Amino- and Guanidinoacylryanodines: Basic Ryanodine Esters with Enhanced Affinity for the Sarcoplasmic Reticulum Ca²⁺-Release Channel

Koert Gerzon,^{*,†} Rod A. Humerickhouse,[†] Henry R. Besch, Jr.,[†] Keshore R. Bidasee,[†] Jeffrey T. Emmick,[†] Roger W. Roeske,[†] Zhenping Tian,[†] Luc Ruest,[‡] and John L. Sutko[§]

Department of Pharmacology and Toxicology and Department of Biochemistry and Molecular Biology, Indiana University, School of Medicine, Indianapolis, Indiana 46202, Departement de Chimie, University of Sherbrooke, Sherbrooke, Quebec J1K2R1, Canada, and Department of Pharmacology, University of Nevada, Reno School of Medicine, Reno, Nevada 89537

Received January 14, 1993

Amino- and guanidinoacyl esters of ryanodine were prepared to evaluate the effect of basicity on the binding affinity of these derivatives for the sarcoplasmic reticulum Ca²⁺-release channel (SR CRC). In the presence of DCC and DMAP Cbz- β -alanine reacts with ryanodine in CH₂Cl₂ to give O_{10eq} -Cbz- β -alanylryanodine (3a), which on hydrogenolysis yields the β -alanyl ester (4a). N,N'bis-Cbz-S-methylthiourea reacts with 4a to yield β -N,N'-bis-Cbz-guanidinopropionylryanodine (5a). O_{10ea} - β -guanidinopropionylryanodine (6a) is obtained on hydrogenolytic deprotection of 5a. The binding affinity of β -alanine ester (4a) and its glycyl congener (4b) is 2-3-fold greater, and that of the β -guanidinopropionyl ester (6a) and its acetyl congener (6b) 3–6-fold greater, than that of ryanodine. The effect of ryanodine on SR Ca²⁺ flux is of a biphasic nature: nanomolar levels open (activate) the channel, while micromolar levels close (deactivate) it. The base-substituted esters 4a and 6a both display a unidirectional effect: they only open the channel. An understanding of ryanodine's mode of action and the design of effective SR CRC activating and deactivating ryanoids for possible therapeutic application are major research objectives.

Chemical alteration of natural products¹⁻⁵ at times leads to *improvement*, only rarely—as in the case of Etoposide⁵—to innovation of therapeutic effectiveness. The latter achievement, indeed, presents a paradigm.

In recent years the two principal bioactive alkaloids, ryanodine $(1)^6$ and 9,21-dehydroryanodine (2),⁷ isolated from the tropical shrub Ryania speciosa Vahl (Figure 1), have been studied extensively because of their characteristic role as modulators of the sarcoplasmic reticular Ca²⁺-release channel (SR CRC, ryanodine receptor) of striated muscle;^{8,9} their insecticidal activity and mammalian toxicity had previously been established.^{9a,10}

Evidence has very recently been presented for the presence of ryanodine receptors in brain¹¹ and other cells,¹² including neutrophils.^{12c} It appears then a worthy challenge to attempt and translate the complex, high-potency in vitro actions and the in vivo toxicity9a,10 of these ryanoids into therapeutic effectiveness for cardiac disease or perhaps alternate pathologies. Clearly, the semisynthesis of altered rvanodines¹³ and evaluation of their in vitro pharmacological activity is an attractive starting point. Determining the relative binding affinity (RBA) of these ryanoids may assist in mapping the binding domain. In addition, some of these derivatives may serve as molecular probes.¹⁴ for example in affinity chromatography^{14a} and in photoaffinity labeling^{14b} of the ryanodine receptor. Also, the search for putative endogenous effectors of this receptor in the mammalian system will be facilitated by the availability of suitable derivatives (vide infra).

To explore the effect of base substitution on ryanoid binding we availed ourselves of ryanodine's accessible, secondary C10-hydroxyl function¹⁰ to prepare O_{10eq}-aminoand -guanidinoacyl esters.^{14b,c} Here we report the semisynthesis and preliminary pharmacological evaluation of O_{10ec} - β -alanyl- (4a), -glycyl- (4b), - β -guanidinopropionyl-

(6a), -guanidinoacetylryanodine (6b), the Cbz-protected intermediates (3a, 3b, 5a, 5b) and the anhydro-ryanodine ester (7). Some of these esters have been shown to possess enhanced affinity for the receptor, relative to ryanodine (1), and to display a unidirectional effect on Ca^{2+} -release channels.

