# Synthesis and Biological Activity of New HMG-CoA Reductase Inhibitors. 3.1,2 Lactones of 6-Phenoxy-3,5-dihydroxyhexanoic Acids

H. Jendralla, E. Granzer, B. v. Kerekjarto, R. Krause, U. Schacht, E. Baader, W. Bartmann,\* G. Beck, A. Bergmann, K. Kesseler, G. Wess, L.-J. Chen,† S. Granata, J. Herchen, H. Kleine, H. Schüssler, and K. Wagner

Hoechst AG, Postfach 800320, D-6230 Frankfurt/M. 80, Germany. Received September 6, 1990

A group of 43 optically active sodium carboxylates (11a-qq and the corresponding lactones 4 were prepared from respective phenols 8 according to Schemes I-III. Phenols 8 were synthesized from commercially available compounds according to Schemes IV-IX. A number of these HMG-CoA reductase inhibitors 11 exceeded mevinolin's activity in vitro (Tables II and III). Selected lactones 4 effectively inhibited hepatic "de novo" cholesterol synthesis in rats in vivo (Table IV). After po administration to rabbits, 4ff(11ff), 4hh, and notably 11jj reduced plasma cholesterol levels more potently than mevinolin (1) (Table V). Whereas 4ff(11ff) displayed the slight superiority expected according to in vitro data, 4hh and 11jj were considerably more potent than expected. Each of these compounds had only moderate activity after po administration to dogs (Table VI). Compound di-11ii, a hybrid of the structural elements of probucol (60) and HMG-CoA reductase inhibitors, after po administration to rats decreased serum lipoproteins and increased HDL/LDL ratio better than probucol (Table VII). HMG-CoA reductase inhibitor 11ll and phenolic building blocks 8, notably 8jj and 8kk, inhibited LDL oxidation in vitro (Table VIII). Chemical structure-activity relationships (Table IX) and the pharmacological profile of phenoxy-type inhibitors 11 diverged from those of known HMG-CoA reductase inhibitors.

The fungal metabolite mevinolin (1) is a potent inhibitor of cholesterol biosynthesis at the level of the major rate-limiting enzyme 3-hydroxy-3-methylglutaryl-CoA reductase (HMG-CoA reductase)<sup>3</sup> and lowers plasma cholesterol levels in man,<sup>4</sup> thus diminishing the risk of coronary heart disease.<sup>5</sup>

A plethora of work has been directed toward the preparation of synthetic analogues 2. In 2 (A-B =  $\mathrm{CH_2CH_2}$  or  $\mathrm{CH}$ — $\mathrm{CH}$ ), the stereochemically complex hexahydronaphthalene moiety of 1 has been replaced by suitably substituted, achiral, aromatic moieties  $\mathrm{R^{6-16}}$  and open-chain moieties  $\mathrm{R^{.17,18}}$  Compounds 2 that exceeded the activity of mevinolin (1) in vitro and in vivo have been described. 1.2,9,10,11,19

Comparatively little effort has been directed toward the modification of the "bridging-unit" A-B of analogues 2. This may be due to an early report of Stokker et al. 6a that discouraged this modification. From these data it was concluded 6a that replacement of the trans-ethenyl bridge with the ethinyl and oxymethylene groups resulted in loss of activity. However it should be noticed that these data were limited and that 3 might be an unsuitable model compound, due to its low affinity to the active site of the enzyme. This assumption was confirmed, when we ob-

tained highly efficacious compounds of type 2 bearing an ethinyl bridge (A-B = C $\equiv$ C, IC<sub>50</sub>  $\leq$  10 nM).<sup>21</sup>

- (1) Part 1 of this series: Beck, G.; Kesseler, K.; Baader, E.; Bartmann, W.; Bergmann, A.; Granzer, E.; Jendralla, H.; von Kerekjarto, B.; Krause, R.; Paulus, E.; Schubert, W.; Wess, G. J. Med. Chem. 1990, 33, 52.
- (2) Part 2 of this series: Jendralla, H.; Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Granzer, E.; von Kerekjarto, B.; Kesseler, K.; Krause, R.; Schubert, W.; Wess, G. J. Med. Chem. 1990, 33, 61.
- (3) Hoffman, W. F.; Alberts, A. W.; Anderson, P. S.; Chen, J. S.; Smith, R. L.; Willard, A. K. J. Med. Chem. 1986, 29, 849.
- Smith, R. L.; Willard, A. K. J. Med. Chem. 1986, 29, 849.
  (4) (a) Mol, M. J. T. M.; Erkelenz, D. W.; Gevers Leuven, J. A.; Schouten, J. A. Lancet 1986, 936. (b) Drugs Future 1987, 12, 437.
- (5) LRC-CPPT J. Am. Med. Assoc. 1984, 251, 351 and 365.
- (6) (a) Stokker, G. E.; Hoffman, W. F.; Alberts, A. W.; Cragoe, E. J., Jr.; Deana, A. A.; Gilfillan, J. L.; Huff, J. W.; Novello, F. C.; Prugh, J. D.; Smith, R. L.; Willard, A. K. J. Med. Chem. 1985, 28, 347. (b) Hoffman, W. F.; Alberts, A. W.; Cragoe, E. J., Jr.; Deana, A. A.; Evans, B. E.; Gilfillan, J. L.; Gould, N. P.; Huff, J. W.; Novello, F. C.; Prugh, J. D.; Rittle, K. E.; Smith, R. L.; Stokker, G. E.; Willard, A. K. J. Med. Chem. 1986, 29, 159. (c) Stokker, G. E.; Alberts, A. W.; Anderson, P. S.; Cragoe, E. J., Jr.; Deana, A. A.; Gilfillan, J. L.; Hirshfield, J.; Holtz, W. J.; Hoffman, W. F.; Huff, J. W.; Lee, T. J.; Novello, F. C.; Prugh, J. D.; Rooney, C. S.; Smith, R. L.; Willard, A. K. J. Med. Chem. 1986, 29, 170.
- (7) Stokker, G. E.; Alberts, A. W.; Gilfillan, J. L.; Huff, J. W.; Smith, R. L. J. Med. Chem. 1986, 29, 852.
- (8) Sandoz, patent WO 84/02131, 1984.
- (9) Sandoz, patent WO 86/00307, 1986.
- (10) Sandoz, patent WO 86/07054, 1986.
- (11) Sandoz, European Application EP-A-0221025, 1987.
- (12) Sandoz, European Application EP-A-0265640, 1988.
- (13) Hoechst, European Application EP-A-0324347, 1989.
- (14) Roth, B. D.; Ortwine, D. F.; Hoefle, M. L.; Stratton, C. D.; Sliskovic, D. R.; Wilson, M. W.; Newton, R. S. J. Med. Chem. 1990, 33, 21.
- (15) Prugh, J. D.; Alberts, A. W.; Deana, A. A.; Gilfillian, J. L.; Huff, J. W.; Smith, R. L.; Wiggins, J. M. J. Med. Chem. 1990, 33, 758.
- (16) Bayer, European Application EP-A-0352575, 1990.
- (17) Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Jendralla, H.; Kesseler, K.; Wess, G.; Schubert, W.; Granzer, E.; von Kerekjarto, B.; Krause, R. Tetrahedron Lett. 1988, 29, 929.
- (18) Bristol-Myers, Deutsche Offenlegungsschrift DE 3805801, 1988.
- (19) Bayer, European Application EP-A-0325130, 1989.
- (20) The potency of 3 (A-B = (E)-CH=CH and OCH<sub>2</sub>) is about 10<sup>-3</sup> and 10<sup>-4</sup>, respectively, compared with the most active compounds in the present report.

<sup>†</sup>Visiting scientist from Shanghai Institute of Pharmaceutical Industry, China.

CO<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>

Scheme I.a Preferred Synthesis of Optically Pure HMG-CoA Reductase Inhibitors 4 ("Chiral Pool" Approach Based on L-(-)-Malic Acid)

°(a) CH<sub>3</sub>SO<sub>2</sub>Cl/pyridine/0 °C; (b) CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Cl/pyridine/0 °C; (c) NaI, acetone, reflux; (d)  $7a/K_2$ CO<sub>3</sub>/DMSO/60-90 °C; (e) 2 N HCl/EtOH/THF/25 °C; (f) CF<sub>3</sub>CO<sub>2</sub>H/25 °C; (g) NaHCO<sub>3</sub>/  $Na_2CO_3 \rightarrow pH 7$ ; (h) 1 N NaOH/EtOH/25 °C.

In this paper we will report phenoxy-type inhibitors 4. Some compounds analogous to 4, based on  $\beta$ -quinolinols,  $\beta$ -naphthols, and thiophenols, are reported for comparison.

Thiophenoxy analogues (2:  $A-B = SCH_2$ ) were briefly investigated. In the few cases studied, they were found to be slightly less active than identically substituted phenoxy analogues 4.22,23

### Chemistry

The  $\beta$ -hydroxy lactones 4 and their corresponding  $\beta, \delta$ dihydroxy sodium carboxylates 11 were prepared from phenols 8 according to Schemes I-III. The "chiral pool" approach, as depicted in Scheme I, is the preferred mode of synthesis. The diastereomerically and optically pure synthon 6 was conveniently prepared in seven steps from commercially available, inexpensive L-(-)-malic acid (5).24

(21) Hoechst, European Application EP-A-0361273, 1990.

(22) (a) Hoechst, Deutsche Offenlegungsschrift DE 3632893, 1988. (b) Hoechst, Deutsche Offenlegungsschrift DE 3929913, 1990.

(23) Inhibitors 2 with a fluoroethenyl bridge (2: A-B = FC-CH) were more potent than their unfluorinated analogues (A-B = HC=CH). Baader, E.; Bartmann, W.; Beck, G.; Below, P.; Bergmann, A.; Jendralla, H.; Kesseler, K.; Wess, G. Tetrahedron Lett. 1989, 30, 5115.

Wess, G.; Kesseler, K.; Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Jendralla, H.; Bock, K.; Holzstein, G.; Kleine, H.; Schnierer, M. Tetrahedron Lett. 1990, 31, 2545.

Scheme II. Synthesis of Optically Enriched HMG-CoA Reductase Inhibitors 4 via Asymmetric Aldol Addition

8 
$$\stackrel{\text{a,b}}{=}$$
  $\stackrel{\text{R}^2}{=}$   $\stackrel{\text{R}^6}{=}$   $\stackrel{\text{c-d}}{=}$   $\stackrel{\text{R}^8}{=}$   $\stackrel{\text{CHO}}{=}$   $\stackrel{\text{R}^8}{=}$   $\stackrel{\text{CHO}}{=}$   $\stackrel{\text{R}^8}{=}$   $\stackrel{\text{CHO}}{=}$   $\stackrel{\text{R}^8}{=}$   $\stackrel{\text{CHO}}{=}$   $\stackrel{\text{CHO}}$ 

^(a) BrCH<sub>2</sub>CH(OEt)<sub>2</sub>/K<sub>2</sub>CO<sub>3</sub>/DMSO/90 °C; (b) 2 N HCl/acetone/65 °C; (c) LDA/(S)-(-)-CH<sub>3</sub>CO<sub>2</sub>CH(Ph)CPh<sub>2</sub>OH<sup>28</sup>/THF/-90 °C; (d) NaOCH<sub>3</sub>/CH<sub>3</sub>OH/25 °C; (e) CH<sub>3</sub>CO<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>)/ LDA/-70 °C  $\rightarrow$  -20 °C; (f) Et<sub>3</sub>B/CH<sub>3</sub>OH/NaBH<sub>4</sub>/-80 °C; (g) 1 N NaOH/EtOH/25 °C; (h) HCl/H<sub>2</sub>O; (i) NEt<sub>3</sub>/ClCO<sub>2</sub>Et/THF/-5 °C; (j)  $CF_3CO_2H/25$  °C; (k)  $NaHCO_3/Na_2CO_3 \rightarrow pH^7$ ; (l) Pd/C/

18

Scheme IIIa,b Alternative Synthesis of Optically Pure HMG-CoA Reductase Inhibitors 4 ("Chiral Pool" Approach Based on  $\alpha$ -D-(+)-Glucose;  $R^9 = t$ -BuPh<sub>2</sub>Si)

R90 CCH<sub>3</sub> a, b

HO 20<sup>32</sup>

R90 CCH<sub>3</sub> c, d, then e or f

R10

21: R10 = OTs

22: R10 = I

23: R11 = OCH<sub>3</sub>

24: R11 = OH

25: R11 = 
$$=$$
 O

a(a) p-CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Cl/pyridine/CH<sub>2</sub>Cl<sub>2</sub>/5 °C; (b) NaI/acetone/ reflux; (c) 8b/K<sub>2</sub>CO<sub>3</sub>/DMSO/60 °C; (d) CH<sub>3</sub>CO<sub>2</sub>H/H<sub>2</sub>O/THF/reflux; (c)  $CrO_3/D$ MidO-100 (d)  $CrO_3/D$ MidO-100 (e)  $CrO_3/D$ MidO-100 (n)  $CrO_3/D$ Mi thiophenol 8pp were reacted according to the same scheme.33

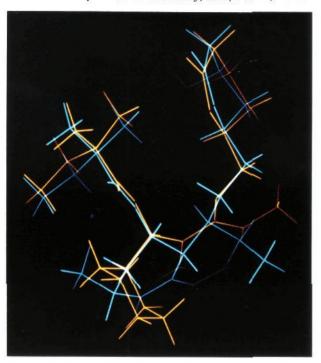


Figure 1. Superposition of synvinolin and compound 4r.

#### Scheme IV.ª Synthesis of Phenols 8a-i

$$R^{2} \xrightarrow{R^{4}} 26$$

$$R^{2} \xrightarrow{R^{4}} 27$$

$$R^{12} \xrightarrow{B^{12}} \xrightarrow{B^{12}} R^{12} \xrightarrow{B^{12}} R^{14} \xrightarrow{B^{12}} R^{12} \xrightarrow{B^{13}} R^{12} \xrightarrow{B^{14}} R^{12} \xrightarrow{B^{15}} R^{12} \xrightarrow{B^{15}} R^{15} \xrightarrow{B^{15}} R^{15$$

 $^{\rm a}$  (a) AlCl $_{\rm 3}/\sim$  150 °C; (b) R $^{\rm 13-}{\rm Br/Mg/THF};$  (c) Pd/C/H $_{\rm 2}/\sim$  AcOH/(concentrated HCl)/25 °C.

Ester 6 was supplied with several different leaving groups to give mesylate 7a, tosylate 7b, iodide 7c, and the unstable triflate 7d. Of these compounds, only mesylate 7a proved to be suitable for a clean, stereochemically homogeneous coupling reaction with phenols 8 to give 9 in 75–90% yield.<sup>25</sup> Cleavage of the acetonide protecting group fol-

#### Scheme V.a Synthesis of Phenols 8j-o

° (a)  $C_6H_5N(CH_3)_2/160$  °C; (b)  $BzCl/K_2CO_3/DMF/75$  °C; (c) 9-BBN/THF/25 °C  $\rightarrow$  reflux; (d) EtOH/2 N NaOH/ $H_2O_2/25$  °C  $\rightarrow$  reflux; (e)  $SiO_2$  chromatography; (f) TsCl/pyridine/25 °C; (g) NaI/acetone/reflux; (h) p-R<sup>18</sup>- $C_6H_4OH/K_2CO_3/DMSO/50$  °C; (i)  $Pd/C/H_2/AcOH/EtOAc$  or concentrated HCl/25 °C/20 min; (j)  $Pd/C/H_2/EtOAc/12$  h.

lowed by a combined cleavage of the *tert*-butyl ester and lactonization gave the diastereomerically and optically pure lactones 4 in 60–75% overall yield based on phenols 8.

Scheme II summarizes the syntheses of optically enriched lactones 4 via asymmetric aldol addition<sup>26,27</sup> of the dianion generated from (S)-(-)-phenyl (2-hydroxy-2,2-diphenylethyl)acetate [(S)-(-)-HYTRA]<sup>28</sup> and 2 equiv of LDA to substituted phenoxyacetaldehydes 13.

We have shown recently that the addition of this dianion to 3-pyrrol-3-ylpropenals 17 proceeds with high asymmetric induction, leading to the  $\beta,\delta$ -dihydroxyheptenoic esters 19 with optical purities of more than 92% ee.2 Similar results were obtained in the addition of this dianion to chiral β-alkoxyaldehydes.<sup>29</sup> Therefore, we expected a similar degree of stereoselectivity for the addition of the dianion of HYTRA to  $\alpha$ -phenoxyacetaldehydes 13. However, the indicated 3(S)-hydroxy isomer 14 exceeded its undesired 3(R) diastereomer by only less than 82:18 (59–64% de, HPLC). The diastereomeric excess of 14 was not significantly improved by recrystallization, nor by a transmetalation of the lithium dianion of HYTRA with MgBr<sub>2</sub><sup>27</sup> and a lower reaction temperature<sup>27</sup> in the asymmetric aldol addition. The disappointing stereoselectivity was corroborated by analyses of methyl ester 15 [59-65% ee, HPLC on chiral column and <sup>1</sup>H NMR (Eu(hfc)<sub>3</sub>) analyses]. Recrystallization of 15 did not significantly improve its optical purity.

<sup>(25)</sup> In the reaction of 7c with phenol 8gg, there was obtained 39% of 9 and 51% of a diastereomer besides 8% of unreacted 7c. The 270-MHz <sup>1</sup>H NMR of the isolated diastereomer does not allow for an unequivocal decision between an isomerization at the β- or the δ-carbon atom.

<sup>(26)</sup> Braun, M.; Devant, R. Tetrahedron Lett. 1984, 25, 5031.

<sup>(27)</sup> Devant, R.; Mahler, U.; Braun, M. Chem. Ber. 1988, 121, 397.

<sup>(28)</sup> Commercially available from Merck-Schuchardt, Germany.

<sup>(29)</sup> Mahler, U.; Devant, R.; Braun, M. Chem. Ber. 1988, 121, 2035.

## Scheme VI.a Synthesis of Phenols 8p-8z

°(a)  $C_6H_{11}COCl/CH_2Cl_2/25$  °C; (b)  $AlCl_3/150$  °C; (c)  $NaBH_4/CH_3OH/H_2O/25$  °C; (d) three steps, Scheme III; (e) HPLC separation of the two diastereomers; (f)  $(CH_3CH_2C(CH_3)_2CO)_2O/toluene/DMAP/110$  °C; (g)  $(n-Bu)_4NF/CH_8CH_2C(CH_3)_2CO_2H/THF/20$  °C; (h)  $LiAlH_4/AlCl_3/Et_2O/reflux$ ; (i)  $Et_2NSF_3/toluene/0$  °C  $\rightarrow$  25 °C; (j)  $p-FC_6H_4MgBr/THF/O \rightarrow$  25 °C; (k) 2 N HCl; (l)  $Pd/C/H_2/CH_3OH/25$  °C; (m) 48% aqueous  $HF/CH_3CH_2C(CH_3)_2CN/25$  °C/20-50 h; (n)  $(n-Bu)_4NF/AcOH/THF/25$  °C.

Reaction of methyl ester 15 with 4 equiv of the enolate of tert-butyl acetate yielded tert-butyl  $\beta$ -keto- $\delta(S)$ -hydroxy carboxylate 16. Highly stereoselective reduction of the keto group<sup>30</sup> was conducted with triethylborane and sodium borohydride to give tert-butyl  $\beta(R)$ - $\delta(S)$ -dihydroxy carboxylate 10. Saponification of ester 10 with sodium hydroxide in ethanol/water gave sodium carboxylate 11, which was converted to  $\beta$ -hydroxy lactone 4 via the mixed anhydride with ethyl chloroformate.

Alternatively, ester 10 can be transformed to  $\beta$ -hydroxy lactone 4 with trifluoroacetic acid (vide supra). The diastereomeric purity of 4 was >96.5% (HPLC), the ratio of enantiomers 81: 19 (62% ee, <sup>1</sup>H NMR/Eu(hfc)<sub>3</sub> analysis), in agreement with the optical purities of precursors 14 and 15 (vide supra). During the early phase of the investigation, <sup>31</sup> lactones 4 and sodium carboxylates 11 were prepared according to the chiral pool approach outlined in

Scheme III. Alcohol  $20^{32}$  was converted to the corresponding iodide 22 via tosylate 21. Nucleophilic substitution of iodide 22 by the substituted phenol,  $\beta$ -naphthol,  $\beta$ -quinolinol, or thiophenol  $8^{33}$  ( $K_2CO_3/DMSO$ ) gave the protected lactol ether 23, which was hydrolyzed to lactol 24, and then oxidized to lactone 25, and the silyl protecting group was removed to give  $\beta$ -hydroxy lactone 4. Although this approach served to produce optically pure 4 with a wide variety of substituents  $R^2-R^6$  in good yield, it cannot compete with the method depicted in Scheme I due to the lengthy synthesis (13 steps) of 22.

Physical data and chemical yields of lactones 4 and corresponding sodium carboxylates 11 are collected in Table I. Additionally this table outlines which of the three procedures (Schemes I-III) was used to prepare 4 and 11 from the respective building block 8. Lactone 4ff was prepared according to all three procedures, allowing for a direct comparison of their efficiencies.

Substituted phenols, analogous  $\beta$ -naphthols,  $\beta$ -quinolinols, and thiophenols were synthesized as depicted in Schemes IV-IX.<sup>33</sup> Most of the synthetic steps correspond

<sup>(30) (</sup>a) Kathawala, F. G.; Prager, B.; Prasad, K.; Repic, O.; Shapiro, M. J.; Stabler, R. S.; Widler, L. Helv. Chim. Acta 1986, 69, 803.
(b) Narasaka, K.; Pai, F.-C. Tetrahedron 1984, 40, 2233.

<sup>(31)</sup> The first compounds 4 with potent in vitro and (partially) in vivo activity were prepared in 1985. Preliminary report: Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Granzer, E.; Jendralla, H.; von Kerekjarto, B.; Kesseler, K.; Krause, R.; Wess, G. International Symposium on Cholesterol Control and Cardiovascular Diseases: Prevention and Therapy, Milan, July 7-9, 1987, Abstract book, p 133.

