^{a}$ $\mu M$	LTI	) <sub>4</sub> <sup>b</sup>	OA	
no.	Х	Y	R1	n	$\mathbb{R}^2$	mp	, °C	formula <sup>d</sup>	meth	% yield	5-LO	CO	% inh	$N^c$	% inh	N
1a	CHCH	CH <sub>2</sub> O	OH	2	CH <sub>3</sub>	187-	-189 C <sub>21</sub> I	$H_{20}N_2O_5$	f	69	1.4	40	97*	8	76*	13
1 <b>b</b>	CHCH	$CH_2O$	н	$^{2}$	$CH_3$	121-		$H_{20}N_2O_4$	f	58	28	$NA^{g}$	86*	3	43	-6
2	$\mathbf{S}$	$CH_2O$	н	<b>2</b>	$CH_3$	129-	-130 C <sub>19</sub> I	$H_{18}N_2O_4S$	2	83	15.7	NA	41*	6	63*	6
3	0	$CH_2O$	н	2	$CH_3$	104-	-105 C <sub>19</sub> I	$H_{18}N_2O_5$	2	66	69.7	NA	3	<b>2</b>	-65	2
4	$NCH_3$	$CH_2O$	н	<b>2</b>	$CH_3$	136-		$H_{21}N_3O_4$	$^{2}$	69	7.2	NA	83*	6	60*	
5	$\mathbf{S}$	$CH_2O$	$\mathbf{H}$	1	$CH_3$	109-		$H_{16}N_2O_4S$	$^{2}$	64	17.0	NA	77*	6	58*	(
6	NCH <sub>3</sub>	$CH_2O$	$\mathbf{H}$	1	$CH_3$	167-	-169 C <sub>19</sub> I	$H_{19}N_{3}O_{4} \cdot 1/_{10}H_{2}O$	2	81	NA		60*	6	5	
7	$\mathbf{s}$	$CH_2O$	OH	<b>2</b>	$CH_3$	123-	-125 C <sub>19</sub> I	$H_{18}N_2O_5S$	7	82	0.36	NA	79*	6	6	
8	$\mathbf{S}$	$CH_2O$	$CH_3$	<b>2</b>	$CH_3$	107-		$H_{20}N_2O_4S-1.4HCl$	8	63	21	NA	21	3	19	;
9	$\mathbf{s}$	$CH_2O$	$\mathbf{H}$	<b>2</b>	Н	191-		$H_{16}N_2O_4S \cdot 1/_2H_2O_4$	9	99	NA		57*	3		
10	NCH <sub>3</sub>	$CH_2O$	$\mathbf{H}$	<b>2</b>	н	213 -		$H_{19}N_{3}O_{4}$	9	86	NA		30	3		
11	$\mathbf{S}$	0	н	<b>2</b>	$CH_3$	78-	-82 C <sub>18</sub> I	$H_{16}N_2O_4S$	11	97	27.1	NA	-2	3	7	
								<u>~</u>			in vi PM		in vi	vo (50	mg/kg id)	
									synth		$IC_{50}^{a}$		LTD	· · ·	OA	
	no.	R		$\mathbb{R}^1$	n	$\mathbb{R}^2$	mp, °C	formula <sup>d</sup>	$meth^e$	% yield	<u>5-L0</u>	co	% inh	N <sup>c</sup>	% inh	Ν
12				н	2	$CH_3$	112-113	C <sub>20</sub> H <sub>19</sub> NO <sub>5</sub>	12	27	17.9	NA	-8	2	12	2
13			<b>\</b>	Н	2	$CH_3$	164–165	$C_{19}H_{20}N_2O_5$	2	73	NA		25	2	7	2
14			ö	н	2	$CH_3$	97-98	$C_{17}H_{18}N_2O_9$	2	86	NA		-1	2		
	V 5901) <sup>12</sup> 171,883) <sup>13</sup>										0.3 18.9	NA NA	44.2 92.6*	3 6	60.2* 79.7*	6 6