Chemistry

Basic esters (4a, 4b, 6a, 6b) of ryanodine (1) were prepared according to Scheme I. O_{10eq} -Cbz- β -alanylryanodine $(3a)^{13b,c}$ was synthesized from 1 and N-Cbz- β alanine by the method of Neises and Steglich.¹⁵ Hydrogenolysis of 3a in the presence of Et₃N gives O_{10eq} - β alanylryanodine (4a), 13b,c which reacts with N, N'-bis-Cbz-S-methylthiourea¹⁶ under mild conditions to yield O_{10eo} - β -N,N'-bis-Cbz-guanidinopropionylryanodine (5a). Hydrogenolysis (Et₃N) of **5a** affords O_{10eq} - β -guanidinopropionylryanodine (6a).

In the presence of aqueous HCl hydrogenolysis of 3a, in addition to ester 4a, yields a byproduct characterized





0022-2623/93/1836-1319\$04.00/0

© 1993 American Chemical Society

[†] Indiana University.

[‡] University of Sherbrooke. [§] University of Nevada.

Scheme I. Semisynthesis of O_{10eq} -Guanidino- and -Aminoacyl Esters of Ryanodine



as β -alanyl-anhydro-ryanodine (7).

The preparation of the guanidinoacetyl ester (6b) similarly proceeds from 1 and Cbz-glycine via O_{10eq} -Cbz-



glycyl- (3b), O_{10eq} -glycyl- (4b), and O_{10eq} -N,N'-bis-Cbzguanidinoacetylryanodine (5b); hydrogenolysis (Et₃N) of 5b affords the O_{10eq} -guanidinoacetate of 1 (6b).

The basic esters 4 and 6 of 1 are shown to be of pharmacological interest (vide infra); in view of the relatively greater abundance of dehydroryanodine (2),^{8,14b} preparation of the corresponding basic esters of 2 is now underway.¹⁷ Compared to 1 and 2, other known natural ryanoids,¹⁸ the less polar "ryania diterpene esters" A, B, C_1, C_2 , and D, ^{18a} the more polar "Ryania diterpene esters" E and F, ^{18b,c} ryanodyl 3-pyridine-3-carboxylate,^{18d} and 18-hydroxyryanodine¹⁰ are minor constituents and are not readily available for chemical alteration.

Pharmacological Results and Discussion

A cursory comparison of the relative binding affinity (RBA) of the esters 3-6 with that of 1 (Table I) reveals a 3-6-fold enhanced affinity of the basic esters 4 and 6, a

Table I. Relative Binding Affinity of Ryanoids and O_{10eq} -Ryanodine Esters for the Sarcoplasmic Reticular Ca²⁺-Release Channel^a

_		$IC_{50} (nM) \pm SD$
1	ryanodine	6.2 ± 0.4
2	dehydroryanodine	8.9 ± 1.2
	O _{10ec} -Ryanodine esters	
3a	$Cbz-\beta$ -alanyl ^b	5.9
3b	Cbz-glycyl	4.8 ± 0.6
4 a	β -alanyl	2.6 ± 0.4
4b	glycyl	1.8 ± 0.4
5a	β -N,N'-bis-Cbz-guanidinopropionyl ^b	43.6
5b	N.N'-bis-Cbz-guanidinoacetyl ^b	135.6
6a	β -guanidinopropionyl	1.1 ± 0.1
6b	guanidinoacetyl	1.8 ± 0.2
	Ry-hemi-succinate	>1000.0
	N-methyl Ry-succinamidate (CH ₂ HNOCCH ₂ CH ₂ COO ₁₀ -Ry)	49.4 ± 2.7
7	β -alanyl-anhydro-ryanodine ^b	149.0
	anhydro-ryanodine ^c	>1000.0

^a IC₅₀ is the concentration of the unlabeled ryanodine or ryanodine ester at which 50% of the high affinity binding sites on the skeletal SR CRC/Ry receptor are occupied. ^b Single experiment. ^c Reported for anhydro-Ryanodine: IC₅₀ > 10 μ M. (Waterhouse, et al. J. Med. Chem. 1987, 30, 710–716. ^d Gerzon, et al. Reg. Meet. Am. Chem. Soc., Indianapolis, 1991, Abstr. MEDI 342.

slightly enhanced affinity of the mono-Cbz protected ester 3, and a 7-20-fold lesser affinity of the di-Cbz protected esters 5a and 5b.

