Neurosteroid Analogues: Structure-Activity Studies of Benz[e]indene Modulators of GABAA Receptor Function. 1. The Effect of 6-Methyl Substitution on the Electrophysiological Activity of 7-Substituted Benz[e]indene-3-carbonitriles

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The effect of 6-methyl substitution on the ability of 7-(2-hydroxyethyl) benz[e] indene-3-carbonitriles to potentiate GABA-mediated chloride current and to directly gate a chloride current in the absence of GABA in cultured rat hippocampal neurons was investigated. Structurally analogous steroid 17-carbonitriles that either contained or did not contain a 19-methyl group were also investigated. Compounds were evaluated at 1 \(\mu M \) for their ability to potentiate GABA-mediated currents and at 10 µM for current activation in the absence of GABA. The benz[e]indene 3(R)-carbonitriles and analogous steroid 17α -carbonitriles had no effects in either assay. The benz[e]indene-3(S)carbonitriles and analogous steroid 17β -carbonitriles were active in both assays. Relative to the 6-unsubstituted benz[e]indene 3(S)-carbonitrile, the following effects of 6-methyl substituents were observed: a 6(a)-methyl group increased both activities; a 6(e)-methyl group decreased both activities; and 6,6-dimethyl substitutents had opposing effects so that both activites remained similar to those of the 6-unsubstituted compound. The activities of the steroid 17β-carbonitriles were not affected significantly by the presence or absence of a 19-methyl group. A conformational analysis using molecular modeling methods was also performed for the benz[e]indene 3Scarbonitriles and the steroid 17β -carbonitriles. The ability of the different 6-methyl substituents to differentially effect the conformations of the flexible benz[e]indenes and the inability of the steroid 19-methyl group to alter the conformations of the rigid steroid 17β -carbonitriles are suggested to explain the results.

The major inhibitory neurotransmitter in the mammalian central nervous system is γ -aminobutyric acid (GABA). Binding of this neurotransmitter to its recognition site on the postsynaptic GABAA receptor/chloride channel complex results in the opening of the channel and, in most neurons, the passage of chloride ions into the neuron. This increase in intracellular chloride concentration makes the membrane potential of the postsynaptic cell more negative thereby decreasing its ability to respond to an excitatory stimulus.

There are additional binding sites for other ligands on the GABAA complex. The most studied of these are the benzodiazepine, 1,2 barbiturate,3 picrotoxin,4 and steroid5-9 binding sites. Ligands for these sites may either modify the response of the complex to GABA or, in the case of the barbiturates and steroids, alter the gating properties of the channel in the absence of GABA. Compounds acting at these sites either to facilitate or to produce an increase in chloride flux may have anxiolytic, sedative-hypnotic, anticonvulsant, muscle relaxant, and anesthetic properties.

We are interested in the development of new ligands for the steroid binding site on the GABAA complex. In this regard, we recently reported that benz[e] indenes 1 and 2 are ligands for the site. 10,11 A comparison of the electrophysiological effects of benz[e]indene 1 and steroid 3 (a prototypic ligand for the steroid binding site 12) in cultured rat hippocampal neurons showed that benz[e]indene 1 was more potent than steroid 3 (EC₅₀ = 0.2 ± 0.03 and 1.2 \pm 0.2 μ M, respectively) at increasing currents mediated by 1 μ M GABA, but less potent than steroid 3 at gating current in the absence of GABA. For this latter effect, the threshold for gating of current by steroid 3 is 1 μ M. whereas this gating threshold was found to be above 10 μ M (in some cells it was above 100 μ M) for benz[e]indene 1. Thus, if the observed dose-response relationships for the two effects caused by steroid 3 are used as reference standards, these parameters are shifted in opposite directions for benz[e]indene 1. We believe, for the reasons discussed below, that the shifted dose-response relationships observed for benz[e]indene 1 have significant practical importance.

