4.30 (H, d, J = 12 Hz), 4.75 (H, d, J = 12 Hz), 5.50 (H, m), 5.80 (H, d of d, J = 10, 5 Hz), 6.00 (H, d, J = 10 Hz), 7.0 (2 H, m), 7.32 (2 H, m). Continued elution with methylene chloride-acetone (10:1, v:v) yielded the desilylated 5f (0.72 g, 1.67 mmol, 13%).

4(R)-(tert-Butyldimethylsiloxy)-6(R)-[2-[8(S)-[(4fluorobenzyl)oxy]-1,2,6,7,8,8a(S)-hexahydro-2(S),6(R)-dimethyl-1(S)-naphthyl]ethyl]-3,4,5,6-tetrahydro-2H-pyran-2-one (6f). Solid chromium trioxide (2.22 g, 22.2 mmol) was added at room temperature under a nitrogen atmosphere to a mechanically stirred solution of pyridine (3.51 g, 44.4 mmol) in methylene chloride (130 mL). The resulting mixture was stirred for 0.5 h, then added via a dropping funnel a solution of 5f (1.87 g, 3.45 mmol) in methylene chloride (20 mL). The resulting mixture was stirred at room temperature for 0.5 h, then quickly filtered through a pad of silica gel on a Buchner filter, washed, and rinsed with portions of methylene chloride-ether. The combined washings and filtrate were washed successively with diluted hydrochloric acid and 5% sodium bicarbonate, dried over MgSO4, and filtered. The filtrate was evaporated to leave a residue which was purified by flash chromatography on a silica gel column. Elution of the column with methylene chloride followed by methylene chloride-acetone (50:1, v:v) provided 6f (1.16 g, 2.14 mmol; 62%) as a solid: NMR (CDCl₃) δ 1.16 (3 H, d, J = 7 Hz), 2.52 (2 H, d, J = 4 Hz), 3.97 (H, m), 4.30 (H, d, J = 12 Hz), 4.72

(H, d, J = 12 Hz), 5.52 (H, m), 5.75 (H, d of d, J = 10, 5 Hz), 6.00 (H, d, J = 10 Hz), 7.0 (2 H, m), 7.3 (2 H, m).

6(R) - [2 - [8(S) - [(4 - Fluorobenzyl)oxy] - 1, 2, 6, 7, 8, 8a(S) - hexahydro-2(S), 6(R)-dimethyl-1(S)-naphthyl]ethyl]-4(R)hvdroxy-3.4.5.6-tetrahvdro-2H-pyran-2-one (2f). Acetic acid (3.20 mL, 55 mmol) was added to a stirred solution of 6f (6.8 g, 12.5 mmol) in THF (20 ml) followed by the addition of a solution of tetra-n-butylammonium fluoride (1 M in THF, 42 mL, 42 mmol). The resulting mixture was stirred at room temperature for 18 h, poured into cold water, and extracted with ether. The ethereal extract was washed successively with water and 5% sodium bicarbonate, dried, filtered, and evaporated to afford a residue. Purification of the residue on a silica gel column eluted with methylene chloride-acetone (10:1, v:v) provided 2f (5.3 g, 12.4 mmol, 99%) as a solid. Analytical sample was obtained via recrystallization from ether-hexane: mp 100–103 °C; NMR (CDCl₃) δ 0.86 (3 H, d, J = 7 Hz), 1.17 (3 H, d, J = 7 Hz), 2.59 (2 H, m), 3.90 (H, m), 4.30 (H, d, J = 12 Hz), 4.71 (H, d, J = 12 Hz)Hz), 5.53 (H, m), 5.76 (H, d of d, J = 10, 5 Hz), 6.01 (H, d, J =10 Hz), 7.03 (2 H, t), 7.33 (2 H, m). Anal. (C₂₆H₃₃FO₄) C, H.

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Naphthosultam Derivatives: A New Class of Potent and Selective 5-HT₂ Antagonists

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A series of 2-(aminoalkyl)naphth[1,8-cd]isothiazole 1,1-dioxides was synthesized and examined in various receptor binding tests. Most compounds demonstrated high affinity for the 5-HT₂ receptor with moderate to high selectivity. A member of this series, compound 24 (RP 62203), displays high 5-HT₂ receptor affinity ($K_i = 0.26$ nM), which is respectively more than 100 and 1000 times higher than its affinity for α_1 ($K_i = 38$ nM) and D₂ ($K_i > 1000$ nM) receptors. This compound is a potent orally effective and long lasting 5-HT₂ antagonist in the mescaline-induced head-twitches test in mice and rats.

Serotonin (5-HT) may be involved in thermoregulation, appetite, pain, sleep, sexual behavior, and behavioral disorders such as depression and anxiety.¹

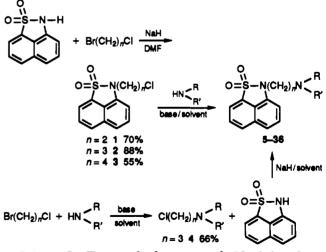
Recent progress in 5-HT research was stimulated by the discovery of multiple 5-HT receptors. The 5-HT₂ subtype has been the most completely characterized due to the availability of more and more selective antagonists.²

Ritanserin³ was the first compound identified with a reasonable separation of 5-HT₂ and non 5-HT effects. Clinical studies with ritanserin indicate that this 5-HT₂ antagonist may be effective in anxiety or disthymic disorders.⁴

We describe here a series of naphtho[1,8-cd]isothiazole 1,1-dioxide (naphthosultam) derivatives, incidentally discovered and structurally unrelated to ritanserin. These compounds were evaluated in vitro for their binding affinity to rat 5-HT₂, α^1 , and D₂ receptors. We describe the structure-activity relationships within this series that led to the discovery of RP 62203 (compound 24). Its pharmacological profile is compared with that of ritanserin.

Chemistry

The synthetic pathway for the preparation of naphthosultam derivatives⁵ listed in Tables I-IV is shown Scheme I



in Scheme I. Two methods were used. Naphthosultam was condensed with a ω -bromochloroalkane in the presence

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⁽¹⁾ Neuropharmacology of Serotonin; Green, A. R., Ed.; Oxford Univ. Press: Oxford, 1985.

⁽²⁾ Janssen, P. A. J. "Pharmacology of Potent and Selective S2-Serotoninergic Antagonists", J. Cardiovasc. Pharmacol. 1985, 7 (Suppl. 7), S2-S11.

Table I. In Vitro Binding Data of

4-Phenyl-1,2,3,6-tetrahydropyridine Derivatives 5-10 and Ritanserin

	A(CH ₂) ₃ N	$\neg \bigcirc$		
		IC	₅₀ (nM)	a
compd	Α	5-HT2 ^b	α_1^b	D_2^b
5	°≥s-N-	1.0	192	1000
6		9.8	580	>100
7		5.1	75	275
8		10	35	80
9	°≥s−N ^{CH} 3	>100		
10	°=s-N'	10	10	110
ritanserin		1.7	37	23

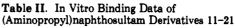
^a IC_{50} values are the mean of at least 2 determinations each with 6 concentrations of test compounds in triplicate. Standard deviations were always smaller than 10%. ^bSee Experimental Section.

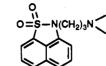
of NaH to give 1-3. These derivatives were reacted with the appropriate amine in the presence of a base (NaHCO₃, K_2CO_3 , or triethylamine) to give the expected compounds 5 and 11-36. An alternative approach was adopted for the synthesis of compounds 6-10: condensation of 1-bromo-3-chloropropane with 4-phenyl-1,2,3,6-tetrahydropyridine in the presence of potassium carbonate gives 4, which was reacted with an amide or a sulfonamide in the presence of sodium hydride to give 6-10. Yields were moderate but not optimized.