<sup>*a*</sup> All IC<sub>50</sub> values were calculated by nonlinear regression analysis and were significant at the p < 0.05 level. <sup>*b*</sup> Starred results (\*) are statistically significant with the two-tail Student's t tests (p < 0.05). <sup>*c*</sup> N = number of animals. <sup>*d*</sup> All compounds had elemental analysis (C, H, N) within ±0.4 of theoretical value. <sup>*e*</sup> Experimental Section compound number. <sup>*f*</sup> See ref 1. <sup>*g*</sup> NA <50% inhibition at 100  $\mu$ M.

Table II. Inhibition of Rat PMN 5-LO and GP Bronchospasm



						<u></u>		in vitro PMN:		in vivo (50 mg/kg id)			
								IC <sub>50</sub> ,ª		LTD	4 <sup>b</sup>	OAb	,
no.	х	$\mathbf{Z}$	$\mathbb{R}^3$	mp, °C	formula <sup>d</sup>	synth meth <sup>e</sup>	% yield	5-LO	CO	% inh	$N^c$	% inh	N
18	S	H,	CO <sub>2</sub> CH <sub>3</sub>	99-102	$C_{20}H_{18}N_2O_4S$	18	62	14.0	NA	67*	3		
19	NCH <sub>3</sub>	$H_{2}$	$CO_2CH_3$	131 - 133	$C_{21}H_{21}N_{3}O_{4}$	18	53	$NA^{f}$		26	<b>2</b>		
<b>20</b>	0 Č	$\tilde{H_{9}}$	$CO_2CH_3$	107 - 109	$C_{20}H_{18}N_2O_5$	18	61	30.0	NA	15	2		
21	CHCH	$H_{2}$	$CO_{2}CH_{3}$	101-104	$C_{22}H_{20}N_2O_4 \cdot 1/10H_2O$	18	39	19.1	NA	89*	6	29	6
<b>22</b>	s	ດ້	H	175-176	$C_{18}H_{14}N_2O_3S$	22	56	NA		6 <b>8</b> *	6	18	6
23	NCH <sub>3</sub>	0	н	150 - 152	$C_{19}H_{17}N_{3}O_{3}$	22	49	NA		79*	3		
24	СНСЙ	0	н	159 - 160	$C_{20}H_{16}N_2O_3$	22	32	NA		88*	6	26	6

<sup>a</sup>All IC<sub>50</sub> values were calculated by nonlinear regression analysis and were significant at the p < 0.05 level. <sup>b</sup>Starred results (\*) are statistically significant with the two-tail Student's t test (p < 0.05). <sup>c</sup>N = number of treated animals. <sup>d</sup>All compounds had elemental analysis (C, H, N) within ±0.4 of theoretical value. <sup>e</sup>Experimental Section compound number. <sup>f</sup>NA <50% inhibition at 100  $\mu$ M.

Table III. Ames Test Results for Compounds 1a, 1b, 21, and  $24^a$ 

Table IV. Physical Data for Key Intermediates<sup>a</sup>

		mean no. of revertants <sup><math>b</math></sup>				
test article	<b>a</b> mount per plate	without activation	with activa- tion <sup>c</sup>			
vehicle control <sup>d</sup>	0.2 mL	14	13			
positive control <sup>e</sup>	0.1 mg	2710	738			
la	0.005 mg	15	13			
	0.015	9	10			
	0.05	13	16			
	0.15	31	12			
	0.5	943	716			
1 <b>b</b>	0.05 mg	24	15			
	0.15	25	15			
	0.5	696	566			
	1.5	2930	2767			
	5.0	2258	2879			
vehicle control	0.2 mL	7	12			
positive control	0.1 mg	1376	634			
<b>2</b> 1	0.02 mg	10	12			
	0.06	13	10			
	0.2	16	10			
	0.6	421	15			
	2.0	475	84			
vehicle control	0.2  mL	8	5			
positive control	0.1 mg	1350	1103			
24	0.005 mg	6	5			
	0.015	9	7			
	0.05	12	7			
	0.15	43	8			
	0.5	155	45			