Schulz and Macdonald¹³ have provided evidence for a hypothesis to explain why phenobarbital is useful as an anticonvulsant, whereas pentobarbital is useful as an anesthetic. This hypothesis correlates the anticonvulsant activity of a barbiturate with its ability to potentiate GABA-mediated chloride currents and the anesthetic activity with its ability to gate chloride currents at the GABA, receptor complex in the absence of GABA. Since both dose-response curves are close together for pentobarbital, the anesthetic effect of this drug will be manifest at nearly all doses so this drug is useful primarily as an anesthetic. On the other hand, these dose-response curves are more widely separated for phenobarbital and it can be

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used at doses where its GABA potentiating effects are dominant so that the drug is a useful anticonvulsant.

Application of this hypothesis to ligands for the steroid binding site on the GABAA complex predicts that a compound like benz[e]indene 1 should be more like phenobarbital than pentobarbital with regard to its biological activity, and in vivo studies with this compound are in progress to address this prediction. The study reported here is concerned with further evaluating the importance of structural differences between steroids and benz[e]indenes in relation to their effects on the above discussed dose-response relationships. Specifically, we have studied the effect of the 19-methyl group in steroids and the analogous methyl groups at C^6 in benz[e]indenes on the electrophysiological activities of the compounds at the GABAA receptor complex present in cultured rat hippocampal neurons.

Chemistry

Other investigators have shown that both steroids 3 and 4, which differ only in their 17β -substituents, have very similar activities as potentiators of muscimol-stimulated ³⁶Cl-uptake in rat brain synaptoneurosomes. ¹⁴ Likewise. we have shown that benz[e]indenes 1 and 2, which differ similarly in their analogous 3S substituents, have comparable potencies as potentiators of 1 µM GABA-mediated chloride current in cultured rat hippocampal neurons.11 Thus, it seems likely that congeneric steroids and benz-[e] indenes having a nitrile group instead of an acetyl group at these same positions will have high biological activities. Accordingly, we chose to evaluate analogues of benz[e]indene 2 and steroid 4. The decision is of practical significance since the nitrile-substitued compounds are more readily prepared.

The desired benz[e]indenes 2 and 34-39 were prepared from either 17β -acetoxy- 5α -estran-3-one (5) or 17β -acetoxy- 5α -androstan-3-one (6) as indicated in Schemes I and III. The reaction sequence $5 \rightarrow 7 \rightarrow 9 \rightarrow 11 \rightarrow 13 \rightarrow 15$ \rightarrow 17 \rightarrow 19 (Scheme I) has been reported previously. 15 In general, this sequence is useful for the preparation of benz[e]indenes having either equatorial substituents or no substituents at C^6 . The analogous reaction sequence $6 \rightarrow 8 \rightarrow 10 \rightarrow 12 \rightarrow 14 \rightarrow 16 \rightarrow 18 \rightarrow 20$, which is reported here, is of general use for the preparation of benz[e]indenes having an axial methyl substituent with or without equatorial substitutents at C^6 .

The Δ^2 -enol acetate 8 was prepared by reacting steroid 6 with isopropenyl acetate in the presence of p-TsOH as described previously.¹⁶ Although the literature does not indicate that the isomeric Δ^3 -enol acetate is formed in this reaction, the proton NMR spectrum of our reaction product indicated (by integration of the peak area of the vinyl protons) that it was a ca. 8:2 mixture of the Δ^2 - and Δ^3 -enol acetates. This mixture of isomeric enol acetates was reacted with ozone in dichloromethane-HOAc at -78 °C. Following reduction with dimethyl sulfide and hydrolysis, the crude product was reduced with NaBH4 and purified by recrystallization to give pure hydroxy acid 10 in 61% yield.