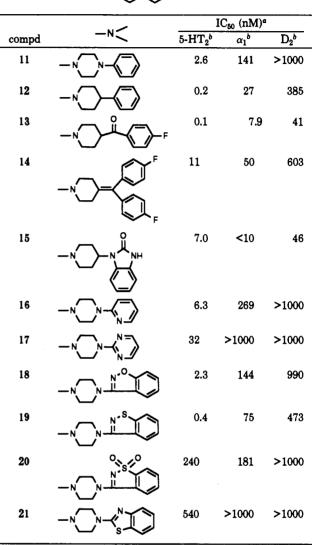
Results and Discussion

In Vitro Studies. Compounds 5–37 and ritanserin were evaluated in vitro in various receptor binding tests using rat brain membranes and suitable [³H] ligands: ketanserin (5-HT₂),⁶ prazosin (α_1),⁷ spiperone (D₂).⁸ IC₅₀ values are

- (3) "Ritanserin", Drugs of the Future 1986, 11, 391-393.
- (4) Reyntgens, A.; Gelderf, Y. F.; Hoppenbrouwers, M.-L. J. A.; Vanden Bussche, G. "Thymosthemic effect of ritanserin (R55667) centrally acting serotonin-S2 receptor blocker", Drug Dev. Res. 1986, 8, 205-211.
- (5) Rhone Poulenc Sante, EP 350403.
- (6) Leysen, J. E.; Niemegeers, C. J. E.; Van Nueten, J. M.; Laduron, P. M. "[³H]Ketanserin (R41468), a selective ³H-ligand for serotonin 2 receptor binding sites. Binding properties, brain distribution, and functional role", *Mol. Pharmacol.* 1982, 21, 301-314.
- (7) Greengrass, P.; Bremner, R. "Binding characteristics of ³Hprazosin to rat brain α-adrenergic receptors", Eur. J. Pharmacol. 1979, 55, 323-326.



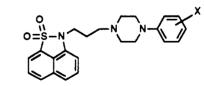




^aSee footnotes to Table I. ^bSee Experimental Section.

Table III. In Vitro Binding Data of

[(Phenylpiperazinyl)propyl]naphthosultam Derivatives 22-32

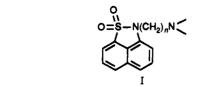


		IC ₅₀ (nM) ^a					
compd	х	5-HT ₂ ^b	α_1^b	D_2^b			
22	2-F	3.0	52	>1000			
23	3-F	18	688	>1000			
24	4-F	0.5	103	>1000			
25	4-C1	5.0	>1000	>1000			
26	4-Br	7.0	>1000	>1000			
27	4-0H	0.3	110	508			
28	4-OCH ₃	25	>1000	>1000			
29	4-CH ₃	4.0	700	>1000			
30	4-CF ₃	275	>1000	>1000			
31	4-NO ₂	25	>1000	>1000			
32	$4-CO_2C_2H_5$	200	>1000	>1000			

^aSee footnotes to Table I. ^bSee Experimental Section.

 Table IV. In Vitro Binding Data of (Aminoalkyl)naphthosultam

 Derivatives 33-36



		IC ₅₀ (nM) ^a						
compd	_N_	n	5-HT ₂ ^b	α_1^b	D_2^b			
33		2	6.3	98	>100			
34	-N_	4	2.1	7.8	55			
35	-N_N-K-F	2	1.0	64				
36	-N_N-F	4	3.7	11	300			

^aSee footnotes to Table I. ^bSee Experimental Section.

shown in Tables I–IV. In addition, compound 24 and ritanserin were tested for their 5- HT_{1A} and H_1 receptor binding affinities, using [³H]-8-OH-DPAT⁹ and [³H]mepyramine¹⁰ as ligands, respectively. K_i values for 24 and ritanserin are compared in Table V.

Among compounds 5–10, the naphthosultam derivative 5 was shown to be the most interesting compound with regard to its in vitro potency and selectivity for the 5-HT₂ receptor. In particular, replacement of the sulfonyl group in 5 by a carbonyl group (6) resulted in a 10-fold decrease in 5-HT₂ receptor affinity and opening the sultam ring in 5 gave compounds 7-9 with reduced potency and loss of selectivity. Replacement in 5 of the amine moiety 4phenyl-1,2,3,6-tetrahydropyridine by 4-phenylpiperidine or 4-phenylpiperazine does not substantially affect the selectivity (compounds 11, 12). However, affinity for the 5-HT₂ receptor and selectivity are largely determined by the nature of the aromatic group attached to the cyclic amine (compounds 13-21). Incorporation of the amine moiety present in the structure of ketanserin (13), ritanserin (14), and benperidol (15) resulted in loss of selectivity in comparison with the parent molecules 5, 11, and 12. In the phenylpiperazine series (compounds 22-32), 4-fluoro (compound 24) and 4-hydroxy (compound 27) substituents are optimal. Alternative substituents gave compounds with reduced potency (compounds 25, 26, 29-32) and in some bases reduced selectivity (compounds 11, 22, 23, 28). In the 4-phenyl-1,2,3,6-tetrahydropyridine as well as in the phenylpiperazine series (compounds 33-36), the alkylene side chain was optimal with n = 3.

In Vivo Studies. For the most potent naphthosultam derivatives found in vitro, central 5-HT₂ antagonistic activity in vivo was assessed by the ability of the compounds

to antagonize mescaline-induced head twitches in mice and rats.¹¹ RP 62203 (compound 24) was the compound with the optimum combination of selectivity in the in vitro assays and in vivo potency. As shown in Table V, RP 62203 is a potent and long lasting orally active 5-HT₂ antagonist. It displays a higher receptor selectivity than ritanserin with a 5-HT₂ receptor affinity ($K_i = 0.26$ nM) that is respectively more than 100 and 1000 times higher than its affinity for α_1 ($K_i = 38$ nM) and D₂ ($K_i > 1000$ nM) receptors.

Conclusion

RP 62203 (compound 24) has been identified as a novel potent and selective $5-HT_2$ antagonist. It is a member of a series of naphthosultam derivatives of formula I. (See formula in Table IV.)

High affinity was obtained with compounds possessing as a terminal amine moiety one of the following group: 4-phenylpiperazine, 4-phenylpiperidine, or 4-phenyl-1,2,3,6-tetrahydropyridine. Selectivity for the 5-HT₂ receptor versus α_1 and D₂ receptors required a propylene chain (n = 3). In the phenylpiperazine series, 4-fluoro (compound 24) and 4-hydroxy (compound 28) substituents led to the optimum combination of in vitro potency and selectivity. Due to its in vivo activity, compound 24 has been selected for clinical evaluation.

Experimental Section

Melting points were recorded on a Köfler apparatus and were uncorrected. ¹H NMR spectra were recorded on a Brücker WM (250 MHz), a Brücker WP 200 (200 MHz), or a Brücker AM 400 (400 MHz) instrument. IR spectra were taken on a Perkin-Elmer Model 938G or 580B instrument. Mass spectra were recorded on a Finnigan 3300 spectrometer. Unless otherwise noted, where elemental analyses are indicated by the symbols of the elements, the results are within 0.4% of the theoretical values. In some cases, crude products were purified by flash column chromatography on silica gel (0.04–0.063 mm supplied by Merck) before recrystallization.