<sup>a</sup> None of the test compounds produced a significant amount of revertants in strains TA98, TA100, TA1535, and TA1538, either in the presence or absence of the metabolic activation system. <sup>b</sup> Three plates per treatment; two plates in cases of toxic response or contamination. <sup>c</sup>Activation provided by S-9 rat liver homogenate. <sup>d</sup> Me<sub>2</sub>SO-d<sub>6</sub>. <sup>e</sup>Without activation, 9-aminoacridine; with activation, 2-aminoanthracene.

interaction that is alleviated in the constrained pyrrolidinone ring.

Pyrrolidine-2,5-dione 24 was determined to be a metabolite of 1b.<sup>10</sup> Apparently, compound 1b is hydrolyzed to the corresponding acid, which is interconvertible with the cyclic imide 24. Compound 24 was prepared and was found to be an inhibitor of  $LTD_4$ -induced bronchospasm. As expected, both benzthiazole 22 and benzimidazole 23 were also  $LTD_4$  antagonists in vivo. None of the pyrrolidinedione-containing compounds were active as 5-LO inhibitors.

(10) Tio, C. Internal Correspondence, Wyeth Laboratories, Inc.

x mp, °C R formula<sup>b</sup>  ${}^{\mathrm{S}}_{\mathrm{S}}$  $NO_2$ 156 - 158 $C_{14}H_{10}N_{2}O_{3}S$  $NH_2$ 118-120  $C_{14}H_{12}N_2OS$ C<sub>15</sub>H<sub>13</sub>N<sub>3</sub>O<sub>5</sub> NCH<sub>3</sub>  $NO_2$ 183-185  $C_{15}H_{15}N_{3}O^{1}/_{4}H_{2}O$ NCH.  $NH_{2}$ 149-153 188-189 C12H13NO4 25

<sup>a</sup> (Chloromethyl)benzimidazole and (chloromethyl)benzthiazole are known compounds; see ref 14 and 15, respectively. <sup>b</sup>All compounds had elemental analysis (C, H, N) within  $\pm 0.4$  of theoretical value.

The most potent 5-LO inhibitor compound 7 (IC<sub>50</sub> of 0.36  $\mu$ M in vitro) was not active in vivo against OA challenge. Indeed, no significant correlation between inhibition of 5-LO and inhibition of OA-or LTD<sub>4</sub>-induced bronchospasm in vivo was seen, which is consistent with our earlier study.

In Tables I and II there are 11 compounds that had significant inhibitory activity against  $LTD_4$ -induced bronchospasm (2, 4–7, 9, 18, and 21–24). Only three compounds had significant inhibitory activity against OA-induced bronchospasm (2, 4, and 5). This may be due to the differences in the absorption, distribution, or metabolism of the compounds or the difference with exogenously applied  $LTD_4$  vs. endogenously generated  $LTD_4$ .

Compounds 2 and 4 were chosen as biological leads and studied further in the Ames assay. Results obtained for compounds 1a and 1b are reported herein for the first time and are included to furnish some insight into their mechanism of action as mutagens. Compound 21 was examined in the Ames assay because it is a potent  $LTD_4$ antagonist that could not be readily metabolized to a hydroxamic acid, and compound 24 was examined because it is a metabolite of 1b that may have been responsible for the observed mutagenic activity.

The assay used is essentially the one Ames recommends for general mutagenicity testing.<sup>3</sup> Three standard tester strains of *Salmonella typhunium* bacteria, TA1535, TA1537, and TA1538, are used in combination with two R factor *S. typhunium* strains, TA100 and TA98. Experimental compounds were examined in the absence and presence of S-9 rat liver homogenates with elevated microsomal enzyme activity.