Treatment of compound 10 with diazomethane yielded (94%) the corresponding methyl ester 12, which was oxidized to compound 14 with PCC.17 Compound 14, which was obtained as a crude oil, was not purified and characterized because it rapidly converted upon standing into multiple products. Hence, as soon as compound 14 was prepared, it was immediately reacted with isopropenyl Scheme I^a

^a(a) TBDMS-OTf, NEt₃ for 5, isopropenyl acetate, TsOH for 6; (b) O₃, NaBH₄; (c) CH₂N₂; (d) PCC; (e) isopropenyl acetate, TsOH; (f) O3; (g) Wilkinson's catalyst; (h) HSCH2CH2SH, BF3-EtOEt; (i)

acetate-p-TsOH and converted into the stable enol acetate 16 (an 8:2 mixture of E/Z isomers as determined by proton NMR analysis). The overall yield for the oxidation and enol acetylation steps was 36%.

The conversion of enol acetate 16 into either compounds 20 or 23 via aldehyde 18 was carried out without purification of aldehyde 18 because this aldehyde, like aldehyde 14, was unstable. Hence, ozonolysis of compound 16 in dichloromethane-HOAc at -78 °C followed by decarbonylation of the resulting aldehyde 18 using Wilkinson's catalyst18-20 in benzonitrile at 160 °C gave the 6(a)-methylsubstituted benz[e]indene 20 (18% from 16). Alternatively, ozonolysis of compound 16 followed by treatment of aldehyde 18 with ethanedithiol-boron trifluoride etherate²¹ gave thioacetal 23 (88% from 16). Desulfurization²² of compound 23 gave the 6,6-dimethyl-substituted benz-[e]indene 25 (77%). The 6(e)-methyl-substituted benz-[e]indene 24 was prepared from aldehyde 17 via thioacetal

Scheme II

22 using a similar thioacetalization (85%)/desulfurization (80%) sequence.

We also considered preparing benz[e]indenes 24 and 25 from compounds 11 and 12, respectively, using Raney nickel in refluxing toluene to effect a dehydroxymethylation reaction as described by Krafft et al.²³ However, when this reaction was tried with compound 12 (Scheme II), the product was found to be the seven-membered ring lactone 21 (93%). No investigations were made into altered reaction conditions that might favor dehydroxymethylation over lactonization.

The remaining steps in the synthesis of benz[e]indenes 2 and 34-39 are shown in Scheme III. Diester compounds 19, 20, 24, and 25 were reduced by DIBALH²⁴ in toluene at 0-5 °C to their corresponding diols 26-29 in high yields (93-98%). The secondary hydroxy group of each of these diols was oxidized selectively with 5.25% NaOCl in glacial AcOH²⁵ to yield compounds 30-33 in moderate yields (64-76%). Reaction of the keto groups of compounds 30, 32, 33 with $1.0\,\mathrm{M}\,t$ -BuOK in dimethoxyethane and tosylmethyl isocyanide (TosMIC)26,27 gave a mixture of the corresponding diastereomeric 3(S)-carbonitriles (2, 37, and 39)and 3(R)-carbonitriles (34, 36, and 38) (27–63% yields); whereas, compound 33 yielded only the 3(S)-carbonitrile 35 (36%). The final purification and/or separation of benz[e]indenes 2 and 34–39 was accomplished by HPLC on silica as described in the Experimental Section.

The stereochemical assignments for the nitrile groups of compounds 2 and 34-39 were based on the ¹H NMR spectra of the compounds. Because of the dihedral angles involved, the coupling constants of the C3 protons of benz[e]indenes having the 3S configuration would be expected to be approximately the same for vicinal coupling to both of the adjacent protons on C2. By contrast, the coupling constants of the C3 protons of benz[e]indenes having the 3R configuration would be expected to be unequal for vicinal coupling to the adjacent protons on C². Consequently, nitriles 2, 35, 37, and 39 whose ¹H NMR spectra showed a resonance at $\delta 2.25 \pm 0.3$ as an apparent triplet (J = 9.6 Hz) were assigned as the 3(S)-carbonitriles. Compounds 34, 36, and 38 whose ¹H NMR spectra showed a resonance at δ 2.53 \pm 0.3 as a doublet of doublets (J =7.0, 2.0 Hz) were assigned as the 3(R)-carbonitriles. The

 13 C chemical shifts of the carbons of the nitrile groups also correlate well with the stereochemistry of the nitrile groups. The 3(S)-carbonitrile carbons are observed at δ 121.2 \pm 0.2 and the 3(R)-carbonitrile carbons are observed at δ 122.2 \pm 0.2.