Preparation of the Halogenoalkyl Derivatives 1-4. 2-(2-Chloroethyl)-2H-naphth[1,8-cd]isothiazole 1,1-Dioxide (1). To 4.8 g (0.10 mol) of sodium hydride (50% dispersion in Vaseline) in 50 mL of DMF, under argon, was slowly added 20.5 g (0.10 mol) of 1,8-naphthosultam in 250 mL of DMF at 25 °C. The reaction mixture was stirred at this temperature for 1 h. To this solution was added 8.8 mL (0.11 mol) of 1-bromo-2-chloroethane, and the mixture was left for 15 h at room temperature. After evaporation to dryness under reduced pressure, the residue was purified by flash chromatography on silica gel, eluting with 7:3 dichloromethane/cyclohexane, to give 20.7 g (77%) of 1 as a solid (mp 96 °C): NMR (CDCl₃, 300 MHz) δ 3.92 (t, J = 7 Hz, 2 H, CH₂Cl), 4.18 (t, J = 7 Hz, 2 H, NCH₂), 6.82 (d, J = 7.5 Hz, 1 H, H Ar), 7.44 (d, J = 7.5 Hz, 1 H, H Ar), 7.53 (t, J = 7.5 Hz, 1 H, H Ar), 7.71 (t, J = 7.5 Hz, 1 H, H Ar), 7.94 (d, J = 7.5 Hz, 1 H, H Ar), 8.04 (d, J = 7.5 Hz, 1 H, H Ar); MS, m/z 267 (M⁺), 218 (M⁺ -CH₂Cl), 154 (C₁₁H₈N⁺).

Compounds 2-4 were prepared in a similar manner.

2-(3-Chloropropyl)-2*H*-naphth[1,8-*cd*]isothiazole 1,1-dioxide (2) was prepared in 69% yield as a solid (after flash chromatography on silica gel with 6:4 dichloromethane/cyclohexane as eluent, mp 78 °C) from 1,8-naphthosultam, 1-bromo-3-chloropropane, and sodium hydride: NMR (CDCl₃, 300 MHz) δ 2.37 (q, J = 6.5 Hz, 2 H, CH₂), 3.74 (t, J = 6.5 Hz, 2 H, CH₂Cl), 4.04 (t, J = 6.5 Hz, 2 H, NCH₂), 6.82 (d, J = 7.5 Hz, 1 H, H Ar), 7.43 (d, J = 7.5 Hz, 1 H, H Ar), 7.52 (t, J = 7.5 Hz, 1 H, H Ar), 7.71 (t, J = 7.5 Hz, 1 H, H Ar), 7.93 (d, J = 7.5 Hz, 1 H, H Ar), 8.04 (d, J = 7.5 Hz, 1 H, H Ar); IR (KBr) 1305 + 1170 + 1140 (SO₂); MS, m/z 281 (M⁺), 218 (M⁺ - C₂H₄Cl), 154 (C₁₁H₈N⁺).

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⁽¹⁰⁾ Chang, R. S. L.; Tran, V. T.; Snyder, S. H. "Heterogeneity of histamine H₁-receptors: species variations in [⁸H]-mepyramine binding of brain membranes", J. Neurochem. 1979, 32, 1653-1663.

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Table	V.	In	Vitro and	in	Vivo	Activities	of	Compound	24	and	Ritanserin
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						mescaline-induced head twitch ^b					
		binding	¹ : K _i (1	nM)			ice mg/kg)	duration of	rats ED ₅₀ (mg/kg)		
compd	5-HT ₂	5-HT _{1A}	α_1	D_2	H ₁	8C	po	action, h	BC	po	
								1	0.65 [0.38–1.12]	0.99 [0.63–1.55]	
24	0.26	70	38	>1000	25	0.16 [0.08–0.34]	0.43 [0.08–2.28]	6	0.45 [0.22–0.89]	2.38 [1.08-5.25]	
								16	5.91 [3.06–11.43]	14.2 [10.7–18.8]	
								1	0.38 [0.24–0.62]	2.55 [1.22-5.33]	
ritanserin	0.69	>1000	29	22	6	0.29 [0.08–1.07]	0.09 [0.04–0.23]	6	0.55 [0.34–0.93]	1.54 [0.74-3.20]	
								16	>5	18.41 [9.17–36.97]	

^aSee footnotes to Table I. ^bED₅₀ values were calculated with 95% confidence limit.

2-(4-Chlorobutyl)-2H-naphth[1,8-cd]isothiazole 1,1-dioxide (3) was prepared in 83% yield as a yellow oil (after flash chromatography on silica gel with 3:7 dichloromethane/cyclohexane as eluent) from 1-bromo-4-chlorobutane, 1,8-naphthosultam, and sodium hydride: NMR (CDCl₃, 200 MHz) δ 1.9-2.25 (m, 4 H, CCH₂CH₂C), 3.65 (t, J = 6.5 Hz, 2 H, CH₂Cl), 3.9 (t, J= 7 Hz, 2 H, NCH₂), 6.87 (d, J = 7.5 Hz, 1 H, H Ar), 7.48 (d, J= 8.5 Hz, 1 H, H Ar), 7.57 (dd, J = 8.5 and 7.5 Hz, 1 H, H Ar), 7.77 (dd, J = 8 and 7.5 Hz, 1 H, H Ar), 7.99 (d, J = 7.5 Hz, 1 H, H Ar), 8.1 (d, J = 8 Hz, 1 H, H Ar).

1-(3-Chloropropyl)-4-phenyl-1,2,3,6-tetrahydropyridine (4). A mixture of 6 mL (60.6 mmol) of 1-bromo-3-chloropropane, 5.1 g (32 mmol) of 4-phenyl-1,2,3,6-tetrahydropyridine in 60 mL of acetonitrile, and 97 g of potassium carbonate was stirred for 20 h at 25 °C. After filtration of the precipitate and evaporation under vacuum of the organic phase, the residue was purified by flash chromatography on silica gel with 5:5 cyclohexane/ethyl acetate as eluent to give 5 g (66%) of 4 as an oil: NMR (CDCl₃, 250 MHz) δ 2.00 (m, 2 H, CCH₂C), 2.55 (m, 4 H, NCH₂CH₂C=), 2.7 (t, 2 H, CCH₂N), 3.1 (bs, 2 H, NCH₂C=), 3.6 (t, 2 H, ClCH₂), 6.00 (bs, 1 H, CH==), 7.18 (bt, 1 H, C₆H₅), 7.28 (bt, 2 H, C₆H₅), 7.35 (bd, 2 H, C₆H₅).

Preparation of 2-(Aminoalkyl)naphth[1,8-cd]isothiazole 1,1-Dioxides 5 and 11-35. 2-[3-(4-Phenyl-1,2,3,6-tetrahydropyridyl)propyl]-2H-naphth[1,8-cd]isothiazole 1,1-Dioxide (5). To 2.8 g (0.01 mol) of 2 and 1.4 mL of triethylamine was added 1.6 g (0.01 mol) of 4-phenyl-1,2,3,6-tetrahydropyridine in 30 mL of toluene. The reaction mixture was stirred 8 h at reflux and then for 15 h at room temperature. After filtration of the precipitate and evaporation to dryness under reduced pressure, the residue was purified by flash chromatography on silica gel with ethyl acetate as eluent. Recrystallization from 2-propand gave 1.5 g (37%) of 5 as a solid (mp 88 °C): NMR (CDCl₃, 200 MHz) δ 2.02 (m, 2 H, CCH₂C), 2.45-2.7 (m, 6 H, NCH₂ and NCH₂CH₂ cycle), 3.1 (m, 2 H, NCH₂ cycle) 3.94 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.17 (m, 1 H, CH=), 7.11-8.28 (m, 11 H, H Ar); IR (KBr) 2815 + 2770 + 2740 (NCH₂), 1310 + 1170 + 1140 (SO₂); MS, m/z 404 (M⁺). Anal. (C₂₄H₂₄N₂O₂S) C, H, N. S.

Compounds 11-35 were prepared in a similar manner.