## 404 Journal of Medicinal Chemistry, 1987, Vol. 30, No. 2

Quinolines 1a, 1b, 21, and 24 demonstrated reactivity in strain TA1537 (Table III). Quinoline 24 gave a variable response, usually one per dose of a three-plate assay at higher doses. Mutagenic activity in tester strain TA1537 suggests that these compounds cause frameshift mutations and implies that these compounds are acting as intercalating agents and not as electrophilic agents.

Benzthiazole 2 and benzimidazole 4 were inactive in all of the tester strains both in the absence and presence of microsomal activation at dose ranges of 0.005-0.5 mg and 0.05-5.0 mg per plate, respectively.<sup>11</sup>

We have described the synthesis and biological activity of novel benzoheterocyclic[(methoxyphenyl)amino]oxoalkanoic acid esters as inhibitors of rat PMN 5-LO and as inhibitors of  $LTD_4$ - and OA-induced bronchospasm in the GP. Our initial goals of retaining the in vivo  $LTD_4$  antagonist activity of 1 while eliminating mutagenic activity were successful in compounds 2 and 4. Since the combination of activity constitutes a novel therapeutic approach with potential for the treatment of asthma and related allergies, further investigation with these compounds is warranted.

#### **Experimental Section**

Melting points were determined on a Thomas-Hoover apparatus and are uncorrected. Spectra were recorded for all compounds and were consistent with assigned structures. NMR spectra were recorded on a Varian XL-300 at 300 MHz, a Varian XL-100 at 100 MHz, or a Varian FT-80A at 80 MHz. Mass spectra were recorded on a Kratos MS-25. IR spectra were recorded with Perkin-Elmer 299 infrared spectrophotometer. Elemental analysis were recorded with a Perkin-Elmer 240C elemental analyzer, and all compounds were within 0.4% of theoretical value.

4-[[3-(2-Benzthiazolylmethoxy)phenyl]amino]-4-oxobutanoic Acid Methyl Ester (2). (A) 2-(Chloromethyl)benzthiazole. To a solution of 2-aminothiophenol (7.5 g, 60 mmol) in methylene chloride at 0 °C was added methyl chloroacetimidate hydrochloride<sup>7</sup> (8.6 g, 60 mmol). The reaction mixture was allowed to warm to room temperature while being stirred overnight. The mixture was washed with water (3×), dried (MgSO<sub>4</sub>), and concentrated to an oil. The oil was distilled [120-135 °C (0.5 mmHg)] to give 7.8 g (71% yield) of product.

(B) 3-(2-Benzthiazolylmethoxy)nitrobenzene. A mixture of 2-(chloromethyl)benzthiazole (17.6 g, 95 mmol), 3-nitrophenol (13.2 g, 95 mmol), cesium carbonate (30 g), sodium carbonate (10 g), potassium iodide (0.5 g), and acetone (600 ml) was heated at reflux overnight. The mixture was filtered and the resulting solution was partially concentrated. A crystalline solid formed, which was filtered and dried, giving 22.8 g (84% yield) of product, mp 157–158 °C.

(C) 3-(2-Benzthiazolylmethoxy)aniline. To a suspension of 3-(2-benzthiazolylmethoxy)nitrobenzene (18.6 g, 65 mmol) in ethanolic hydrochloric acid was added powdered iron. The reaction was stirred overnight at room temperature. After neutralization with saturated aqueous sodium bicarbonate, the mixture was extracted with methylene chloride ( $3\times$ ). The extract was dried (MgSO<sub>4</sub>) and concentrated to give 15.1 g (90% yield) of product, mp 117-120 °C.