A similar synthetic strategy was used for the preparation of the desired steroid 17-carbonitriles (Scheme IV). Although steroid 41 is commercially available, it is prohibitively expensive. Since potassium tri-sec-butylborohydride in tetrahydrofuran (K-Selectride) has been shown to selectively and stereospecifically reduce 3,17-diketosubstituted androgens to 3α -hydroxy-17-keto-substituted products, ^{28,29} we investigated the reduction of 19-norsteroid 40 with this reagent. Even though the absence of a 19methyl group increases the accessibility of K-Selectride to the β -face of 19-norsteroid 40, the only reduction product obtained was the 3α -hydroxy-19-norsteroid 41 (86%). The ¹H NMR and ¹³C NMR spectra of crude compound 41 showed the resonances of the proton on C3 as an apparent triplet at δ 4.06 and C³ at δ 66.19. No resonances for the corresponding proton (m, δ 3.5) and carbon (δ 71.26) resonances of an authentic sample of 3β -hydroxy- 5α estran-17-one were observed.

The reaction of 19-norsteroid 41 with TosMIC yields (52%) a 1.2:1 mixture of the 17β -carbonitrile 43 and the 17α -carbonitrile 44, respectively. As reported previously, ¹⁴ reaction of steroid 42 with TosMIC yields (59%) a 1.5:1 mixture of the 17β -carbonitrile 4 and the 17α -carbonitrile 45, respectively. These diastereomeric mixtures were separated by HPLC on silica as reported in the Experimental Section. As described above for the analogous benz[e]indenes (2 and 34-39), the configurations of the 17-carbonitrile groups of compounds 4 and 43-45 were assigned from the ¹H NMR and ¹³C NMR spectra of the compounds.

Electrophysiology

Voltage clamp recordings were obtained from cultured postnatal rat hippocampal neurons using whole-cell patch clamp methods. Tach compound was evaluated at 1 μ M for its ability to potentiate 1 μ M GABA-mediated currents and also evaluated at 10 μ M for its ability to directly gate a current in the absence of GABA (Table I). The two responses were studied at a 10-fold difference in compound concentration so that structural modifications having effects on each activity could be readily identified. The concentrations selected for these experiments were based on our previous experience with compounds 1-3. Recordings that are representative of the responses observed for the benz[e] indenes are shown for compounds 2, 34,

Scheme IIIa

19,20,24,25
$$\xrightarrow{a}$$

C

RI. RI. RI. H

26 R = H, RI = H

27 R = CH₃, RI = H

28 R = H, RI = CH₃

29 R = CH₃, RI = CH₃

30 R = H, RI = CH₃

31 R = CH₃, RI = CH₃

32 R = H, RI = CH₃

33 R = CH₃, RI = CH₃

34 R = H, RI = H

25 R = H, RI = H

35 R = CH₃, RI = H

36 R = H, RI = CH₃

37 R = H, RI = CH₃

38 R = CH₃, RI = CH₃

39 R = CH₃, RI = CH₃

^a(a) DIBALH; (b) 5.25% NaOCl, HOAc; (c) TosMIC, t-BuOK.

