

Communications to the Editor

Acetohydroxamic Acids as Potent, Selective, Orally Active 5-Lipoxygenase Inhibitors

Sir:

In the search for novel inhibitors of the 5-lipoxygenase (5-LO) pathway of arachidonic acid metabolism, we have had occasion to look at several series of hydroxamic acids, two examples of which are described within.

The 5-LO pathway leads to several compounds with extremely potent biological activities: leukotriene B₄ (LTB₄) has been shown¹ to be a potent chemotactic agent *in vivo*, while the peptido leukotrienes LTC₄ and LTD₄ are powerful bronchoconstrictors² and lead to an increase in vascular permeability.³ Furthermore, elevated levels of leukotrienes have been found⁴ in certain disease states such as asthma, rheumatoid arthritis, and psoriasis.

We⁵ and others have developed compounds that inhibit the 5-LO or 5-LO and cyclooxygenase (CO) pathways of arachidonic acid metabolism, while the alternative approach of leukotriene antagonists has also been extensively investigated.⁶ Unfortunately, many of the compounds so far developed suffer from toxicity problems or lack of oral bioavailability. More recently, several groups have prepared analogues of arachidonic acid,⁷ 5-HETE,⁸ and 15-HETE⁹ which contain a hydroxamic acid moiety. In these examples, the hydroxamic acid portion of the molecule is thought to bind to Fe³⁺ at the catalytic site of the enzyme.

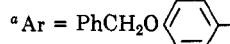
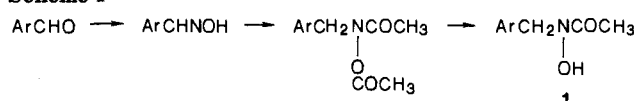
On this basis, a number of hydroxamic acids were prepared as potential 5-LO inhibitors. Of these, compounds 1 (BW A137C), *N*-[4-(benzyloxy)benzyl]acetohydroxamic acid, and 2 (BW A4C), *N*-[(*E*)-3-(3-phenoxyphenyl)prop-2-enyl]acetohydroxamic acid, were found to be potent, selective inhibitors of human leukocyte 5-LO (Table I) and to demonstrate significant oral bioavailability in animals. Compound 1 was prepared from the oxime of 4-(benzyl-

Table I. In Vitro Inhibition of 5-LO and CO from Human Polymorphs

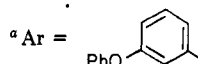
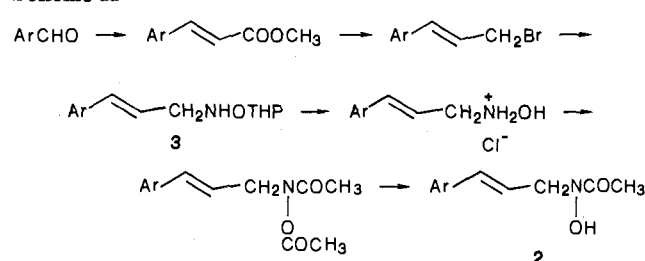
compd	mp, °C	IC ₅₀ , μM: ^a 5-LO	CO
1	121-122	0.77 ± 0.16 (9) ^b 0.2-2.3 ^c	22 ± 4 (9)
2	84	0.14 ± 0.03 (7) ^b 0.06-0.36 ^c	3.2 ± 0.8
4	124-125	0.05 ± 0.01 (3) ^b	5 ± 3 (3)

^a Homogenates of human polymorphs were preincubated with inhibitor (added in DMSO) for 5 min at 37 °C before initiating reaction by addition of arachidonic acid and CaCl₂ (final concentrations 5 μM and 2 mM, respectively). After a further 5 min, incubation reaction was stopped by boiling, and LTB₄ and TXB₂ were assayed by RIA.¹⁵ ^b Mean ± SEM for (*n*) experiments. ^c Minimum and maximum IC₅₀'s.

Scheme I^a



Scheme II^a



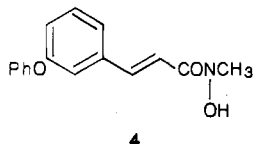
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oxy)benzaldehyde by reduction with sodium cyanoborohydride in acetic acid followed by *in situ* treatment with acetic anhydride; selective removal of the *O*-acetyl group then gave 1 in high yield (Scheme I).

