

5-HT₃ Receptor Antagonists. 3. Quinoline Derivatives Which May Be Effective in the Therapy of Irritable Bowel Syndrome

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A series of quinolinecarboxylic acid derivatives has been previously described as a new class of 5-HT₃ receptor antagonists due to deviation of a carbonyl moiety from the plane of an aromatic ring in their minimum-energy conformations. These derivatives were evaluated in a wrap-restraint stress-induced defecation model in rats. Reference compounds, ondansetron (1), granisetron (2), and YM060 (4), potently inhibited a stress-induced increase in stools excreted from fed rats (ID₅₀ = 0.27, 0.12, and 0.0052 mg/kg, po, respectively). However, quinoline derivatives exhibited different activities depending on structural class. 4-Hydroxyquinoline-3-carboxylic acid derivatives 5 and 6a possess high affinity for the 5-HT₃ receptor (K_i = 6.1 and 1.5 nM, respectively) and exhibit potent activity in the Bezold-Jarisch (B-J) reflex test (ED₅₀ = 0.0017 and 0.000 10 mg/kg, iv, respectively), but they did not effectively inhibit the increase in fecal pellet output at the dose of 1 mg/kg, po. On the other hand, most of 1-substituted 2-oxoquinoline-4-carboxylates 10 showed less potent activity in the B-J reflex test than 1 or 2 but inhibited restraint stress-induced defecation more potently than 1 or 2. The ID₅₀ value of *endo*-8-methyl-8-azabicyclo[3.2.1]oct-3-yl 1-isobutyl-2-oxo-1,2-dihydro-4-quinolinecarboxylate 10e was 0.013 mg/kg, po. With respect to the selected compounds 6a and 10e, effects on 5-HT- and thyrotropin-releasing hormone (TRH)-induced defecation, castor oil-induced diarrhea and wrap-restraint stress-induced colonic propulsion in rats were examined. These 5-HT₃ receptor antagonists did not effectively inhibit castor oil-induced diarrhea, which has been reported not to be mediated via the 5-HT₃ receptor. Although 10e showed 800-fold decreased potency compared with 4 in the B-J reflex test, 10e exhibited activity as potent as 4 in 5-HT- and TRH-induced defecation assays; 10e exhibited 7-fold increased potency compared with 4 in wrap-restraint stress-induced colonic propulsions. From these results, 10e appears to interact selectively with 5-HT₃ receptors in the gastrointestinal system and might be effective in the therapy of irritable bowel syndrome (IBS).

Introduction

5-Hydroxytryptamine (5-HT) is a biogenic amine that mediates a variety of physiological actions by binding to distinct cell-surface receptors.¹ 5-HT receptors have been classified into 5-HT₁, 5-HT₂, 5-HT₃, and 5-HT₄ subtypes on the basis of their pharmacological and physiological responses.²⁻⁵ Recently, the 5-HT₃ receptor has attracted considerable attention, and our understanding of this receptor has increased dramatically over the past few years because of the discovery and widespread availability of potent and selective antagonists.¹ These antagonists include ondansetron (1),⁶ granisetron (2),⁷ and tropisetron (3),⁸ which have been shown to be effective in the control of cancer chemotherapy-induced emesis, an event suggested to be modulated by the 5-HT₃ receptor in the area postrema.⁹