(D) Compound 2. To a solution of 3-(2-benzthiazolylmeth-

- (11) Benzthiazole 2 initially appeared to be active in tester strain TA1537. However, upon retesting in a liquid suspension assay, which permits treatment of the bacteria followed by removal of the test article during the incubation period, this compound was also found to be inactive.
- (12) Musser, J. H.; Chakraborty, U.; Scortino, S.; Gordon, R. J.; Khandwala, A.; Neiss, E. S.; Pruss, T. P.; Inwegen, R.; Weinryb, I.; Coutts, S. M. J. Med. Chem., in press.
- (13) Fleisch, J. H.; Rinkema, L. E.; Haisch, K. D.; Swanson-Bean, D.; Goodson, T.; Ho, P. P. K.; Marshall, W. S. J. Pharmacol. Exp. Ther. 1985, 233, 148.
- (14) King, F. E.; Acheson, R. M.; Spensley, P. C. J. Chem. Soc. 1949, 1401.
- (15) Kristinsson, H. Synthesis 1979, 102.

oxy)aniline (14.1 g, 55 mmol) and triethylamine (5.6 g) in tetrahydrofuran (250 mL) at room temperature was slowly added a solution of 3-carbomethoxypropionyl chloride (8.3 g, 55 mmol) in tetrahydrofuran. The reaction mixture was stirred for 1 h. The mixture was filtered through a pad of Celite and silica gel, and the solvent was removed in vacuo, giving a solid. The solid was recrystallized from ethyl acetate, giving 17 g (83% yield) of product, mp 129–130 °C.

In a like manner as above with the appropriate combination of 2-aminothiophenol, 2-aminophenol, or N-methyl-2phenylenediamine and 3-carbomethoxypropionyl chloride or 2-carbomethoxyacetyl chloride, compounds 3-6 were prepared. (See Table I for yield and melting point.)

Also as above with 1-acetanilinyl chloride (prepared from aniline and chloroacetyl chloride) and 2-(chloromethyl)benzthiazole, compound 13 was prepared.

Compound 14 was prepared as above except with use of 2-(chloromethyl)pyridine.

4-[[3-(2-Benzthiazolylmethoxy)phenyl]hydroxyamino]-4-oxobutanoic Acid Methyl Ester (7). (A) [3-(2-Benzthiazolylmethoxy)phenyl]hydroxylamine. To a solution of 3-(2-benzthiazolylmethoxy)nitrobenzene (1.6 g, 5.6 mmol) (see Experimental Section for compound 2, part B) in tetrahydrofuran with 10% palladium on carbon was slowly added hydrazine hydrate (0.4 mL). The reaction mixture was stirred overnight at room temperature. The mixture was filtered through Celite and the solvent was removed in vacuo to give 1.3 g (87% yield) of product, mp 130-134 °C.

(B) Compound 7. Compound 7 was prepared by the method for compound 2, part D, with [3-(2-benzthiazolylmethoxy)phenyl]hydroxylamine. A white solid was obtained, mp 123-125 °C.

4-[N-Methyl[3-(2-benzthiazolylmethoxy)phenyl]amino]-4-oxobutanoic Acid Methyl Ester (8). (A) N-Methyl-3-(2-benzthiazolylmethoxy)aniline. A mixture of 3-(2-benzthiazolylmethoxy)aniline (3.8 g, 14.88 mmol), prepared as in the Experimental Section for compound 2, part C, triethyl orthoformate (25 mL), and 5-10 drops of trifluoroacetic acid was heated at reflux for 6 h. The mixture was concentrated, and the residual oil was dissolved in ethanol and treated with sodium borohydride (1.5 g) at 0 °C. The reaction mixture was allowed to warm to room temperature while being stirred overnight. The mixture was then refluxed for 3 h and concentrated. The remaining oil was dissolved in methylene chloride and washed with water, dried (MgSO<sub>4</sub>), and concentrated. The remaining oil was crystallized from hexanes, giving 1.8 g (45% yield) of product, mp 93-95 °C.