The enhanced binding affinity of the positively charged esters 4 and 6 to the receptor indicates that an anionic function may be proximate to the ryanodine binding site. That a negative charge does reside adjacent to the C-10 region is supported by the minimal binding affinity (IC₅₀ > 1.0 μ M) noted for the negatively charged O_{10eq} -succinate of 1, -OOCCH₂CH₂COO₁₀-ryanodine.^{13a,c,19} Furthermore, CH₃HNOCCH₂CH₂COO₁₀-ryanodine, the corresponding, noncharged N-methylamide, regains receptor affinity; this

Amino- and Guanidinoacylryanodines

amide exhibits 24% of the affinity of 1. While the enhanced binding of the base-substituted esters (4a,b and 6a,b) implicates a negative locus adjacent to the binding site, adequate binding of the mono-Cbz esters (3a,b) reveals a degree of steric tolerance at the region of the C10equatorial hydroxyl. A low RBA of the two bulky bis-Cbz-guanidino esters (5a,b) suggests this steric tolerance to be of limited scope. Clearly, the RBA relationships among the ryanodines (1 and 2) and esters 3-6 reflects, among others, the effect of basicity and/or dimension (amino vs guanidino, n = 1 vs n = 2) of the substituent. To be able to assign a relative contribution factor to the one or the other parameter a larger series of derivatives would be necessary.

Ryanodine exhibits two opposing effects on the sarcoplasmic reticular Ca²⁺-release channel (SR CRC).^{8,9} Measuring the passive flux of ${}^{45}Ca^{2+}$ across the SR CRC (see the Experimental Section) allows quantitation of Ca²⁺ efflux from skeletal SR vesicles as a means of assessing both opening (activating) and closing (deactivating) effects of 1 and its esters in a single assay.^{8,9,18c} In this assay 1 activates the SR CRC at low micromolar concentrations, activation reaching its maximum at about 30 μ M; concentrations higher than 30 μ M induce deactivation of the channel. An altered-unidirectional-pharmacological profile, differing from the biphasic profile of 1, was observed for esters 4a and 6a. The activation curve for the β -alanyl ester (4a) is shifted to the left-toward lower concentrations-of that of 1, maximal activation being reached at a concentration of 10 μ M. This 3-fold-lowered shift parallels the 2-3-fold greater RBA for this ester (4a).

A perhaps more significant aspect, however, of the interaction of the β -alanyl ester (4a) with the receptor involves its overall effect on the channel.²⁰ Whereas 4a, like 1, induces full opening of the SR CRC, higher concentrations of 4a, up to 1 mM, exhibit none of the closing action seen with high concentrations of 1. In preliminary experiments the guanidino ester (6a), which has an RBA for the receptor about 6 times greater than that of 1, also appears to be purely an activator of the SR CRC.²⁰ Thus, while the base-substituted esters (4a, 6a) retain the ability to open this channel, they entirely lack the ability of 1 to close the SR CRC.

Whether the biphasic pharmacological activity profile of 1 can be attributed (a) to 1 interacting at two different, independent binding sites,^{8,9f} (b) to different conformations of the ryanodine molecule interacting at a single binding site,^{14b} or (c) to a combination of these (and perhaps other) factors^{18c} are questions remaining to be answered in fiture studies. The selective channel activator action of the basesubstituted esters 4a and 6a at the SR CRC may help to elucidate the mechanism of ryanodine's action.

Work in our and other laboratories pursues the chemical alteration of the ryanodines. In addition to the present C10-hydroxyl esters, the C9–C21 double bond in dehydroryanodine,^{18b} and the pyrrolecarboxylic acid moiety^{18d} are among the sites at which chemical alteration possibly may yield pharmacologic information. An understanding of ryanodine's mode of action and the design of effective SR CRC activating and deactivating agents for possible therapeutic application remain major research objectives.

Experimental Section

Melting points (uncorrected) were determined in open capillary tubes with a Mel-Temp apparatus, Laboratory Devices. Elemental analysis was performed by Midwest Micro Lab, Indianapolis, IN. Proton magnetic resonance (¹H NMR) spectra were obtained in CD_3OD or $CDCl_3$ solution on a Bruker AM 250 instrument; the proton chemical shift values are reported in ppm (δ) relative to internal tetramethylsilane. Mass spectra were determined on Vacuum Generators' VG CAB CSE instruments using glycerol with lithium iodide, giving ions (M⁺ + 7), except for the guanidino ester derivatives (**6a**,**b**) which give M⁺ + 1 ions. Analytical TLC was performed on 0.25-mm silica gel plates (Merck, Kieselgel 60, 230–400 mesh) using system CMaM (CHCl₃/CH₃OH/aqueous 40% CH₃NH₂, 98/2/0.2) or, for the guanidino esters, system BAWP (1-butanol/acetic acid/water/pyridine, 4/1/2/1, by volume).^{16b}