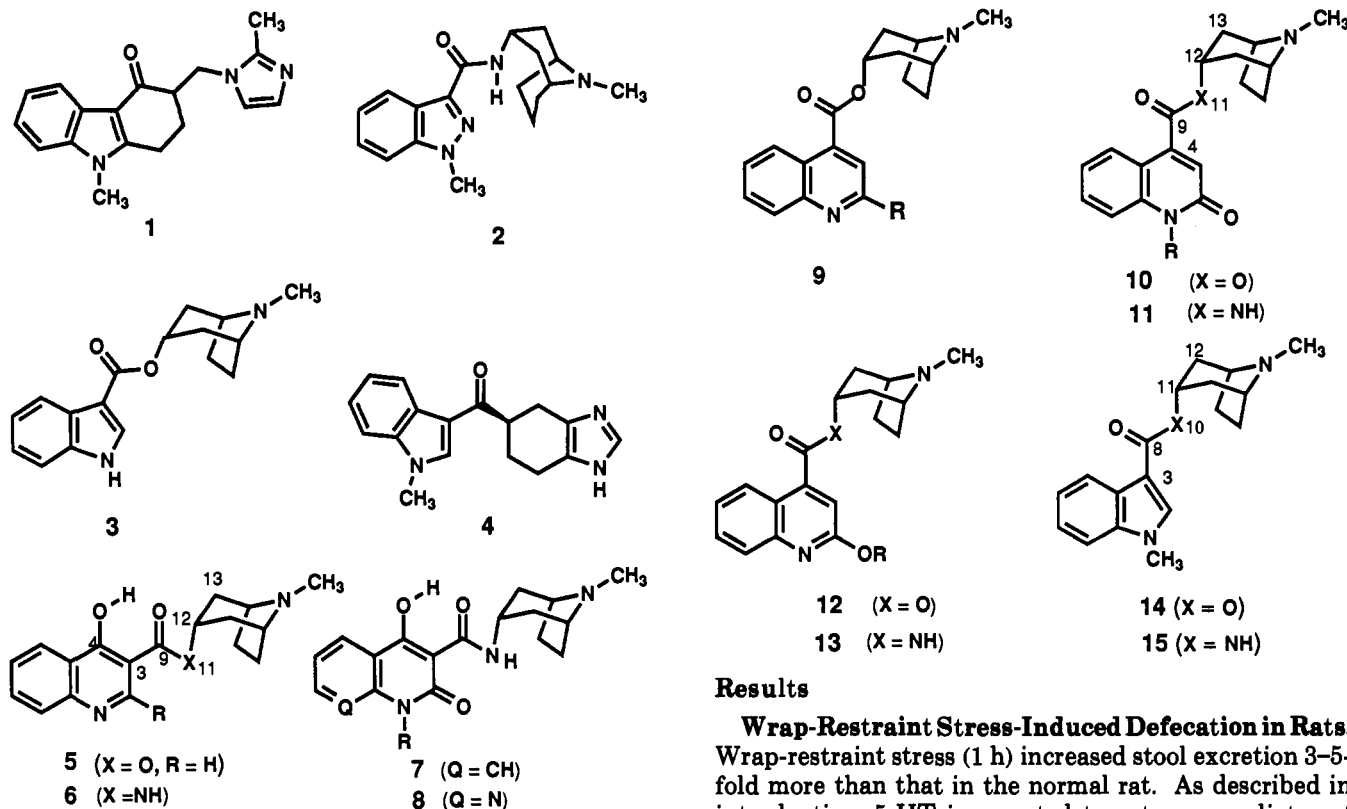
Approximately 90% of endogenous 5-HT is estimated to be within the gastrointestinal tract, and the 5-HT₃ receptor was originally found in the periphery.^{10,11} The role of 5-HT₃ receptors in controlling gastrointestinal contractility *in vitro* is well established. 5-HT or the selective 5-HT₃ receptor agonist, 2-methyl-5-HT, stimulates contractile responses of guinea pig ileum^{12,13} or colon^{14,15} which are blocked by 5-HT₃ receptor antagonists. Furthermore, the fact that immobilization stress results in an increase in plasma and brain 5-HT concentration in

rats indicates the possibility that 5-HT acts as a mediator of gut dysfunction caused by stress. Recently, *in vivo* examination also showed the 5-HT acted as a mediator of restraint stress-, 5-HT-, or thyrotropin-releasing hormone (TRH)-induced gut dysfunction through the 5-HT₃ receptor.^{10,11,16} A potent and selective 5-HT₃ receptor antagonist, YM060 (4)¹⁵ was reported to inhibit restraint (cage) stress-, 5-HT-, and TRH-induced defecation and diarrhea.¹⁶ A recent study by Williams et al. indicates that there are similarities between the intestinal effects of wrap-restraint stress in rats and intestinal symptoms associated with stress and irritable bowel syndrome (IBS) in humans.¹⁷ Thus, wrap-restraint may be an appropriate animal model in which to study stress-related intestinal dysfunction. On the other hand, stress-induced transit of solid residue through the proximal colon has been demonstrated to be a determinant of stool weight in patients with diarrhea-predominant IBS.¹⁸ In fact, ondansetron (1) is reported to slow colonic transit in healthy humans and is under clinical trial as a drug for diarrhea predominant IBS.^{18b-d} Thus, it is of great interest for new drug development to determine whether 5-HT₃ receptor antagonists inhibit wrap-restraint stress-induced defecation and/or accelerated transit through the colon.

We have recently reported novel quinoline derivatives as 5-HT₃ receptor antagonists (5-13), of which binding affinities did not correlate with *in vivo* activities [Bezold-Jarisch (B-J) reflex test].^{19,20} The selective 5-HT₃ receptor antagonists reported to date may be represented by a

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combination of an aromatic group and a carbonyl-containing moiety connected to a basic amine. The carbonyl group is coplanar to an aromatic group, and interatomic distances in the 5-HT₃ receptor antagonist pharmacophore are in adequate ranges (O in a carbonyl group–an aromatic center, ca. 3–4 Å; O in a carbonyl group–N in a basic center, ca. 5 Å; N in a basic center–an aromatic center, ca. 7–8 Å).^{21–24} Molecular modeling studies using QUANTA 3.2/CHARMM 21.3 revealed that although the interatomic distances in the pharmacophores of our quinoline derivatives were similar to the reported values, the carbonyl moiety in the minimum-energy conformers of 7c (R = CH₃), 10a (R = CH₃), and 11d (R = CH₃) deviated from the plane of an aromatic ring.^{19,20} The preceding 5-HT₃ receptor antagonists such as 14 and 15 possessed planar conformation.^{19,20} Therefore, we speculated that our quinoline derivatives might interact with the 5-HT₃ receptor in a different way from that of the reported 5-HT₃ antagonists.

This paper describes the effects of these quinoline derivatives on wrap-restraint stress-induced defecation and transit through the colon and their possibility in the therapy of IBS.