(B) Compound 8. Compound 8 was prepared by the method of compound 2, part D, but with N-methyl-3-(2-benzthiazolyl-methoxy)aniline. A white solid was obtained, mp 107-110 °C.

4-[[3-(2-Benzthiazolylmethoxy)phenyl]amino]-4-oxobutanoic Acid (9). Compound 2 (2.2 g, 5.9 mmol) was dissolved in THF (30 mL) and treated with aqueous potassium hydroxide (1.0 g in 30 mL of  $H_2O$ ). The reaction mixture was stirred overnight. The THF was removed in vacuo and 1 N HCl was added to give a pH of 2. A white precipitate formed, which was filtered and dried to give 2.1 g (99% yield) of product, mp 191–193 °C.

In the same manner with compound 4 as starting material, compound 10 was prepared, mp 213-214 °C.

4-[[3-(2-Benzthiazolyloxy)phenyl]amino]-4-oxobutanoic Acid Methyl Ester (11). (A) 3-(2-Benzthiazolyloxy)nitrobenzene. A mixture of 2-chlorobenzthiazole (17.0 g, 0.1 mol) and 3-nitrophenol (13.9 g, 0.1 mol) was heated at 140 °C for 6 h. After cooling, the remaining oil was crystallized from diisopropyl ether, giving 16.8 g (62% yield) of product, mp 86-88 °C.

(B) Compound 11. Compound 11 was prepared by the method of compound 2, parts C and D, but with 3-(2-benzthiazolyloxy)nitrobenzene. A white solid was obtained, mp 78-82 °C.

<sup>4-[[3-(2-</sup>Benzofuranylmethoxy)phenyl]amino]-4-oxobutanoic Acid Methyl Ester (12). (A) 2-(Bromomethyl)benzofuran. A mixture of 2-methylbenzofuran (2.0 g, 15.1 mmol), N-bromosuccinimide (2.7 g, 15.1 mmol), and carbon tetrachloride was heated at reflux overnight. The reaction mixture was filtered and the solvent was removed in vacuo, giving 3.8 g (52% monobrominated) of product (62% yield).

(B) Compound 12. Compound 12 was prepared by the method for compound 2, parts B-D, but with 2-(bromomethyl)benzofuran. A white solid was obtained, mp 112-113 °C.

N-[3-(2-Benzthiazolylmethoxy)phenyl]-2-oxopyrrolidine-4-carboxylic Acid Methyl Ester (18). (A) N-(3-Hydroxyphenyl)-2-oxopyrrolidine-4-carboxylic Acid. A mixture of 3-aminophenol (10.9 g, 0.1 mol) and itaconic acid (13.0 g, 0.1 mol) was heated to 120-130 °C for 5 min. After cooling, a solid formed, giving 21.5 g (97% yield) of product, mp 214-215 °C.

(B) N-(3-Hydroxyphenyl)-2-oxopyrrolidine-4-carboxylic Acid Methyl Ester. A solution of N-(3-hydroxyphenyl)-2-oxopyrrolidine-4-carboxylic acid (21.0 g, 94.9 mmol) in methanol with p-toluenesulfonic acid (0.1 g) was heated to reflux. The reaction mixture was refluxed for 4 days while water was removed with a Soxhlet extractor filled with 3A molecular sieves. The mixture was cooled and a solid formed, which was filtered and dried, giving 9.5 g (42% yield) of product, mp 178–180 °C.

(C) Compound 18. A mixture of 2-(chloromethyl)benzthiazole (see compound 2, part A) (3.12 g, 17 mmol), N-(3-hydroxyphenyl)-2-oxopyrrolidine-4-carboxylic acid methyl ester (4.0 g, 17 mmol), cesium carbonate (5.3 g, 17 mmol), sodium carbonate (1.8 g), potassium iodide (0.1 g), and acetone (200 mL) was heated at reflux overnight. The mixture was filtered and the resulting solution concentrated to an oil. The oil was triturated with ether, forming a solid, which was filtered and dried to give 4.0 g (62% yield), mp 99-102 °C.