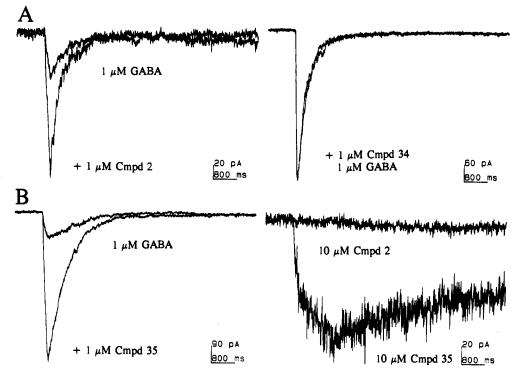


Figure 1. Effects of benz[e] indenes on GABA receptor-mediated currents in cultured rat hippocampal neurons: (A) The traces show the response of two neurons to 1 μ M GABA in the absence and presence of 1 μ M 2 (left) and 1 μ M 34 (right). (B) The left trace shows the response of a neuron to 1 μ M GABA in the absence and presence of 1 μ M 35. The right traces show the response of a neuron to $10 \mu M$ 2 and $10 \mu M$ 35. The prolonged nature of the current activated by 35 is typical of currents gated by benz[e] indenes (or steroids) at concentrations where these agents activate a response. All traces were obtained at a holding potential of -60 mV and GABA and/or the benz[e]indenes were applied for 200 ms.

Scheme IVa HO, 41 R = H 42 R = CH₃ 44 R = H 43 R = H 45 R = CH₃ 4 R = CH₃

^a(a) K-Selectride; (b) TosMIC, t-BuOK.

and 35 in Figure 1. For the steroids, similar representative recordings are shown for compounds 4 and 45 in Figure 2.

The benz[e]indene-3(R)-carbonitiriles 34, 36, and 38 and the steroid 17α -carbonitriles 44 and 45 were inactive as either potentiators (even at concentrations of 10 μ M) or direct activators of current. The benz[e]indene-3(S)carbonitriles 2, 35, 37, and 39 and the steroid 17β carbonitriles 4 and 43 were all potentiators of GABAmediated currents. Benz[e]indene-3(S)-carbonitrile 37 was not an effective potentiator at $1 \mu M$, but it was effective when evaluated at 10 μ M. For potentiation of GABAmediated current the potency of the compounds increased as follows: $37 < 39 \simeq 2 < 35 \simeq 43 \simeq 4$. Of these compounds, only 10 μ M benz[e]indene 35 and 10 μ M steroids 43 and 4 directly gated a current. At $1 \mu M$, steroids

Table I. Electrophysiological Effects of Benz[e]indenes and Steroids on GABAA Receptor Function

compd	N^a	compd (1 µM) potentiation % response relative to current produced by GABA ^b	compd (10 µM) gated current ^c
34	4	$NR^{d,e}$	NR
36	4	NR°	NR
38	4	NRe	NR
44	8	NR ^e	NR
45	4	NR ^e	NR
2	12	255 ± 8^{f}	NR
35	6	382 ± 47	26 ± 4
35	5		$NR (1 \mu M)^g$
37	5	NR	NR
37	5	$244 \pm 56 (10 \mu\text{M})^h$	
39	10	220 ± 17	NR
43	7	372 ± 49	38 ± 5
43	7		$15 \pm 4 \ (1 \ \mu M)^g$
4	10	443 ± 55	49 ± 4
4	6		$22 \pm 5 \ (1 \ \mu M)^g$

 $^{a}N =$ Number of cells examined. b To calculate the % response. the magnitude of the peak current produced by 1 μ M GABA plus 1 μM compound was normalized with respect to the peak current produced by 1 μ M GABA alone on the same cell. A % response of 100% reflects no change in the current compared to 1 μM GABA alone. At the foot of the dose-response curve in cultured postnatal rat hippocampal neurons is a GABA concentration of $1 \mu M$. These experiments were conducted at $-60\,\mathrm{mV}$ and compounds were applied by pressure ejection for 200 ms. The compound gated current reflects the peak current directly gated by 10 µM compound in the absence of GABA compared to the response obtained from the same cell in response to 1 µM GABA alone. d NR denotes no response. Compounds that increased current by $\leq 6\%$ of the response to $1 \mu M GABA$ in the same cells were considered to give no response. e This compound at 10 μ M also did not potentiate 1 μ M GABA-mediated currents. If Values are the mean \pm SEM. If This table entry indicates the direct gating effect of this compound at 1 μ M. ^h This table entry indicates the potentiating effect of this compound at 10 µM on currents mediated by 1 μ M GABA.