Chemistry

Synthetic methods of most of the compounds have been previously described.^{19,20} Compound 9a was prepared as shown in Scheme I. Isatin (16) was reacted with 4-methyl-2-oxopentane to give the carboxylic acid 17a, which was then treated with SOCl₂ followed by reaction with tropine to afford the target compound 9a. Compound 9b was similarly prepared from 2-phenylquinoline-4-carboxylic acid (17b) (Scheme II). The physical and analytical data of the newly synthesized compounds are listed in Table I. Ondansetron (1),²⁵ granisetron (2),²⁶ and YM060 (4)²⁷ were prepared by using the reported procedures, respectively.

Results

Wrap-Restraint Stress-Induced Defecation in Rats.

Wrap-restraint stress (1 h) increased stool excretion 3–5-fold more than that in the normal rat. As described in introduction, 5-HT is reported to act as a mediator of restraint (cage) stress-induced gut dysfunction through the 5-HT₃ receptor.¹⁶ Thus, our quinoline-type 5-HT₃ antagonists were evaluated in a wrap-restraint stress-induced defecation model. The results are shown in Tables II and III along with data of the binding assay and the B-J reflex test.^{19,20}

Oral administration (1 h treatment) of 1, 2, and 4 dose-dependently inhibited wrap-restraint stress-induced increases in stools excreted from fed rats. The ID₅₀ values of 1, 2, and 4 were 0.27, 0.12, and 0.0052 mg/kg, respectively (Table II). Although 4-hydroxyquinoline-3-carboxylic acid derivatives (5–7) possess high affinity for the 5-HT₃ receptor and exhibit potent activity in the B-J reflex test, they did not effectively inhibit the increases in fecal pellet output at the dose of 1 mg/kg. The ID₅₀ value of 6a (KF 20170) was 1.4 mg/kg, which was 270-fold larger than that of 4.

Subsequently, we evaluated quinoline-4-carboxylic acid derivatives in this model. 2-Substituted 4-quinolinecarboxylates 9a and 9b also did not effectively inhibit fecal pellet output at the dose of 1 mg/kg (Table II). However most of 1-substituted 2-oxo-1,2-dihydroquinoline-4-carboxylates (10b–10i) inhibited wrap-restraint stress-induced defecation more potently than ondansetron or granisetron (Table III). Among them, compounds with a branched alkyl chain at the 1-position exhibited the most potent activity (10c, 10e, and 10g). On the other hand, the amide derivatives (11a–11c), with dramatically decreased binding affinity for the 5-HT₃ receptor and activity in the B-J reflex test, did not effectively inhibit wrap-restraint stress-induced defecation. Further, 2-alkoxyquinoline-4-carboxylates (12a–12f), which possess high affinity for the 5-HT₃ receptor and exhibit moderate activity in the B-J reflex test, did not effectively inhibit defecation. 2-Alkoxyquinoline-4-carboxamide derivatives (13a–13c), which possess low affinity for the 5-HT₃ receptor, did not effectively inhibit defecation at the dose of 1 mg/kg.

Scheme I

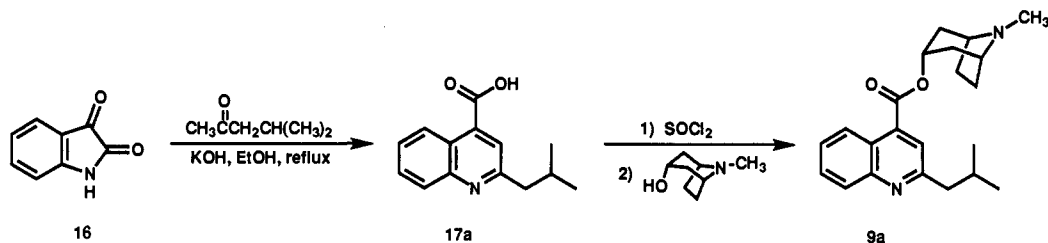
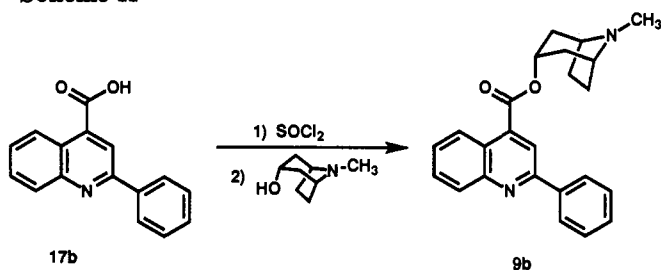


Table I. 2-Substituted Derivatives

compd	structure	yield, %	mp, °C	formula	anal. ^a
9a		20	185.5–188.0	C ₂₂ H ₂₈ N ₂ O ₂ ·C ₄ H ₄ O ₄ ^b ·2.0H ₂ O	C, H, N
9b		30	148.0–149.0	C ₂₄ H ₂₄ N ₂ O ₂ ·2HCl·2.0H ₂ O	C, H, N

^a Analyses for the elements indicated were within $\pm 0.4\%$ of the theoretical values. ^b C₄H₄O₄, fumaric acid.