 O_{10eq} -Cbz- β -alanylryanodine (3a) was prepared by a modification of the method of Neises and Steglich.¹⁵ A stirred CH₂-Cl₂ solution containing equimolar amounts of ryanodine (1), Cbz- β -alanine, and DCC with catalytic amounts of DMAP was allowed to react for 8 h at 4 °C. Termination of the reaction with H_2O and column chromatography (system CMaM) gave a 75% yield of the crystalline, protected ester (3a), mp 178-180 °C. TLC: R_f (CMaM) = 0.5. MS (FAB, LiI): M⁺ + 7 = 705. Calcd $(\dot{C}_{36}H_{46}N_2O_{12})$ mol. wt. = 698. ¹H NMR (CD₃OD): δ 7.32 (m, 5 H, phenyl aromatic protons), 7.06, 6.87, 6.23 (three double doublets, pyrrole hydrogens), 5.58 (s, 1 H, H-C3), 5.31 (d, 1 H, H_{ax} -C10), 5.07 (s, 2 H, PhC H_2 O), 3.45 (t, 2 H, NHC H_2), 2.59 (m, 1 H, H-C13), 1.40 (s, CH₃-C1), 1.11 (d, 3 H, CH₃-C13), 0.90 (s, 3 H, CH₃-C5), 0.82 (d, 3 H, CH₃-C9), and 0.74 (d, 3 H, CH₃-C13). The designation of the product (3a) as the O_{10eq} -ryanodine ester (3a) is supported by the observation that the doublet for the C_{10ax} -hydrogen, present in the spectrum of 1 at 3.94 ppm, in the above spectrum of 3a is shifted downfield to 5.31 ppm (cf. ref 10).

 O_{10eq} -β-Alanylryanodine (4a). Ester 3a was hydrogenolyzed under hydrogen (40 lb/in.²) with Pd-C (10%) in ethanolic solution containing equimolar Et₃N. Filtration of the catalyst, evaporation of the solvents under reduced pressure, and trituration of the solid residue with a pentane/ethyl ether mixture (9/1) yields crystalline β-alanylryanodine (4a), mp 182-184 °C. TLC (CMaM): $R_f = 0.22$. MS (FAB, LiI): M⁺ + 7 = 571. Calcd (C₂₈H₄₀N₂O₁₀) mol. wt. = 564. ¹H NMR (CD₃OD): δ 7.03, 6.87, and 6.23 (three double doublets; pyrrole hydrogens), 5.58 (s, 1 H, HC3), 5.4 (d, 1 H, HC10), 3.0 (t, 2 H, H₂NCH₂), 2.60 (t, 2 H, CH₂CO), 2.56 (d, H_b) and 1.94 (d, H_a) (AB pattern, H₂C14), 2.26 (m, 1 H, HC13), 2.10 (m, 1 H, HC9), 1.40 (s, 3 H, CH₃-C1), 1.03 (d, 3 H, CH₃-C13), 0.89 (s, CH₃-C5), 0.85 (d, 3 H, CH₃-C9), and 0.74 (d, 3 H, CH₃-C13).

O_{1θeq}-β-N,N-Bis-Cbz-guanidinopropionylryanodine (5a). The β-alanyl ester (4a, 100 mg, 0.17 mmol) in a stirred DMF solution (1 mL) containing Et₃N (0.2 mmol) was allowed to react with N,N'-bis-(Cbz)-S-methylisothiourea¹⁶ (200 mg, 0.55 mmol) at rcom temperature for 18 h. Removal of DMF under reduced pressure (0.1 mmHg) at 35 °C and column chromatography of the oily residue on SILICAR gel (system CMaM) gave the ester 5a (100 mg). TLC (CMaM): $R_f = 0.72$. MS (FAB, LiI) M⁺ + 7 = 881. Calcd (C₄₅H₅₄N₄O₁₄) mol. wt. = 874. ¹H NMR (CD₃OD) relevant peaks: δ 7.5–7.3 (m, 10 H, 2 Ph groups), 5.6 (s, 1 H, H-C3), 5.35 (d, 1 H, H-C10), 5.26 (s, 2 H, ArCH₂O), 5.12 (s, 2 H, PhCH₂O), 3.72 (m, 2 H, CH₂NH), 2.70 (m, 2 H, CH₂CO).