2-[3-(4-Phenyl-1-piperazinyl) propyl]-2H-naphth[1,8cd]isothiazole 1,1-dioxide (11) was prepared in 54% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 126 °C) from 2 and 1-phenylpiperazine: NMR (CDCl₃, 200 MHz) δ 2.16 (q, J = 7 Hz, 2 H, CCH₂C), 2.52 (t, J = 7 Hz, 2 H, NCH₂), 2.56 (m, 4 H, NCH₂ cycle), 3.25 (m, 4 H, PhNCH₂), 3.99 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.8-8.09 (m, 11 H, H Ar); IR (KBr) 2825 + 2785 (NCH₂), 1310 + 1175 + 1145 (SO₂); MS, m/z 407 (M⁺), 343 (M⁺ - SO₂). Anal. (C₂₃H₂₅N₃O₂S) C, H, N, S.

2-[3-(4-Phenyl-1-piperidinyl) propyl]-2H-naphth[1,8-cd]isothiazole 1,1-dioxide (12) was prepared in 57% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 125 °C) from 2 and 4-phenylpiperidine: NMR (CDCl₃, 200 MHz) δ 1.85 (m, 4 H, CH₂ piperidine), 2-2.3 (m, 4 H, CCH₂C and NCH₂ piperidine), 2.5 (m, 1 H, CCHC), 2.6 (t, J = 7 Hz, 2 H, NCH₂), 3.1 (t, 2 H, NCH₂ piperidine), 3.97 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.9 (m, 11 H, H Ar); IR (KBr) 2810 + 2770 + 2740 (NCH₂), 1310 + 1175 + 1135 (SO₂). Anal. (C₂₄H₂₆N₂O₂S) C, H, N. S.

2-[3-[4-(4-Fluorobenzoyl)-1-piperidinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (13) was prepared in 24% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 132 °C) from 2, triethylamine as base, sodium iodide, and (4-fluorobenzoyl)-4-piperidine: NMR (CDCl₃, 200 MHz) δ 1.87 (m, 4 H, CH₂ piperidine), 2-2.25 (m, 4 H, CCH₂C and NCH₂ piperidine), 2.54 (t, *J* = 7 Hz, 2 H, NCH₂), 3.03 (m, 2 H, NCH₂ piperidine), 3.23 (m, 1 H, CHC=O), 3.94 (t, *J* = 7 Hz, 2 H, SO₂NCH₂), 6.87-8.1 (m, 10 H, H Ar); IR (KBr) 2805 + 2870 (NCH₂), 1675 (C=O), 1310 + 1165 + 1135 (SO₂); MS, *m/z* 452 (M⁺), 388 (M⁺ - SO₂). Anal. (C₂₅H₂₅FN₂O₃S) C, H, N, S.

2-[3-[4-[Bis(4-fluorophenyl)methylene]-1-piperidinyl]propyl]-2*H*-naphth[1,8-*cd*]isothiazole 1,1-dioxide (14) was prepared in 67% yield as an oil (after flash chromatography on silica gel with dichloromethane as eluent, hydrochloride, mp 137 °C) from 2, triethylamine as base, and 4-[bis(4-fluorophenyl)methylene]piperidine: NMR (DMSO, 200 MHz) δ 2.1-2.7 (m, 6 H, CCH₂C and CH₂ piperidine), 2.9-3.6 (m, 6 H, NCH₂ and NCH₂ piperidine), 4.0 (t, *J* = 7 Hz, 2 H, SO₂NCH₂), 7.1-8.33 (m, 14 H, H Ar), 10.75 (m, 1 H, NH⁺Cl⁻); IR (KBr) 2800-2000 (NH⁺), 1315 + 1170 + 1140 (SO₂); MS, *m/z* 530 (M⁺), 466 (M⁺ - SO₂). Anal. (C₃₁H₂₈F₂N₂O₂S·HCl) C, H, N, S.

2-[3-[4-(2-Oxo-1-ben zimida zolinyl)-1-piperidinyl]propyl]-2H-naphth[1,8-cd]isothiazole 1,1-dioxide (15) was prepared in 7% yield as a solid (after flash chromatography on silica gel with 95:5 dichloromethane/ethanol as eluent and recrystallization from acetonitrile, mp 253 °C) from 2, triethylamine as base, and 1-(4-piperidinyl)-2-benzimidazolinone: NMR (DMSO, 250 MHz) δ 1.64 (m, 2 H, CH₂ piperidine), 2.0 (m, 4 H, CH₂ piperidine and CCH₂C), 2.33 (m, 2 H, NCH₂ piperidine), 2.5 (m, 2 H, NCH₂), 3.0 (m, 2 H, NCH₂ piperidine), 3.96 (t, J = 7 Hz, 2 H, SO₂NCH₂), 4.15 (m, 1 H, NCH), 6.9-8.32 (m, 10 H, H Ar), 10.86 (m, 1 H, NH); MS, m/z 462 (M⁺), 398 (M⁺ - SO₂). Anal. (C₂₅H₂₆N₄O₃S) C, H, N, S. C: calcd, 64.91; found, 63.4.

2-[3-[4-(2-Pyridinyl)-1-piperazinyl]propyl]-2H-naphth-[1,8-*cd*]isothiazole 1,1-dioxide (16) was prepared in 33% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 101 °C) from 2, triethylamine as base, and 1-(2-pyridyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.17 (q, 2 H, CCH₂C), 2.5–2.7 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.6 (m, 4 H, NCH₂ piperazine), 3.98 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.6–8.12 (m, 10 H, H Ar); IR (KBr) 1590 + 1560 + 1555 + 1480 + 1435 (2-pyridyl), 1310 + 1170 + 1135 (SO₂); MS, m/z 408 (M⁺). Anal. (C₂₂H₂₄N₄O₂S) C, H, N, S.

2-[3-[4-(2-Pyrimidy1)-1-piperaziny1]propy1]-2H-naphth-[1,8-cd]isothiazole 1,1-dioxide (17) was prepared in 63.5% yield as a solid (after flash chromatography on silica gel with 95:5 ethyl acetate/methanol as eluent and recrystallization from acetonitrile, mp 112 °C) from 2, triethylamine as base, and 1-(2-pyrimidy1)piperazine: NMR (CDCl₃, 200 MHz) δ 2.15 (m, 2 H, CCH₂C), 2.5-2.7 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.86 (t, J = 5.5 Hz, 4 H, NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.5–8.32 (m, 9 H, H Ar); IR (KBr) 1545 + 1510 (pyrimidine), 1310 + 1170 + 1140 (SO₂); MS, m/z 409 (M⁺), 345 (M⁺ – SO₂). Anal. (C₂₁-H₂₃N₅O₂S) C, H, N. S.

2-[3-[4-(1,2-Benzisoxazol-3-yl)-1-piperazinyl]propyl]-2Hnaphth[1,8-cd]isothiazole 1,1-dioxide (18) was prepared in 4% yield as a solid (after flash chromatography on silica gel with dichloromethane as eluent and recrystallization from acetonitrile, mp 163 °C) from 2, triethylamine as base, and 4-(1,2-benzisoxazol-3-yl)-1-piperazine in DMF: NMR (CDCl₃, 200 MHz) δ 2.15 (q, J = 7 Hz, 2 H, CCH₂C), 2.63 (t, J = 7 Hz, 2 H, NCH₂), 2.7 (m, 4 H, NCH₂ piperazine), 3.63 (m, 4 H, NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.86-8.09 (m, 10 H, H Ar); IR (KBr) 1610 + 1530 + 1450 (1,2-benzisoxazole); MS, m/z 448 (M⁺), 383 (M⁺ - SO₂). Anal. (C₂₄H₂₄N₄O₃S) C, H, N, S.