Scheme II



5-HT- and TRH-Induced Defecation, Castor Oil-Induced Diarrhea and Wrap-Restraint Stress-Induced Colonic Propulsion in Rats. With respect to the selected compounds (1, 2, 4, 6a, and 10e), effects on 5-HT- and TRH-induced defecation^{16,28} and castor oil-induced diarrhea²⁹ were examined (Table IV). An increase in pellet weight was induced in a dose-dependent manner by subcutaneous administration of 5-HT at doses ranging from 1 to 10 mg/kg. Compounds 10e and 4 significantly inhibited 5-HT (3 mg/kg)-induced increase in fecal pellet output with the ID₅₀ values of 0.018 and 0.013 mg/kg, po, respectively. Compounds 6a, 1, and 2 also inhibited 5-HT-induced defecation with relatively high doses (ID₅₀ = 0.49–0.69 mg/kg, po).

Subcutaneous administration of TRH at 1 mg/kg resulted in an increase in defecation with a 3-h pellet output weight (2–3-fold more stool weight than that in the normal). All compounds showed significant prevention of TRH-induced increases in the excreted stool weight of fed rats. Compound 4 exhibited the most potent activity.

Oral administration of castor oil (1 ml/rat) caused diarrhea. All compounds inhibited castor oil-induced diarrhea with higher doses (IE₅₀ = 0.79–1.4 mg/kg) than those in other models.

Further, the ability to inhibit wrap-restraint stress-induced colonic propulsion was examined by measuring the time required to expel a Teflon ball which was inserted about 3 cm into the rat colon.³⁰ Compounds 1 and 6a did not effectively inhibit this colonic propulsion. However, compounds 10e and 4 potentially inhibited the propulsion

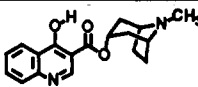
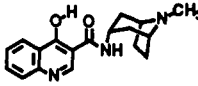
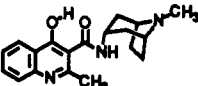
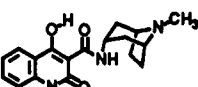
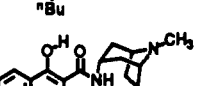
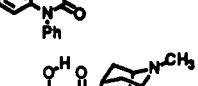
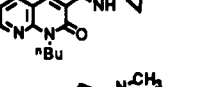
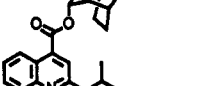
with the ID₅₀ values of 0.010 and 0.069 mg/kg, po, respectively.

Discussion

Stress is a complex reaction characterized by the activation of both hormonal and neuronal systems. In humans, stress commonly results in gastrointestinal disorders such as IBS, in association with changes in gastrointestinal motility, gastric secretion, and digestive transit.^{17,31,32} Wrap-restraint stress resulted in altered small and large intestinal transit but did not result in diarrhea.¹⁷ No erosions developed after 60 min of wrap-restraint stress. In contrast, cold restraint, which is a method classically used in the study of stress-induced ulcer formation, resulted in gastric erosions. Thus wrap-restraint stress is a mild, nonulcerogenic stressor that reproduces the symptoms associated with stress-related intestinal dysfunction in humans.

Most 1-substituted 2-oxo-1,2-dihydroquinoline-4-carboxylates inhibited wrap-restraint stress-induced defecation. The inhibitory activity of compound 10e (R = *i*-Bu) was 10–20-fold more potent than that of 1 or 2 and comparable to that of 4 in this model. The present studies appear to confirm the important role of 5-HT and the 5-HT₃ receptor in the wrap-restraint stress. Although compound 6a has been proved to show the most potent activity in the B-J reflex test (rats, iv) and the cisplatin-induced emesis (dogs, id),²⁰ 6a or its derivatives exhibited weak activities in the inhibition of defecation. These results greatly contrast with those of 1, 2, 4, and 10 (Table II). We have previously reported that activity in the B-J reflex test of compounds 6, 10, 11, 12, and 13 did not have good correlation with the K_i values in the 5-HT₃ receptor binding assay.^{19,20} The result in Table III indicates that inhibitory activity of these compounds in a wrap-restraint stress-induced defecation model did not have good correlation with activity in the B-J reflex test. Since this discrepancy might partly be explained by the difference in their pharmacokinetics, selected compounds were further studied in several defecation and diarrhea models.

Table II. 5-HT₃ Receptor Binding Affinity, Inhibition of B-J Reflex, and Inhibition of Wrap-Restraint Stress-Induced Defecation for Compounds 1, 2, 4, 5, 6a,b, 7a,b, 8, and 9a,b