<sup>(32)</sup> Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Fehlhaber, H.-W.; Jendralla, H.; Kesseler, K.; Saric, R.; Schüssler, H.; Teetz, V.; Weber, M.; Wess, G. Tetrahedron Lett. 1988, 21, 2563.

Table I: Physical Properties and Yields of β-Hydroxy Lactones 4 and of Corresponding β,δ-Dihydroxy Sodium Carboxylates 11

no.	R²	R <sup>3</sup>	R <sup>4</sup>	R <sup>5</sup>	R <sup>6</sup>	mp,ª °C	$R_{f}$ (solvent) <sup>b</sup>	% yield <sup>c</sup> (p <b>rocedure</b> ) <sup>d</sup>	formula*	no.f	dec, °C
le .	CH <sub>3</sub>	Н	CH <sub>3</sub>	Н	$CH(p-C_6H_4F)_2$	140-1418	0.19 (1)	73 (3)	C27H26F2O4	lla	h
lb	Cl	H	Cl	Н	$CH(p-C_6H_4F)_2$	amorphous	0.20(1)	84 (3)	C <sub>25</sub> H <sub>20</sub> Cl <sub>2</sub> F <sub>2</sub> O <sub>4</sub>	11 <b>b</b>	237- <b>239</b>
<b>l</b> c	CH <sub>3</sub>	H	Cl	Н	$CH(p-C_6H_4F)_2$	amorphous	0.10(1)	83 (3)	C <sub>28</sub> H <sub>23</sub> ClF <sub>2</sub> O <sub>4</sub>	11c	h
<b>4d</b>	CH <sub>3</sub>	Н	CH <sub>3</sub>	H	$CH(p-C_6H_4F)(p-C_6H_4OCH_3)$	h	0.12 (1)	<b>85 (3</b> )	C <sub>28</sub> H <sub>29</sub> FO <sub>5</sub>	11 <b>d</b>	232-234
le	CH <sub>3</sub>	H	CH <sub>3</sub>	Н	$CH(p-C_6H_4F)(m-C_6H_4CF_3)$	h	0.16 (1)	<b>79 (3</b> )	C28H26F4O4	lle	<b>236–237</b>
if	CH <sub>3</sub>	H	CH <sub>3</sub>	H	$CH(p-C_6H_4F)CH_3$	110–112 <sup>i</sup>	0.21 (1)	<b>45 (3</b> )	C <sub>22</sub> H <sub>25</sub> FO <sub>4</sub>	11 <b>f</b>	h
lg	CH <sub>3</sub>	H	CH <sub>3</sub>	H	CH(p-C <sub>6</sub> H <sub>4</sub> F)i-Bu	h	0.24 (1)	81 (3)	C <sub>25</sub> H <sub>31</sub> FO <sub>4</sub>	1 <b>1g</b>	h
i <b>h</b>	Cl	H	CH <sub>3</sub>	H H H	$CH(m-CH_3-p-F-C_6H_3)_2$	77-78	0.26 (1)	<b>81 (3</b> )	C <sub>28</sub> H <sub>27</sub> ClF <sub>2</sub> O <sub>4</sub>	1 <b>1ĥ</b>	h
li	Cl	Н	Cl	H	$CH(m-CH_3-p-F-C_6H_3)_2$	62-63	0.26 (1)	<b>88 (3</b> )	$C_{27}H_{24}Cl_2F_2O_4$	11i	h
lj	Cl	Н	Cl	H	$CH(p-C_6H_4F)[(CH_2)_2O(p-C_6H_4F)]$	glass	0.19 (1)	<b>86 (3</b> )	$C_{27}H_{24}Cl_2F_2O_5$	11 <b>j</b>	h
(k	CH <sub>3</sub>	H	Cl	H	$(CH_2)_3O(p-C_6H_4F)$	98-99	0.19 (1)	<b>67 (3</b> )	C22H24ClFO5	11 <b>k</b>	h
11	CH <sub>3</sub>	Н	Н	H	$(CH_2)_3O(p-C_6H_4F)$	h	0.19 (1)	<b>72 (3)</b>	C <sub>22</sub> H <sub>25</sub> FO <sub>5</sub>	1 <b>1</b> 1	h
l m	CH <sub>3</sub>	Н	Cl	H H	$(CH_2)_3O(p-C_6H_4Cl)$	70–71	0.19 (1)	<b>55 (3</b> )	$C_{22}H_{24}Cl_2O_5$	11 <b>m</b>	h
ín.	c-C <sub>5</sub> H <sub>9</sub>	H	Cl	Н	$(CH_2)_3O(p-C_6H_4F)$	glass	0.20(1)	<b>79 (3</b> )	$C_{28}H_{30}ClFO_5$	lln	h
lo	c-C <sub>5</sub> H <sub>9</sub>		H	H H	$(CH_2)_3O(p-C_6H_4F)$	glass	0.20 (1)	<b>94 (3</b> )	$C_{28}H_{31}FO_{5}$	llo	h
P	Cl	Н	Cl	H	CO-c-C <sub>6</sub> H <sub>11</sub>	h	0.16 (1)	<b>88 (3</b> )	$C_{19}H_{22}Cl_2O_5$	11 <b>p</b>	h
r	CH <sub>3</sub>	Н	CH <sub>3</sub>	H	$(S)$ -CH(c- $\overline{C_6}$ H <sub>11</sub> )O <sub>2</sub> CC(CH <sub>3</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	glass	0.19 (1)	90 (3)	$C_{27}H_{40}O_6$	11r	h
le	CH <sub>3</sub>	H	CH <sub>3</sub>	H H	(R)-CH(c-C <sub>6</sub> H <sub>11</sub> )O <sub>2</sub> CC(CH <sub>3</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	glass	0.17 (1)	<b>92 (3</b> )	$C_{27}H_{40}O_6$	118	h
t	CH <sub>3</sub>	Н	CH <sub>3</sub>	Н	CH <sub>2</sub> -c-C <sub>6</sub> H <sub>11</sub>	1 <b>23</b>	0.20 (1)	<b>68 (3</b> )	$C_{21}H_{30}O_4$	11 <b>t</b>	h
u	Cl	Н	Cl	H	CH <sub>2</sub> -c-C <sub>6</sub> H <sub>11</sub>	92 <del>-9</del> 3#	0.16 (1)	81 <b>(3</b> )	$C_{19}H_{24}Cl_2O_4$	11 <b>u</b>	h
l♥	CH <sub>3</sub>	H	Cl	H	CH <sub>2</sub> -c-C <sub>6</sub> H <sub>11</sub>	8 <del>9-</del> 91	0.20 (1)	78 <b>(3</b> )	C <sub>20</sub> H <sub>27</sub> O <sub>4</sub> Cl	11 <b>v</b>	h
W	CH <sub>3</sub>	H	CH <sub>3</sub>	H	(S)-CH(c-C <sub>6</sub> H <sub>11</sub> )NHCOC(CH <sub>3</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	amorphous	0.08 (1)	<b>83 (3</b> )	C27H41NO5	11 <b>w</b>	h
X	CH <sub>3</sub>	Н	CH <sub>3</sub>	H	(R)-CH(c-C <sub>6</sub> H <sub>11</sub> )NHCOC(CH <sub>3</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	199	0.07 (1)	<b>73 (3</b> )	$C_{27}H_{41}NO_5$	11x	h
y	Cl	Н	Cl	H	$CF = c - C_6 H_{10}$	glass	0.18 (1)	84 (3)	C <sub>16</sub> H <sub>21</sub> Cl <sub>2</sub> FO <sub>4</sub>	11 <b>y</b>	h
Z	CH <sub>3</sub>	H	CH <sub>3</sub>	H	$CH-c-C_6H_{11}(p-C_6H_4F)$	foam	0.20 (1)	90 (3)	C <sub>27</sub> H <sub>33</sub> FO <sub>4</sub>	11 <b>z</b>	h
lee .	i-Pr	Н	Ph	H	CH <sub>2</sub> -c-C <sub>6</sub> H <sub>11</sub>	glass	0.26 (1)	84 (3)	$C_{28}H_{36}O_4$	llaa	h
bb	Cl	Н	Cl	H	Ph	amorphous	0.11 (1)	<b>75 (3)</b>	$C_{16}H_{16}Cl_2O_4$	11 <b>bb</b>	h
ec	Cl	Н	Cl	$OCH_{2}^{-}$ $(p-C_{6}H_{4}F)$	Н	amorphous	0.31 (2)	87 (3)	$C_{19}H_{17}Cl_2FO_5$	llcc	h
dd		-CH=	=СНСН=СН-	H	$CH(p-C_0H_4F)_2$	171-173	0.23 (1)	88 (3)	$C_{32}H_{30}F_2O_4$	11 <b>dd</b>	h
00	CH <sub>3</sub>	-CH=	=CHCH=CH-	Н	$CH(p-C_6H_4F)_2$	177-179	0.16 (1)	67 (3)	C <sub>30</sub> H <sub>26</sub> F <sub>2</sub> O <sub>4</sub>	11ee	h
ff	i-Pr	H	i-Pr	H	p-C <sub>6</sub> H <sub>4</sub> F	145-146	0.25 (1)	83 (3)	C <sub>24</sub> H <sub>29</sub> FO <sub>4</sub>	11 <b>ff</b>	243-245
off <sup>j</sup>	i-Pr	H	i-Pr	Н	p-C <sub>6</sub> H <sub>4</sub> F	63-65	0.15 (4)	74 (2)	C <sub>28</sub> H <sub>29</sub> FO <sub>5</sub>	11 <b>ff<sup>i,k</sup></b>	230-233
<b>ff</b> j.	i-Pr	H	i-Pr	Н	p-C <sub>6</sub> H <sub>4</sub> F	142-144	0.26 (1)	68 (2)	C <sub>24</sub> H <sub>29</sub> FO <sub>4</sub>		
ff	i-Pr	H	i-Pr	Н	p-C <sub>6</sub> H <sub>4</sub> F	143-145	0.24 (1)	81 <b>(1</b> )	C <sub>24</sub> H <sub>29</sub> FO <sub>4</sub>		
SE.	i-Pr	H	<i>t-</i> <b>B</b> u	H	p-C <sub>6</sub> H <sub>4</sub> F	178-180	0.28 (1)	75 <b>(3</b> )	$C_{25}H_{31}FO_4$	11gg	<b>256-25</b> 9
i <b>hh</b>	i-Pr	H	p-C <sub>6</sub> H <sub>4</sub> F	H	p-C <sub>6</sub> H <sub>4</sub> F	190-192	0.23 (1)	82 (3)	$C_{27}H_{26}F_2O_4$	11 <b>hh</b>	235-237
ibb	i-Pr	Н	p-C <sub>6</sub> H <sub>4</sub> F	H	p-C <sub>6</sub> H <sub>4</sub> F	189-191	0.22 (1)	82 (1)	$C_{27}H_{26}F_2O_4$	11 <b>hh</b>	235-237

Scheme VII.<sup>a</sup> Synthesis of Phenols and β-Naphthols 8aa-ee

NO <sub>2</sub> R <sup>21</sup>	d R222
R <sup>20</sup> R <sup>21</sup> a, b 37: H H  c, b 38: <i>i</i> -Pr H  c, b 39: <i>i</i> -Pr CH <sub>2</sub> H	40: R <sup>22</sup> = NH <sub>2</sub> e.f - 8aa: R <sup>22</sup> = OH
R <sup>2</sup> OH OH	CI OH OR <sup>23</sup>
R <sup>2</sup> R <sup>4</sup> 9	CI 42: R <sup>23</sup> = H 8cc: R <sup>23</sup> = CH <sub>2</sub> — F
OH CO₂CH₃ - 43	R <sup>24</sup> R <sup>25</sup> R <sup>24</sup> R <sup>25</sup> H OH  H H H H H H H H H H H H H H H H

a (a) i-PrMgCl/-70 °C; (b) DDQ/-40 °C → 25 °C; (c) c-Hex-CH<sub>2</sub>MgBr/-70 °C; (d) Raney Ni/H<sub>2</sub>; (e) NaNO<sub>2</sub>/HBr; (f) H<sub>2</sub>O/reflux; (g) Cl<sub>2</sub>/AcOH/20 °C/3 h; (h) p-FC<sub>8</sub>H<sub>4</sub>CH<sub>2</sub>Br/NaOH/DMF/100 °C/4 h; (i) 4 equiv p-FC<sub>6</sub>H<sub>4</sub>MgBr/THF/25 °C; (j) Pd/C/H<sub>2</sub>/AcOH/HCl; (k) CHCl<sub>3</sub>/NaOH/H<sub>2</sub>O/75 °C; (l) 1 equiv Na/CH<sub>3</sub>OH; (m) 4 equiv i-PrBr/toluene/reflux; (n) [(CH<sub>3</sub>OCH<sub>2</sub>CH<sub>2</sub>-O)<sub>2</sub>AlH<sub>2</sub>]Na (Red-Al)/xylene/135 °C.

to well-established chemistry; however some features deserve comment.

When ketone 8p (Scheme VI) was treated with (diethylamido)sulfur trifluoride (DAST), replacement of the carbonyl oxygen by two geminal fluorine atoms was expected. However, vinyl fluoride 8y was obtained instead, independent of the excess of DAST used. Attempts to achieve the addition of hydrogen fluoride to the double bond of 8y were unsuccessful using dry HF or synthetic equivalents. However, when 8y was transformed to  $\beta$ -hydroxy lactone 4y according to Scheme III, and then was treated with dry HF, the addition occurred without difficulty to give the desired geminal difluoride 4qq (Scheme X). We attribute the atypical behavior of 8p's carbonyl to the proximity of the free phenolix hydroxyl group.

In studies with 1 it has been demonstrated that the sterically demanding ester group contributes significantly to biological activity. While the alcohol resulting from hydrolysis of  $\alpha$ -methylbutanoic acid ester 1 has retained only 0.1–0.2% 35 of mevinolin's activity, replacement of the

w H	M		Co				ıct	ase	e I	nh	ibit	or.
185-210	(225) 250-300"	192-195	193-196	239-240	239-242	ų	ų	4	ų	4		hexane (CH)/eth
mono-11ii	di-11ii	11jj	1133	11kk°	111k	11mm <sup>p</sup>	llnn <sup>p</sup>	1100	11pp <sup>e</sup>	1199		nm: (1) evelo
CAHEFORS,	CraH,0F,010S2	C31H36F2O5S	C27H28F204S	C33H47FO7	CaH.FO,	NO.	· •	$C_{22}H_{21}NO_4$	Cz.HzoClzF2OsS	C19H22CIF2O4"	C21H29C1O4	ver thickness 0.25 mm: (
75 (1)	73 (1)	81 (1)	85 (1)	76 (1)	71 (1)	82 (3)	83 (3)	83 (3)	83 (3)	<b>4</b> 0	<b>49</b> 2	Merck. la
0.20 (5)	0.35 (6)	0.18 (3)	0.26 (1)	0.20 (4)	0.20 (4)	0.09 (1)	0.12 (1)	0.24 (2)	0.21 (1)	0.33 (7)	0.21 (1)	silica gel 60Fer
glass	glass	glass	108-110	61–63	120-122	132-134	124-128	155-157	8,00	glass	glass	e noted bTT.C plates
p-C,H,F	p-C,H,F	p-C,H,F	p-C,H,F	i-Pr	p-C,H,F	p-C,H,F	p-CHF	CH,	CH(p-C,H,F),	CF.c-C,H,1	CHCl-c-CeH11	nd: not recrustellized unless otherwise
H	H	H	H	p-C,H,F	.Pr	ď	. a	a	H,	Н	н	fied common
SC(CH.),SR"	SC(CH,),SR"	S(p-C,H,F)	S(p-C,H,F)	. OAc	· OAc	-CHCH-CH-	-CH=CHCH=CH-	СН—СНСН—СН-	ಶ	ວ	$CH_3$	ographically nur
H	r. H	H Y								<b>H</b>	Н, Н	m chromos
iil-onom	di-10ii i-Pr						4nn <sup>p</sup> i-1		4pp' Cl			Ohtained fro

80%. Prepared from 11ff via mixed anhydride. "See Scheme X. "Turns dark at 225 °C, melts with decomposition at 250-300 °C. "Acetate saponified; see Scheme X. Compounds contain 1 equiv of NaOAc. P.Quinolinoxy subunit; atom carrying R<sup>5</sup> in formulae 4 and 11 is N. "N: calcd, 3.42; found, 2.93. 'Thiophenoxy subunit; atom A in formula 2 is S. "Sinters at >55 °C. "75% yield after freeze-drying of aqueous solution. "Based on 4y, after chromatographic purification. See Scheme X. "No elemental analysis. Fully characterized by 1H, 13C, 19F NMR, and

(see supplementary material). "Based on 25q, after chromatographic purification. See Scheme X.

4 or 10 >95% unless otherwise noted. Substituents R2-R6 are the same as for 4. Recrystallized from i-Pr<sub>2</sub>O/n-hexane. And determined. Recrystallized from EA/CH. 162% ee. Yield

acetate (EA) 1:1, (2) 100% EA, (3) CH/EA 2:1, (4) CH/EA 3:1, (5) toluene/EA 7:1, (6) CHCl<sub>3</sub>/CH<sub>3</sub>OH 9:1, (7) CH<sub>2</sub>Cl<sub>2</sub>/EA 4:1.

<sup>d</sup> Procedures 1, 2, and 3 refer to Schemes I, II, and III, respectively.

precursor 10, 16, or 25.

Analytical results were within ±0.4% of the theoretical value unless noted. 'Yield from

<sup>c</sup> Isolated yields after chromatography on silica gel, based on

<sup>(33)</sup> For uniformity of Tables and Schemes phenols, β-naphthols, β-quinolinols, and thiophenols were all assigned compound no. 8.

<sup>34)</sup> Middleton, W. J. J. Org. Chem. 1975, 40, 574.

<sup>(35) (</sup>a) Endo, A. J. Med. Chem. 1985, 28, 401. (b) Lee, T.-J. Trends Pharmacol. Sci. 1987, 8, 442.

Scheme VIII.<sup>a</sup> Pd(0)-Catalyzed Arylation of Phenols. Synthesis of Phenols 8ff-ll

a (a) Quinoline/cat. 2CuO·Cr<sub>2</sub>O<sub>3</sub>/190 °C; (b) Br<sub>2</sub>/CCl<sub>4</sub>/10 °C; (c) BzCl/K<sub>2</sub>CO<sub>3</sub>/2-butanone/reflux; (d) Mg/cat. I<sub>2</sub>/THF/reflux; (e) p-FC<sub>6</sub>H<sub>4</sub>I/2 mol % (PPh<sub>3</sub>)<sub>4</sub>Pd/THF 20 °C → 55 °C; (f) Raney Ni/EtOH/25 °C/filtration; (g) Pd/C/H<sub>2</sub> (1 bar)/EtOH/25 °C; (h) CH<sub>3</sub>OC-(CH<sub>3</sub>)<sub>3</sub>/ZrCl<sub>4</sub>/CH<sub>2</sub>Cl<sub>2</sub>/0 °C; (i) I<sub>2</sub>/KI/H<sub>2</sub>O/EtNH<sub>2</sub>/25 °C; (j) 3 equiv FC<sub>6</sub>H<sub>4</sub>MgBr/1 mol % (PPh<sub>3</sub>)<sub>4</sub>Pd/THF/25 °C → reflux; (k) 4 equiv FC<sub>6</sub>H<sub>4</sub>MgBr/1 mol % (PPh<sub>3</sub>)<sub>4</sub>Pd/THF/reflux; (l) BrN(CH<sub>3</sub>)<sub>2</sub>/CCl<sub>4</sub>/−10 °C → 0 °C; (m) NaSCN/Br<sub>2</sub>/CH<sub>3</sub>OH/10 °C; (n) LiAlH<sub>4</sub>/THF/reflux; (o) (CH<sub>3</sub>O)<sub>2</sub>C(CH<sub>3</sub>)<sub>2</sub>/cat. TsOH·H<sub>2</sub>O/benzene/reflux; (p) p-FC<sub>6</sub>H<sub>4</sub>MgBr/THF/40 °C; (q) 1.3 equiv Ac<sub>2</sub>O/2.0 equiv NEt<sub>3</sub>/1 mol % DMAP/CH<sub>2</sub>Cl<sub>2</sub>/−18 °C/2 days; (r) 4 equiv Ac<sub>2</sub>O/pyridine/25 °C; (s) 1.1 equiv LiOH/H<sub>2</sub>O/CH<sub>3</sub>O(CH<sub>2</sub>)<sub>2</sub>OCH<sub>3</sub>/25 °C/3 days; (t) chromatographic separation from 8kk.

 $\alpha$ -methylbutanoic acid ester by an  $\alpha, \alpha$ -dimethylbutanoic acid ester (to give synvinolin) potentiates the activity by a factor of 2.5.3 Molecular models of synvinolin and the respective S configurated ester  $4\mathbf{r}$  may be superimposed (Figure 1). A good matching of the structures can be achieved, if the molecular models are fitted so as to minimize the distances of corresponding atoms in the lactone and ester moieties.

In the superposition (Figure 1) the aromatic ring of 4r mimics the cyclohexenyl moiety of synvinolin. The omethyl substituent of 4r is spatially situated between the methyl group and the bridge methylene (A = CH<sub>2</sub>) of synvinolin. To avoid elaborate protecting/deprotecting strategies of the phenol component during the preparation of ester 4r, we did not synthesize the respective phenol 8r, but converted 8q to the  $\beta$ -silyloxy lactone 25q instead. Treatment of 25q with  $\alpha,\alpha$ -dimethylbutanoyl chloride<sup>3</sup> did not produce the expected mixture of diastereomeric esters 25r/25s, but gave the diastereomeric chlorides 25rr (Scheme X), which were converted to 4rr. However, refluxing of a toluene solution of the HPLC-separated diastereomeric alcohols 25q and  $\alpha,\alpha$ -dimethylbutanoic anhydride<sup>36</sup> in the presence of DMAP furnished esters 25r

and 25s (Scheme VI), which were converted to hydroxy lactones 4r and 4s.