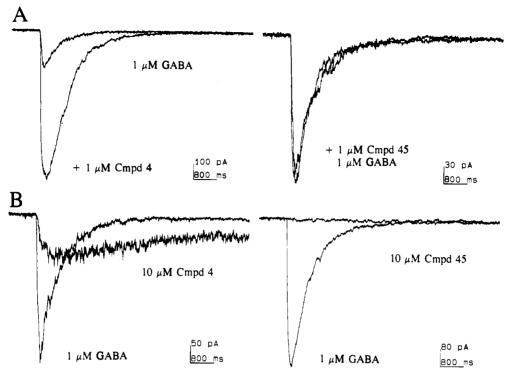


Figure 2. A steroid 17β -carbonitrile potentiates a GABA-mediated response and directly gates a chloride currents whereas its 17α carbonitrile congener does not: (A) The traces show the response of two neurons to 1 µM GABA in the absence and presence of 1 μM 4 (left) and 1 μM 45 (right). (B) The traces show the response of two neurons to 1 μM GABA or 10 μM 4 (left) and 1 μM GABA or 10 μ M 45 (right). Note the prolonged nature of the steroid-gated current compared to current gated by 1 μ M GABA. All recording were done at -60 mV and agents were applied for 200 ms.

43 and 4 still had the ability to directly activate a current, whereas benz[e]indene 35 did not. The order of potency for the compounds displaying channel activating activity in the concentration range of 1-10 μ M was 35 < 43 \simeq 4.

Discussion

Previous studies have demonstrated that the steroids which potentiate muscimol-stimulated ³⁶Cl⁻ uptake in rat brain synaptoneurosomes and/or which possess anesthetic activity are 17β -substituted. The corresponding 17α substituted analogues of these steroids are devoid of these activities. 12,14 We found that steroids 43 and 4, which are 17β-carbonitriles, are effective potentiators of GABAmediated current. These steroids also directly gate a chloride current. The corresponding steroid 17α -carbonitriles, 44 and 45, had neither activity. Thus, our electrophysiological results for steroids 4 and 43-45 establish that both potentiation of GABA-mediated current and the direct gating of a chloride current is significant only for steroids having an appropriate 17-substituent in the B-configuration. Similar results were found for the benz-[e]indenes. The benz[e]indene 3(R)-carbonitriles (34, 36, and 38), which are the analogues of the steroid 17α carbonitriles, also had no effect on either of the measured activities. The benz[e]indene 3(S)-carbonitriles (2, 35, 37, and 39), which are the analogues of the steroid 17β carbonitriles, were active compounds, and their effects were further modified by the \tilde{C}^6 substitution pattern.

The concept that methyl group substitution at the benz[e]indene C⁶ position might have electrophysiological consequences occurred to us because the initially prepared benz[e]indenes lacked two structural features found in previously well-studied anesthetic steroids. These benz-[e]indenes lacked the two carbons in the steroid A-ring (C¹ and C²), and they lacked the 19-methyl group. Thus, we reasonably questioned how the potency of benz[e]indene 2 as a potentiator of GABA current and/or a direct activator of chloride current would be altered by methyl group substitution at C⁶.