2-[3-[4-(1,2-Benzisothiazol-3-yl)-1-piperaziny] propy]-2H-naphth[1,8-cd]isothiazole 1,1-dioxide (19) was prepared in 24% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from methyl ethyl ketone, mp 172-173 °C) from 2, triethylamine as base, and 4-(1,2-benzisothiazol-3-yl)-1-piperazine in DMF: NMR (CDCl₃, 200 MHz) δ 2.16 (q, J = 7 Hz, 2 H, CCH₂C), 2.64 (t, J = 7 Hz, 2 H, NCH₂), 2.72 (m, 4 H, NCH₂ piperazine), 3.6 (m, 4 H, NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.89–8.1 (m, 10 H, H Ar); IR (KBr) 1560 + 1425 (1,2-benzisothiazole), 1310 + 1170 + 1135 (SO₂); MS, m/z 464 (M⁺), 432 (M⁺ - SO₂). Anal. (C₂₄-H₂₄N₄O₂S₂) C, H, N. S.

2-[3-[4-1,1-Dioxo-1,2-benzisothiazol-3-yl)-1-piperaziny]propy]-2*H*-naphth[1,8-*cd*]isothiazole 1,1-dioxide (20) was prepared in 51% yield as an oil (after flash chromatography on silica gel with 97:3 dichloromethane/methanol as eluent, dihydrochloride, mp >260 °C) from 2, triethylamine as base, sodium iodide, and 4-(1,1-dioxo-1,2-benzisothiazol-3-yl)-1-piperazine in DMF: NMR (DMSO, 200 MHz) δ 2.27 (m, 2 H, CCH₂C), 3.2-3.7 (m, 10 H, H and NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 4.7 (m, 1 H, NH⁺Cl⁻), 7.55-8.4 (m, 10 H, H Ar), 11.2 (m, 1 H, NH⁺Cl⁻); MS, m/z 496 (M⁺), (M⁺ - SO₂). Anal. (C₂₄H₂₄N₄O₄S₂·2HCl) C, H, N, S.

2-[3-[4-(2-Benzothiazolyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (21) was prepared in 46% yield as a solid (after flash chromatography on silica gel with 98:2 dichloromethane/methanol as eluent and recrystallization from acetonitrile, mp 195 °C) from 2, triethylamine as base, and 1-(2-benzothiazolyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.14 (q, J = 7 Hz, 2 H, CCH₂C), 2.5-2.7 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.69 (m, 4 H, NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.85-8.09 (m, 10 H, H Ar); IR (KBr) 1560 + 1535 + 1445 (benzothiazole), 1310 + 1170 + 1135 (SO₂); MS, m/z 464 (M⁺). Anal. (C₂₄H₂₄N₄O₂S₂) C, H, N, S.

2-[3-[4-(2-Fluorophenyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (22) was prepared in 45% yield as a solid (after flash chromatography on silica gel with dichloromethane as eluent and recrystallization from acetonitrile, mp 112 °C) from 2, triethylamine as base, and 1-(2-fluorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.13 (q, J = 7 Hz, 2 H, CCH₂C), 2.6 (t, J = 7 Hz, 2 H, NCH₂), 2.67 (m, 4 H, NCH₂ piperazine), 3.15 (m, 4 H, NCH₂ piperazine), 3.97 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.86-7.97 (m, 10 H, H Ar); MS, m/z 425 (M⁺), 361 (M⁺ - SO₂). Anal. (C₂₃H₂₄FN₃O₂S) C, H, N, S.

2-[3-[4-(3-Fluorophenyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (23) was prepared in 37% yield as a solid (after flash chromatography on silica gel with 5:5 dichloromethane/ethyl acetate as eluent and recrystallization from acetonitrile, mp 103 °C) from 2, triethylamine as base, and 1-(3-fluorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.13 (q, J = 7 Hz, 2 H, CCH₂C), 2.6 (t, J = 7 Hz, 2 H, NCH₂), 2.67 (m, 4 H, NCH₂ piperazine), 3.15 (m, 4 H, NCH₂ piperazine), 3.97 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.86-7.97 (m, 10 H, H Ar); MS, m/z425 (M⁺), 361 (M⁺ - SO₂). Anal. (C₂₃H₂₄FN₃O₂S) C, H, N, S.

2-[3-[4-(4-Fluorophenyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (24) was prepared in 89% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 95–97 °C) from 2, triethylamine as base, and 1-(4-fluorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.17 (q, J = 7 Hz, 2 H, CCH₂C), 2.64 (t, J = 7 Hz, 2 H, NCH₂), 2.67 (m, 4 H, NCH₂) piperazine), 3.19 (m, 4 H, NCH₂ piperazine), 3.99 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.8–8.09 (m, 10 H, H Ar); MS, m/z 425 (M⁺), 361 (M⁺ – SO₂). Anal. (C₂₃H₂₄FN₃O₂S) C, H, N, S.

2-[3-[4-(4-Chlorophenyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (25) was prepared in 36% yield as a solid (after recrystallization from acetonitrile and then methyl ethyl ketone, mp 120 °C) from 2, triethylamine as base, sodium iodide, and 1-(4-chlorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.13 (q, J = 6.5 Hz, 2 H, CCH₂C), 2.5-2.7 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.2 (m, 4 H, NCH₂ piperazine), 3.97 (t, J = 6.5 Hz, 2 H, SO₂NCH₂); MS, m/z 441 (M⁺), 377 (M⁺ -SO₂). Anal. (C₂₃H₂₄ClN₃O₂S) C, H, N, S.

2-[3-[4-(4-Bromophenyl)-1-piperazinyl]propyl]-2Hnaphth[1,8-cd]isothiazole 1,1-dioxide (26) was prepared in 44% yield as a solid (after flash chromatography with ethyl acetate and recrystallization from acetonitrile, mp 149 °C) from 2, triethylamine as base, and 1-(4-bromophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.13 (q, J = 6.5 Hz, 2 H, CCH₂C), 2.5-2.7 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.2 (m, 4 H, NCH₂ piperazine), 3.97 (t, J = 6.5 Hz, 2 H, SO₂NCH₂); MS, m/z 485 (M⁺), 421 (M⁺ - SO₂). Anal. (C₂₃H₂₄BrN₃O₂S) C, H, N, S.

2-[3-[4-(4-Hydroxyphenyi)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (27) was prepared in 19% yield as a solid (after flash chromatography on silica gel with 50:50 dichloromethane/ethyl acetate as eluent and recrystallization from acetonitrile, mp 185 °C) from 2, triethylamine as base, and 1-(4-hydroxyphenyl)piperazine dihydrobromide in DMF: NMR (DMSO + ϵ CD₃CO₂D, 200 MHz) δ 2.13 (m, 2 H, CCH₂C), 2.7-2.9 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.08 (m, 4 H, NCH₂ piperazine), 3.96 (t, *J* = 7 Hz, 2 H, SO₂NCH₂), 6.68 (m, 10 H, H Ar); IR (KBr) 3200 + 2200 (OH), 1305 + 1170 + 1135 (SO₂); MS, *m/z* 423 (M⁺), 359 (M⁺ - SO₂). Anal. (C₂₃H₂₅N₃O₃S) C, H, N, S.

2-[3-[4-(4-Methoxyphenyl)-1-piperazinyl]propyl]-2Hnaphth[1,8-cd]isothiazole 1,1-dioxide (28) was prepared in 64% yield as a solid (after flash chromatography on silica gel with ethyl acetate as eluent and recrystallization from acetonitrile, mp 160 °C) from 2, triethylamine as base, and 1-(4-methoxyphenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.14 (q, J = 7 Hz, 2 H, CCH₂C), 2.6 (t, J = 7 Hz, 2 H, NCH₂), 2.65 (m, 4 H, NCH₂) piperazine), 3.14 (m, 4 H, NCH₂ piperazine), 3.78 (s, 3 H, OCH₃), 3.97 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.8-8.08 (m, 10 H, HAr); IR (KBr) 1300 + 1175 + 1150 + 1140 (SO₂), 1240 (C--O), 2835 + 1035 (OCH₃); MS, m/z 437 (M⁺), 373 (M⁺ - SO₂). Anal. (C₂₄-H₂₇N₃O₃S) C, H, N, S.