 O_{10eq} - β -Guanidinopropionylryanodine (6a). Hydrogenolysis of 5a (75 mg) with Pd-C (10%) in ethanol containing Et₃N, as described above for the hydrogenolysis of 3a, followed by column chromatography (System CCaM), gave the crystalline guanidino ester (6a, 35 mg). TLC (BAWP): $R_f = 0.73$. MS (FAB, LiI): M⁺ + 1 = 607. Calcd (C₂₉H₄₂N₄O₁₀) mol. wt. = 606. Anal. Calcd C, 57.43; H, 6.93; N, 9.24. Found; C, 51.78; H, 7.05; N, 8.29.

O_{10eq}-β-Alanyl-anhydro-ryanodine (7). Cbz-β-alanylryanodine (3a, 0.1 mmol) was hydrogenolyzed using Pd-C (10%) in EtOH containing HCl (1 mL, 0.1 N). Filtration of the catalyst and evaporation of solvents gave a mixture of β-alanyl-anhydroryanodine (7) and β-alanylryanodine (4a) as the HCl salts. Conversion to the free bases (aqueous Na₂CO₃, CHCl₃) and column chromatography on SILICAR using CHCl₃ and then CHCl₃/CH₃OH/aqueous 40% CH₃NH₂ mixtures (92/2/0.2, 96/ 4/0.4, and 94/6/0.6), gave β-alanyl-anhydro-ryanodine (7) and ester 4a. TLC (CMaM): $R_f = 0.4$. MS (FAB, Li1): M⁺ + 7 = 553. Calcd (C₂₈H₃₈N₂O₉) mol. wt. = 546. ¹H NMR (CD₃OD): δ 7.06, 6.87, 6.26 (three doublets for pyrrole hydrogens), 6.17 (q, 1 H, HC3), 5.63 (d, 1 H, HC10), 3.42 (d, H_b) and 2.59 (d, H_a) (AB pattern, H_2 C14), 2.71 (m, 1 H, HC13), 3.07 (t, 2 H, H_2 NC H_2), 2.61 (t, 2 H, CH_2 CO), 2.05 (m, 1 H, HC9), 1.82 (d, 3 H, CH_3 C1), 1.1 (d, 3 H, CH_3 -C13), 0.99 (d, 3 H, CH_3 -C9), 0.96 (s, 3 H, CH_3 -C5), 0.91 (d, 3 H, CH_3 -C13).

Designation of this byproduct as O_{10eq} - β -alanyl-anhydroryanodine (7) is based on the observation that the C3 proton, present as a singlet in the spectrum of the β -alanyl ester (4a) at 5.58 ppm, in that of the β -alanine anhydro-ester (7) appears as a quartet at 6.17 ppm. This C3 proton appears in the spectrum of 1 as a singlet at 5.63 ppm and as a quartet at 6.17 ppm in that of anhydro-ryanodine;^{6b} a similar C3 proton shift obtains between ryanodol (singlet at 4.11 ppm) and anhydro-ryanodol (quartet at 4.70).^{6d}

Cbz-glycylryanodine (3b). This ester was prepared from Cbz-glycine and 1, analogously to **3a**. TLC (CMaM): $R_f = 0.46$. MS (FAB, glycerol): M⁺ + 23 = 707.3. Calcd (C₃₅H₄₄N₂O₁₂) mol. wt. = 684. ¹H NMR (CDCl₃): δ 5.40 (d, 1 H, C10-H), 5.14 (s, 2 H, PhCH₂O).

Glycylryanodine (4b). Ester 4b was prepared by hydrogenolysis of 3b in the presence of Et₃N. TLC (CMaM): $R_f =$ 0.23. Calcd for C₂₇H₃₈N₂O₁₀ mol. wt. = 550. No physicochemical data available because of small sample size.

Di-Cbz-guanidino acetylryanodine (5b). Ester 5b, analogously to 5a, was obtained from the reaction of 4b and N,N'-bis-Cbz-S-methylisothiourea.¹⁶ TLC (CMaM): $R_f = 0.6$. Calcd (C₄₄H₅₂N₄O₁₄) mol. wt. 860. No physicochemical data due to small sample size.

 O_{10eq} -Guanidinoacetylryanodine (6b). Hydrogenolysis of 5b in the presence of Et₃N, as in the preparation of 6a, gave 6b. TLC (BAWP): $R_f = 0.8$. MS (FAB, LiI): M⁺ + 1 = 593. Calcd (C₂₈H₄₀N₄O₁₀) mol. wt. = 592. While the amino esters (4a,b) peak with lithium iodide in the mass spectrum at M⁺ + 7, the more basic guanidino esters (6a and 6b) peak at M⁺ + 1.