The preparation of 2 proved somewhat more problematic; reduction of the appropriate unsaturated oxime, as above, gave the unsaturated hydroxylamine, which is unstable at the pH of the reaction medium. Although 2 could be obtained by this route, extensive purification was required and yields were low. A preferable synthesis is shown in Scheme II. Reaction of 3-phenoxybenzaldehyde under standard conditions (pyridine, malonic acid, piperidine) gave the cinnamic acid, which was esterified, reduced (DIBAL), and converted to the (*E*)-allylic bromide (1:1 Et₂O/hexane, 48% HBr). Reaction with 3 equiv of *O*-tetrahydropyranyloxyamine¹⁰ in DMF gave the

monoalkylated hydroxylamine **3** together with about 10% bisalkylated product. Hydrolysis of the crude reaction mixture with concentrated HCl in MeOH gave the deprotected hydroxylamine hydrochloride, which was purified by crystallization (EtOAc). Acetylation followed by O-deacetylation, as above, gave **2** in good overall yield. Acetylation of **3** followed by deprotection (PPTS, MeOH) gave less pure product.

Both **1** and **2** have so far been found to be devoid of toxicity problems and to be nonmutagenic¹¹ in the Ames Salmonella test. Furthermore, **1** and **2** selectively inhibit the ex vivo Ca²⁺ ionophore stimulated production of LTB₄ in whole rat blood for well over 6 h after a single oral dose of 50 mg/kg; compound **2**, in fact, has an ED₅₀ at 6 h of 9 mg/kg. In contrast, compound **4**, which is structurally



similar to and in vitro is equipotent with **2** shows no ex vivo activity at 6 h after 50 mg/kg orally in rats. It should be noted that **4** is structurally related to the hydroxamic acid based inhibitors recently disclosed by several other groups.¹²

Compound **2** has also demonstrated its ability to block the "leukotriene-dependent" anaphylactic bronchospasm¹³ in anesthetized guinea pigs in a dose-related manner. In the 6-h carrageenin sponge implant model of inflammation,¹⁴ **2** selectively inhibits the formation of LTB₄ over PGE₂ in the sponge exudates with an ED₅₀ of 2.6 mg/kg. This inhibition was accompanied by a decrease in the number of leukocytes in the sponge exudate, but there was no direct correlation between the two values. Further extensive biological observations with compounds **1** and **2** will appear in due course.¹⁵

Thus, with the development of potent, selective, orally active inhibitors of 5-LO, it should be possible to determine the relevance of lipoxygenase products in human disease states.

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Registry No. **1**, 106328-28-3; **1** (O-acetyl deriv), 106328-89-6; **2**, 106328-57-8; **2** (O-acetyl deriv), 112270-90-3; **3**, 112270-88-9; 5-LO, 80619-02-9; 4-(benzyloxy)benzaldehyde oxime, 76193-67-4; 3-phenoxybenzaldehyde, 39515-51-0; malonic acid, 141-82-2; methyl 3-[(4-phenoxy)phenyl]propenoate, 87087-33-0; 3-bromo-1-[(4-phenoxy)phenyl]propene, 112270-87-8; N-hydroxy-3-(4-phenoxyphenyl)-2-propenamine hydrochloride, 112270-89-0.

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**9-(trans-2',trans-3'-Dihydroxycyclopent-4'-enyl)
Derivatives of Adenine and 3-Deazaadenine:
Potent Inhibitors of Bovine Liver
S-Adenosylhomocysteine Hydrolase**

Sir:

Neplanocin A (NpcA, Chart I), a cytotoxic, cyclopentenyl analogue of adenosine, is a naturally occurring antitumor antibiotic, which was isolated from the bacterium *Ampullariella regularis*.¹⁻⁴ NpcA possesses antitumor activity in vivo against murine L1210 leukemia in mice^{2,5} and antiviral activity in cell culture against vaccinia virus,⁶ herpes simplex-1,⁷ herpes simplex-2,⁷ and vesicular stomatitis virus.⁷ Our laboratory has shown that NpcA is a potent, irreversible inhibitor of S-adenosylhomocysteine (AdoHcy) hydrolase (E.C. 3.3.1.1) isolated from bovine liver⁶ and *Alcaligenes faecalis*.⁸ This enzyme, which catalyzes the reversible hydrolysis of AdoHcy to adenosine and homocysteine, is the only metabolic route for the removal of AdoHcy in eukaryotic cells.⁹ Subsequently, inhibition of AdoHcy hydrolase by NpcA in eukaryotic cells (e.g., mouse L929 and neuroblastoma N2a cells) leads to elevation of cellular levels of AdoHcy and inhibition of S-adenosylmethionine (AdoMet) dependent methylations.^{10,11} The inhibition of AdoHcy hydrolase has been correlated with the antiviral activity of NpcA,¹² and

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