2-[3-[4-(4-Methylphenyl)-1-piperazinyl]propyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (29) was prepared in 43% yield as a solid (after flash chromatography on silica gel with 80:20 dichloromethane/ethyl acetate as eluent and recrystallization from acetonitrile, mp 136 °C) from 2, triethylamine as base, and 1-(4-methylphenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.14 (q, J = 7 Hz, CCH₂C), 2.29 (s, 3 H, CH₃), 2.6 (t, J = 7 Hz, 2 H, NCH₂), 2.65 (m, 4 H, NCH₂ piperazine), 3.19 (m, 4 H, NCH₂ piperazine), 4.0 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.87-8.08 (m, 10 H, H Ar); MS, m/z 421 (M⁺), 357 (M⁺ - SO₂). Anal. (C₂₄H₂₇N₃O₂S) C, H, N, S.

2-[3-[4-[4-(Trifluoromethyl)phenyl]-1-piperazinyl]propyl]-2H-naphth[1,8-cd]isothiazole 1,1 dioxide (30) was prepared in 39% yield as a solid (after flash chromatography on silica gel with ethyl acetate and recrystallization from acetonitrile, mp 153 °C) from 2, triethylamine as base, and 1-[4-(trifluoromethyl)phenyl]piperazine: NMR (CDCl₃, 200 MHz) δ 2.15 (q, J = 7 Hz, 2 H, CCH₂C) 2.5-2.7 (m, 6 H, NCH₂ and NCH₂ pi perazine), 3.82 (m, 4 H, NCH₂ piperazine), 3.98 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.84-8.08 (m, 10 H, H Ar); MS, m/z 475 (M⁺), 411 (M⁺ - SO₂). Anal. (C₂₄H₂₃F₃N₃O₂S) C, H, N, S. C: calcd, 60.61; found, 60.1.

2-[3-[4-(4-Nitrophenyl)-1-piperazinyl]propyl]-2Hnaphth[1,8-cd]isothiazole 1,1-dioxide (31) was prepared in 36% yield as a solid (after flash chromatography on silica gel with 80:20 dichloromethane/ethyl acetate as eluent and recrystallization from methyl ethyl ketone, mp 186 °C) from 2, triethylamine as base, and 1-(4-nitrophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.14 (q, J = 7 Hz, 2 H, CCH₂C), 2.64 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.46 (m, 4 H, NCH₂ piperazine), 3.99 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.85-8.14 (m, 10 H, H Ar); IR (KBr) 1325 (SO₂ 2-[3-[4-[4-(Ethoxycarbonyl)phenyl]-1-piperazinyl]propyl]-2*H*-naphth[1,8-*cd*]isothiazole 1,1-dioxide (32) was prepared in 33% yield as a solid (after flash chromatography on silica gel with ethyl acetate and recrystallization from acetonitrile, mp 139 °C) from 2, triethylamine as base, and 1-[4-(ethoxycarbonyl)phenyl]piperazine: NMR (CDCl₃, 300 MH2) δ 1.38 (t, J = Hz, 3 H, CH₃), 2.16 (q, J = 6.5 Hz, 2 H, CCH₂C), 2.55-2.75 (m, 6 H, NCH₂ and NCH₂ piperazine), 3.38 (m, 4 H, NCH₂ piperazine), 3.98 (t, J = 6.5 Hz, 2 H, SO₂NCH₂), 4.34 (q, J = 7 Hz, 2 H, CO₂CH₂), 6.85-8.05 (m, 10 H, HAr); IR (KBr) 1700 (C==0), 1320 + 1170 + 1140 (SO₂), 1285 (OCO); MS, m/z 479 (M⁺), 415 (M⁺ - SO₂). Anal. (C₂₈H₂₉N₃O₄S) C, H, N, S. C: calcd, 65.11; found, 64.2.

2-[2-(4-Phenyl-1,2,3,6-tetrahydropyridyl)ethyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (33) was prepared in 26% yield as a solid (after flash chromatography on silica gel with dichloromethane as eluent and recrystallization from acetonitrile, mp 106 °C) from 1, triethylamine as base, and 4-phenyl-1,2,3,6tetrahydropyridine: NMR (CDCl₃, 200 MHz) δ 2.64 (m, 2 H, C=CCH₂ cycle), 2.91 (m, 2 H, NCH₂), 3.04 (t, J = 7.5 Hz, 2 H, NCH₂), 3.35 (m, 2 H, NCH₂), 4.1 (t, J = 7.5 Hz, 2 H, SO₂NCH₂), 6.11 (m, 1 H, CH=C), 6.86-8.09 (m, 11 H, HAr); MS, m/z 390 (M⁺). Anal. (C₂₃H₂₂N₂O₂S) C, H, N, S.

2-[4-(4-Phenyl-1,2,3,6-tetrahydropyridyl)butyl]-2*H*naphth[1,8-*cd*]isothiazole 1,1-dioxide (34) was prepared in 18% yield as a solid (after flash chromatography on silica gel with ethyl acetate and recrystallization from acetonitrile, mp 101 °C) from 3, triethylamine as base, and 4-phenyl-1,2,3,6-tetrahydropyridine: NMR (CDCl₃, 200 MHz) δ 1.7 (m, 2 H, CCH₂CC), 2.02 (m, 2 H, CCCH₂C), 2.5-2.8 (m, 6 H, NCH₂ and C=CCH₂ cycle), 3.2 (m, 2 H, NCH₂), 3.9 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.10 (m, 1 H, CH=C), 6.8-8.07 (m, 11 H, HAr); MS, m/z 418 (M⁺), 354 (M⁺ -SO₂). Anal. (C₂₆H₂₆N₂O₂S) C, H, N, S. C: calcd, 71.74; found, 72.3.

2-[2-[4-(4-Fluorophenyl)-1-piperazinyl]ethyl-2H-naphth-[1,8-cd]isothiazole 1,1-dioxide (35) was prepared in 23% yield as a solid (after flash chromatography on silica gel with 80:20 dichloromethane/ethyl acetate as eluent and recrystallization from diisopropyl ether, mp 133 °C) from 1, triethylamine as base, and 1-(4-fluorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 2.34 (t, J = 7 Hz, 2 H, NCH₂), 2.67 (m, 4 H, NCH₂ piperazine), 3.19 (m, 4 H, NCH₂ piperazine), 4.02 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.8-8.1 (m, 10 H, H Ar); MS, m/z 411 (M⁺), 347 (M⁺ - SO₂). Anal. (C₂₂H₂₂FN₃O₂S) C, H, N, S.

2-[4-[4-(4-Fluorophenyl)-1-piperazinyl]butyl]-2Hnaphth[1,8-cd]isothiazole 1,1-dioxide (36) was prepared in 41% yield as a solid (after flash chromatography on silica gel with 50:50 dichloromethane/ethyl acetate and recrystallization from acetonitrile, mp 110 °C) from 3, triethylamine as base, and 1-(4fluorophenyl)piperazine: NMR (CDCl₃, 200 MHz) δ 1.85 (m, 2 H, CCH₂CC), 2.17 (m, 2 H, CCCH₂C), 2.62 (t, J = 7 Hz, 2 H, NCH₂), 2.67 (m, 4 H, NCH₂ piperazine), 3.19 (m, 4 H, NCH₂ piperazine), 3.95 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.8-8.08 (m, 10 H, H Ar); MS, m/z 439 (M⁺), 375 (M⁺ - SO₂). Anal. (C₂₄H₂₆F-N₃O₂S) C, H, N, S.