In an attempt to transform benzylic alcohol 25q (Scheme VI) to the corresponding fluoride, it was stirred with 48% aqueous hydrogen fluoride in acetonitrile. Unexpectedly, the corresponding diastereomeric acetamides were obtained. Obviously, the benzylic cation generated from 25q had been trapped by acetonitrile, resulting in the formation of a carbiminium ion, which then had been trapped by water. The reaction was repeated, utilizing 2,2-dimethylbutyronitrile, to give diastereomeric amides 25w and 25x, which could be separated chromatographically. Surprisingly, the silyl protection of the  $\beta$ -hydroxy lactone moiety of 25w,x was fully retained under the reaction conditions. Additionally, the reaction does not seem to respond to sterical hindrance. Considering the forcing conditions necessary to esterify 25q to 25r,s or desacylmevinolin to synvinolin,36 the smooth formation of 25w,x at 25 °C is remarkable. Assignments of absolute config-

 <sup>(36) (</sup>a) Hirama, M.; Iwashita, M. Tetrahedron Lett. 1983, 24, 1811.
 (b) Girotra, N. N.; Wendler, N. L. Tetrahedron Lett. 1982, 23, 5501

°(a) Isatine/EtOH/6 N aqueous KOH/reflux; (b) concentrated aqueous HCl/0 °C; (c)  $CH_2N_2/Et_2O/0$  °C; (d) LiAlH<sub>4</sub>/Et<sub>2</sub>O; (e) Pd/C/H<sub>2</sub>/AcOH, concentrated aqueous HCl; (f) 5 equiv CH<sub>3</sub>MgI/THF/25 °C; (g) 67% aqueous HJ/ref P/130-150 °C; (h) CH<sub>3</sub>COCH<sub>2</sub>OH/AcOH/H<sub>2</sub>O/cat. H<sub>2</sub>SO<sub>4</sub>/reflux/4 h; (i) (CH<sub>3</sub>)<sub>2</sub>NCSCl/DMF/80 °C; (j) 275 °C/30 min; (k) LiAlH<sub>4</sub>/Et<sub>2</sub>O.

uration to the benzylic carbons (\*) of esters 25r,s and amides 25w,x are tentative.<sup>37</sup>

Aliphatic alkyl groups can be introduced into the ortho or para position of substituted nitrobenzenes by the method of Bartoli and Kienzle. Accordingly, we obtained 39 from p-nitrobiphenyl (37; Scheme VII). However, an attempt to introduce a p-fluorophenyl substituent into 38 in an analogous fashion failed. Nitro compound 39 was converted to the corresponding phenol 8aa via reduction and deamination. An isopropyl substituent could be introduced into the activated ortho position of  $\beta$ -naphthol 45 by a nucleophilic substitution reaction with isopropyl bromide to give 8dd. Analogous reactions with methyl iodide or methyl p-toluenesulfonate did not give significant amounts of the ortho-methylated naphthol 8ee. Therefore 8ee had to be prepared via Reimer-Tiemann formylation and reduction of the aldehyde 46 with Red-Al. Al

In studies with HMG-CoA reductase inhibitors 2 containing heterocyclic aromatic moieties R, we obtained very potent compounds, when a p-fluorophenyl substituent and an isopropyl substituent were present in the aromatic heterocycle R in the two ortho positions to the bridge A-B.<sup>1,2</sup> Consequently, we were highly interested to prepare

requisite phenoxy analogues 4. However, phenols of this special substitution pattern were unknown, because no general method had been reported for the direct arylation of phenols. 42,43 Despite the high tolerance of oxygen functionality in palladium-catalyzed bond formation,4 the methodology of palladium(0)-catalyzed couplings45,46 had not been extended to the arylation of phenols. Scheme VIII summarizes the syntheses of the desired phenols 8ff-11 based on this principle. Whereas 8ff was prepared via the coupling of a metallorganic phenol component (Grignard of 50) with a halobenzene, the reactivity of the components was reversed to obtain the double coupling product 8hh or the ortho-arylation product 57: halogenated phenols were coupled with arylmagnesium halides. 8gg was prepared according to both variants. Zirconium-(IV)-induced para-tert-butylation of 52 was achieved, following the method of Sartori et al.47 The palladium-(0)-catalyzed arylation of phenols provided the means to prepare molecules that are hybrids of the structural elements of the established antiatherosclerotic agent probucol (60) and of HMG-CoA reductase inhibitors. Probucol has been recently demonstrated to potently inhibit the oxidation of LDL particles in vivo, thus interfering with a strongly atherogenic process<sup>48-50</sup> in addition to its moderate hypocholesterolemic action.51-53 From the chemical viewpoint, the reasons for probucol's selective and efficacious antioxidative (free radical trapping) activity are (a) p-hydroxymercaptobenzene moieties (hydroquinone analogue), the hydroxy groups each being shielded by two bulky o-tert-butyl substituents; (b) the prevailing transport of plasma-probucol in LDL particles, because of its highly lipophilic properties. 51,52

The bulky ortho substituents are essential, since they still allow probucol to transfer the phenolic hydrogen atom to a free radical, but they inhibit the resulting phenoxyl radical to propagate the radical chain reaction due to steric inhibition. Since an isopropyl and a p-fluorophenyl substituent still exert appreciable shielding to the neighboring hydroxyl, we reasoned that probucol's tert-butyl groups may be exchanged with these substituents, without loosing the ability to inhibit LDL oxidation. This was born out by experiment. Both equivalent phenol groups of 8ii can be coupled with synthon 7a according to Scheme I. We prepared the monocoupling product mono-9ii as well as the dicoupling product di-9ii and converted them to the

(43) Jendralla, H.; Chen, L.-J. Synthesis 1990, 827.

(46) Negishi, E. I. Acc. Chem. Res. 1982, 15, 340.

(48) Carew, T. E.; Schwenke, D. C.; Steinberg, D. Proc. Natl. Acad. Sci. U.S.A. 1987, 84, 7725.

(50) Schwarz, G. Selecta 1987, 42, 2556.

(53) Beynen, A. C. Artery 1987, 14, 113.

<sup>(37)</sup> Assignments are based on the better biological potency and on the larger R<sub>f</sub> value (silica, cyclohexane/ethyl acetate) of 25r and 25w.

<sup>(38) (</sup>a) Bartoli, G. Acc. Chem. Res. 1984, 17, 109. (b) Kienzle, F. Helv. Chim. Acta 1978, 61, 449.

<sup>(39)</sup> Organikum, 10th ed.; VEB Verlag der Wissenschaften: Berlin, 1971; p 588.

<sup>(40)</sup> Wynberg, H. Chem. Rev. 1960, 60, 169.

<sup>(41)</sup> Cerny, M.; Malek, J. Collect. Czech. Chem. Commun. 1970, 35, 3079.

<sup>(42)</sup> Bode, K. D. In Houben-Weyl, 4th ed.; Georg Thieme Verlag: Stuttgart, 1977; Vol. VI/1c, p 1084.

<sup>(44) (</sup>a) Heck, R. F. Acc. Chem. Res. 1979, 12, 146. (b) Patel, B. A.; Ziegler, C. B.; Cortese, N. A.; Plevyak, J. E.; Zebovitz, T. C.; Terpko, M.; Heck, R. F. J. Org. Chem. 1978, 43, 2941.

<sup>(45)</sup> Heck, R. F. Palladium Reagents in Organic Syntheses; Academic Press: New York, 1985.

<sup>(47)</sup> Sartori, G.; Bigi, F.; Casiraghi, G.; Casnati, G.; Chiesi, L.; Ar-decini, A. Chem. Ind. 1985, 762.

<sup>(49)</sup> Kita, T.; Nagano, Y.; Yokode, M.; Ishii, K.; Kuma, N.; Ooshima, A.; Yoshida, H.; Kawai, C. Proc. Natl. Acad. Sci. U.S.A. 1987, 84, 5928.

<sup>(51)</sup> Eder, H. A.; Bilheimer, D. W.; Steinberg, D.; Davignon, F.; Yamamoto, A.; et al. Dujovne, C. A.; et al. Miettinen, T. A.; et al. Kuo, P. T.; et al. A Symposium: New Developments in the Treatment of Hypercholesterolemia—Probucol. Am. J. Cardiology 1986, 57, 1Hff.

<sup>(52)</sup> Strandberg, T. E.; Vanhanen, H.; Miettinen, T. A. Gen. Pharmacol. 1988, 19, 317.

<sup>(54)</sup> Pryor, W. A.; Strickland, T.; Church, D. F. J. Am. Chem. Soc. 1988, 110, 2224.

## Scheme X<sup>4</sup>

 $^{a}$  (a) dry HF(l)/0 °C/5 h; (b) SiO<sub>2</sub> chromatography; (c) 3 equiv CH<sub>3</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>2</sub>COCl/3 equiv DMAP/5 equiv pyridine/CH<sub>2</sub>Cl<sub>2</sub>/25 °C/1 h; (d) (n-Bu)<sub>4</sub>NF·3H<sub>2</sub>O/AcOH/THF/0 °C; (e) 1.2 equiv 7a/2.4 equiv K<sub>2</sub>CO<sub>3</sub>/HMPT/cat. 18-crown-6/80 °C/12 h; (f) removal of di-9ii by SiO<sub>2</sub> chromatography; (g) 2 N aqueous HCl/THF/EtOH/25 °C/16 h; (h) 2 equiv aqueous 1 N NaOH/EtOH/25 °C/3 h; (i) 4.0 equiv 7a/8.0 equiv K<sub>2</sub>CO<sub>3</sub>/HMPT/cat. 18-crown-6/80 °C/12 h; (j) removal of mono-9ii by SiO<sub>2</sub> chromatography; (k) 1.65 equiv 7a/3.3 equiv K<sub>2</sub>CO<sub>3</sub>/DMSO/cat. 18-crown-6/85 °C/12 h.

respective sodium carboxylates mono-11ii and di-11ii.

Taking advantage of their slightly different steric shielding, the two hydroxy groups of hydroquinone 61 could be selectively acetylated (Scheme VIII). In the DMAP-catalyzed reaction at -18 °C of 61 with 1.3 equiv of acetic anhydride, 8kk was obtained in 70% yield, with only a trace of its regioisomer 811. Diacetate 62, obtained with 4 equiv of acetic anhydride at 25 °C, was monosaponified with moderate selectivity to give 811 and 8kk in a ratio of 2:1, the latter being removed by chromatography. 8kk and 811 were converted to sodium carboxylates 11kk

and 111l, respectively, according to Scheme I. The acetoxy protecting group was saponified by sodium hydroxide in parallel to the tert-butyl ester, as outlined in Scheme X.  $\beta$ -Quinolinols 8mm and 8nn (Scheme IX) were prepared by a condensation reaction of 63 with isatine as the key step, in analogy to the method of Kaslow et al. 55 In 800 the aliphatic and the aromatic substituent have changed places. It was obtained by condensation of  $\alpha$ -hydroxy-

Table II. Inhibition of Solubilized Rat Liver HMG-CoA Reductase in Vitro

no.	IC <sub>50</sub> , <sup>b</sup> nM	rel <sup>c</sup> pot.	no.	IC <sub>50</sub> , <sup>b</sup> nM	rel <sup>c</sup> pot.
11a	70	10	11y	95	9
11 <b>b</b>	50	13	11 <b>z</b>	95	8
11c	70	10	11 <b>aa</b>	400	2
11 <b>d</b>	100	7	11 <b>bb</b>	190	5
11e	90	7	11cc	<b>26</b> 0	2 5 3
11 <b>f</b>	160	5	11 <b>dd</b>	380	2
11g	140	5 5	11ee	900	1
11 <b>h</b>	270	2	11 <b>ff</b>	2.7	300
11i	1000	1	11gg	400	2
11j	150	4	11 <b>hh</b>	17	43
11k	80	10	mono-11iid	30	15
111	250	3	di-11i <b>i</b> d	10	37
11 <b>m</b>	50	15	11jj	6	113
11 <b>n</b>	>1000	<1	11 <b>kk</b>	4	171
11 <b>o</b>	>1000	<1	11ll	<b>2</b> 0	34
11 <b>p</b>	190	4	11 <b>mm</b>	1000	1
11 <b>r</b>	60	12	11nn	50	16
11 <b>s</b>	500	1	1100	>1000	<1
11t	100	9	11pp	950	1
11 u	27	31	11qq	85	9
11v	75	12	4rre	>1000	<1
11w	320	2	mevinolin $(1)^f$	8	100
11 <b>x</b>	470	1			

<sup>a</sup>The assay system described in ref 1 was used. <sup>b</sup>IC<sub>50</sub> values were determined by using four or five concentrations of each inhibitor. For estimation of relative inhibitory potencies, mevinolin was assigned a value of 100. The IC50 value of test compound was compared with that of mevinolin, corrected for the somewhat different molecular weight. <sup>d</sup>See Scheme X. Lactone form. Sodium carboxylate form.

acetone with o-aminobenzophenone.<sup>56</sup> Phenol 8b was transformed to the corresponding thiophenol 8pp (Scheme IX) via a Newman-Kwart rearrangement.<sup>57</sup>

### Biological Results

The optically pure sodium salts 11 were evaluated for their ability to inhibit solubilized, partially purified rat liver HMG-CoA reductase in vitro (Table II) and to inhibit cellular HMG-CoA reductase in cultures of hepatic cells (HEP G2, a human hepatoma cell line), as determined by the inhibition of the incorporation of sodium [14C]acetate into cholesterol (Table III). The latter method has both advantages and disadvantages compared with the inhibition of isolated lyophylized enzyme according to Table II. The cell culture test (Table III) incorporates the phenomena of drug uptake into the living cells and the distribution of the drug into the diverse cell compartments, as well as intracellular drug metabolism and possibly drug's toxicity to the cells. It allows for recognition of inhibition of cholesterol biosynthesis, regardless of the specific enzyme that is inhibited. HMG-CoA reductase inhibition may be observed, as well as inhibition of any other enzyme that is relevant on the biosynthetic pathway that leads to cholesterol. In contrast, inhibition of the isolated enzyme in vitro (Table II) is specific for HMG-CoA reductase and does not include the phenomena of uptake, distribution, elimination, and toxicity of the drug. In conclusion, the cell test treats cholesterol biosynthesis inhibition on a higher level, but the enzyme test is more specific and allows for a classification of the inhibitor type and a distinction of competitive and noncompetitive inhibitors.

Selected compounds were evaluated for their ability to inhibit hepatic de novo cholesterol synthesis in male rats after po administration, as determined by the inhibition

Table III. Inhibition of Acetate Incorporation into Cholesterol in HEP G2 Cellsa

no.b	IC <sub>50</sub> , nM	rel <sup>d</sup> pot.
mevinolin (1)	50	100
11 <b>r</b>	12000	<1
11s	1700	3
11 <b>w</b>	15000	<1
11 <b>x</b>	10000	<1
11aa	100	50
11 <b>d</b> d	3500	2
11ee	20000	<1
4ff	1.8	250
11ff	30	150 (170)
11gg	>500	<10
11 <b>hh</b>	55	90
mono-11ii	>1700	<3
di-11 <b>i</b> i	>1000	<5
11jj	0,5	10000
11kk	$4.1 (7.1)^h$	1200 (700)
11]]	$5.6 (28)^h$	900 (180) <sup>h</sup>
11mm	>5000	<1
11nn	250	20
1100	>5000	<b>&lt;</b> 1
11qq	2500	$ar{f 2}$
4rr	6500	< <u>1</u>

<sup>a</sup> Assay described in ref 1. <sup>b</sup> For structure see Table I. <sup>c</sup> IC<sub>50</sub> values varied somewhat for different batches of cells. Mevinolin sodium salt averaged  $IC_{50} = 50$  nM and was used in every run as an internal standard. The measured IC's for test compounds 11 were corrected for deviations of mevinolin's IC from its average value. d Mevinolin was assigned a value of 100. Potencies were obtained by comparison of test compounds 11 with the internal standard mevinolin. 'Sodium dihydroxy carboxylate form, optically pure. Lactone form of mevinolin IC<sub>50</sub> = 4.6 nM. Determined with a rat hepatocyte cell culture. Lowest activity determined in a set of experiments with different batches of HEP G2 cells.

Table IV. Inhibition of Hepatic Cholesterol De Novo Synthesis in Vivo (Rat, Orally, 5 mg/kg of body wt)a

no.	% cholesterol de novo synthesis	rel pot.
no drug	100.0	
4a	71.6	33
4b	65.6	40
4m	25.2	87
4u	36.4	74
4ff	11.4	103
4gg	95.7	6
4hh	84.5	18
mevinolin $(1)^b$	14.0	100

<sup>&</sup>lt;sup>a</sup> Assay described in ref 58d. <sup>b</sup> Lactone form, optically pure.

of the incorporation of sodium [14C]octanoate58 into hepatic cholesterol (Table IV).

Selected compounds were further evaluated for their ability to decrease plasma cholesterol levels in normolipemic NZW rabbits (Table V) and male beagle dogs (Table VI) after po administration. All these tests were also conducted under the same experimental conditions with mevinolin. The respective results are included in Tables II-VI.

Sodium carboxylates mono-11ii and di-11ii, intended to be hybrids of antioxidants and HMG-CoA reductase inhibitors (vide supra and infra), were additionally eval-

<sup>(56)</sup> Fehnel, E. A. J. Heterocycl. Chem. 1967, 4, 566.

Newman, M. S.; Hetzel, F. W. Org. Synth. 1971, 51, 139. Kwart, H.; Omura, H. J. Am. Chem. Soc. 1971, 93, 7250.

<sup>(58) (</sup>a) Dietschy, J. M.; McGarry, J. D. J. Biol. Chem. 1974, 249, 52. (b) Anderson, J. M.; Dietschy, J. M. J. Lipid Res. 1979, 20, 740. (c) Stange, E. F.; Dietschy, J. M. J. Lipid Res. 1983, 24, 72. (d) Dietschy, J. M.; Spady, D. K. J. Lipid Res. 1984, 25, 1469.

<sup>(59)</sup> Granzer, E.; Mauz, O.; Weiser, G.; Müller, K. R. 7th International Symposium on Drugs Affecting Lipid Metabolism, Milan, May 28-31, 1980; Abstract Book, p 187.

Windholz, M., Ed. The Merck Index, 10th ed.; Merck & Co., Inc.: Rahway, NJ, 1983; no. 2340, p 338.

**Table V.** Hypocholesterolemic Activity in Vivo (NZW Rabbits, n = 5 or 6, Po)<sup>a</sup>

no.	dosage, mg/kg body wt per day	serum total cholesterol <sup>b</sup>
4a <sup>c</sup>	50	-19
11bd	75	-25°
4m <sup>c</sup>	50	-2
4u <sup>c</sup>	50	-11
4ff°	10	-30
11 <b>ff</b> <sup>d</sup>	10	-32
	20′	-37
4gg <sup>c</sup>	10	-15
4hh <sup>c</sup>	10	-28
	20′	-34
11jj <sup>d</sup>	5	-46
11kk <sup>d</sup>	5	-12
mevinolin (1)c	10	-25
(-,	20'	-27

<sup>a</sup>Experimental protocol described in ref 1. With a mean value of serum total cholesterol of 40 mg/dL the error limits (SEM) in the experiments varied between 1.3 and 3.2 mg/dL. <sup>b</sup> Percent change relative to control group after oral treatment for 10 days. Continued treatment did not lead to a further decrease of serum total cholesterol levels with any of the compounds listed. <sup>c</sup> Lactone form. <sup>d</sup> Sodium carboxylate form. <sup>e</sup> Four of five rabbits died on continued treatment between 10th and 14th application. <sup>f</sup> 20 mg/kg of body wt per day were administered over a period of 10 mg/kg of body wt per day (that is on 11th – 20th day of the total treatment period). The value given in the table was determined on the 20th day of the total treatment period.

Table VI. Hypocholesterolemic Activity in Vivo (Male Beagle Dogs. n = 4, Po)<sup>a</sup>

no.	dosage, mg/kg body wt per day	duration of treatment, days	serum LDL-cholesterol <sup>b</sup>
11ff°	10	14	-10
	25	14	-21
4hh <sup>d</sup>	30	21	+3
11 <b>hh</b> ¢	25	14	+3
4jj <sup>d</sup>	5	14	-9
••	30	14	-21
mevinolin $(1)^d$	10	19	-20

°For experimental protocol see Experimental Section. With a mean value of serum LDL-cholesterol of 310  $\mu$ g/mL the error limits (SEM) in the experiments were  $44 \pm 5 \mu$ g/mL. Percent change relative to control group. °Sodium carboxylate form. d Lectore form.

uated for their effects on serum lipoproteins and other metabolic parameters after subchronic (7 days) po administration to male rats. The data, together with those of clofibrate and probucol, are compiled in Table VII.

Those phenols 8 that possess the structural elements prerequisite for antioxidative/free-radical inhibiting activity and HMG-CoA reductase inhibitors (lactone 4 and sodium carboxylates 11) prepared from building blocks 8

were evaluated for their ability to inhibit microsomal lipid peroxidation and cupric sulfate induced LDL oxidation in vitro (Table VIII).

Examples given in Table IX demonstrate that structure-activity relationships (SAR) for phenoxy-type inhibitors (2, A-B = OCH<sub>2</sub>) in some aspects diverge from those of similar systems (2, A-B = CH<sub>2</sub>CH<sub>2</sub> or CH—CH).

#### Discussion

Compounds 11a-e, carrying a diphenylmethyl substituent in the ortho position of the phenolic core, inhibited solubilized rat liver HMG-CoA reductase with 7-13% of mevinolin's potency. Different substitution of the remaining ortho and para position of the phenolic core (CH<sub>3</sub>) or Cl) and of the core of the phenyl substituents (F, CH<sub>3</sub>, OCH<sub>3</sub>) had only small effects on the solubilized liver enzyme. Lactones 4a and 4b inhibited hepatic cholesterol de novo synthesis in rats with 33 and 40%, respectively, of mevinolin's potency, after po administration. The tendency of highly lipophilic chloro(fluoro)-substituted compounds to considerably surpass in vivo (Table IV) the activity expected from their inhibition of solubilized enzyme (Table II) is reflected in compounds with other substitution patterns, too (notably 4m and 4u). They inhibited cholesterol synthesis in vivo with 87 and 74%, respectively, of mevinolin's activity, though possessing only 15 and 31%, respectively, of mevinolin's potency to inhibit solubilized HMG-CoA reductase.