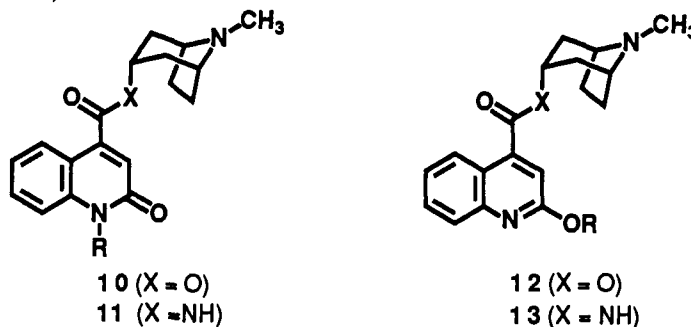
compd	structure	binding affinity ^a K _i ^b nM	B-J reflex		defecation	
			ED ₅₀ ^c (mg/kg, iv) at 5 min	95% confidence interval	ID ₅₀ ^d (mg/kg, po)	95% confidence interval
5		6.1 ± 0.2	0.0017	0.0007–0.0042	(13)	
6a		1.5 ± 0.22	0.00010	0.0000–0.0002	1.4	1.3–1.6
6b		7.2	0.015	0.0098–0.024	(41)	
7a		0.48 ± 0.041	0.29	0.18–0.46	(61)	
7b		1.4 ± 0.23	0.073	0.050–0.14	(48)	
8		6.4	0.29	0.16–0.52	(35)	
9a		0.69 ± 0.06	0.0056		(28)	
9b		(71) ^e	0.34	0.26–0.46	(30)	
1 (ondansetron)		7.6 ± 0.59	0.021	0.015–0.028	0.27	0.23–0.31
2 (granisetron)		2.1 ± 0.26	0.019	0.0056–0.098	0.12	0.10–0.13
4 (YM060)		0.071	0.000089	0.000015–0.00030	0.0052	0.0041–0.0063

^a [³H]Quipazine-labeled 5-HT₃ receptor sites in HG108-15. ^b K_i value was determined as described in the previous paper.¹ Results are expressed as mean values ± SEM after individual measurement (number of determinations is 3). ^c ED₅₀, i.e. the dose of antagonists causing 50% reduction of serotonin effect, was calculated by linear-regression analysis (number of rats is 4). ^d ID₅₀, i.e. the dose of antagonists causing 50% inhibition of restraint stress-induced increase in defecation. Number in a parentheses is inhibition percent at 1 mg/kg, po. ^e Numbers in parentheses are inhibition percents at 10 nM.

5-HT has been reported to cause an increase in the number of fecal pellets excreted from rats via stimulation of the 5-HT₃ receptors. TRH activates colonic transit via a vagally mediated serotonergic mechanism.²⁸ Thus, compounds 1, 2, 4, 6a, and 10e clearly inhibited defecation induced by 5-HT and TRH (Table IV). On the other hand, castor oil-induced diarrhea has been suggested not to be mediated by 5-HT and/or 5-HT₃ receptors.¹⁶ Consequently, all 5-HT₃ receptor antagonists did not effectively inhibit castor oil-induced diarrhea.

Compound 6a showed moderate activity in three models (5-HT, TRH, and diarrhea), and its activity was as potent as that of 1 or 2, but much less potent than that of 4. On the contrary, compound 10e exhibited activity as potent as that of 4 in 5-HT- and TRH-induced defecation assays, and 7-fold more potent activity than that of 4 in inhibition of colonic propulsion, although 10e showed nearly an 800-fold decreased potency compared with 4 in the B-J reflex test. This difference could not be explained by simple difference in their oral absorption because oral administration of 10e potently inhibited the B-J reflex, and its

intravenous administration inhibited colonic repulsion as potently as oral administration. Detailed studies will be published elsewhere. The pharmacological specificity of compounds 4, 6a,²⁰ and 10e for 5-HT₃ recognition sites was examined by investigating them in a wide variety of binding assays. These include assays for serotonin receptors (5-HT_{1A}, [³H]-8-OH-DPAT, rat hippocampus), dopamine receptors (D₁, [³H]SCH23390, rat striatum; D₂, [³H]spiperone, rat striatum), adenosine receptors (A₁, [³H]-CHA, guinea pig cerebral cortex; A₂, [³H]NECA, rat striatum), histamine receptors (H₁, [³H]pyrilamine, guinea pig cerebellum), and adrenaline receptors (α₁, [³H]-WB4104, [³H]clonidine, rat cerebral cortex; β, [³H]-dihydroalprenolol, rat cerebral cortex). The apparent affinities, measured as IC₅₀ (concentration of 50% inhibition) values, for 4, 6a, and 10e were larger than 10 μM, demonstrating that these compounds were inactive at displacing binding to a number of recognition sites. Further, 4, 6a, and 10e did not exhibit 5-HT₄ agonistic and antagonistic activity at 10 μM in the rat oesophageal

Table III. 5-HT₃ Receptor Binding Affinity, Inhibition of B-J Reflex, and Inhibition of Wrap-Restraint Stress-Induced Defecation for Compounds 10a–10k, 11a–11c, 12a–f, and 13a–c