Pharmacological Methods. The relative binding affinity (RBA) of ryanodine (1), dehydroryanodine (2) and esters 3–7 listed in Table I were determined using a competition assay, as described.^{8,9b,c,18c} Briefly, rabbit skeletal SR membrane vesicles^{9c-f} were incubated in the presence of 6.7 nM tritiated ryanodine and increasing concentrations of the unlabeled esters, to competitively displace the [³H]ryanodine. The IC₅₀ value of each of the esters was determined from the respective displacement curve using the computer binding analysis program EBDA/ligand.²¹

Passive flux of ${}^{45}Ca^{2+}$ through the SR Ca^{2+} -release channel^{9e,f} was used to evaluate the ability of the basic esters **4a** and **6a** to open (activate) and close (deactivate) the channel. Briefly, junctional SR vesicles from rabbit skeletal muscle were passively loaded with ${}^{45}Ca^{2+}$ in the presence and in the absence of the esters. Ca^{2+} efflux was then initiated by diluting the vesicles 100-fold into a low Ca^{2+} concentration. ${}^{45}Ca^{2+}$ remaining in the vesicles was determined after a 3-s efflux period by termination and filtration of the vesicles followed by scintillation counting.

Acknowledgment. The authors gratefully acknowledge the valuable support given by Dr. John Occolowitz, the Lilly Research Laboratories, in providing the mass spectrographic evaluation of several of the esters reported here and by Sangyeol Kwon in the preparation of the SR membrane vesicles. We thank Tracy Welty and Bruce Henry for excellent help with the manuscript and Phil Wilson and Gary Schmitt for valuable assistance with the illustrations. Kurt Besch is gratefully remembered for providing the pure ryanodine used. This work was supported in part by a Glaxo Cardiovascular Discovery Grant and by the Showalter Trust.

References

- Flynn, E. H. Cephalosporins and Penicillins. Chemistry and Biology; Academic Press: New York, 1972.
- (2) Kirst, H. A.; Sides, G. D. New Directions for Macrolide Antibiotics; Structural Modifications and in vitro Activity. Antimicrob. Agents Chemother. 1989, 33, 1413–1418. Pharmacokinetics and Clinical Efficacy. Ibid. 1989, 33, 1419–1422.
- Efficacy. *Ibid.* 1989, 33, 1419–1422.
 (3) Arcamone, F. The Development of New Antitumor Anthracyclines, In Anticancer Agents Based on Natural Product Models, Cassidy, J. M., Douros, J. D., Eds.; Academic Press: New York, 1980; Chapter 1, pp 1–38.