Replacements in the bulky ortho substituent of the p-fluorophenyl by alkyl groups decreased enzyme inhibition (Table II, compare 11f and 11g with 11a). Surprisingly, the activity dropped considerably when the p-fluorophenyl group was replaced by a p-fluoro-m-methylphenyl group (compare 11h and 11i with 11c and 11b), indicating strict steric requirements in the enzyme remote from the space probed by reported inhibitors.<sup>8-19</sup>

The bulky ortho substituent  $R^6$  of 11a-i was replaced by the sterically undemanding but extended 3-phenoxy-propyl substituent. The resulting compounds 11k and 11m inhibited the solubilized enzyme with 10 and 15%, respectively, and the cholesterol synthesis in vivo with 87% (4m) of mevinolin's potency.

Unexpectedly, the p-fluorophenyl group was no longer tolerated in the benzylic ( $\alpha$ -) position of the 3-phenoxypropyl substituent (Tables II and III, compare 11j with 11b,c,k,m). This result implies that the 3-phenoxypropyl substituent has less conformational freedom in the enzyme-inhibitor complex than the bulkier and rigid p-fluorophenyl group of 11b or 11c. The presence of a para substituent  $R^4$  was essential for adequate inhibitory potency (see 111). The second ortho substituent  $R^2$  may not be larger than isopropyl (compare 11n,o with 11k).

Compound 11p with a cyclohexylcarbonyl substituent  $R^6$  had low activity.  $\alpha,\alpha$ -Dimethylbutanoic acid esters 11r

Table VII. Effect on Serum Lipoproteins and Other Metabolic Parameters after Subchronic (7 Days) Oral Administration to Male Rats  $(n = 10)^a$ 

		% ch	ange relativ	e to control	group		
	dosage, mg/kg	, to	al cholester	ol	liver	body	cholesterol
test compound	of body wt per day	VLDL	LDL	HDL	weight	weight	HDL/LDL <sup>b</sup>
mono-11ii	30	+12	-26	+13	+3	+5	1.27
	100	+13	-16	-3	+1	+5	1.15
di-11ii	100	-23	-52	+9	-1	+5	2.25
probucol (60)°	30	+4	-24	-11	+5	+5	0.97
clofibrate (71)d	100	-40	-19	-29	+15	+8	0.88
no druge	<del>-</del>					+4	$1.00^{b}$

<sup>&</sup>lt;sup>e</sup>For protocol see Experimental Section. With a mean value of serum total cholesterol of 87 mg/dL the error limits in the experiments were  $4.6 \pm 0.8$  mg/dL. <sup>b</sup> Value of control group was normalized to 1.00. <sup>e</sup>Formula: Scheme VIII. <sup>d</sup> Formula:  $p\text{-ClC}_8H_4OC(CH_3)_2CO_2C_2H_5$  (71). <sup>e</sup>Control group.

Table VIII. Inhibition of Microsomal Lipid Peroxidation and of Cu<sup>2+</sup>-Catalyzed LDL Oxidation (in Vitro)<sup>a</sup>

	inhibn o	f microsomal lipid p	oeroxidn	inhibi	of Cu <sup>2+</sup> -cat. LDL	oxidn
no.	IC <sub>50</sub> , μΜ	% at 10 <sup>-5</sup> M	rel <sup>c</sup> pot.	IC <sub>50</sub> , μM	% at 10 <sup>-5</sup> M	rel <sup>c</sup> pot.
mono-11ii	2.7		4	-		
di-11 <b>ii</b>		<10	≪0.1			
4jj				>10	23	<0.1
4jj 8jj	3.0	95	3	1.0		11
11jj		<10	≪0.1	>10	41	0.1
8kk	2.8	98	4	0.15		73
11kk		<10	≪0.1	6.0		2
811	4.8	83	2			
11ll		27	<0.1	1.1		10
terbuficine <sup>59</sup> (72) <sup>d</sup>	0.1		100°	0.11		100°
probucol (60)e				0.5		22

<sup>a</sup> Assays described in Experimental Section. <sup>b</sup> For definition of R<sup>2</sup>-R<sup>6</sup> and structures of 4 and 11, see Table I. <sup>c</sup> Terbuficine (72) was assigned a value of 100. <sup>d</sup> Formula:

Formula: Scheme VIII.

and 11s of the corresponding alcohol were prepared with the intention to simulate the ester group of synvinolin (vide supra); however this met with only moderate success (11r). The corresponding amides 11w and 11x had low activity. We concluded that polar functionality (keto, ester, amide) is not tolerated in the  $\alpha$ -position of the cyclohexylmethyl substituent  $R^6$ , and indeed obtained reasonably active compounds 11t-y (Tables II and III) when this functionality was omitted. Within this series, again, o,p-dichloro substitution (11u) was best and led to 74% of mevinolin's in vivo activity (Table IV).

Introduction of one or two fluorine atoms or of p-fluorophenyl into the  $\alpha$ -position of the cyclohexylmethyl substituent  $\mathbb{R}^6$  worsened activity slightly, leading to compounds (11y,qq,z) comparable to o-diphenylmethyl substitution (e.g. 11b). Replacement of the phenolic oxygen by sulfur diminished the activity (11pp).

There are no "optimal" substituents R<sup>2</sup>-R<sup>6</sup> per se. All the substituents are highly interdependent as regards to their optimal shape. For example, the ortho (R<sup>2</sup>) substituent isopropyl has been found highly effective in combination with the ortho (R<sup>6</sup>) substituent p-fluorophenyl and the para (R4) substituent (substituted) phenyl or isopropyl in several five- and six-membered heterocyclic systems. 1,2,8-14,16,19 This SAR was roughly but not perfectly reflected in the phenoxy systems 11ff-ll (vide infra). However, the R<sup>6</sup> substituents cyclohexylmethyl or bis(pfluorophenyl)methyl, which had been found effective together with R<sup>2</sup> substituents such as methyl or chloro (vide supra), could not be successfully combined with R<sup>2</sup> = isopropyl (11aa,dd). A meta/para-condensed aromatic ring, well-tolerated in the case of o-isopropyl, o'-(p-fluorophenyl) substitution (see 77,61 Table IX), was not tolerated in the case of o-methyl, o'-[bis(p-fluorophenyl)methyl] substitution (compare 11a with 11ee). Up to this point, the reported SAR results are qualitatively

consistent with the purely steric model proposed by Roth et al. 14 for systems with a two-carbon bridge (2: A-B =  $CH_2CH_2$  or (E)-CH=CH). However, we obtained unequivocal evidence that SAR for phenoxy systems 11 partially diverge from this model and that electronic effects cannot always be neglected. The most striking examples are collected in Table IX. In the case of pyridine-based inhibitor 731 substitution of the p-isopropyl by the tertbutyl group gave 74,1 three times more active in enzyme inhibition and more potent in vivo (rabbit po). The same substitution performed on the sterically equivalent, highly efficacious phenoxy system 11ff gave 11gg, virtually inactive in vitro and in vivo. Sterically equivalent pyrrole,<sup>2</sup> indole,61 and pyridine systems1 (Table IX) had highly consistent inhibitory activity. Substitution of o-methyl for o-isopropyl uniformly increased activity by a factor of 12-13 and gave compounds with about 3 times mevinolin's potency in vitro. The activity of sterically equivalent quinolinoxy system 11mm comparably increased, when o-methyl was replaced by isopropyl (11nn), but both compounds are less active than expected based on purely steric considerations. A comparison of dichlorophenoxy compound 11bb with sterically equivalent 80<sup>6c</sup> (Table IX) similarly leads to the conclusion that the electronic effects on efficacy of HMG-CoA reductase inhibition cannot be neglected.

Interesting activities and pharmacological profiles were obtained for the group of compounds 11ff-11. 11ff inhibited HMG-CoA reductase with 3 times the potency of mevinolin (1). In the cell test lactone 4ff and sodium salt 11ff were 2.5 and 1.5 times, respectively, more active than the corresponding forms of mevinolin. 4ff inhibited hepatic cholesterol "de novo" synthesis in vivo (rat, po, Table IV) equipotent with mevinolin. 4ff and 11ff decreased plasma cholesterol levels (rabbit, po, Table V) equipotently with mevinolin at 10 mg/kg per day and with 150% of mevinolin's efficacy at 20 mg/kg per day. Thus, 11ff was the first phenoxy-type inhibitor to be successful in a chronic in vivo experiment since 4a, 4m, and 4u were not significantly active even in the high dose of 50 mg/kg per

<sup>(61)</sup> Kathawala, F. G.; Scallen, T.; Engstrom, R. G.; Weinstein, D. B.; Schuster, H.; Stabler, R.; Kratunis, J.; Wareing, J. R.; Jewell, W. F.; Widler, L.; Wattanasin, S. 8th International Symposium on Atherosclerosis, Rome, Oct 9-13, 1988; p 445.

Table IX. Examples for Divergence of SAR, Induced by the Ether Bridge

R³	no.	IC <sub>50</sub> , <sup>a</sup> nM	in vivo	(rabbit, po) <sup>b</sup>	no.	IC <sub>50</sub> , <sup>a</sup> nM	in vivo (1	abbit, po) <sup>b</sup>
i-Pr t-Bu	73° 74°	3° 1°	very pot more po	tent tent than 73	11ff 11gg	2.7 400	very poten not signific	t cantly active
	но₄	CO <sub>2</sub> Na OH	НО	CO₂Na ,OH	HO CO <sub>2</sub> Ni	HO	CO <sub>2</sub> Na	
		75, 76	R_N 7	7	11mm,nn			
R	no.	IC <sub>50</sub> , a nM	no.	IC <sub>50</sub> , <sup>a</sup> nM	no.	IC <sub>50</sub> , a nM	78, 79 no.	IC <sub>50</sub> , a nM
CH <sub>3</sub> i-Pr	75 <sup>d,e</sup> 76 <sup>d</sup>	35 <sup>d-f</sup> 3 <sup>d</sup>	77#	2.5 <sup>f,h</sup> (6.9) <sup>g,i</sup>	11mm 11nn	1000 50	<b>7</b> 8° 7 <b>9</b> °	38° 3°
		но	CO <sub>2</sub> Na OH		но	CO <sub>2</sub> Na OH		
		CI			CI	J f		
			Ci 11 <b>bb</b>		Ci 80 (70	00679 <sup>/</sup> )		

<sup>a</sup> Inhibition of solubilized HMG-CoA reductase. Test according to Table II. <sup>b</sup> Test according to Table V. <sup>c</sup> Reported in ref 1. <sup>d</sup> Reported in ref 2. <sup>e</sup> Additional methyl substituent in position 5 of pyrrole. <sup>f</sup> Corrected for inactive enantiomer. <sup>e</sup> Reported in ref 61. <sup>h</sup> Resynthesized in Hoechst AG (1986) and used in test according to Table II for comparison. <sup>f</sup> Difference of reported and measured IC<sub>50</sub> value probably due to differences in preparation of solubilized, partially purified enzyme. <sup>f</sup> Reported in ref 6c.

day (Table V), despite possessing 33-87% of mevinolin's activity in the *acute* in vivo experiment (Table IV). With 75 mg/kg per day 11b was significantly active, but toxic, in the rabbit experiment.

Surprising data were obtained with the o,p-bis(p-fluorophenyl)-substituted system 11hh. Its enzyme inhibition was reproducibly only 43% of mevinolin's, considerably less than expected by analogy to heterocyclic systems with a two-carbon bridge<sup>1,2</sup> (compare 76<sup>2</sup> and 79, Table IX). 11hh had 90% of mevinolin's potency in the cell test (Table III), only 18% in the acute in vivo (Table IV) test, but 100–150% in the chronic in vivo experiment (Table V). Thus 4hh decreased plasma cholesterol levels in rabbits as potently as the chemically very similar compound 4ff, despite exhibiting only 14% of its in vitro and acute in vivo activity.

This enhanced chronic in vivo activity was even more pronounced with 11jj, which differs from 11hh only by the introduction of a sulfur atom in para position. 11jj was only slightly superior to mevinolin in vitro (Table II), but 100 times more active in the cell test, repeatedly (Table III). It reduced plasma cholesterol levels (rabbit po) by 46% at a dosage of only 5 mg/kg per day (Table V), thus being 4-6 times more efficacious than mevinolin. Its

phenolic building block 8jj (a potential metabolite of 11jj; vide infra) inhibited LDL oxidation in vitro with half of probucol's activity (Table VIII). 11kk (0,0'-diisopropyl substitution) exceeded the enzyme-inhibiting activity of its regioisomer 11ll by a factor of 5. 11kk had 7-12 times mevinolin's potency in the cell test, but was not significantly active in the rabbit experiment. Its phenolic building block 8kk inhibited LDL oxidation 3 times more than probucol, while 11kk itself had only 9% of probucol's potency. Regioisomeric 11ll inhibited LDL oxidation with half of probucol's activity.

Compounds mono-11ii and di-11ii were only moderately active HMG-CoA reductase inhibitors. This is attributed to an excessive bulk of their para substituents R<sup>4</sup> (compare 11gg). However, both compounds reduced LDL-cholesterol after po administration to male rats (Table VII). Compounds that exclusively act via HMG-CoA reductase inhibition are usually inactive in this model, due to a fast, intensive enzyme induction that compensates for the inhibition.<sup>62</sup> An improved HDL/LDL ratio, notably for di-11ii, was observed.

<sup>(62)</sup> Endo, A.; Tsujita, Y.; Kuroda, M.; Tanzawa, K. Biochim. Biophys. Acta 1979, 575, 266.

Unfortunately, all tested phenoxy-type inhibitors had only moderate activity after po administration to male beagle dogs (Table VI). This result was unexpected, since for inhibitors based on heterocyclic systems (carrying the same substituents) in vitro activity correlated well with the in vivo activity in both rabbits and dogs.<sup>1,2</sup>

The reason for the decreased oral activity of phenoxytype inhibitors 11ff, 4hh (or 11hh), and 4jj in the dog (Table VI), as opposed to their excellent oral activities in the rabbit (Table V), the rat (Table IV), and in vitro (Tables II and III), was not elucidated. However, since investigations of HMG-CoA reductases of several different animal species did not indicate relevant species differences of the active site,  $^{63}$  we presume that a fast metabolic deactivation takes place in the dog. According to the generally accepted mechanism, aryl ethers are metabolically cleaved by an oxidative O-desalkylation. Accordingly, phenoxy-type inhibitors (2, A = O) would be oxidized in the  $\alpha$ -position (B in 2). The resulting hemiacetal would be hydrolyzed to give the corresponding phenol 8, which possesses the desired ability to inhibit LDL oxidation (Table VIII).

In summary, the introduction of an oxygen atom into bridge position A of general formula 2 induced some pronounced and surprising changes of the pharmacological profile. Chemical structure-activity relationships of respective compounds 4 diverged considerably from those of HMG-CoA reductase inhibitors with carbon-carbon bridges (A-B = CH<sub>2</sub>CH<sub>2</sub> or CH=CH). The plasma cholesterol lowering activity (rabbit) of some compounds of general formula 4 was considerably stronger than would be expected on the basis of the in vitro potency of these compounds. Although several compounds 4 were highly efficacious in the rabbit, they had only moderate activity in male beagle dogs, whereas heterocyclic inhibitors<sup>1,2</sup> and mevinolin (1) exhibited comparable activity in both animal species. Some compounds of general formula 4 exceeded the activity of clofibrate and probucol to reduce plasma LDL-cholesterol levels after subchronic, oral administration to male rats, an activity not generally seen for compounds that exclusively act via HMG-CoA reductase inhibition.62 Several compounds of general formula 4 inhibited cholesterol biosynthesis and oxidation of LDL (at least in vitro), thus interfering with two atherogenic processes simultaneously.

Lactones 4ff and 4hh, as well as sodium carboxylates 11ff and notably 11jj, exceeded mevinolin's potency in vitro and in vivo (rat, rabbit), but not in the dog.<sup>65</sup> The surprisingly low subchronic toxicity<sup>66</sup> of these compounds renders them attractive as backups of HR 780,<sup>1</sup> which is currently in clinical studies.

(64) Mutschler, E. Arzneimittelwirkungen. Lehrbuch der Pharmakologie und Toxikologie, 5th ed.; Wissenschaftliche Verlagsgesellschaft: Stuttgart, 1986; p 21.

(65) A referee suggested that the differences seen between dog and other species might not be evident when the more sensitive model of cholestyramine primed dogs is used. Respective studies are ongoing and will be reported elsewhere.

(66) Following a referee's suggestion, data will be reported elsewhere.

## **Experimental Section**

For general remarks see ref 1. All starting materials were commercially available unless indicated otherwise. Compounds  $6,^{24}$  8ff,  $^{43}$  8hh,  $^{43}$  8ii,  $^{43}$  8jj,  $^{43}$  20,  $^{32}$  48-51,  $^{43}$  53,  $^{47}$  56,  $^{43}$  57-59,  $^{43}$  61,  $^{43}$   $\alpha,\alpha$ -dimethylbutanoic anhydride (needed for preparation of 25r and 25s), and  $\alpha,\alpha$ -dimethylbutyronitrile (needed for preparation of 25w and 25x) were prepared as described in the literature. Compounds 54, 55, and 8gg were obtained in analogy to the descriptions given for 8ff, 8hh, and 57.

(3R,5S)-6-[(Methylsulfonyl)oxy]-3,5-O-isopropylidene-3,5-dihydroxyhexanoic Acid tert-Butyl Ester (7a). Methanesulfonyl chloride (238 g, 2.08 mol) was added dropwise at 0-5 °C within 1 h to the solution of 6 (360 g, 1.39 mol) in  $CH_2Cl_2$  (1.5 L) and pyridine (1.5 L), and the mixture was stirred for 30 min at 0 °C. The solvents were removed in vacuo. The residue was dissolved in toluene (0.5 L) and washed twice with water, saturated NaHCO<sub>3</sub> solution, and with brine. The organic phase was dried (MgSO<sub>4</sub>), evaporated in vacuo, redissolved in toluene, and re-evaporated. The residual oil was seeded with crystals of 7a, leading to a quick crystallization. The solid was stirred with cyclohexane (1 L), suction-filtered, and washed with cyclohexane. It was dried in vacuo to give 421 g (1.25 mol, 90% yield) of colorless crystals, mp 76-78 °C. Anal.  $(C_{14}H_{26}SO_7)$  C, H, S.

(3R,5S)-6-[(p-Tolylsulfonyl)oxy]-3,5-O-isopropylidene-3,5-dihydroxyhexanoic Acid tert-Butyl Ester (7b). p-Toluenesulfonyl chloride (5.8 g, 30.5 mmol) was added in portions at 0-5 °C to the solution of 6 (4.0 g, 15.4 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and pyridine (50 mL). The mixture was stirred for 2.5 h at 0 °C and 0.5 h at 20 °C. The solvent was evaporated in vacuo. The residue was redissolved in toluene and washed twice with water, saturated NaHCO<sub>3</sub> solution, and brine. The organic phase was dried, concentrated in vacuo, and filtered through 50 g of silica with CH<sub>2</sub>Cl<sub>2</sub>/EtOAc 30:1. The solvent of the filtrate was evaporated in vacuo to give 5.0 g (12.1 mmol, 78% yield) of colorless crystals (mp 90-93 °C), which were washed with hexane, dried in vacuo, and stored at -25 °C. Anal. (C<sub>20</sub>H<sub>30</sub>SO<sub>7</sub>) C, H, S.

(3R,5S)-6-Iodo-3,5-O-isopropylidene-3,5-dihydroxyhexanoic Acid tert-Butyl Ester (7c). NaI (12.5 g, 83.4 mmol) was added to the solution of tosylate 7b (4.6 g, 11.1 mmol) in acetone (250 mL). The mixture was refluxed (18 h), NaI (7.0 g, 46.7 mmol) was added, and reflux was resumed (6 h), leaving 20% of unreacted 7b according to TLC. The solvent was evaporated, the residue dissolved in toluene and was washed twice with water and then with brine. The organic phase was dried and concentrated in vacuo, and the residue was chromatographed through 200 g of silica with cyclohexane/EtOAc 4:1, giving 500 mg of unreacted 7b and 3.4 g (9.2 mmol, 83% yield) of the title compound as a glass. Anal. (C<sub>13</sub>H<sub>23</sub>IO<sub>4</sub>) C, H, I.

In a series of experiments aimed to prepare the corresponding triflate 7d, no stable product of reasonable lipophilicity could be obtained from 6 and triflic anhydride in pyridine.

Coupling of Phenols 8 with Mesylate 7a To Give Acetonides 9. General Procedure. DMSO or HMPA were employed as the solvent, the latter in case of phenols 8 containing an oxidation-sensitive thio functionality or hydroquinone moieties.

K<sub>2</sub>CO<sub>3</sub> powder (1.0 mol) was added to a solution of phenol 8 (0.5 mol) and mesylate 7a (0.5 mol) in the solvent (1.5 L). 18-Crown-6 (100 mg) was added and the mixture was heated to 60-85 °C, depending on the steric shielding of phenol 8 (reaction time ≈ 12 h). Reaction progress was monitored by TLC (100% toluene or cyclohexane/EtOAc 5:1) until 8 had disappeared. [With sterically hindered 8 it is preferable to add additional 7a (0.25 mol) after 12 h and to continue the heating for 12 h.] Ice (2 kg), saturated NaHCO<sub>3</sub> solution (1 L), and Et<sub>2</sub>O (1 L) were added and the mixture was extracted with Et<sub>2</sub>O (3 × 1 L). The combined extracts were washed with NaHCO3 solution and with brine, dried (MgSO<sub>4</sub>), and concentrated to dryness in vacuo. The residue generally crystallized, when triturated with CH<sub>3</sub>OH and scraped: 60-85% yield. Products from very sterically hindered phenols 8 were less pure and needed to be purified by silica chromatography (cyclohexane/EtOAc 10:1 + 0.1% NEt<sub>3</sub>): 45-60% yield. 9ff (solvent DMSO, 75% yield, mp 73-74 °C), 9hh (DMSO, 85%,

<sup>(63) (</sup>a) Luskey, K. L.; Stevens, B. J. Biol. Chem. 1985, 260, 10271.
(b) Liscum, L.; Finer-Moore, J.; Stroud, R. M.; Luskey, K. L.; Brown, M. S.; Goldstein, J. L. J. Biol. Chem. 1985, 260, 522.
(c) Chin, D. J.; Gil, G.; Russell, D. W.; Liscum, L.; Luskey, K. L.; Basu, S. K.; Okayama, H.; Berg, P.; Goldstein, J. L.; Brown, M. S. Nature 1984, 308, 613. (d) Basson, M. E.; Thorsness, M.; Finer-Moore, J.; Stroud, R. M.; Rine, J. Mol. Cell Biol. 1988, 8, 3797. (e) Woodward, H. D.; Allen, J. M. C.; Lennarz, W. J. J. Biol. Chem. 1988, 263, 18411.