compd	R	binding affinity ^a K _i , ^b nM	B-J reflex		defecation	
			ED ₅₀ ^c (mg/kg, iv) at 5 min	95% confidence interval	ID ₅₀ ^d (mg/kg, po)	95% confidence interval
10a	CH ₃	2.6 ± 0.20	0.18	0.085–0.51	(47)	
10b	CH ₃ CH ₂	1.1 ± 0.07	0.022	0.018–0.026	0.052	0.046–0.057
10c	(CH ₃) ₂ CH	0.32 ± 0.045	0.081	0.044–0.15	0.040	0.033–0.048
10d	CH ₃ (CH ₂) ₂	0.45 ± 0.009	0.33	0.12–0.92	0.17	0.16–0.19
10e	(CH ₃) ₂ CHCH ₂	0.47 ± 0.021	0.033	0.022–0.047	0.013	0.011–0.014
10f	CH ₃ (CH ₂) ₃	0.68 ± 0.036	0.041	0.015–0.11	0.14	0.12–0.16
10g	(CH ₃) ₂ CH(CH ₂) ₂	1.5 ± 0.0	0.059	0.044–0.078	0.041	0.035–0.047
10h	CH ₃ (CH ₂) ₄	0.96 ± 0.09	0.059	0.044–0.081	0.073	0.066–0.080
10i	CH ₃ (CH ₂) ₅	1.1 ± 0.07	0.086	0.067–0.11	0.066	0.057–0.076
10j	C ₆ H ₅ CH ₂	1.3 ± 0.05	0.15	0.12–0.20	(22)	
10k	C ₆ H ₅	0.51 ± 0.012	0.044	0.035–0.057	(59)	
11a	CH ₃ (CH ₂) ₂	67 ± 16	0.35	0.27–0.46	(12)	
11b	CH ₃ (CH ₂) ₃	60 ± 4.9	0.37	0.26–0.55	(47)	
11c	C ₆ H ₅	63 ± 9	0.19	0.14–0.26	(14)	
12a	CH ₃	0.58 ± 0.015	0.42	0.21–0.82	(45)	
12b	CH ₃ CH ₂	0.88 ± 0.025	0.070	0.049–0.12	(36)	
12c	(CH ₃) ₂ CHCH ₂	0.44 ± 0.064	0.24	0.18–0.31	(36)	
12d	CH ₃ (CH ₂) ₃	0.39 ± 0.020	0.20	0.11–0.31	(64)	
12e	CH ₃ (CH ₂) ₄	0.86 ± 0.10	1.13	0.93–1.39	(30)	
12f	CH ₃ (CH ₂) ₅	3.2 ± 0.26	7.68	3.48–16.9	(23)	
13a	CH ₃ (CH ₂) ₂	42 ± 11	1.54	0.90–2.63	(29)	
13b	(CH ₃) ₂ CH	53 ± 19	1.86	1.13–3.04	(39)	
13c	CH ₃ (CH ₂) ₃	19 ± 25	0.54	0.43–0.74	(43)	
1 (ondansetron)		7.6 ± 0.59	0.021	0.015–0.028	0.27	0.23–0.31
2 (granisetron)		2.1 ± 0.26	0.019	0.0056–0.098	0.12	0.10–0.13
4 (YM060)		0.071 ± 0.0071	0.000089	0.000015–0.00030	0.0052	0.0041–0.0063

^{a–d} See footnotes a–d in Table II.**Table IV.** Effects of 6a, 10e, and Reference Compounds on 5-HT- and TRH-Induced Defecation, Castor Oil-Induced Diarrhea, and Wrap-Restraint Stress-Induced Colonic Propulsion

compd	ID ₅₀ ^a [mg/kg, po]			ID ₅₀ ^b [mg/kg, po] inhibition of colonic propulsion
	inhibition of defecation		inhibition of diarrhea castor oil	
	5-HT	TRH		
6a (KF 20170)	0.69 (0.63–0.76)	0.053 (0.044–0.063)	0.79 (0.52–1.3)	1.8 (1.7–1.9)
10e (KF 18259)	0.018 (0.016–0.021)	0.021 (0.019–0.025)	1.4 (0.95–2.3)	0.010 (0.0092–0.011)
1 (ondansetron)	0.53 (0.48–0.59)	0.31 (0.26–0.36)	1.4 (1.2–1.7)	1.6 (1.5–1.7)
2 (granisetron)	0.49 (0.44–0.56)	0.074 (0.066–0.082)	0.91 (0.74–1.1)	0.74 (0.67–0.82)
4 (YM060)	0.013 (0.011–0.015)	0.0051 (0.0046–0.0057)	1.1 (0.77–1.9)	0.069 (0.065–0.074)

^a ID₅₀, i.e. the dose of antagonists causing 50% inhibition of increase in defecation and castor oil-induced diarrhea. Numbers in parentheses are 95% confidence intervals. ^b ID₅₀, i.e. the dose of antagonists causing 50% inhibition of decrease in the time of expulsion of a Teflon ball. Numbers in parentheses are 95% confidence intervals.

tunica muscularis mucosae.³³ Thus, 4, 6a, and 10e were identified to be selective 5-HT₃ antagonists.

The B-J reflex is known to be mediated by reflex stimulation of the vagus nerve following activation of the sensory nerve located in the right ventricle wall.¹⁹ Wrap-restraint stress-induced defecation and colonic propulsion could be mediated by the 5-HT₃ receptor in the gastrointestinal system. Therefore, compound 10e seems to interact selectively with the 5-HT₃ receptor in the gastrointestinal system and might be different from the reported 5-HT₃ receptor antagonists, including 4, which are considered to interact with both of the 5-HT₃ receptor

in the gastrointestinal system and that in the cardiovascular system. Considering the structure of compound 10e, its carbonyl moiety seems to deviate from the plane of an aromatic ring in the minimum-energy conformer, similarly as in the case of compound 10a.¹⁹ The carbonyl moiety of 11a also deviates from the planarity, but a series of 11 showed decreased affinity for the 5-HT₃ receptor and did not inhibit stress-induced defecation. Thus this deviation might be important for 10 as a 5-HT₃ antagonist in the gastrointestinal system. Compound 10e may belong to a novel class of 5-HT₃ receptor antagonists. Different properties of 5-HT₃ receptor antagonists in these models

suggested that there are multiple subtypes of the 5-HT₃ receptor. In fact, the existence of such multiple 5-HT₃ receptor molecular species has been suggested by several groups.^{34,35}