- (4) Gerzon, K. Dimeric Catharanthus Alkaloids. In Anticancer Agents Based on Natural Product Models; Cassidy, J. M., Douros, J. D., Eds.; Academic Press: New York, 1980; Chapter 8, pp 271-317.
- (5) Stähelin, H. F.; von Wartburg, A. The Chemical and Biological Route from Podophyllotoxin Glucoside to Etoposide. *Cancer Res.* 1991, 51, 5-15.
- (6) (a) Rogers, E. F.; Koniuszy, F. R.; Shavel, J., Jr.; Folkers, K. Plant Insecticides. I. Ryanodine. A New Alkaloid from Ryania Speciosa Vahl. J. Am. Chem. Soc. 1948, 70, 3086-3088. (b) Kelly, R. B.; Whittingham, D. J.; Wiesner, K. The Structure of Ryanodine. Can. J. Chem. 1951, 29, 905-911. (c) Wiesner, K. The Structure of Ryanodine. Adv. Org. Chem. 1972, 8, 295-316. (d) Deslongchamps, P.; Ruest, L.; et al. The Total Synthesis of (+)-Ryanodol. I-IV. Can. J. Chem. 1990, 68, 115-192.
- (7) Waterhouse, A. L.; Holden, I.; Casida, J. E. 9,21-Didehydoryanodine: a New Principal Toxic Constituent of the Botanical Insecticide Ryania. J. Chem. Soc., Chem. Commun. 1984, 1265– 1266.
- (8) Jones, L. R.; Besch, H. R., Jr.; Sutko, J. L.; Willerson, J. T. Ryanodine-induced Stimulation of Ca²⁺ Uptake by Cardiac Sarcoplasmic Reticulum Vesicles. J. Pharmacol. Exp. Ther. 1979, 209, 48-55.
- (a) Jenden, D. J.; Fairhurst, A. S. The Pharmacology of Ryanodine. (9) Pharmacol. Rev. 1969, 21, 1-25. (b) Sutko, J. L.; Willerson, J. T.; Templeton, G. H.; Jones, L. R.; Besch, H. R., Jr. Ryanodine: Its Alterations of Cat Papillary Muscle Contractile State and Responsiveness to Inotropic Interventions and a Suggested Mechanism of Action. J. Pharmacol. Exp. Ther. 1979, 209, 37-47. (c) Besch, H. R., Jr.; Jones, L. R.; Watanabe, A. M. Intact Vesicles of Canine Cardiac Sarcolemma: Evidence from Vectorial Properties of Na+,K+-ATPase. Circ. Res. 1979, 39, 586-589. (d) Fleischer, S.; Ogunbunmi, E. M.; Dixon, M. C.; Fleer, E. A. M. Localization of Ca²⁺ Release Channels with Ryanodine in Junctional Terminal Cisternae of Sarcoplasmic Reticulum of Fast Skeletal Muscle. Proc. Natl. Acad. Sci. U.S.A. 1985, 82, 7256–7259. (e) Meissner, G. Ryanodine Activation and Inhibition of the Ca²⁺-Release Channel of Sarcoplasmic Reticulum. J. Biol. Chem. 1986, 261, 6300-6306. (f) Lattanzio, F. A., Jr.; Schlatterer, R. G.; Nicar, M.; Campbell, K. P.; Sutko, J. L. The Effects of Ryanodine on Passive Calcium Fluxes across Sarcoplasmic Reticulum Membranes. J. Biol. Chem. 1987, 262. 2711-2718.
- (10) Waterhouse, A. L.; Pessah, I. N.; Francini, A. O.; Casida, J. E. Structural Aspects of Ryanodine Action and Selectivity. J. Med. Chem. 1987, 30, 710-716.
- (11) (a) Ashley, R. H. Activation and Conductance Properties of Ryanodine-Sensitive Calcium Channels from Brain Microsomal Membranes Incorporated into Planar Lipid Bilayers. J. Membrane Biol. 1989, 111, 179-189. (b) McPherson, P. S.; Campbell, K. P. Solubilization and Biochemical Characterization of the High Affinity ³H-Ryanodine Receptor from Rabbit Brain Membranes. J. Biol. Chem. 1990, 265, 18454-18460. (c) Walton, P. D.; Airey, J. A.; Sutko, J. L.; Beck, C. F.; Mignery, G. A.; Sudhof, T. C.; Deerinck, T. J.; Ellisman, M. H. Ryanodine and Inositol Triphosphate Receptors Co-exist in Avian Cerebellar Purkinje Neurons. J. Cell Biol. 1991, 113, 1145-1157. (d) Witcher, D. H.; Strifler, B. A.; Jones, L. R. Cardiac-specific Phosphorylation Site for Multifunctional Ca²⁺/Calmodulin-dependent Protein Kinase is Conserved in the Brain Ryanodine Receptor. J. Biol. Chem. 1992, 267, 4963-4967.
- (12) (a) Shoshan-Barmatz, V.; Pressley, T. A.; Higham, S.; Kraus-Friedmann, N. Characterization of High-affinity Ryanodine-binding Sites of Rat Liver Endoplasmic Reticulum. Biochem. J. 1991, 276, 41-46. (b) Staudermann, K. A.; Murawski, K. The Inositol 1,4,5-Triphosphate-forming Agonist Histamine Activates a Ryanodine-sensitive Ca²⁺-release Mechanism in Bovine Adrenal Chromaffin Cells. J. Biol. Chem. 1991, 266, 19150-19153. (c) Rardon, D. P.; Krause, P. C. Purification of a Ryanodine-Sensitive Channel Protein from Human Neutrophils. In Excitation-Contraction Coupling in Skeletal, Cardiac, and Smooth Muscle; Frank, G. B., Bianchi, C. P., ter Keurs, Henk E. D., Eds.; Adv. Exp. Med. Biol. 1992, 311, 405-406.
- (13) (a) Gerzon, K.; Besch, H. R., Jr.; Humerickhouse, R. A. Receptor Binding of Ryanodine- and Dehydroryanodine-O_{10eq}-N-acylaminoacylates. *Reg. Meeting Am. Chem. Soc.*, Indianapolis, IN, **1991**, May 29-31, Abstr. MEDI 342. (b) Gerzon, K.; Bidasee, K. R.; Besch, H. R., Jr.; Humerickhouse, R. A.; Ruest, L.; Sutko, J. L. O₁₀-β-Alanyl-ryanodine, a Key Intermediate for Ryanodine Molecular Probes, 203rd Nat. Meet. Am. Chem. Soc., San Francisco, April 6, 1992, Abstr. MEDI 138. (c) Gerzon, K.; Humerickhouse, R. A.; Besch, H. R., Jr. Novel Ester Derivatives of Ryanodine and Dehydroryanodine. U.S. Patent Pending.
 (14) (a) Sutko, J. L.; et al. Unpublished results using biotinoyl-β-alanine-
- (14) (a) Sutko, J. L.; et al. Unpublished results using biotinoyl-β-alanineryanodine in receptor affinity chromatography studies. (b) Bidasee, K. R.; Gerzon, K.; Kwon, S.; Hummerickhouse, R. A.; Emmick, J. T.; Besch, H. R., Jr. Photo-Affinity Labeling of the Calcium Release Channel of the Sarcoplasmic Reticulum of Rabbit Skeletal Muscle with an Azido-Aryl Derivative of Ryanodine. Manuscript to be