<sup>(67)</sup> Hasek, R. H.; Elam, E. U.; Martin, J. C. J. Org. Chem. 1961, 26, 1822.

mp 114-115 °C), mono-9ii (HMPA, 48% besides 20% of di-9ii, mp 49-52 °C), di-9ii (HMPA, 8 equiv  $K_2CO_3$ , 4 equiv 7a, 55% besides 15% of mono-9ii, mp 47-51 °C), 9jj (HMPA, 74%, oil), 9kk (DMSO, 52%, 152-154 °C), and 9ll (DMSO, 53%, 145-147 °C) were produced.

tert-Butyl  $\beta$ ,  $\delta$ -Dihydroxy Carboxylates 10. General Procedure. A solution of acetonide 9 (0.5 mol) in EtOH (2.5 L), THF (1.25 L), and 2 N hydrochloric acid (280 mL) was stirred for 1 day at ambient temperature. The solution was adjusted to pH 7 with solid NaHCO<sub>3</sub>. Organic solvents were removed in vacuo, and the residue was divided between Et<sub>2</sub>O and water. The aqueous phase was extracted with Et<sub>2</sub>O. The combined extracts were washed with water and then with brine, dried (MgSO<sub>4</sub>), and evaporated in vacuo. 10ff (95% yield, an oil that slowly crystallized), 10hh (99%, colorless solid); mono-10ii, di-10ii, 10jj, 10kk, and 10ll (see Table I) were produced.

Saponification of tert-Butyl Esters 10 To Give Sodium Carboxylates 11. General Procedure. 1 N NaOH (10 mL, 10 mmol; in the case of mono-10ii, di-10ii, 10kk, and 10ll, 20 mL, 20 mmol) was added to the solution of ester 10 (10 mmol) in EtOH (75 mL). The reaction mixture was stirred until TLC (CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH 10:1) indicated disappearance of 10 (usually 2-3 h). The solvent was removed in vacuo. The residue was repeatedly redissolved in CH<sub>3</sub>OH or toluene and evaporated to dryness each time. It was washed with i-Pr<sub>2</sub>O and then with Et<sub>2</sub>O and dried in vacuo to give a colorless solid; >95% yield. 11kk and 11ll prepared according to this procedure contain 1 equiv of NaOAc.

Cyclization of tert-Butyl Esters 10 To Give Lactones 4. General Procedure. Trifluoroacetic acid (55 mL) was added at 0 °C to the solution of ester 10 (100 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (200 mL). The mixture was stirred for 2 h at 20 °C. TLC (cyclohexane/EtOAc 1:1) indicated complete transformation. NaHCO<sub>3</sub> (50 g) was added portion wise at 0 °C. The mixture was poured into pH 7 buffer and extracted with EtOAc (3 × 100 mL). The extracts were washed with pH 7 buffer, water, and with brine. They were dried (MgSO<sub>4</sub>), and the solvent was evaporated in vacuo. The residue was either recrystallized (i-Pr<sub>2</sub>O/EtOAc 2:1) or purified by chromatography (see Table I).

Coupling of Phenol 8gg with Iodide 7c To Give 9gg and a Stereoisomer. K<sub>2</sub>CO<sub>3</sub> powder (540 mg, 3.9 mmol) and hydroquinone (10 mg) were added to a solution of 8gg (557 mg, 1.95 mmol) and iodide 7c (725 mg, 1.95 mmol) in DMSO (50 mL). The mixture was stirred under argon at 52-55 °C for 3 h. (toluene/EtOAc 30:1) indicated only small amounts of iodide 7c, and unreacted phenol 8gg and a new spot (product) were present in a ratio of about 1:1. Chromatography (200 g silica, toluene) removed 8gg (300 mg) and gave an oil, which according to HPLC (LiChrosorb Si 60, 10 μm, methylcyclohexane/EtOAc 22:1, 40 °C, 1.0 mL/min, detect 254 nm) consisted of two product peaks ( $t_{\rm R}$ 5.56 and 6.08 min, 39 and 51%) and of 7c ( $t_R$  7.56 min, 8%). The products were separated by preparative HPLC and characterized by 270-MHz <sup>1</sup>H NMR and MS. Comparison of the NMR with those of compounds 9 (obtained from mesylate 7a) suggested that the product of smaller  $t_R$  was 9gg, whereas the more polar product was a stereoisomer of 9gg.

1,1-Diethoxy-2-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]ethane (12ff).  $K_2CO_3$  powder (243 g, 1.76 mol) and then bromoacetaldehyde diethyl acetal (191 g, 0.97 mol) were added to a solution of 2,4-diisopropyl-6-(4-fluorophenyl)phenol (8ff; 240 g, 0.88 mol) in DMSO (2 L; dried by passage through basic alumina activity I, immediately before use). The mixture was heated for 10 h to 90 °C. It was cooled and poured into ice/water (5 L). It was extracted with Et<sub>2</sub>O (4 × 1 L). The combined extracts were washed with brine and dried, and the solvent was evaporated in vacuo.  $K_2CO_3$  powder (1 g) was added to the residue, and it was distilled in vacuo (without column) to give, after a small prerun, the title compound (298 g, 87% yield) as a colorless oil (bp 148–152 °C/2 ×  $10^{-4}$  bar). Anal.  $(C_{24}H_{33}FO_3)$  C, H, F.

[2,4-Diisopropy]-5-(4-fluorophenyl)phenoxy]acetaldehyde (13ff). 2 N HCl (0.9 L) was added to the solution of acetal 12ff (287 g, 0.74 mol) in acetone (2.75 L). The mixture was stirred for 5 h at 65 °C and then allowed to cool to 20 °C overnight. The acetone and part of the water was evaporated in vacuo. The residue was taken up in  $\rm Et_2O$ . The organic phase was separated and washed with water, saturated NaHCO<sub>3</sub> solution, and with brine. The organic phase was dried (MgSO<sub>4</sub>) and the solvent was

evaporated in vacuo. The residue was distilled in vacuo (without column) to give, after a small prerun, the title compound (198 g, 85% yield) as a colorless oil (bp 140–143 °C/1 × 10<sup>-4</sup> bar), which quickly crystallized. Anal. ( $C_{20}H_{23}FO_2$ ) C, H, F.

(S)-2-Hydroxy-1,2,2-triphenylethyl (3S)-Hydroxy-4-[2,4diisopropyl-6-(4-fluorophenyl)phenoxy]butanoate (14ff). (a). A 1.6 M solution of n-BuLi in hexane (381 mL, 610 mmol) was added at -70 °C under N<sub>2</sub> to a solution of diisopropylamine (90 mL, 636 mmol) in THF (700 mL). The solution was stirred for 30 min at 0 °C. The resulting LDA solution was added via a Flex-needle<sup>68</sup> through septa to a suspension (-70 °C) of (S)-(-)-phenyl (2-hydroxy-2,2-diphenylethyl)acetate<sup>28</sup> (93.0 g, 280 mmol) in THF (900 mL). The mixture was allowed to warm up to 0 °C and stirred for 30 min at this temperature. The resulting yellow-orange clear solution was cooled to -90 °C and a precooled solution of [2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]acetaldehyde (13ff; 80.0 g, 254 mmol) in THF (250 mL) was added dropwise via a Flex-needle at such a rate that the temperature did not climb above -80 °C. The reaction mixture was stirred for 1 h at -90 °C. The cold mixture was poured into saturated aqueous NH<sub>4</sub>Cl solution (2 L) and stirred for 1 h with warming to 25 °C. It was extracted with Et<sub>2</sub>O (3  $\times$  1 L), the combined extracts were washed with 20% NaCl solution and dried, and the solvent was evaporated in vacuo. The residue was triturated with n-pentane (2 L) and stirred for 30 min. The solid was suction filtered, washed with n-pentane and dried in vacuo to give a colorless solid (159 g, 246 mmol, 97% yield). HPLC indicated 60% de. <sup>1</sup>H NMR [Eu(hfc)<sub>3</sub>] analysis of methyl ester 15ff prepared from this sample (vide infra) indicated 65% ee. It was generally observed that optical induction as analyzed by NMR was indicated to be 4-5% in excess of the HPLC value. An analytical sample was obtained by recrystallization from warm i-Pr<sub>2</sub>O, mp 169–171 °C. Anal. ( $C_{42}H_{43}FO_5$ ) C, H, F. HPLC of this sample still indicated 60% de; <sup>1</sup>H NMR [Eu(hfc)<sub>8</sub>] analysis of 15ff showed 65% ee. Modification of reaction conditions [(a) addition of aldehyde 13ff at -75 °C, or at -110 °C in the presence of 2-methylbutane; (b) transmetalation of the lithium enolate with 2 molar equiv of freshly prepared MgI<sub>2</sub> or MgBr<sub>2</sub>] consistently gave 14ff with 55-60% de, which was not significantly changed by recrystallization from warm acetone/hexane.

Methyl (3S)-Hydroxy-4-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]butanoate (15ff). Sodium (6.6 g, 287 mmol) was dissolved in CH<sub>3</sub>OH (300 mL). This solution (271 mL, containing 260 mmol of NaOCH<sub>2</sub>) was added dropwise within 10 min to a solution of 14ff (520 mmol) in CH<sub>3</sub>OH (2.5 L). The mixture was stirred for 90 min at ambient temperature. Glacial acetic acid (17.2 g, 286 mmol) was added dropwise with ice cooling. Solvents were removed in vacuo. The residue was redissolved in Et<sub>2</sub>O and washed with 20% NaCl solution and then with saturated NaHCO<sub>3</sub> solution and with brine. The solution was dried (MgSO<sub>4</sub>) and the solvent was evaporated in vacuo. n-Hexane (3 L) was added to the residue and the suspension was stirred extensively (90 min). The precipitated 1,1,2-triphenylethane-1,2-diol (150 g) was filtered off and washed with hexane. The filtrate was evaporated in vacuo to leave colorless crystals [192 g, 495 mmol, 95% yield, mp 72-74 °C, 65% ee (1H NMR/Eu-(hfc)3]. A sample was dissolved in a small amount of warm methanol and kept in a freezer to give crystals that had mp 76-77 °C and 66% ee. Anal.  $(C_{23}H_{29}FO_4)$  C, H, F.

tert-Butyl (5S)-Hydroxy-3-oxo-6-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]hexanoate (16ff). tert-Butyl acetate (232.4 g, 2.0 mol) was added dropwise at -70 °C under N<sub>2</sub> to a solution of LDA (2.05 mol) in THF/hexane (1:1, 2.5 L). After 1 h at -70 °C, the solution of methyl ester 15ff (194 g, 0.5 mol) in THF (1.0 L) was added (10 min) and the mixture was stirred for 2.5 h at -20 °C. The cold yellow solution was poured into 20% aqueous NH<sub>4</sub>Cl solution (5 L), leading to decolorization. The mixture was stirred for 10 min and then extracted with Et<sub>2</sub>O (2 × 2 L). The extracts were washed with brine, saturated NaHCO<sub>3</sub> solution, and brine and dried (MgSO<sub>4</sub>). The solvent was evaporated in vacuo to leave a pale-yellow oil (274 g, >100%) that slowly crystallized. A pure sample was obtained by chromatog-

<sup>(68)</sup> Commercially available from Aldrich Chemical Co.; Milwau-

raphy (SiO<sub>2</sub>, cyclohexane/EtOAc 3:1). Anal. ( $C_{28}H_{37}FO_5$ ) C, H, F.

tert-Butyl 3(R),5(S)-Dihydroxy-6-[2,4-diisopropyl-6-(4fluorophenyl)phenoxy]hexanoate (10ff). Triethylborane (560 mL of a 1 M solution in THF) was added dropwise at 20 °C to a solution of CH<sub>3</sub>OH (375 mL) in THF (1.5 L). The solution was stirred for 1 h. It was cooled to -78 °C. A solution of crude tert-butyl ester 16ff (236 g, 0.5 mol) in THF (800 mL) was added dropwise and the solution was stirred for 1 h at -70 to -75 °C. NaBH<sub>4</sub> (24.7 g, 0.65 mol) was added at once and stirring was continued for 3 h at this temperature. The cold mixture was poured into a 20% aqueous NH<sub>4</sub>Cl solution (3 L) and extracted with Et<sub>2</sub>O (3  $\times$  1 L). The extracts were washed with brine and dried, and the solvent was evaporated in vacuo. The residual oil was redissolved several times in wet methanol and this solvent was evaporated in vacuo at <20 °C. A methanolic solution was allowed to stand at 0 °C overnight. TLC (toluene/Et<sub>2</sub>O 6:1) indicated the successful conversion of the nonpolar boron ester of the diol to free diol 10ff ( $R_f = 0.11$ ). Pure 10ff was obtained after chromatography through silica (3.6 kg) with toluene/Et<sub>2</sub>O 6:1 as a colorless solid (175 g, 75% yield, mp 64–65 °C). Anal.  $(C_{28}H_{39}FO_5)$  C, H, F.

Sodium 3(R),5(S)-Dihydroxy-6-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]hexanoate (11ff). 1 N NaOH (245 mL) was added dropwise at 0-5 °C to a solution of tert-butyl ester 10ff (118.5 g, 0.25 mol) in EtOH (300 mL). The solution was stirred for 4 h at 25 °C. Solvents were removed in vacuo. Toluene was added several times and the solvent was evaporated in vacuo each time. n-Pentane (1 L) was added and the suspension was stirred for 30 min. The solid was collected by suction filtration (88 g, 80% yield, mp 230–233 °C dec). Anal. ( $C_{24}H_{30}FO_{5}Na$ ) C, H F

HPLC (Nucleosil 7  $C_{18}$ ,  $H_2O/CH_3CN$  60:40 + 0.1%  $NH_4OAc$ , 40 °C, 1.0 mL/min, detect. 260 nm) indicated the presence of the title compound (94.9%,  $t_R$  11.87 min, identical with material obtained from 10ff) and a homogeneous, slightly more polar impurity (5.0%,  $t_R$  10.00 min, most likely the 3(S) diastereomer of 11ff, due to incomplete stereoselectivity of the  $Et_3B/NaBH_4$  reduction). A 13-g portion (12% yield) of 11ff, containing 8.0% of the (diastereomeric) impurity, was obtained from the mother liquor.

4(R)-Hydroxy-6(S)-[[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]methyl]tetrahydro-2H-pyran-2-one (4ff), Prepared from Sodium Carboxylate 11ff According to Scheme II. 2 N hydrochloric acid (50 mL) was added at 0 °C to a solution of sodium carboxylate 11ff (22.0 g, 50 mmol) in water (800 mL). The precipitated carboxylic acid was immediately extracted twice with EtOAc. The combined extracts were washed twice with brine and then dried and evaporated to dryness in vacuo. The residue was dried in vacuo (20.92 g, 50 mmol, mp 97-100 °C). It was dissolved in THF (250 mL) and NEt<sub>3</sub> (5.57 g, 55 mmol) was added at 0-5°C. The mixture was stirred for 10 min at 0 °C and then cooled to -10 °C, and ethyl chloroformate (5.43 g, 50 mmol) was added dropwise. The mixture was stirred for 1 h at -5 °C. A 20% NaCl solution (1 L) was added. The solution was extracted with Et<sub>2</sub>O (3 × 300 mL). The combined extracts were washed with brine and dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo. The residue was chromatographed with toluene/EtOAc 3:1 + 0.03% NEt<sub>3</sub> through silica (2 kg). Small nonpolar impurities were removed and the last product-containing fractions were cutoff to reduce the amount of diastereomer. The solvent was removed in vacuo, and the residue was washed with a small amount of cold  $Et_2O$  and then with *n*-pentane. The solid was dried in vacuo to give a colorless powder (13.7 g, 34.2 mmol, 68% yield, mp 142–144 °C). Anal. ( $C_{24}H_{29}FO_4$ ) C, H, F.

HPLC (LiChrosorb SI 60 Merck, n-hexane/cyclohexane/1,2-dimethoxyethane 50:100:45, 40 °C, 1.0 mL/min, detect. 254 nm) indicated a chemical purity of the title compound ( $t_R$  19.4 min) of 96.8%. The major impurity ( $t_R$  22.95 min, probably 4(S)-hydroxy diastereomer) was 1.2%. For <sup>1</sup>H NMR determination of enantiomeric purity 2 × 5.5 mg of 11ff was dissolved in CD<sub>2</sub>Cl<sub>2</sub> in two NMR tubes. (+)-Eu(hfc)<sub>3</sub> (15 mg) and 25 mg of (-)-Eu-(hfc)<sub>3</sub>, respectively, were added, and the 400-MHz spectra were recorded. Sufficient differentiation of the signals of enantiomers were obtained for four sets of signals that in the absence of Eu(hfc)<sub>3</sub> were located at (1)  $\delta$  = 7.50 ppm (2 H, AA' part of

AA'XX'Y, 2,6-H of FC<sub>8</sub>H<sub>4</sub>), (2)  $\delta$  = 7.10 ppm (1 H, d, 3-H of 2-(4-FC<sub>6</sub>H<sub>5</sub>)-4,6-(i-Pr)<sub>2</sub>C<sub>6</sub>H<sub>2</sub>O), (3)  $\delta$  = 7.00 ppm (1 H, d, 5-H of 2-(4-FC<sub>6</sub>H<sub>5</sub>)-(4,6-i-Pr)<sub>2</sub>C<sub>6</sub>H<sub>2</sub>O, (4)  $\delta$  = ? (2 H, d with fine coupling, methylene group).

In the case of (+)-Eu(hfc)<sub>3</sub> signals of the minor enantiomer were shifted upfield relative to the corresponding signals of the major enantiomer. The indicated sets of signals (major/minor) were located at (1)  $\delta = 8.58/8.51$ , (2) 7.58/7.53, (3) 7.48/7.43, (4) 6.16/6.06 ppm.

In the case of (-)-Eu(hfc)<sub>3</sub> signals of the minor enantiomer were shifted downfield relative to the corresponding signals of the major enantiomer. The indicated sets of signals (major/minor) were located at (1)  $\delta = 8.54/8.62$ , (2) 7.55/7.59, (3) 7.45/7.49, (4) 6.18/6.26 ppm.

The ratio of integrals (major/minor, average from three recorded integrals) was  $81:19\pm3$ , regardless which of the indicated signal sets was used and regardless of the absolute configuration of Eu(hfc)<sub>3</sub>. This corresponds to 62% ee of 11ff.

2(RS)-Methoxy-4(R)-(tert-butyldiphenylsiloxy)-6(S)-[[(p-tolylsulfonyl)oxy]methyl]tetrahydropyran (21). p-Toluenesulfonyl chloride (112 g, 587 mmol) was added portionwise to a solution of alcohol  $20^{32}$  (120 g, 300 mmol) in  $CH_2Cl_2$  (0.5 L) and pyridine (0.5 L). The mixture was stirred for 3 h at ambient temperature. Solvents were removed in vacuo. The residue was redissolved in toluene and washed twice with water, twice with saturated NaHCO<sub>3</sub> solution, and once with brine. The organic phase was dried (MgSO<sub>4</sub>), and the solvent was evaporated in vacuo. The residue was chromatographed through silica (1.6 kg) with toluene/EtOAc 10:1. Unreacted p-TsCl was eluted first, followed by the title compound (163 g, yield 98%) as a nearly colorless glass.

2(RS)-Methoxy-4(R)-(tert-butyldiphenylsiloxy)-6(S)-(iodomethyl)tetrahydropyran (22). NaI (418 g, 2.79 mol) was added to the solution of tosylate 21 (193 g, 0.35 mol) in acetone (dried over  $\rm K_2CO_3$ ; 4.0 L). The mixture was refluxed for 24 h, allowed to cool, and filtered. The solvent of the filtrate was removed in vacuo. The residue was divided between Et<sub>2</sub>O and water. The organic phase was washed with water, twice with 5% aqueous NaHSO<sub>3</sub> solution, with saturated NaHCO<sub>3</sub> solution, and then with water. It was dried (Na<sub>2</sub>SO<sub>4</sub>) and the solvent was removed in vacuo. The residue was chromatographed through silica (2.4 kg) with CH<sub>2</sub>Cl<sub>2</sub>. 22 was obtained (160 g, 90% yield) as a colorless glass. Anal. (C<sub>23</sub>H<sub>31</sub>IO<sub>3</sub>Si) C, H, I.