In conclusion, wrap-restraint stress as well as exogenous 5-HT or TRH administration caused significant alterations in bowel function in rats. KF 18259 (10e), which possesses high affinity for the 5-HT₃ receptor but showed moderate activity in the B-J reflex test, inhibited potently wrap-restraint stress-, 5-HT-, TRH-induced defecation and wrap-restraint stress-induced colonic propulsion. Since 10e might be a selective 5-HT₃ antagonist in the gastrointestinal system, this class of 5-HT₃ receptor antagonists might be effective in the therapy of IBS.

Experimental Section

Proton nuclear magnetic resonance spectra (¹H NMR) were recorded on a JEOL JNM GX-270 FT NMR or a Hitachi R-90H FT NMR spectrometer with Me₄Si as an internal standard, and mass spectra were recorded on a JMS-SX102 instrument. Melting points were determined with a Büchi-510 melting point apparatus and are uncorrected. Infrared spectra (IR) were recorded on a JASCO IR-810 spectrometer. Elemental analysis were performed by the analytical department of our laboratories.

Chemistry. *endo*-8-Methyl-8-azabicyclo[3.2.1]oct-3-yl 2-Isobutyl-4-quinolinecarboxylate Fumarate (9a). To a mixture of KOH (44.0 g, 784.2 mmol) and EtOH (400 mL) was added isatin (16; 22.1 g, 150.2 mmol) and subsequently 4-methyl-2-oxopentane (40 mL, 319.5 mmol) in EtOH (100 mL), followed by heating under reflux for 30 min. After cooling and concentration of the mixture under reduced pressure, Et₂O was added to the residue followed by extraction with H₂O. Then, dilute HCl was added to acidify the solution. The precipitated crystals were collected by filtration, washed with H₂O, and dried to give compound 17a (13.3 g, 39%): ¹H NMR (DMSO-*d*₆) δ 8.63 (d, 1 H, *J* = 8.5 Hz), 8.05 (d, 1 H, *J* = 8.2 Hz), 7.81 (s, 1 H), 7.50–7.85 (m, 2 H), 2.86 (d, 2 H, *J* = 7.0 Hz), 2.21 (m, 1 H), 0.93 (d, 6 H, *J* = 6.6 Hz). SOCl₂ (20 mL) was added to the compound described above (2.81 g, 12.3 mmol) followed by stirring. Excess SOCl₂ was removed under reduced pressure, and anhydrous THF (50 mL) was added (solution A). A 15% *n*-BuLi-hexane solution (8.8 mL, 14.2 mL) was added to a mixture of tropine (2.00 g, 14.2 mmol) and anhydrous THF (6 mL) at 0 °C followed by stirring for a further 15 min. Then, solution A was dropwise added under an argon atmosphere. The mixture was stirred at 0 °C for 1 h and at room temperature for a further 2.5 h. To this mixture was added a small quantity of MeOH and H₂O followed by extraction with CHCl₃. The organic layer was dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure, and the residue was purified by silica gel column chromatography with CHCl₃-MeOH (10:1) as eluent. To this product were added *i*-PrOH (40 mL) and fumaric acid (0.26 g, 2.24 mmol), and the mixture was stirred at room temperature. Hexane (15 mL) was added to this solution at room temperature with stirring, and the precipitated crystals were collected by filtration and dried to give compound 9a as the fumarate (1.16 g, 20%): IR (KBr) 3410 (br), 2958, 1715, 1694, 1590, 1367, 1273, 1254, 1201, 1158, 1079, 1024, 981, 970, 928, 823, 798, 774, 651, 633, 550 cm⁻¹; MS *m/z* 352 (M⁺); ¹H NMR (DMSO-*d*₆) δ 8.63 (d, 1 H, *J* = 7.7 Hz), 8.07 (d, 1 H, *J* = 7.7 Hz), 7.83 (s, 1 H), 7.81 (m, 1 H), 7.71 (m, 1 H), 5.32 (m, 1 H), 3.69 (m, 2 H), 2.90 (d, 2 H, *J* = 7.3 Hz), 2.59 (s, 3 H), 2.00–2.65 (m, 9 H), 0.96 (d, 6 H, *J* = 6.6 Hz). Anal. (C₂₂H₂₈N₂O₂·C₄H₄O₄) C, H, N.

endo-8-Methyl-8-azabicyclo[3.2.1]oct-3-yl 2-Phenyl-4-quinolinecarboxylate Dihydrochloride (9b). The product in a free form obtained in a similar manner to that described above was dissolved in CHCl₃, and to this solution was added EtOAc saturated with HCl. The mixture was poured into cold Et₂O with stirring, and the precipitated crystals were collected by filtration and dried to give compound 9b as the hydrochloride (30%): MS *m/z* 372 (M⁺); ¹H NMR (DMSO-*d*₆) δ 11.13 (br s, 1 H), 8.65 (d, 1 H, *J* = 7.8 Hz), 8.50 (s, 1 H), 8.24–8.36 (m, 2 H), 8.22 (d, 1 H, *J* = 7.8 Hz), 7.90 (m, 1 H), 7.75 (m, 1 H), 7.51–7.68

(m, 3 H), 5.38 (m, 1 H), 3.93 (m, 2 H), 2.68 (d, 3 H, *J* = 4.9 Hz), 2.05–2.90 (m, 8 H). C₂₄H₂₄N₂O₂·2HCl.