Preparation of the 4-Phenyl-1,2,3,6-tetrahydropyridine Alkyl Derivatives 6-10. N-[3-(4-Phenyl-1,2,3,6-tetrahydropyridyl)propyl]benz[cd]indol-2(1H)-one (6). To 1.3 g (27 mmol) of sodium hydride (50% dispersion in Vaseline) in 10 mL of DMF, under argon, was slowly added 4.5 g (26.6 mmol) of benz[cd]indol-2(1H)-one in 25 mL of DMF at 25 °C. The reaction mixture was stirred at 100 °C for 30 min and then cooled to room temperature. To this solution was added 9.3 g (39.5 mmol) of 4 in 20 mL of DMF. The mixture was stirred at reflux for 2 h. After evaporation to dryness under reduced pressure, the residue was purified by flash chromatography on silica gel with ethyl acetate as eluent to give 7.4 g (75.5%) of 6 (oxalate, mp 138 °C): NMR (DMSO, 250 MHz) δ 2.19 (m, 2 H, CCH₂C), 2.73 (m, 2 H, C=CCH₂ cycle), 3.18 (m, 2 H, NCH₂), 3.32 (m, 2 H, NCH₂ cycle), 3.77 (m, 2 H, NCH₂ cycle), 4.04 (t, J = 6 Hz, C(0)NCH₂), 6.18 (m, 1 H, CH=C), 7.30-8.23 (m, 11 H, H Ar); IR (KBr) 1700 (CO); MS, m/z 368 (M⁺). Anal. (C₂₅H₂₄N₂O-C₂H₂O₄) C, H, N.

N-[3-(4-Phenyl-1,2,3,6-tetrahydropyridyl)propyl]-N-(1naphthyl)methanesulfonamide (7) was prepared in 37% yield as a solid (after flash chromatography on silica gel with 98:2 dichloromethane/ethanol and recrystallization from ethyl acetate, mp 162 °C) from 4, N-(1-naphthyl)methanesulfonamide, and sodium hydride: NMR (CDCl₃, 250 MHz) δ 1.86 (m, 2 H, CCH₂C), 2.5–2.7 (m, 6 H, NCH₂ and C—CCH₂ cycle), 3.05 (s, 3 H, SO₂CH₃), 3.12 (m, 2 H, NCH₂ cycle), 3.9 (t, J = 7.5 Hz, 2 H, SO₂NCH₂), 6.02 (m, 1 H, CH—C), 7.2–8.22 (m, 12 H, H Ar); IR (KBr) 1335 + 1145 (SO₂); MS, m/z 420 (M⁺). Anal. (C₂₅H₂₈N₂O₂S) C, H, N, S. C: calcd, 71.40; found, 70.7.

N-[3-(4-Phenyl-1,2,3,6-tetrahydropyridyl)propyl]-Nphenylbenzenesulfonamide (8) was prepared in 56% yield as a solid (after flash chromatography on silica gel with 98:2 dichloromethane/ethanol as eluent and recrystallization from diisopropyl ether, mp 112 °C) from 4, N-phenylbenzenesulfonamide, and sodium hydride: NMR (CDCl₃, 250 MHz) δ 1.75 (m, 2 H, CCH₂C), 2.45-2.7 (m, 6 H, CH₂NCH₂CH₂), 3.08 (m, 2 H, NCH₂C=C), 3.68 (t, J = 7 Hz, 2 H, SO₂NCH₂), 6.03 (m, 1 H, CH=C), 7.05-7.65 (m, 15 H, H Ar). Anal. (C₂₈H₂₈N₂O₂S) C, H, N, S.

N-Methyl-*N*-[3-(4-phenyl-1,2,3,6-tetrahydropyridyl)propyl]-1-naphthalenesulfonamide (9) was prepared in 68% yield as an oil (after flash chromatography on silica gel with ethyl acetate, oxalate mp 157 °C) from 4, *N*-methyl-1-naphthalenesulfonamide, and sodium hydride: NMR (oxalate salt) (DMSO, 250 MHz) δ 1.97 (m, 2 H, CCH₂C), 2.73 (m, 2 H, NCH₂ piperidine), 2.87 (s, 3 H, NCH₃), 3.01 (m, 2 H, NCH₂), 3.3 (m, 4 H, NCH₂ piperidine and SO₂NCH₂), 3.74 (m, 2 H, NCH₂C=C), 6.2 (m, 1 H, CH=C), 7.25-8.75 (m, 13 H, H Ar). Anal. (C₂₅H₂₈N₂O₂S· C₂H₂O₄) C, H, N, S.

6-[3-(4-Phenyl-1,2,3,6-pyridyl)propyl]-6*H*-dibenz[*ce*]-1,2thiazine 5,5-dioxide (10) was prepared in 48% yield as an oil (after flash chromatography on silica gel with 98:2 dichloromethane/ethanol as eluent, oxalate mp 170 °C) from 4, 6*H*-dibenz[*ce*]-1,2-thiazine 5,5-dioxide, and sodium hydride: NMR (oxalate salt) (DMSO, 250 MHz) δ 1.94 (m, 2 H, CCH₂C), 2.63 (m, 2 H, NCH₂ piperidine), 2.90 (m, 2 H, NCH₂), 3.11 (m, 2 H, NCH₂ piperidine), 3.55 (m, 2 H, NCH₂C=C), 3.98 (t, *J* = 7 Hz, 2 H, SO₂NCH₂), 6.13 (m, 1 H, CH=C), 7.25-8.35 (m, 13 H, H Ar). Anal. (C₂₆H₂₆N₂O₂S,C₂H₂O₄) C, H, N, S. **Biological Evaluation. Membrane Preparation.** Male

Biological Evaluation. Membrane Preparation. Male Sprague-Dawley rats (200-250 g) were killed by decapitation and their brains rapidly removed on ice. The relevant brain structures were dissected out and homogenized in 10 volumes of ice-cold Tris-HCl buffer (50 mM; pH 7.6). The homogenate was centrifuged for 10 min at 5000g at 4 °C and the supernatant discarded. The pellet was washed by resuspension in seven volumes of the same buffer and recentrifugation. The final pellet was then resuspended in five volumes of Tris-HCl buffer, distributed in 1-mL aliquots in plastic vials (Nunc), and frozen at -80 °C until required. For the histamine receptor preparation, cerebellar membranes were prepared in the same way from male guinea pigs (250 g, Dunkey-Hartley). Protein content was measured according to the BCA method¹² using bovine serum albumin as the standard. $5HT_2$ Binding Assay. The affinity of the different compounds

 $5HT_2$ Binding Assay. The affinity of the different compounds for $5HT_2$ receptors was assessed by using the [³H]ketanserin binding assay described by Leysen et al.⁶ Aliquots (1 mL) of cortical membranes at a protein concentration of 0.15 mg mL⁻¹ in Tris-HCl buffer containing bovine serum albumin, 2 mg/mL, were incubated for 15 min at 37 °C with the compound of interest or with 10⁻⁶ M methysergide (to define the nonspecific binding) in the presence of [³H]ketanserin (final concentration 0.4 nm). The binding interaction was terminated by filtration across Whatman GF/B glass fiber filters using a Skatron cell harvester and followed by two washes with 2.5 mL of ice-cold buffer. The radioactivity retained on the filters was determined by liquid scintillometry in 4.5 mL of Ready-Gel scintillant (Beckman).

The specific binding was defined as that displaceable by 10^{-6} M methysergide. The percentage of the specific binding obtained with each concentration of the test compound was calculated and the IC₅₀ (concentration that inhibits 50% of the specific binding) determined by nonlinear regression analysis using an iterative

⁽¹²⁾ Smith, P. K.; Krohn, R. I.; Hermanson, G. T.; Mallia, A. K.; Gartner, F. H.; Provenzano, M. D.; Fujimoto, E. K.; Gocke, N. M.; Olson, B. J.; Klenk, D. C. "Measurement of protein with bicinchoninic acid", Anal. Biochem. 1985, 150, 76-85.