Coupling of Phenols 8 with Iodide 22 To Give Lactol Ethers 23. Typical Procedure: 2(RS)-Methoxy-4(R)-[tert-butyldiphenylsiloxy]-6(S)-[[2,4-diisopropyl-6-(4fluorophenyl)phenoxy]methyl]tetrahydropyran (23ff). Phenol 8ff (27.2 g, 0.1 mol) was added to the suspension of K<sub>2</sub>CO<sub>3</sub> powder (27.6 g, 0.2 mol) and hydroquinone (50 mg) in DMSO (250 mL). The mixture was stirred for 45 min at 25 °C. The solution of iodide 22 (61.1 g, 0.12 mol) in DMSO (250 mL) was added, and the mixture was stirred for 4 h at 50-55 °C. TLC (cyclohexane/EtOAc; first development 9:1, second development 15:1) indicated the complete disappearance of iodide 22  $(R_f 0.51)$ , but still some unreacted phenol 8ff  $(R_i, 0.71)$ , besides the title compound 23ff  $(R_f 0.61)$ . The mixture was allowed to cool. A 20% NaCl solution (1 L) and Et<sub>2</sub>O (1 L) were added. The organic phase was separated and the aqueous phase was extracted with Et<sub>2</sub>O  $(3 \times 500 \text{ mL})$ . The combined extracts were washed with water (2 × 100 mL) and with brine and dried, and the solvent was evaporated in vacuo. The residue was chromatographed through silica (1.6 kg) first with cyclohexane/toluene 1:2, then toluene, and then toluene/EtOAc 30:1, to give 23ff (51.0 g, 78% yield) as a colorless glass.

In analogous reactions 23a-p,g (3 h, 70 °C), 23t-v,y-cc,ee (1 h, 40 °C), 23gg (8 h, 55 °C), and 23hh,oo were obtained in 66-83% yield. 23mm,nn (2 h, 50 °C) were obtained in 55-58% yield. 23dd (5 h, 40 °C, careful  $O_2$  exclusion) was obtained in 35% yield. 23pp (1 h, 50 °C) was obtained in 65% yield.

2(RS)-Hydroxy-4(R)-[tert-butyldiphenylsiloxy]-6(S)-[[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]methyl]tetra-hydropyran (24ff). A solution of 23ff (50.0 g, 76.3 mmol) in THF (4.5 L), H<sub>2</sub>O (4.5 L), and glacial acetic acid (6.3 L) was stirred for 26 h at 80-85 °C bath temperature. TLC (cyclohexane/EtOAc 9:1) indicated that only little unreacted 23ff was left. The solvents were removed in vacuo. The residue was redissolved repeatedly

in toluene (3  $\times$  2 L), and the solvent was removed in vacuo each time. The residue was chromatographed through silica (2 kg) with cyclohexane/EtOAc 12:1 to obtain 24ff (42.2 g, 86% yield) as an amorphous, colorless powder. Anal. ( $C_{40}H_{49}FO_4Si$ ) C, H, F.

In analogous reactions 24a-c (3 days), 24f,g, 24h,i (3 days), and 24k-q,t-v,y,bb-ee,gg were obtained in 60-80% yield. 24hh and 24aa (2 days, 75 °C bath) were obtained in 85 and 92% yield, respectively. 24d,e,j,z were obtained in 33-50% yield besides 31-38% of the requisite unreacted educts 23. 24mm-pp were obtained in 55-65% yield (besides 10-25% of recovered educts 23), when acetic acid was substituted by trifluoroacetic acid. A representative procedure is as follows: A solution of 2300 (3.8 g, 6.15 mmol) in THF (80 mL), H<sub>2</sub>O (40 mL), and CF<sub>3</sub>CO<sub>2</sub>H (40 mL) was stirred for 4 h at 60 °C. At 20 °C the pH was adjusted to 5.5 with 4 N NaOH (115 mL). The solvents were removed in vacuo, and the residue was extracted with ether. The extract was washed with saturated NaHCO3 solution and with brine and dried (Na<sub>2</sub>SO<sub>4</sub>). The solvent was removed in vacuo, and the residue was chromatographed through silica (200 g) with cyclohexane/ EtOAc 2:1 to give 2400 (2.2 g, 59% yield) as a colorless, amorphous

Oxidation of Lactol 24 to Lactone 25 with N-Iodosuccinimide. Typical Procedure: 4(R)-(tert-Butyldiphenylsiloxy)-6(S)-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxymethyl]tetrahydropyran-2-one (25ff). Tetra-n-butylammonium iodide (19.25 g, 52.1 mmol) was added with cooling and stirring (10 °C) to the solution of lactol 24ff (33.4 g, 52.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2.5 L). N-iodosuccinimide (46.9 g, 208.4 mmol) was added. The mixture was stirred with exclusion of light (covered with alumina foil) for 1 h at 10 °C and 20 h at ambient temperature. TLC (cyclohexane/EtOAc 9:1) indicated complete disappearance of 24ff  $(R_f 0.11)$  and the clean formation of the product (R, 0.23). The mixture was washed with water (1 L), with 5% aqueous NaHSO<sub>3</sub> solution (3  $\times$  1 L), and water (0.5 L). The organic phase was dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo. The residue was filtered through silica (2 kg) with cyclohexane/EtOAc 92:8, to obtain 25ff (33.0 g, yield 99%) as a colorless glass. Anal. (C<sub>40</sub>H<sub>47</sub>FO<sub>4</sub>Si) C, H, F.

In analogous reactions 25q,dd,ee,gg,hh,mm-pp were obtained in 82-98% yield, generally as colorless, amorphous solids.

Oxidation of Lactol 24 to Lactone 25 with CrO<sub>3</sub> on Celite. Typical Procedure: 4(R)-[tert-Butyldiphenylsiloxy]-6-(S)-[[2,4-dichloro-6-(cyclohexylidenefluoromethyl)phenoxy]methyl]tetrahydropyran-2-one (25y). Powdered chromium trioxide (8.6 g, 86 mmol) was added at 15-20 °C to a suspension of Celite (5 g) in CH<sub>2</sub>Cl<sub>2</sub> (100 mL). A solution of pyridine (13.6 g, 172 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added dropwise. The mixture was stirred for 20 min at 20-25 °C. A solution of lactol 24y (5.7 g, 8.6 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (60 mL) was added dropwise. The suspension was stirred for 1 h at 20-25 °C. TLC indicated complete disappearance of 24y and clean formation of the product. CH<sub>2</sub>Cl<sub>2</sub> (400 mL) and Celite (100 g) was added and the mixture was suction filtered. The solid was washed with CH<sub>2</sub>Cl<sub>2</sub> (100 mL). The combined filtrates were filtered through Celite again, and then the solvent was removed in vacuo. The residue was dissolved in a minimal amount of cyclohexane/EtOAc 1:1, given onto a column of silica (400 g), and flash chromatographed with cyclohexane/EtOAc 9:1. 25y (5.0 g, yield 88%) was obtained as an amorphous colorless solid. Anal. (C<sub>35</sub>H<sub>40</sub>Cl<sub>2</sub>F<sub>2</sub>SiO<sub>4</sub>) C, H, Cl.

In analogous reactions 25a-p,t-v,y-cc were obtained in 82-90%

yield, generally as colorless, amorphous solids.

Deprotection of β-Silyloxy Lactones 25 To Give β-Hydroxy Lactones 4. Typical Procedure: 4(R)-Hydroxy-6(S)-[[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]methyl]tetra-hydropyran-2-one (4ff). Glacial acetic acid (11.65 g, 194.1 mmol), then tetra-n-butylammonium fluoride trihydrate (45.92 g, 145.6 mmol), was added at 20 °C to a stirred solution of 25 (31.0 g, 48.5 mmol) in THF (1.5 L). The mixture was stirred for 4 h at ambient temperature and then allowed to stand overnight. The solvent was removed in vacuo. Et<sub>2</sub>O (1 L) and water (0.5 L) were added to the residue. The aqueous phase was extracted with Et<sub>2</sub>O (2 × 0.5 L) and the combined ethereal solutions were washed with water (0.1 L) and brine, dried (MgSO<sub>4</sub>), and evaporated in vacuo. The residue was dissolved in toluene and the solvent was evaporated in vacuo. The residue was chromatographed through silica

(2 kg) with cyclohexane/EtOAc 1:1 to give 4ff (16.2 g, 83% yield) as colorless crystals (mp 145–146 °C). Anal. ( $\rm C_{24}H_{29}FO_4$ ) C, H, F

For yields of other compounds of general formula 4, obtained by analogous reactions, see Table I; for spectra, see Table X of the supplementary material. 4r and 4s were prepared according to another procedure (vide infra).

Saponification of  $\beta$ -Hydroxy Lactones 4 To Give  $\beta$ , $\delta$ -Dihydroxy Sodium Carboxylates 11. Typical Procedure: Sodium 3(R),5(S)-Dihydroxy-6-[2,4-diisopropyl-6-(4-fluorophenyl)phenoxy]hexanoate (11ff). Aqueous NaOH (1 N, 18.4 mL, 18.4 mmol) was added dropwise at 10 °C to a solution of 4ff (7.0 g, 17.5 mmol) in EtOH (800 mL). The solution was stirred 1 h at 20–25 °C. TLC (CHCl<sub>3</sub>/CH<sub>3</sub>OH 8:2) indicated complete disappearance of 11ff and a homogeneous product ( $R_f$  0.26). The solvent was removed in vacuo. The residue was dissolved in ethanol and the solvent was evaporated in vacuo. The residue was stirred for 10 min in n-pentane (50 mL) and then suction filtered, and the washing was repeated. The solid was dried in vacuo to obtain 11ff (7.2 g, 94% yield) as a colorless amorphous powder (mp 243–245 °C dec). Anal. ( $C_{24}H_{30}FO_5$ Na) C, H, F. HPLC (vide supra) indicated a purity of >99.5%.

For other compounds of general formula 11, see Table I; for spectra, see Table X (supplementary material).

Preparation of o-Benzoylphenols 27 by Fries Rearrangement. Typical Procedure: 1-[(4-Fluorobenzoyl)-oxy]-2,4-dichlorobenzene (26b). 2,4-Dichlorophenol (350 g, 2.16 mol) was added dropwise at 20 °C to the solution of NaOH (109.2 g, 2.72 mol) in  $\rm H_2O$  (0.94 L). The mixture was stirred for 30 min. At 15–25 °C (cooling) 4-fluorobenzoyl chloride (500 g, 3.16 mol) was added dropwise. The mixture was stirred for 30 min at ambient temperature. TLC indicated quantitative disappearance of starting materials. The mixture was diluted with  $\rm H_2O$  (2 L). The solid was filtered and washed with cold 10% aqueous NaOH solution (2 L) and then with  $\rm H_2O$  (2 × 2 L). The solid was suspended in EtOH (2 L), stirred for 30 min, suction-filtered, washed with EtOH, and dried in vacuo at 50 °C to give a colorless solid (608 g, 99% yield, mp 128–129 °C). Anal. ( $\rm C_{13}H_7Cl_2FO_2$ ) C, H, Cl, F.

2,4-Dichloro-6-(4-fluorobenzoyl)phenol (27b). Ester 26b (608 g, 2.14 mol) and AlCl<sub>3</sub> (712 g, 5.35 mol) were well mixed and slowly heated to 150 °C (inner temperature). The mixture was stirred for 2 h. TLC (CHCl<sub>3</sub>) indicated disappearance of 26b. The mixture was allowed to stand at 25 °C overnight. It was dissolved in EtOAc (4 L) with warming and then hydrolyzed at <30 °C (external cooling) with 2 N HCl (3 L). The organic phase was washed with 2 N HCl, with H<sub>2</sub>O, and then with brine. It was dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo. The residue was suspended in i-Pr<sub>2</sub>O (1 L) and stirred for 30 min. The solid was suction filtered, washed with i-Pr<sub>2</sub>O, and dried in vacuo to give a slightly yellow solid (494 g, 81% yield, mp 120–123 °C). Anal. (C<sub>13</sub>H<sub>7</sub>Cl<sub>2</sub>FO<sub>2</sub>) C, H, Cl, F.

Preparation of 28 by Grignard Additions to o-Benzoylphenols 27. Typical Procedure: 2,4-Dichloro-6-[bis(4fluorophenyl) hydroxymethyl] phenol (28b). (4-Fluorophenyl)magnesium bromide was prepared from Mg turnings (66.5 g, 2.77 mol) and 4-bromo-1-fluorobenzene (441 g, 2.52 mol) in THF (1.2 L). A solution of ketone 27b (240 g, 0.845 mol) in THF (1 L) was added dropwise. The solvent was removed in vacuo. The residue was suspended in Et<sub>2</sub>O (2.5 L) and 20% aqueous NH<sub>4</sub>Cl solution (3 L) was added dropwise with cooling. The organic phase was separated, and the aqueous phase was extracted with CH2Cl2. The combined organic phases were washed with brine and then dried, and the solvents were evaporated in vacuo. The residue was suspended in i-Pr<sub>2</sub>O (1 L) and stirred for 15 min. The solid was suction filtered, washed with i-Pr<sub>2</sub>O, and dried in vacuo to give slightly yellow crystals (273 g, 85% yield, mp 193–195 °C). Anal.  $(C_{19}H_{12}Cl_2F_2O_2)$  C, H, Cl, F.

Preparation of 8a-i by Hydrogenation of Benzylic Alcohols 28a-i. Typical Procedure: 2,4-Dichloro-6-[bis(4-fluorophenyl)methyl]phenol (8b). Benzylic alcohol 28b (240 g, 0.632 mol) was dissolved in glacial acetic acid (4.8 L) and concentrated HCl (48 mL) with slight warming. After cooling to 20 °C, 10% Pd on charcoal (4 g) was added. The suspension was deoxygenated with N<sub>2</sub>, then saturated with H<sub>2</sub>, and hydrogenated at ambient temperature in a shaking device under 1 bar

of  $H_2$ . A 13.3-L portion of  $H_2$  was consumed within 90 min. The catalyst was filtered off and washed with ethanol. The filtrates were concentrated in vacuo. The residue was dissolved in EtOAc (12 L) and washed with water, saturated NaHCO<sub>3</sub> solution, and brine. The organic phase was dried and the solvent was removed in vacuo to give an oil that was purified by chromatography through silica (6.4 kg) with cyclohexane/CH<sub>2</sub>Cl<sub>2</sub> 4:1 to give 170 g [74% yield, purity > 99.5% (GLC)]. Anal. ( $C_{19}H_{12}Cl_2F_2O$ ) C, H, Cl, F.

Data for 8a-i may be obtained from Table XI of the supplementary material.

Preparation of Phenols 8j–o. Typical Procedure: 1-(4-Fluorophenyl)prop-2-en-1-ol. (4-Fluorophenyl)magnesium bromide was prepared from Mg turnings (13.3 g, 0.55 mol) and 4-bromo-1-fluorobenzene (87.5 g, 0.5 mol) in Et<sub>2</sub>O (650 mL). The Grignard solution was cooled to -10 °C and the solution of acrolein (28 g, 0.5 mol) in Et<sub>2</sub>O (100 mL) was added dropwise. The mixture was stirred for 1 h at -10 °C, and 20% aqueous NH<sub>4</sub>Cl solution (0.5 L) was added dropwise. The organic phase was separated and the aqueous phase was extracted with Et<sub>2</sub>O. The combined extracts were washed with water and dried, and the solvent was evaporated in vacuo. The residue was chromatographed through silica (1 kg) with CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (51 g, 67% yield).

1-(4-Fluorophenyl)-3-bromoprop-1-ene. A solution of PBr<sub>3</sub> (41.7 g, 154 mmol) in toluene (150 mL) was added dropwise at 0-5 °C to a solution of 1-(4-fluorophenyl)prop-2-en-1-ol (47 g, 309 mmol) in toluene (1.2 L). The mixture was washed twice with saturated NaHCO<sub>3</sub> solution and twice with brine. The organic solution was dried and the solvent was evaporated in vacuo. The residue was used immediately for the next step.

1-(4-Fluorophenyl)-3-(2,4-dichlorophenoxy)prop-1-ene (29j). 2,4-Dichlorophenol (50 g, 309 mmol) was added to crude 1-(4-fluorophenyl)-3-bromoprop-1-ene (309 mmol), followed by acetone (1 L) and  $\rm K_2CO_3$  (127 g, 927 mmol). The suspension was refluxed for 4 h. The solvent was removed in vacuo. The residue was dissolved in Et<sub>2</sub>O, washed with H<sub>2</sub>O, and dried, and the solvent was evaporated. The residue was recrystallized from hot petroleum ether, to give a colorless solid (72 g, 79% yield, mp 80-81 °C). Anal. ( $\rm C_{15}H_{11}Cl_2FO$ ) C, H, Cl, F.

2,4-Dichloro-6-[3-(4-fluorophenyl)prop-1-en-3-yl]phenol (30j). N,N-Dimethylaniline (38.1 g, 315 mmol) was added to a solution of 29j (52.3 g, 210 mmol) in diethylene glycol monomethyl ether (600 mL). The mixture was heated for 10 h to 160 °C. It was allowed to cool. Et<sub>2</sub>O (2 L) and petroleum ether (4 L) were added, and the solution was washed twice with 2 N HCl and twice with brine. It was dried, and the solvents were removed in vacuo. The residue was chromatographed through silica (1 kg) with cyclohexane/toluene 1:1 to give an oil (52.4 g, 84% yield).

1-(Benzyloxy)-2,4-dichloro-6-[3-(4-fluorophenyl) prop-1-en-3-yl]benzene (31j).  $K_2CO_3$  (48.5 g, 351 mmol) and benzyl chloride (24.3 g, 192 mmol) were added to the solution of phenol 30j (52.0 g, 175 mmol) in DMF (0.5 L). The suspension was heated for 8 h to 70-75 °C. Water (2 L) and  $Et_2O$  (2 L) were added to the cold suspension. The ether phase was washed twice with 2 N NaOH solution and twice with brine. It was dried, and the solvent was evaporated. An analytical sample was obtained by chromatography (cyclohexane/EtOAc 50:1) of a small portion. The crude product (69.7 g, 100% yield) did not exhibit any impurity on TLC and was used in the next step.

1-(Benzyloxy)-2,4-dichloro-6-[1-(4-fluorophenyl)-3-hydroxypropyl]benzene (32j). A solution of 31j (69.0 g, 179 mmol) was added dropwise to a 0.5 M solution of 9-BBN in THF (525 mL, 262 mmol). The solution was stirred for 30 min at ambient temperature and 90 min under reflux. It was cooled to 5 °C, and EtOH (105 mL), followed by 2 N aqueous NaOH (173 mL), was added dropwise. The mixture was cooled, and 30% aqueous  $H_2O_2$  (45.5 mL) was added dropwise at <30 °C. The mixture was refluxed for 1 h. Toluene (2 L) was added, and the solution was washed with ice/water (3 × 0.5 L) and with brine (0.5 L). The organic phase was dried; the solvents were evaporated in vacuo. The residue was chromatographed through silica (2 kg) with cyclohexane/EtOAc 85:15 to give a colorless glass (56.7 g, 79% yield).

1-(Benzyloxy)-2,4-dichloro-6-[1-(4-fluorophenyl)-3-(to-syloxy)-1-propyl]benzene (33j). p-Toluenesulfonyl chloride

(38.2 g, 200 mmol) was added in portions at 0–5 °C to the solution of 32j (56.7 g, 140 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (150 mL) and pyridine (150 mL). The mixture was stirred for 6 h at 20 °C and allowed to stand at –10 °C overnight. The solvents were evaporated in vacuo. The residue was dissolved in toluene (0.5 L) and washed with H<sub>2</sub>O (2 × 200 mL), saturated NaHCO<sub>3</sub> solution (2 × 200 mL), and brine. The organic phase was dried, and the solvent was evaporated in vacuo. The residue was filtered through silica (1 kg) with CH<sub>2</sub>Cl<sub>2</sub> to give a pale pink oil (71.3 g, 91% yield).

1-(Benzyloxy)-2,4-dichloro-6-[1-(4-fluorophenyl)-3-iodo-1-propyl]benzene (34j). NaI (47.9 g, 319 mmol) was added to the solution of tosylate 33j (71.3 g, 128 mmol) in acetone (1 L). The mixture was refluxed for 4 h. The solvent was removed in vacuo. The residue was dissolved in toluene. The solution was washed twice with water, twice with NaHSO<sub>3</sub> solution, once with NaHCO<sub>3</sub> solution, and then with brine. The organic phase was dried, and the solvent was removed in vacuo. The residue was chromatographed through silica (1 kg) with cyclohexane/EtOAc 9:1 to give an oil (45.6 g, 70% yield).

1-(Benzyloxy)-2,4-dichloro-6-[1-(4-fluorophenyl)-3-(4-fluorophenoxy)-1-propyl]benzene.  $K_2CO_3$  (12.1 g, 87.7 mmol) was added to the solution of 4-fluorophenol (4.9 g, 43.8 mmol) in DMSO (30 mL). A solution of iodide 34j (15.0 g, 29.2 mmol) in DMSO (80 mL) was added at once. The mixture was heated for 4 h to 50 °C. It was cooled, and  $Et_2O$  (0.5 L) and water (0.5 L) were added. The water phase was separated and extracted with  $Et_2O$  again. The combined extracts were washed with  $H_2O$  (2 × 100 mL) and then dried (MgSO<sub>4</sub>), and the solvent was evaporated in vacuo. The residue was chromatographed through silica to give a colorless solid (11.1 g, 76% yield).

2,4-Dichloro-6-[1-(4-fluorophenyl)-3-(4-fluorophenoxy)-1-propyl]phenol (8j).  $N_2$  was bubbled through the solution of the benzyl ether (5.5 g, 11 mmol) in glacial acetic acid (560 mL) and concentrated HCl (5.5 mL). Pd (10%) on charcoal (1.5 g) was added. The suspension was saturated with  $H_2$  and shaken under 1 bar of  $H_2$  at ambient temperature. A 250-mL portion of  $H_2$  was consumed within 20 min. TLC (cyclohexane/EtOAc 9:1) indicated complete transformation of starting material ( $R_f$  0.50) to product ( $R_f$  0.29). The mixture was purged with  $N_2$ . The catalyst was filtered off and washed with ethanol. The filtrates were concentrated in vacuo. Toluene was added, and the solvent was evaporated in vacuo. The residue was chromatographed through silica (450 g) with cyclohexane/EtOAc 92:8 to give a colorless solid (4.5 g, 99% yield). For data see Table XI of the supplementary material.