Biology. **5-HT₃ Receptor Binding Test.** The binding assay was carried out according to the procedure described in the previous papers.^{19,20}

Pharmacology. **Bezold-Jarisch Reflex Test.** The Bezold-Jarisch reflex assay was carried out according to the reported procedure.³⁶ Male Wistar rats (230–330 g, Japan SLC) were anesthetized with urethane (1.25 g/kg, ip), and their tracheas were cannulated. Blood pressure was recorded from the left carotid artery via a saline/heparin-filled pressure transducer from which the heart rate was also continuously monitored. Compounds dissolved in saline were injected intravenously into the exterior jugular vein. The B-J reflex was evoked by a rapid bolus iv injection of 5-HT (30 μg/kg), and consistent responses were established every 12 min before each challenge with 5-HT. Each compound was evaluated at 3–6 doses (*n* = 4 in each dose). The ED₅₀ values, the doses which reduced the 5-HT-induced bradycardia by 50%, and 95% of confidence intervals were calculated by Probit method and Litchfield-Wilcoxon method.

Effects on Wrap-Restraint Stress-Induced Defecation.¹⁷ Adult male Sprague-Dawley (SD) rats weighing 150–200 g were given the test compounds orally. After 60 min, rats were lightly anesthetized with ether, and their forelimbs, upper forelimbs, and thoracic trunk were wrapped in paper tape to restrict, but not prevent, movement. Rats recovered from ether within 2–5 min and immediately moved about in their cages and ate and drank. The wrap-restraint restricted mobility of their forelimbs, which prevented them from grooming the face, upper head, and neck. Normal rats were anesthetized with ether but were not wrapped. The effects of the test compounds were determined by measuring the stool weight with 60 min of stressing. The stool weights of control rats (vehicle treated) and normal rats (without stress) were 0.4–0.6 and 0.1–0.2 g/rat, respectively. The test compounds were evaluated at four doses (*n* = 12 in each dose). The ID₅₀ values and 95% of confidence interval were calculated by Probit method.

Effects on 5-HT-Induced Defecation.¹⁶ Adult male SD rats weighing 150–200 g were given the test compounds orally. After 60 min, 5-HT was administered subcutaneously (3 mg/kg). The effects of the test compounds were determined by measuring the stool weight 60 min after 5-HT administration. The stool weights of control (vehicle treated) and normal rats (without 5-HT) were 0.6–0.8 and 0.2–0.25 g/rat, respectively. The test compounds were evaluated at four doses (*n* = 12 in each dose). The ID₅₀ values and 95% of confidence interval were calculated by Probit method.

Effects on TRH-Induced Defecation.¹⁶ Adult male SD rats weighing 150–200 g were given the test compounds orally. After 60 min, TRH was administered subcutaneously (1 mg/kg). The effects of the test compounds were determined by measuring the stool weight 3 h after TRH administration. The stool weights of control rats (vehicle treated) and normal rats (without TRH) were 0.5–0.55 and 0.2–0.3 g/rat, respectively. The test compounds were evaluated at five doses (*n* = 12 in each dose). The ID₅₀ values and 95% of confidence interval were calculated by Probit method.

Effects on Castor Oil-Induced Diarrhea.²⁹ Adult male SD rats weighing 150–200 g were given the test compounds orally. After 60 min, castor oil was administered orally (1 ml/rat). The effects of the test compounds were determined by measuring the stool weight 90 min after castor oil administration. The stool weights of control rats (vehicle treated) and normal rats (without castor oil) were 1.5–2.0 and 0 g/rat, respectively. The test compounds were evaluated at four doses (*n* = 8 in each dose). The ID₅₀ values and 95% of confidence intervals were calculated by Probit method.

Effects on Wrap-Restraint Stress-Induced Colonic Propulsion.³⁰ Adult male SD rats weighing 150–200 g were given the test compounds orally. After 60 min, rats were lightly anesthetized with ether, stressed by wrapping their forelimbs, upper forelimbs and thoracic trunk in paper tape, and then a Teflon ball (diameter = 3.17 mm) was inserted about 3 cm into the rat colon. The effects of the test compounds were determined by measuring the time required to expel a Teflon ball. The times in control rats (vehicle treated) and normal rats (without wrap

restraint) were 150–300 and 1800–2300 seconds, respectively. The test compounds were evaluated at four doses ($n = 10$ in each dose). The ID_{50} values and 95% of confidence intervals were calculated by Probit method.

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