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curve-fitting procedure to a simple Langmuir isotherm performed on an IBM-PC computer.¹³

5HT_{1A} Binding Assay. The 5HT_{1A} binding assay was performed by the method described by Hall et al.,⁹ using [³H]-8-OH-DPAT as the radioligand and rat hippocampal membranes as the tissue source. Aliquots (0.5 mL) of hippocampal at a protein concentration of 1 mg mL⁻¹ in Tris-HCl buffer were incubated at 37 °C with the compound of interest or with 10⁻⁵ M serotonin. After 5 min, [³H]-8-OH-DPAT (final concentration 1 nM) was added and the incubation continued for a further 10 min. The binding interaction was terminated by filtration as above. The specific binding was defined as that displaceable by 10⁻⁶ serotonin.

 α_1 -Adrenoceptor Binding Assay. This assay was carried out according to the method described by Greengrass and Bremner,⁷ using [³H]prazosin as the radioligand and rat cerebrocortical membranes as the tissue source. Aliquots (1.2 mL) of cortical membranes at a protein concentration of 0.15 mg mL⁻¹ in Tris-HCl buffer containing bovine serum albumin, 2 mg/mL, were incubated for 40 min at 25 °C with the compound of interest or with 10⁻⁶ M phentolamine in the presence of [³H]prazosin (final concentration 0.065 nM). The binding interaction was terminated by filtration as above. The specific binding was defined as that displaceable by 10⁻⁶ M phentolamine.

 D_2 -Dopamine Receptor Binding Assay. Binding to D_2 dopamine receptors was evaluated by using the binding of [³H]spiperone to rat striatal membranes as described by Urwyler and Coward.⁸

Aliquots (1.2 mL) of striatal membranes at a protein concentration of 0.05 mg mL⁻¹ in Tris-HCl buffer containing bovine serum albumin, 2 mg/mL, and NaCl (150 nM), cinanserin (10^{-6} M), and 10^{-6} M sulpiride. After 5 min, [³H]spiroperidol (final concentration 0.05 nM) was added and the incubation continued for a further 40 min. The binding interaction was terminated by filtration as above. The specific binding was defined as that displaceable by 10^{-6} M sulpiride.

 H_1 -Histamine Receptor Binding Assay. The method described by Chang et al.¹⁰ using [³H]mepyramine binding to guinea pig cerebellar membranes was used to assess activity at H_1 -histamine receptors.

Aliquots (1.2 mL) of cerebellar membranes at a protein concentration of 0.35 mg mL⁻¹ in sodium phosphate buffer (50 mM; pH 7.5) were incubated for 30 min at 25 °C with the compound of interest or with 10⁻⁶ M promethazine and with [³H]mepyramine (final concentration 0.7 nM). The binding interaction was terminated by filtration as above. The specific binding was defined as that displaceable by 10⁻⁶ M promethazine.

Mescaline-Induced Head Twitches. In mice,¹⁴ drugs (6

(14) Corne, S. J.; Pickering, R. W. "A possible correlation between drug-induced hallucinations in man and a behavioural response in mice", Psychopharmacologia 1967, 11, 65-78. animals/dose) were administered 60 min (sc) or 90 min (po) before 50 mg kg⁻¹ ip of mescaline. Fifteen minutes after dosing with mescaline, mice were observed during a 2-min period, and the presence or the absence ("all or none" response) of head twitches was noted. The ED₅₀ was defined as the dose of the test compound that protected 50% of mice from head twitches.

In rats, drug activity was studied by using the method described by Niemegers and al.¹¹ The rats (6 animals/dose) were treated with the test compound and 1 h later received an iv dose (20 mg kg⁻¹ of mescaline). Head twitches were counted over a period of 15 min starting immediately after the mescaline injection. The occurrence of less than 3 head twitches was selected as the criterion for drug-induced antagonism. The ED₅₀ was defined as the dose of the drug which protected 50% of rats.

The duration of action was assessed in this method, by treating the rats with the compound to be tested 6 and 16 h (both sc and po) before mescaline. The ED_{50} values (50% of protected rats) were calculated at each time point.

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Registry No. 1, 127625-84-7; 2, 127625-83-6; 3, 127625-85-8; 4, 84404-69-3; 5, 127625-30-3; 6, 134133-51-0; 6 oxalate, 134133-66-7; 7, 134133-52-1; 8, 134133-53-2; 9, 134133-54-3; 9 oxalate, 134133-68-9; 10, 134133-55-4; 10-oxalate, 134133-67-8; 11, 127625-35-8; 12, 127625-36-9; 13, 127625-37-0; 14, 134133-56-5; 15, 127625-59-6; 16, 127625-34-7; 17, 134133-57-6; 18, 127648-05-9; 19, 134133-58-7; 20, 134133-59-8; 21, 134133-60-1; 22, 127625-57-4; 23, 127625-56-3; 24, 127625-29-0; 25, 127625-38-1; 26, 127625-41-6; 27, 127648-03-7; 28, 134133-61-2; 29, 127625-58-5; 30, 134133-62-3; 31, 127625-54-1; 32, 134133-63-4; 33, 127625-39-2; 34, 127625-43-8; 35, 134133-64-5; 36, 134133-65-6; 1,8-naphthosultam, 603-72-5; 1-bromo-2-chloroethane, 107-04-0; 1-phenylpiperazine, 92-54-6; 4-phenylpiperazine, 771-99-3; 4-(4-fluorobenzoyl)piperidine, 56346-57-7; 4-[bis(4-fluorophenyl)methylene]piperidine, 58113-36-3; 1-(4-piperidinyl)-2-benzimidazolinone, 20662-53-7; 1-(2pyridyl)piperazine, 34803-66-2; 1-(2-pyrimidinyl)piperazine, 20980-22-7; 4-(1,2-benzisoxazol-3-yl)-1-piperazine, 87691-89-2; 4-(1,2-benzisothiazol-3-yl)-1-piperazine, 87691-87-0; 4-(1,1-dioxo-1,2-benzisothiazol-3-yl)-1-piperazine, 131540-88-0; 1-(2benzothiazolyl)piperazine, 55745-83-0; 1-(2-fluorophenyl)piperazine, 1011-15-0; 1-(3-fluorophenyl)piperazine, 3801-89-6; 1-(4-fluorophenyl)piperazine, 2252-63-3; 1-(4-chlorophenyl)piperazine, 38212-33-8; 1-(4-bromophenyl)piperazine, 66698-28-0; 1-(4-hydroxyphenyl)piperazine, 56621-48-8; 1-(4-methoxyphenyl)piperazine, 38212-30-5; 1-(4-methylphenyl)piperazine, 39593-08-3; 1-[4-(trifluoromethyl)phenyl]piperazine, 30459-17-7; 1-(4-nitrophenyl)piperazine, 6269-89-2; 1-[4-(ethoxycarbonyl)phenyl]piperazine, 80518-57-6; 4-phenyl-1,2,3,6-tetrahydropyridine, 10338-69-9; benz[cd]indol-2(1H)-one, 130-00-7; N-(1naphthyl)methanesulfonamide, 53715-52-9; N-phenylbenzenesulfonamide, 1678-25-7; N-methyl-1-naphthalenesulfonamide, 71862-34-5; 6H-dibenz[ce]-1,2-thiazine 5,5-dioxide, 1864-33-1.

⁽¹³⁾ McPherson, G. A. "Analysis of radioligand binding experiments. A collection of computer programs for the IBM PC", J. Pharmacol. Meth. 1985, 14, 213-228.