Compounds 19 and 21 were prepared by following the procedures used in the preparation of compound 18 and employing 1-methyl-2-(chloromethyl)benzimidazole, 2-(chloromethyl)benzoxazole and 2-(chloromethyl)quinoline. **N-[3-(2-Benzthiazolylmethoxy)phenyl]pyrrolidine-2,5dione (22).** (A) 4-[(3-Hydroxyphenyl)amino]-4-oxobutanoic Acid Methyl Ester. To an ice-cold solution of 3-aminophenol (21.8 g, 0.2 mol) and triethylamine (21.3 g, 0.2 mol) in THF (250 ml) was added a solution of 3-carbomethoxypropionyl chloride (30.1 g, 0.2 mol) in THF. The reaction mixture was allowed to warm to room temperature and was filtered through a pad of Celite and silica gel. The solvent was removed in vacuo to give a solid. Recrystallization from ethyl acetate gave 39.9 g (88% vield) of product, mp 144-146 °C.

yield) of product, mp 144-146 °C. (B) Compound 22. A mixture of 4-[(3-hydroxyphenyl)amino]-4-oxobutanoic acid methyl ester (1.45 g, 6.5 mmol), 2-(chloromethyl)benzthiazole (1.20 g, 6.5 mmol) (see Experimental Section for compound 2, part A), cesium carbonate (1.0 g), sodium carbonate (0.7 g), potassium iodide (5 mg), and acetone (60 ml) was heated at reflux for 2 h. The reaction mixture was filtered through a pad of Celite and silica gel, and the solvent was removed in vacuo. Recrystallization from acetone gave 1.34 g (56% yield) of product, mp 175-176 °C.

Compounds 23 and 24 were prepared by following the procedure used in the preparation of compound 22 and employing *N*methyl-2-(chloromethyl)benzimidazole and 2-(chloromethyl)quinoline.

**Biological Test Procedures.** Experimental detail for the rat PMN 5-LO and the GP  $LTD_4$ - and OA-induced bronchospasm model are provided in ref 1.

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# Chemical Differentiating Agents. Differentiation of HL-60 Cells by Hexamethylenebis[acetamide] Analogues

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Hexamethylenebis[acetamide] (HMBA) is an agent in clinical trial that induces differentiation of certain types of tumor cells to nonmalignant phenotypes. In an attempt to discover a more potent compound, a number of bis-functionalized amides, imides, and hydrazine derivatives of HMBA were prepared and evaluated in vitro with the HL-60 human promyelocytic leukemia cell line. Among the compounds evaluated, the 5,5-dimethylhydantoin derivative is almost 10 times more potent than HMBA in inducing differentiation. The bis-imide, diacetyl-HMBA, is both more potent and effective than its parent compound. Six of the 16 compounds evaluated cause at least 20% differentiation. An inverse relationship between the degree of differentiation and the percentage of viable cells is described for HMBA and its analogues.

Compounds that induce cancer cells to differentiate to a less malignant phenotype provide an attractive area for the development of new anticancer drugs. Reduced toxicity relative to conventional chemotherapeutic agents is a distinct possibility since the mechanism of antitumor action is not based primarily on cytotoxicity.

The number of compounds that influence cell differentiation and growth characteristics continues to increase. These materials, which include simple organic molecules as well as proteins,<sup>1-3</sup> are thought to influence gene expression. An important aid in the ability to search for agents that induce terminal differentiation in malignant cells occurred when it was discovered that a virus-induced murine erythroleukemia cell line (MELC), when treated with Me<sub>2</sub>SO, expressed many of the features common to terminally differentiated erythroid cells.<sup>4,5</sup> The development of the human HL-60 myeloid leukemia cell line in 1977 provided another important in vitro differentiation system.<sup>6-8</sup> While there are a number of cell lines now

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