2-Methyl-6-[3-(4-fluorophenoxy)-1-propyl]phenol (81). A solution of chlorophenol 8k (240 mg, 0.82 mmol) in EtOAc (40 mL) was purged with N<sub>2</sub>. Pd (10%) on charcoal (40 mg) was added, and the mixture was shaken for 12 h under 1 bar of H<sub>2</sub>. A 20-mL portion of H<sub>2</sub> was consumed during this time. The catalyst was filtered off. The filtrate was concentrated in vacuo. The residue was dissolved in toluene, and the solvent was evaporated in vacuo. The residue was chromatographed through silica with cyclohexane/EtOAc 92:8 to give a colorless solid (153.5 mg, 72% yield).

In analogy, 80 was obtained from chlorophenol 8n in 94% yield. 2,4-Dimethyl-6-(cyclohexylcarbonyl)phenol (8ss). A solution of cyclohexanecarbonyl chloride (55 g, 375 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (25 mL) was added dropwise at 20 °C to 2,4-dimethylphenol (35a; 41.5 g, 340 mmol). The mixture was stirred for 1 h at 20 °C. Then it was warmed to 60 °C, while the CH<sub>2</sub>Cl<sub>2</sub> distilled off. The residue was stirred for 30 min at 60 °C. TLC (100% toluene) indicated complete transformation of 35a  $(R_f\ 0.14)$  to 1-[(cyclohexylcarbonyl)oxy]-2,4-dimethylbenzene  $(R_f\ 0.41)$ . It was cooled to 10 °C. AlCl<sub>3</sub> (49 g, 367 mmol) was added. The mixture was heated for 1 h to 150 °C. TLC indicated complete transformation to 8ss  $(R_f 0.53)$ . The mixture was cooled and then dissolved in EtOAc (500 mL). Ice (200 g) and 4 N HCl (200 mL) was added. The organic phase was washed with 2 N HCl and saturated KHCO<sub>3</sub> solution and dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo. The residue was filtered through silica (700 g) with toluene/cyclohexane 1:1 to obtain a colorless oil (70.9 g) that was distilled in a Kugelrohr apparatus (110 °C/7  $\times$  10<sup>-5</sup> bar) to give a colorless oil (69.9 g, 89% yield). Anal.  $(C_{15}H_{20}O_2)$  C, H.

2,4-Dichloro-6-(cyclohexylcarbonyl)phenol (8p) [4 h, 150 °C, mp 103 °C (from CH<sub>3</sub>OH), 32% yield] and 2-Methyl-4-

chloro-6-(cyclohexylcarbonyl)phenol (8tt) (4 h, 150 °C, 69% yield) were obtained by analogous procedures.

2,4-Dimethyl-6-(cyclohexylhydroxymethyl)phenol (8q). A solution of NaBH<sub>4</sub> (3.3 g, 87.2 mmol) in water (15 mL) was added at 0 °C to the solution of ketone 8ss (20.0 g, 86.2 mmol) in methanol (200 mL). The mixture was stirred for 30 min at 20 °C. TLC (cyclohexane/EtOAc 4:1) indicated complete transformation of 8ss into alcohol 8q. The solvents were removed in vacuo. Water (15 mL) was added dropwise. The pH was adjusted to 4.0 with aqueous NaHSO<sub>4</sub> solution. The mixture was extracted four times with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), and the solvents were removed in vacuo. The residue was filtered through silica with toluene/EtOAc 20:1. The eluent was removed in vacuo and the residue was recrystallized from n-hexane to obtain pale-yellow crystals (17.7 g, 88% yield, mp 116 °C). Anal. (C<sub>15</sub>H<sub>22</sub>O<sub>2</sub>) C, H.

4(R)-(tert-Butyldiphenylsiloxy)-6(S)-[[2,4-dimethyl-6-(cyclohexylhydroxymethyl)phenoxy]methyl]tetrahydropyran-2-one (25q). 23q (74% yield) and 24q (76% yield) were obtained according to the typical procedures given above. Lactone 25q (82% yield) was obtained, utilizing N-iodosuccinimide, according to the typical procedure. The two diastereomers of 25q (stereochemistry of hydroxy group) can be separated by silica chromatography (toluene/EtOAc 95:5) under flash chromatography or, better yet, HPLC conditions. The  $R_f$  value of the R diastereomer is small than that of the S diastereomer.

4(R)-(tert-Butyldiphenylsiloxy)-6(S)-[[2,4-dimethyl-6-[cyclohexyl[( $\alpha$ , $\alpha$ -dimethylbutanoyl)oxy]methyl]phenoxy]methyl]tetrahydropyran-2-one (25r) and (25s). A solution of 25q (S isomer; 350 mg, 0.58 mmol),  $\alpha$ , $\alpha$ -dimethylbutanoic anhydride (310 mg, 1.45 mmol), and 4-(dimethylamino)pyridine (180 mg, 1.47 mmol) in toluene (10 mL) was heated for 48 h to 110 °C. The solvent was evaporated in vacuo. Further volatile components were removed (30 min) in a Kugelrohr distillation apparatus (60 °C/7 ×  $10^{-6}$  bar). The residue was purified by silica chromatography to give 25r (260 mg, 65% yield) as a colorless glass.

In analogy, 25q (R isomer, 293 mg) gave 25s (236 mg, 69% yield) as a colorless glass.

4(R)-Hydroxy-6(S)-[[2,4-dimethyl-6-[cyclohexyl[( $\alpha,\alpha$ -dimethylbutanoyl)oxy]methyl]phenoxy]methyl]tetrahydropyran-2-one (4r) and (4s). A solution of 25r (250 mg, 0.36 mmol),  $\alpha,\alpha$ -dimethylbutanoic acid (90 mg, 0.77 mmol), and tetra-n-butylammonium fluoride trihydrate (600 mg, 1.90 mmol) in THF (20 mL) was kept at 0 °C for 16 h. Toluene (50 mL) was added, and the solvents were removed in vacuo. The residue was dissolved in a small amount of CH<sub>2</sub>Cl<sub>2</sub> and chromatographed through silica gel with cyclohexane/EtOAc 2:1. Toluene was added to the pure product, and the solvent was evaporated in vacuo. The residue was dried in vacuo to obtain 4r (145 mg, 90% yield) as a colorless glass. Anal. ( $C_{27}$ H<sub>40</sub>O<sub>6</sub>) C, H.

In analogy, from 25s (230 mg) there was obtained 4s (140 mg). For spectra of 4r and 4s, see Table X of the supplementary material.

Preparation of Phenols 8t-v by Reduction of Phenolic Ketones. Typical Procedure: 2,4-Dichloro-6-(cyclohexylmethyl)phenol (8u). A solution of NaBH<sub>4</sub> (5.8 g, 153 mmol) in ice/water (50 mL) was added dropwise at 0 °C to the solution of ketone 8p (55.7 g 204 mmol) in CH<sub>3</sub>OH (1 L). The mixture was stirred for 1 h at 20 °C. Solvents were removed in vacuo. Et<sub>2</sub>O (100 mL) was added. 2 N H<sub>2</sub>SO<sub>4</sub> was added at 10 °C until the pH remained acidic. The organic phase was separated, and the aqueous phase was extracted twice with ether. The combined organic phases were dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo to leave a semisolid [55.2 g, crude 2,4-dichloro-6-cyclohexylhydroxymethyl)phenol (8uu)].

A solution of  $AlCl_3$  (189 g, 1.42 mol) in  $Et_2O$  (270 mL) was added at 0 °C to the suspension of  $LiAlH_4$  (26.8 g, 0.71 mol) in  $Et_2O$  (270 mL). A solution of crude alcohol 8uu (55.2 g, 0.20 mol) in  $Et_2O$  (170 mL) was added dropwise. The mixture was purged with  $N_2$ , and the reaction temperature was raised to 80 °C, while the  $Et_2O$  was distilling off. After 15 min at 80 °C gas evolution started and ceased after 30 min. The mixture was stirred for 15 additional minutes at 80 °C. It was cooled and  $Et_2O$  (500 mL) was added. At 0 °C 1 N HCl (500 mL) was added dropwise (caution! highly exothermic,  $H_2$  evolution). The organic phase

was separated and the aqueous phase was extracted with Et<sub>2</sub>O. The combined extracts were washed with 2 N HCl and with brine. They were dried, and the solvent was evaporated in vacuo. The residue was chromatographed through silica first with cyclohexane/toluene 1:1, then with toluene, and then with toluene/EtOAc 9:1 to give 8u (40.7 g, 77% yield) as a glass and unreacted alcohol 8uu (6.5 g, 12%) as a solid (mp 86 °C from n-pentane). 8u was distilled in a Kugelrohr apparatus (100 °C/7 × 10<sup>-4</sup> bar) to get a colorless solid (40.0 g, mp 45 °C). Anal. ( $C_{13}H_{16}Cl_2O$ ) C, H, Cl.

8t and 8y were obtained in 65 and 48% yield, respectively, when the ketones were subjected immediately to the LiAlH $_4$ /AlCl $_3$  reduction, the prior reduction of ketone to alcohol being unnecessary. This shortened procedure with 8p gave alcohol 8uu and less than 10% of 8u.

4(R)-(tert-Butyldiphenylsiloxy)-6(S)-[[2,4-dimethyl-6-[cyclohexyl(1,1-dimethylpropanamido)methyl]phenoxy]-methyl]tetrahydropyran-2-one (25w and 25x). A solution of alcohol 25q (1:1 mixture of diastereomers, 1.06 g, 1.77 mmol) in  $\alpha,\alpha$ -dimethylbutyronitrile (10 mL) and 48% aqueous HF (1 mL) was stirred for 50 h at 25 °C. TLC (cyclohexane/EtOAc 4:1) indicated clean transformation of 25q ( $R_f$  0.18) to 25w ( $R_f$  0.32) and 25x ( $R_f$  0.25). CH<sub>2</sub>Cl<sub>2</sub> (40 mL) and aqueous KHCO<sub>3</sub> solution (10 mL) were added. The organic phase was separated and dried (MgSO<sub>4</sub>), and the solvents and excess reagent (bp 128–129 °C) were removed in vacuo. HPLC (silica, cyclohexane/EtOAc 20:1) gave 25w (546 mg) and 25x (490 mg) as colorless solids.

 $\beta$ -Silyloxy lactones 25w and 25x were deprotected to give 4w and 4x, respectively, according to the typical procedure and Table 1

2,4-Dichloro-6-(cyclohexylidenefluoromethyl)phenol (8y). DAST (9.66 g, 7.9 mL, 60 mmol) was added dropwise with a syringe at 0-5 °C to a solution of ketone 8p (10.8 g, 40 mmol) in toluene (60 mL). The mixture was stirred for 2 h at 25 °C. It was added dropwise at 0 °C into a stirred two-phase mixture of toluene (200 mL) and saturated Na<sub>2</sub>CO<sub>3</sub> solution (200 mL). The mixture was stirred for 20 min. The organic phase was separated, and the aqueous layer was extracted with toluene again. The combined extracts were washed with brine and dried (MgSO<sub>4</sub>), and the solvent was evaporated. The residue was chromatographed through silica (500 g) with cyclohexane/toluene 3:1 to give 8y (5.1 g, yield 47%) and recovered 8p (2.8 g, 26%). 8y: Anal. (C<sub>13</sub>H<sub>13</sub>Cl<sub>2</sub>FO) C, H, Cl.

2,4-Dimethyl-6-[cyclohexylidene(4-fluorophenyl)-methyl]phenol (36). To a Grignard solution, prepared from cyclohexyl chloride (7.15 g, 60.3 mmol) and Mg turnings (1.6 g, 65.8 mmol) in Et<sub>2</sub>O (100 mL) (20-min reflux) was added dropwise a solution of 2,4-dimethyl-6-[(4-fluorophenyl)carbonyl]phenol (4.9 g, 20.1 mmol) in Et<sub>2</sub>O (50 mL). The mixture was stirred for 3 h at 20 °C. HCl (2 N, 40 mL) was added dropwise. The aqueous phase was extracted with Et<sub>2</sub>O (2 × 100 mL). The combined Et<sub>2</sub>O layers were washed with brine and dried, and the solvent was evaporated. The residue was dissolved in toluene (50 mL). p-Toluenesulfonic acid (100 mg) was added, and the mixture was warmed for 2 h to 100 °C. The main product was purified by silica chromatography (cyclohexane) and then Kugelrohr distilled (~130 °C/7 ×  $10^{-5}$  bar) to give a solid (4.4 g, 70% yield).

2,4-Dimethyl-6-[cyclohexyl(4-fluorophenyl)methyl]phenol (8z). A solution of 36 (1.8 g) in CH<sub>3</sub>OH, containing 10% Pd/C (0.5 g) was shaken for 5 h under 1 bar of H<sub>2</sub>. Standard workup gave an oil that was purified by Kugelrohr distillation ( $\sim$ 150 °C/1 × 10<sup>-4</sup> bar) to give 1.6 g (89% yield) of an oil that slowly crystallized. Anal. (C<sub>21</sub>H<sub>26</sub>FO) C, H, F.

3-Isopropyl-4-nitrobiphenyl (38). A 0.7 M solution of isopropylmagnesium chloride in THF (343 mL, 240 mmol) was added at -70 °C within 3 h to the solution of 4-nitrobiphenyl (37; 20 g, 100 mmol) in THF (400 mL). The mixture was stirred for 1 h at -70 °C. 2,3-Dichloro-5,6-dicyano-p-benzoquinone (DDQ; 22.7 g, 100 mmol) was added at once at -40 °C. The mixture was allowed to warm to 20 °C and was stirred for 1 h at this temperature. The reaction mixture was poured into  $H_2O$  (1.2 L) and allowed to stand overnight. The organic solvent was removed in vacuo. The aqueous phase was extracted with EtOAc (3 × 0.5 L). The combined extracts were washed with brine and dried (MgSO<sub>4</sub>), and the solvent was evaporated. The residue (21 g) was chromatographed through silica with cyclohexane/CH<sub>2</sub>Cl<sub>2</sub> 4:1 to

give a pale-red oil (7.9 g, 33% yield).

3-(Cyclohexylmethyl)-5-isopropyl-4-nitrobiphenyl (39). A Grignard solution was prepared from cyclohexylmethyl bromide (4.1 g, 23.2 mmol), Mg turnings (564 mg, 23.2 mmol), and THF (30 mL). This solution was added dropwise at -70 °C within 45 min to a solution of 38 (2.8 g, 11.6 mmol) in THF (30 mL). At -40 °C DDQ (5.27 g, 23.2 mmol) was added at once. The mixture was stirred for 90 min at 20 °C. Toluene and  $\rm H_2O$  was added. After 20 min the aqueous phase was separated and washed with toluene. The combined toluene layers were washed with  $\rm H_2O$  and dried, and the solvent was evaporated in vacuo. The residue (6.2 g) was chromatographed through silica with cyclohexane/toluene 9:1 to give a pale-red oil (2.0 g, 51% yield).

3-(Cyclohexylmethyl)-5-isopropyl-4-aminobiphenyl (40). Nitro compound 39 (1.95 g, 5.78 mmol) was dissolved in a methanolic solution (150 mL) of ammonia (20 g/L). Raney Ni (3 g) was added, and the mixture was shaken for 2 h at 20 °C in a  $H_2$  atmosphere. The catalyst was filtered off and washed with  $CH_3OH$ . The filtrate was evaporated in vacuo. The residue was chromatographed (cyclohexane/toluene 1:2) to give a colorless oil (1.62 g, 91% yield).

2-(Cyclohexylmethyl)-4-phenyl-6-isopropylphenol (8aa). A suspension of amine 40 (1.6 g, 5.2 mmol) in hot 20% aqueous hydrobromic acid (10 mL) was quickly cooled to 0 °C with extensive stirring. The solution of NaNO<sub>2</sub> (450 mg, 6.5 mmol) in  $H_2O$  (1.5 mL) was added dropwise. The mixture was stirred for 15 min at 0 °C. Urea (90 mg, 1.5 mmol) was added, and the mixture was stirred for 10 min at 0 °C and then heated to 100 °C, until the  $N_2$  evolution ceased (1 h). Et<sub>2</sub>O (50 mL) and brine (20 mL) were added. The aqueous phase was extracted with Et<sub>2</sub>O again. The combined ethereal phases were washed with saturated NaHCO<sub>3</sub> solution and then with brine and dried, and the solvent was evaporated. The residue was filtered through silica with cyclohexane/toluene 7:3 to give 8aa (810 mg, yield 50%) as a colorless oil that slowly crystallized. Anal. ( $C_{22}H_{28}O$ ) C, H.

3,5-Dichloro-2-hydroxybiphenyl (8bb). Chlorine was bubbled for 3 h at 20 °C into a solution of 2-hydroxybiphenyl (41, 17.0 g, 0.1 mol) in glacial acetic acid (50 mL). TLC (toluene/cyclohexane 7:3) indicated quantitative conversion of the starting material ( $R_f$  0.18) to the product 8bb ( $R_f$  0.42) and a minor byproduct ( $R_f$  0.53). The reaction mixture was poured onto ice (500 g) and extracted with Et<sub>2</sub>O (3 × 100 mL). The combined extracts were dried, and the solvent was evaporated in vacuo. The residue was dissolved in toluene (2 × 100 mL) and the solvent was evaporated in vacuo each time. The residue was chromatographed through silica (1.8 kg) with cyclohexane/toluene 1:1 (5 L) to obtain pure 8bb (21.57 g, 90% yield). Anal. ( $C_{12}H_8Cl_2O$ ) C, H, Cl.

2,4-Dichloro-5-[(4-fluorobenzyl)oxy]phenol (8cc). A solution of 4,6-dichlororesorcine (42; 17.9 g, 0.1 mol), NaOH pellets (4.0 g, 0.1 mol), and 4-fluorobenzyl bromide (18.9, 0.1 mol) in DMF (50 mL) was heated for 4 h to 100 °C. The mixture was cooled, and  $H_2O$  (150 mL) was added. The mixture was acidified and then extracted with  $E_2O$  (2 × 100 mL). The extracts were washed with water and with brine and dried (MgSO<sub>4</sub>), and the solvent was removed in vacuo. The residue was recrystallized with charcoal from cyclohexane/i-Pr<sub>2</sub>O to give a colorless solid (8.3) that after Kugelrohr distillation (190 °C/7 × 10-6 bar) had mp 105 °C. This solid was 1,3-bis[(4-fluorobenzyl)oxy]-4,6-dichlorophenol. The mother liquor was chromatographed through silica with toluene to give 8cc (4.8 g, 17% yield) as a colorless solid that after recrystallization from hot cyclohexane had mp 75 °C. Anal. ( $C_{13}H_9Cl_2FO_2$ ) C, H, Cl.

 $\gamma$ -[Bis(4-fluorophenyl) hydroxymethyl]- $\beta$ -naphthol (44). A Grignard solution was prepared from 4-bromofluorobenzene (140 g, 0.8 mol) and Mg turnings (21.1 g, 0.88 mol) in THF (600 mL) (60 °C, 90 min). The solution of 2-hydroxy-3-naphthoic acid methyl ester (43; 40.4 g, 0.2 mol) in THF (250 mL) was added dropwise to the Grignard solution. The mixture was stirred for 30 min at ambient temperature. The solvent was evaporated in vacuo. Et<sub>2</sub>O (1 L) was added to the residue, followed by 20% aqueous NH<sub>4</sub>Cl solution (0.5 L). The aqueous phase was extracted with ether (0.5 L). The combined ethereal phases were washed with water and with brine, dried, and evaporated. The residual oil was dissolved at 25 °C in i-Pr<sub>2</sub>O. The same volume of n-pentane was added. The precipitate was suction filtered and washed with n-pentane to give 44 (45 g, mp 173-175 °C). Ad-

ditional 44 (5.5 g, mp 171-173 °C) was obtained from the mother liquor (total yield 50.5 g, 70%).

 $\gamma$ -[Bis(4-fluorophenyl)methyl]- $\beta$ -naphthol (45). A solution of 44 (50 g, 138 mmol) in AcOH (1.2 L) and concentrated HCl (12 mL) was purged with N<sub>2</sub>. Pd (10%) on charcoal (4 g) was added and the mixture was shaken under 1 bar of H<sub>2</sub> at 25 °C until H<sub>2</sub> uptake ceased (3.6 L). The catalyst was filtered off and the filtrate was evaporated in vacuo. The residue was dissolved in toluene (0.5 L) and washed with H<sub>2</sub>O (2 × 0.25 L), with saturated NaHCO<sub>3</sub> solution (0.25 L), and with brine. The organic solution was dried and evaporated in vacuo. The residue was chromatographed through silica (2 kg) with cyclohexane/CH<sub>2</sub>Cl<sub>2</sub> 1:1 to give a very thick oil (39.3 g, yield 82%) that slowly crystallized.

α-Formyl-γ-[bis(4-fluorophenyl)methyl]-β-naphthol (46). A solution of NaOH (12.8 g, 320 mmol) in  $H_2O$  (27 mL) was added dropwise to a solution of 45 (8.0 g, 23.1 mmol) in EtOH (70 mL). The mixture was heated to 70–80 °C and CHCl<sub>3</sub> (8.8 g, 74 mmol) was added dropwise within 80 min. The mixture was stirred for 1 h at 80 °C. The solvent was removed in vacuo.  $H_2O$  (100 mL) was added, hydrochloric acid was added to pH 6, and the mixture was extracted with Et<sub>2</sub>O (3 × 100 mL). The extracts were washed with water, dried, and evaporated in vacuo. The residue was chromatographed through silica (0.5 kg) with cyclohexane/EtOAc 93:7 to give a colorless solid (2.9 g, 34% yield), mp 137–139 °C) and recovered 45 (1.4 g).

2-Isopropyl-4-[bis(4-fluorophenyl)methyl]- $\beta$ -naphthol (8dd). A solution of 45 (4.26 g, 12.3 mmol) in CH<sub>3</sub>OH (15 mL) was added to a solution of Na (282 mg, 12.3 mmol) in CH<sub>3</sub>OH (15 mL). The solvents were evaporated. Toluene (3 × 20 mL) was added to the residue and evaporated to dryness each time. The residue was dried for 1 h in vacuo. The solid was suspended in degassed toluene (50 mL) and heated to reflux with careful O<sub>2</sub> exclusion (argon). 2-Bromopropane (6.2 g, 50.4 mmol) was added dropwise within 30 min. The mixture was refluxed for 24 h. The cold mixture was acidified with 2 N HCl. H<sub>2</sub>O (50 mL) was added. The organic layer was washed with brine and dried, and the solvent was evaporated in vacuo. All workup steps were performed quickly, minimizing O<sub>2</sub> contact. The residue was chromatographed through silica (300 g) with cyclohexane/EtOAc 93:7 to give 8dd (3.7 g, 77% yield). Anal. (C<sub>26</sub>H<sub>22</sub>F<sub>2</sub>O) C, H, F.