submitted to J. Biol Chem. (c) Mais, D. E.; Bowling, N.; Watanabe, A. M. Synthesis and Biochemical Properties of an ¹²⁵I-labelled Ryanodine Derivative. Biochem. Biophys. Res. Commun. 1992, 183, 462-467.

- Neises, B.; Steglich, W. Simple Method for the Esterification of Carboxylic acids. Angew. Chem., Int. Ed. Engl. 1978, 117, 522.
 Nowak, K.; Kania, L. Guanidine derivatives. V. Synthesis of Mono-
- (16) Nowak, K.; Kania, L. Guanidine derivatives. V. Šynthesis of Monoand Diamino Acid Derivatives of S-methyl-isothiourea, *Rocz. Chem.* 1969, 43, 1953–1960. (b) Tian, Z.; Edwards, P.; Roeske, R. W. Synthesis of Optically Pure C.-methyl-arginine. *Int. J. Peptide Protein Res.* 1992, 40, 119–126.
- (17) In the preparation of O_{likeq} -esters we do not use the commercial mixture of 1 and 2 but use pure ryanodine (1) or pure 2 (>97% by HPLC). See for the isolation of ryanoids refs 6, 7, 10, 14b, and 18a,c.
- (18) (a) Ruest, L.; Taylor, D. R.; Deslongchamps, P. Investigation of the Constituents of Ryania Speciosa. Can. J. Chem. 1985, 63, 2840-2843. (b) Welch, W.; et al. The Major Determinants of Tight Binding of Ryanodine to the Vertebrate Skeletal Muscle Receptor. New Natural Ryanoids from Ryania Speciosa Vahl and some Ryanoid Derivatives: 8- and 21-Aminoryanoids. Manuscript in preparation for submission to Biochemistry. (c) Humerickhouse,

R. A.; Besch, H. R., Jr.; Gerzon, K.; Ruest, L.; Sutko, J. L.; Emmick, J. T. Differential Activating and Deactivating Effects Among Natural Ryanodine Congeners on Calcium Release Channels of Sarcoplasmic Reticulum: Evidence for Separation of Effects at Functionally Distinct Sites. Submitted to Molecular Pharmacology. (d) Jefferies, P. R.; Toia, R. F.; Casida, J. E. Ryanodyl 3-(pyridine-3-carboxylate); a novel ryanoid from Ryania Insecticide. J. Nat. Prod. 1991, 54, 1147-1149.

- (19) A conjugate prepared from this ryanodine-hemi-succinate with protein (BSA) constitutes an antigen (ref 13c) designed to elicit anti-ryanodine antibodies which—among other uses (RIA, etc.)—may serve in the search for putative endogenous effectors of the Ry receptor(s) in mammalian systems.
- (20) (a) Emmick, J. T.; Besch, H. R., Jr.; Bidasee, K. R.; Kwon, S.; Gerzon, K. Separating the Dual Actions of Ryanodine on the Sarcoplasmic Reticular Calcium Release Channel. *Biophys. Soc. Meeting*, Washington, D. C. Feb 14-18, 1993, Abstract 383. (b) Emmick, J. T.; Besch, H. R., Jr. Unpublished results.
 (21) McPherson, G. A. Analysis of Radioligand Binding Experiments.
- (21) McPherson, G. A. Analysis of Radioligand Binding Experiments. A Collection of Computer Programs for the IBM PC. J. Pharmacol. Methods 1985, 14, 213–228.