2-Methyl-4-[bis(4-fluorophenyl)methyl]- $\beta$ -naphthol (8ee). A 70% (3.5 M) solution of sodium bis(2-methoxyethoxy)dihydroaluminate (Red-Al) in toluene (6.95 mL, 24.3 mmol) was diluted with xylene (20 mL). This solution was added at 120 °C to the solution of aldehyde 46 (2.6 g, 6.95 mmol) in xylene (100 mL), while the toluene was distilling off. When the reaction temperature reached 135 °C, the distillation head was replaced by a reflux condenser. After 2 h at reflux, the mixture was cooled, diluted with Et<sub>2</sub>O (300 mL), and acidified with 20% aqueous  $H_2$ SO<sub>4</sub>. The aqueous layer was extracted twice with Et<sub>2</sub>O. The combined organic layers were washed twice with saturated NaHCO<sub>3</sub> solution and then with brine. They were dried, and the solvent was evaporated in vacuo. The residue was chromatographed through silica (100 g) with cyclohexane/toluene 1:1 to give 8ee (1.3 g, yield 52%). Anal. (C<sub>24</sub>H<sub>18</sub>F<sub>2</sub>O) C, H, F.

4-Acetoxy-3-(p-fluorophenyl)-2,5,6-rriisopropylphenol (8kk). A solution of triethylamine (1.3 mL) and 4-(dimethylamino)pyridine (60 mg) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was purged with N<sub>2</sub>. Substituted hydroquinone 61<sup>43</sup> (1.55 g, 4.7 mmol) was added, and air contact was minimized. Acetic anhydride (0.62 g, 6.1 mmol, 1.3 equiv) was purged with N<sub>2</sub> and then added to the reaction mixture at -30 °C. The mixture was covered with argon, stoppered, and kept for 3 days in a freezer (-18 °C). The mixture was poured into ice/hydrochloric acid. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were washed with saturated NaHCO<sub>3</sub> solution and with brine. They were dried, and the solvent was removed in vacuo. The residue was chromatographed through silica (150 g) with cyclohexane/toluene 1:1, then with toluene. A colorless solid was obtained. (Mp 192-194 °C, 1.24 g, 70% yield). Anal. (C<sub>23</sub>H<sub>29</sub>FO<sub>3</sub>) C, H, F.

1,4-Diacetoxy-6-(p-fluorophenyl)-2,3,5-triisopropylbenzene (62). Acetic anhydride (3.5 mL, 36.7 mmol, 3.0 equiv) and then pyridine (23 mL) were added at 0 °C to the substituted hydroquinone 61<sup>43</sup> (4.04 g, 12.23 mmol). Reaction was conducted and worked up, and the product was isolated as described for 8kk (vide

supra). A colorless solid was obtained. (Mp 172-173 °C, 3.06 g, 67% yield).

4-Acetoxy-6-(p-fluorophenyl)-2,3,5-triisopropylphenol (811). A solution of lithium hydroxide (114 mg, 4.77 mmol, 1.1 equiv) in water (10 mL) was added to a solution of diacetate 62 (1.8 g, 4.34 mmol) in 1,2-dimethoxyethane (30 mL). The mixture was stirred for 3 days at 25 °C. It was poured into 2 N HCl and extracted with ether. The extract was washed with saturated NaHCO<sub>3</sub> solution and with brine and then dried and concentrated in vacuo. Column chromatography (conditions as with 8kk) gave a colorless solid (mp 174-177 °C, 980 mg, 61% yield), besides 8kk (480 mg, 30% yield). Anal.  $(C_{23}H_{29}FO_3)$  C, H, F.

p-Fluorophenacyl Acetate (63). Bromine (25.7 mL, 0.5 mol) was added dropwise at 0 °C to the solution of p-fluoroacetophenone (69.1 g, 0.5 mol) in Et<sub>2</sub>O (200 mL). After a short induction period, the bromine drops were immediately decolorized to give a pale yellow solution. It was washed with water and dried, and the solvent was evaporated to give an oil (97.0 g, 89% yield) that quickly crystallized. This crude compound was added to a suspension of sodium acetate (123.0 g, 1.5 mol) in glacial acetic acid (0.5 L). The mixture was warmed at 120 °C bath temperature under a reflux condenser to give a clear, colorless solution. After 1 h a colorless precipitate was observed. After 4 h at 120 °C the solvent was removed in vacuo (bath 60 °C). The residue was triturated with Et<sub>2</sub>O (3 × 300 mL) and suction filtered each time. The combined filtrates were washed with saturated NaHCO<sub>3</sub> solution (6 × 100 mL) and then with brine. The solution was dried, and the solvent was evaporated. The residue (yellow solid) was distilled through a 15-cm Vigreux column in vacuo. After a prerun (bp 30-90 °C/1.3  $\times$  10<sup>-4</sup> bar, 4.6 g, yellow oil) there was obtained a colorless oil (bp 91-95 °C/9  $\times$  10<sup>-5</sup> bar, 76.6 g, 71% yield) that crystallized on standing.

2-(4-Fluorophenyl)-3-hydroxyquinoline-4-carboxylic Acid (64). KOH solution (6 N, 300 mL) was added to a solution of isatine (62 g, 0.42 mol) in EtOH (180 mL). The dark solution was heated to reflux, and a solution of p-fluorophenacyl acetate (63) (76 g, 0.387 mol) in EtOH (250-mL) was added dropwise within 30 min. The solution was refluxed for 12 h (bath 90 °C). Then most of the solvent (500 mL) was distilled off (bath 120 °C). The residue was poured into ice (1.8 kg)/concentrated HCl (480 mL). The yellow solid was suction filtered, suspended in hot 70% aqueous EtOH (4 L) and allowed to cool. The golden-yellow solid was suction filtered and dried at 50 °C in vacuo to give crude 64 (88.6 g, 81% yield, mp 206-207 °C dec). This crude product may be further purified via the sodium salt: saturated NaHCO<sub>3</sub> solution (500 mL) was added to the solution of crude 64 (14.2 g, 50 mmol) in EtOAc (250 mL). The mixture was stirred for 2 h at 20 °C. The organic layer was washed with brine and dried, and the solvent was evaporated in vacuo. The residue was recrystallized from CH<sub>3</sub>OH to give a solid (15 g, 98% yield, mp 255 °C dec). This compound was pure by TLC and <sup>1</sup>H NMR.

2-(4-Fluorophenyl)-3-hydroxy-4-(methoxycarbonyl)quinoline (65). An ethereal solution of diazomethane (30 mmol) was added at 0 °C to the suspension of crude, finely ground acid 64 (7.1 g, 25 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 L). The mixture was stirred for 30 min at 0 °C. The excess of CH<sub>2</sub>N<sub>2</sub> was destroyed with some drops of acetic acid. Volatiles were removed in vacuo. The residue was dissolved in CH2Cl2 and washed with saturated NaHCO3 solution and then with water. The organic layer was dried and the solvent was evaporated in vacuo. The residue was chromatographed through silica with toluene/cyclohexane 4:1 to give a crystalline solid (6.0 g, 80% yield, mp 118-120 °C). Anal. ( $C_{17}H_{12}FNO_3$ ) C, H, F, N.

2-(4-Fluorophenyl)-3-hydroxy-4-(hydroxymethyl)quinoline (66). A solution of methyl ester 65 (14.86 g, 50 mmol) in Et<sub>2</sub>O (150 mL) was added dropwise at 0-10 °C to the suspension of LiAlH<sub>4</sub> (1.9 g, 50 mmol) in Et<sub>2</sub>O (150 mL). The mixture was stirred for 30 min, refluxed for 2 h, EtOAc (20 mL) and then H<sub>2</sub>O (10 mL) were added, and then 2 N NaOH solution was added dropwise at 0 °C. EtOAc (50 mL) was added, and the mixture was filtered. The organic phase was dried and then evaporated in vacuo. The residue was chromatographed through silica with EtOAc/cyclohexane 1:2 to give a solid (9.3 g, 69% yield, mp 182-184 °C dec).

2-(4-Fluorophenyl)-3-hydroxy-4-methylquinoline (8mm). A solution of alcohol 66 (3.4 g, 12.6 mmol) in glacial acetic acid (500 mL) and concentrated hydrochloric acid (75 mL) was purged with N2. Pd (10%) on charcoal (3.0 g) was added, and the mixture was shaken for 1 day under 1 bar of H<sub>2</sub>. The catalyst was filtered off and washed with acetic acid. The filtrate was evaporated in vacuo. EtOAc and NaHCO<sub>3</sub> solution were added. The organic phase was separated and washed with brine, dried, and evaporated in vacuo. The residue was chromatographed (EtOAc/cyclohexane 1:4) to give a solid (2.2 g, 69% yield, mp 172-174 °C). Anal. (C<sub>16</sub>H<sub>12</sub>FNO) C, H, F, N.

2-(4-Fluorophenyl)-3-hydroxy-4-(2-hydroxy-2-propyl)quinoline (67). A solution of 65 (4.46 g, 15 mmol) in THF (40 mL) was added dropwise at 35 °C to a Grignard reagent, prepared from Mg turnings (2.0 g, 82 mmol) and methyl iodide (10.65 g, 75 mmol) in Et<sub>2</sub>O (60 mL). The mixture was stirred for 2 h at ambient temperature and then poured into 20% aqueous solution of NH<sub>4</sub>Cl. The organic phase was separated and the aqueous phase was extracted twice with Et<sub>2</sub>O. The combined organic phases were washed with water and then with brine and then dried. The solvent was evaporated in vacuo and the residue was chromatographed through silica (EtOAc/cyclohexane 1:4) to give a solid (4.0 g, 90% yield, mp 180-182 °C).

2-(4-Fluorophenyl)-3-hydroxy-4-isopropylquinoline (8nn). A solution of alcohol 67 (3.8 g, 12.8 mmol) and red phosphorus (3.97 g, 128 mmol, 10 equiv) in 67% aqueous HI (75 mL) was heated for 4 h at 150 °C (bath temperature). Volatiles were removed in vacuo. EtOAc and saturated NaHCO3 was added to the residue. The aqueous phase was extracted with EtOAc. The combined EtOAc phases were washed with aqueous NaHSO<sub>3</sub> solution and with brine and then dried. The solvent was evaporated in vacuo and the residue was chromatographed through silica (EtOAc/cyclohexane 8:1) to give a solid (2.33 g, 65% yield, mp 173-175 °C). Anal. (C<sub>18</sub>H<sub>16</sub>FNO) C, H, F, N.

3-Hydroxy-2-methyl-4-phenylquinoline (800). Glacial acetic acid (100 mL), water (20 mL), and concentrated H<sub>2</sub>SO<sub>4</sub> (1 mL) were added to the mixture of o-aminobenzophenone (68; 19.7 g, 0.1 mol) and hydroxyacetone (technical grade, Aldrich; 7.41 g, 6.85 mL, 0.1 mol). The mixture was refluxed for 4 h and allowed to stand at ambient temperature under N<sub>2</sub> overnight. The mixture was poured into ice-cold 10% NaOH solution (800 mL). The red brown solution was washed with Et<sub>2</sub>O (5  $\times$  100 mL). The aqueous phase (10-20%) was evaporated in vacuo to remove the small amount of dissolved Et<sub>2</sub>O. The aqueous phase was acidified (AcOH) and allowed to stand at 0 °C until the precipitation was complete. The fine solid was suction filtered, washed with Et<sub>2</sub>O, and dried in vacuo to obtain an ockre solid [21.9 g, 93% yield, mp 237-238 °C dec (lit.56 mp 236-237 °C dec, 51% yield)].

O-[4,6-Dichloro-2-[bis(4-fluorophenyl)methyl]phenyl] N,N-Dimethylthiocarbamate (69). Phenol 8b (29.2 g, 80 mmol, 1 equiv) was given portionwise at 0 °C to the suspension of a 50% dispersion of sodium hydride in mineral oil (3.6 g) in DMF (60 mL). The mixture was stirred for 30 min at 25 °C and recooled to 0 °C. The solution of N,N-dimethylthiocarbamic acid chloride (12.4 g, 1.25 equiv) in DMF (20 mL) was added, and the mixture was warmed to 80 °C for 2 h. It was recooled to 20 °C, Et<sub>2</sub>O (500 mL) was added, and the organic solution was washed with water  $(2 \times 250 \text{ mL})$  and with aqueous KHCO<sub>3</sub> solution. It was dried, and the solvent was evaporated in vacuo. The residue was recrystallized from CH<sub>3</sub>OH to give a solid (32.3 g, 89% yield, mp 178-179 °C).

S-[4,6-Dichloro-2-[bis(4-fluorophenyl)methyl]phenyl]N, N-dimethylthiocarbamate (70). Ester 69 (32 g) was heated to 275 °C under  $N_2$  for 30 min. The cold mixture was dissolved in the minimum amount of hot n-hexane. Charcoal (2 g) was added. The mixture was refluxed for 10 min and filtered hot. From the filtrate crystallized a solid (25.5 g, 80% yield, mp 130-131 °C).

4,6-Dichloro-2-[bis(4-fluorophenyl)methyl]thiophenol (8pp). A solution of 70 (6.2 g) in Et<sub>2</sub>O was added dropwise at 0 °C to the suspension of LiAlH<sub>4</sub> (0.8 g) in Et<sub>2</sub>O (50 mL). The mixture was stirred for 90 min at 25 °C. At 0 °C, 2 N H<sub>2</sub>SO<sub>4</sub> was added dropwise to pH 3. The aqueous phase was extracted with Et<sub>2</sub>O. The combined organic phases were dried, and the solvent was evaporated in vacuo. Solvent traces were removed in vacuo to leave a thick oil (5.2 g, 100% yield) that was pure according to TLC and <sup>1</sup>H NMR. Anal. (C<sub>19</sub>H<sub>12</sub>Cl<sub>2</sub>F<sub>2</sub>S) C, H, Cl.

4(R)-Hydroxy-6(S)-[[2,4-dichloro-6-(cyclohexyldifluoromethyl)phenoxy[methyl]tetrahydropyran-2-one (4qq). Monofluorolactone 4y (183 mg) was given at 0 °C to anhydrous HF (1 mL), contained in a polyethylene vessel. The mixture was stirred for 5 h under argon at 0 °C. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL), poured on solid NaF (5 g), and stirred overnight at ambient temperature. It was filtered, and the solid was washed with CH2Cl2. The filtrate was evaporated in vacuo. The residue was chromatographed through silica with CH<sub>2</sub>Cl<sub>2</sub>/EtOAc 4:1 to give a colorless glass (75 mg, 40% yield) that was pure by TLC and <sup>1</sup>H, <sup>13</sup>C, and <sup>19</sup>F NMR (see Table X of the supplementary material).

4(R)-(tert-Butyldiphenylsiloxy)-6(S)-[[2,4-dimethyl-6-(cyclohexylchloromethyl)phenoxy]methyl]tetrahydropyran-2-one (25rr). A solution of  $\alpha,\alpha$ -dimethylbutanoyl chloride (245 mg, 1.8 mmol, 3 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (3 mL) was added at 5 °C to a solution of alcohol 25q (370 mg, 0.6 mmol, 1 equiv) and DMAP (230 mg, 1.9 mmol, 3 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (8 mL) and pyridine (230 mg, 2.9 mmol, 5 equiv). The mixture was stirred for 1 h at 25 °C, then diluted with CH<sub>2</sub>Cl<sub>2</sub> (50 mL), washed with ice/water, and dried (Na<sub>2</sub>SO<sub>4</sub>). The drying agent was washed with toluene (25 mL) containing 1% pyridine. The filtrate was evaporated in vacuo. The residue was chromatographed through silica that had been prewashed with toluene (150 mL), containing 1% pyridine, then with toluene (200 mL). Toluene/EtOAc 98:2 was used as the solvent. A glass (310 mg, 74% yield) was obtained.

Biological Assays. The HMG-CoA reductase inhibition assay (Table II), the assay for inhibition of acetate incorporation into cholesterol in cultures of HEP G2 cells (Table III), and the determination of hypocholesterolemic activity in vivo after po administration of test compounds to NZW-rabbits (Table V) were conducted as described before.1

The inhibition of hepatic cholesterol de novo synthesis in vivo after po administration of test compounds to rats (Table IV) was determined as described in ref 58d.

Hypocholesterolemic Activity in Male Beagle Dogs (Table VI). The dosage of test compound indicated in Table VI (neat) was administered in gelatine capsules (size 000) daily in the afternoon (3:00 p.m.) for the period indicated to four male beagle dogs. A control group, consisting of four male beagle dogs of comparable body weight, obtained empty gelatine capsules. At the end of the treatment period, venous blood was collected (after fasting overnight) in the morning (8:00 a.m.) from the individual animals. The serum lipoproteins of the individual blood samples were separated with a preparative ultracentrifuge (density criteria: 1.006/1.063/1.21). Safety parameters SGOT, SGPT, aP, bilirubine, and creatinine indicated no appreciable pathologic event. The difference of the average of the plasma LDL levels of the treated animals to that of the control group is listed in Table VI.

Effect on Serum Lipoproteins and Other Metabolic Parameters after Subchronic Oral Administration to Male Rats (Table VII). Test compounds dissolved in polyethylene glycol 400 were administered in the morning in the daily dosage indicated in Table VII via gavage to groups (n = 10) of male rats of the strain HOE:WISKf (SPF 71) with initial body weights exceeding 180 g. Only vehicle was administered to the control group. The animals obtained food and water ad libitum. After the last (seventh) administration, animals were fasted. Blood samples were collected 24 h later from each individual animal retroorbitally under a mild ether narcosis. Immediately thereafter, the animals were sacrificed by spinal distorsion. The liver weight, body weight, and total food consumption were determined. Serum total cholesterol content was determined enzymatically from the blood of each individual animal by test combination of Boehringer-Mannheim (CHOD-PAP high-performance method). For determination of serum lipoproteins, the serum of all rats of one group was pooled, and serum lipoproteins were separated with a preparative ultracentrifuge. The following conditions were used for separation of the fractions VLDL, LDL, and HDL:

1	VLDL	<1.006
2	LDL	1.006-1.04
3	$\mathtt{HDL}$	1.04-1.21

The determination of the protein was according to the method of Lowry et al. 69 From the pooled serum of a group, total glycerol was determined (GPO-PAP high-performance method, Boehringer-Mannheim).

Inhibition of Microsomal Lipid Peroxidation (Table VIII, Left Side). Lipid peroxidation was assayed in rat liver microsomal fractions by measuring thiobarbituric acid reactive substances. Microsomal fractions were diluted with 50 mM TRIS-HCl. pH 7.4, containing KCl (100 mM) and MgCl<sub>2</sub> (6.0 mM). Final protein concentration in the incubation mixture amounted to 0.4 mg/mL. Lipid peroxidation was initiated with ADP (2 mM)/ FeCl<sub>3</sub> (10 μM) and a NADPH-regenerating system.<sup>70</sup> Samples were incubated at 37 °C for 30 min in a shaking water bath. Thiobarbituric acid reactive material was determined and the absorbance was measured at 535 nm.71

Inhibition of Cu<sup>2+</sup>-Catalyzed LDL Oxidation (Table VIII. Right Side). LDL was isolated from porcine plasma containing EDTA (1 mg/mL) by sequential ultracentrifugation in salt solutions of NaCl/NaBr between densities of 1.019 and 1.063 g/ mL.72 LDL was then dialyzed against phosphate-buffered saline (160 mM NaCl, 10 mM NaH<sub>2</sub>PO<sub>4</sub>), pH 7.4, and stored under N<sub>2</sub> at 4 °C. Prior to the oxidation process, LDL fractions were diluted with phosphate-buffered saline to the final protein concentration of 0.1 mg/mL, and 2.5-mL aliquots were preincubated with test compounds (25 µL of ethanolic solution) for 1 h at 37 °C capped under N<sub>2</sub>.73 For Cu<sup>2+</sup>-catalyzed oxidation of LDL, 12.5 µL of 1 mM CuSO<sub>4</sub> was added to each sample to attain a concentration of 5 µM Cu<sup>2+</sup>. Incubation was carried out at 37 °C for 2 h under an air atmosphere, and fluorescence intensity was measured at 430 nm with excitation at 365 nm.74

Acknowledgment. We are indebted to Dr. W. Schubert for the preparation of Figure 1, to Dr. H.-W. Fehlhaber, Dr. H. Kogler, and co-workers for <sup>1</sup>H NMR and mass spectra, to Dr. V. Teetz and co-workers for HPLC and GLC analyses, and to Mr. H. Leffringhausen for elemental analyses.

Supplementary Material Available: Spectral data of \(\beta\)hydroxy lactones 4, tert-butyl esters 10, and corresponding  $\beta$ ,  $\delta$ dihydroxy sodium carboxylates 11 are collected in Table X, and physical and spectral data and yield of phenolic building blocks 8 are collected in Table XI (11 pages). Ordering information is given on any current masthead page.

<sup>(69)</sup> Lowry, O. H.; Roseborough, N. J.; Farr, A. L.; Randell, R. J. J. Biol. Chem. 1951, 193, 265.

Scholich, H.; Murphy, M. E.; Sies, H. Biochim. Biophys. Acta 1989, 1001, 256.

<sup>(71)</sup> Buege, J. A.; Aust, S. D. Methods Enzymol. 1978, 52, 302.

<sup>(72)</sup> Havel, R. J.; Eder, H. A.; Bragdon, J. H. J. Clin. Invest. 1955,

<sup>(73)</sup> McLean, L. R.; Hagaman, K. A. Biochemistry 1989, 28, 321.

<sup>(74)</sup> Steinbrecher, U. P. J. Biol. Chem. 1987, 262, 3603.