Because it was not feasible for us to generate complete dose-response curves for both activities of all the compounds, we were not able to determine if the different benz[e]indene C⁶ methyl group substitution patterns affect the dose-response relationships for the two activites in a strictly equivalent manner. However, the results do indicate that each C⁶ substitution pattern caused similar rather than opposing shifts in the dose-response curves for the two activities. Relative to compound 2, benz[e]indene 35, which contains a 6(a)-methyl group, exhibited not only enhanced GABA potentiation at 1 μ M, but also direct gating at 10 μ M. In this respect, benz[e]indene 35 is like steroids 43 and 4. However, unlike these steroids. compound 35 does not directly gate a current at 1 μ M. Thus, the major effect of the 6(a)-methyl group is not to diminish markedly the separation between the doseresponse curves for the two activities of compound 35, but to shift both dose-response curves to the left. This is an important conclusion since it indicates that the presence or absence of a methyl group in this position is not the controlling factor explaining why these dose-response curves are separated more for the 7-substituted benz-[e]indenes than they are for the congeneric anesthetic

Benz[e] indene 37, which contains a 6(e)-methyl group. is ineffective both as a potentiator of GABA currents at 1 μ M and as a direct channel activator at 10 μ M. Compound 37, however, does potentiate GABA currents at $10 \,\mu\text{M}$ indicating that the agent is not inactive, but only less potent than benz[e]indene 2. Clearly, the equatorial substituent has shifted the dose-response curve for potentiation of GABA-mediated current to the right. Most likely, the dose-response curve for direct activation has also been shifted to the right.

The opposing effects of the axial- and equatorial-methyl groups act to nullify each other when both groups are present in the same benz[e]indene. Thus, compound 39. which contains both methyl groups at C6, has electrophysiological activities that are not notably different than those of compound 2. The relative separation of the doseresponse curves for the two types of modulatory effects appears to be unaffected (relative to compound 2) in any major way by this methyl substitution pattern.

The active steroids 43 and 4, which differ by substitution with a 19-methyl group, do not differ significantly in the electrophysiological parameters we measured. This result is consistent with the earlier report that the anesthetic activities of 3α -hydroxy- 5α -pregnane-11,20-dione and 3α hydroxy-19-nor- 5α -pregnane-11,20-dione are nearly identical. 12 Nevertheless, in view of the results discussed for the 6-methylbenz[e]indenes, one might have expected that the presence or absence of a steroid 19-methyl group might have some notable effect on the electrophysiological parameters we measured. A possible reason for why the activities of the benz[e]indenes, but not the steroids, are significantly altered by methyl substituents that are similarly located in each class of compounds is suggested by molecular modeling studies.

All comparisons of three-dimensional structures were carried out on molecules whose minimum energy conformations were first determined by a molecular mechanics program. There are two energetically equivalent global minimum energy conformations for steroid 4. These conformations differ only in the orientation of the hydrogen on the oxygen of the 3α -hydroxyl group. The conformation used for molecular modeling is shown in Figure 3A. The conformation of steroid 4 that was not chosen for modeling studies has the hydrogen of the hydroxyl group rotated ~120° in a counterclockwise direction. The conclusions of the reported modeling study were not changed when this alternate minimum energy conformation of steroid 4 was used as the reference conformation for the comparison of the three-dimensional shapes of the different molecules. This does not infer that the directionality of this hydrogen atom is not important in optimizing the interactions of these molecules with their receptor binding site. It is just that the current studies do not provide any basis for preferring either orientation of the hydroxyl group in the modeling studies.

Figure 3A shows in stick drawings the result obtained when the carbon atoms of the steroid B, C, and D rings, the 18-methyl group, and the atoms in the 17β -carbonitrile group from steroid 4 are superimposed using a least-squares fitting program on the corresponding atoms in steroid 43. Orientation of the steroids relative to each other in this way makes it clear that the alleviation of the 1,3-diaxial interactions between the 19-methyl group and the 6β -, 8β and 11β -axial hydrogens of steroid 4 causes a downward displacement of the A ring. The oxygen atom of the essential 3α -hydroxyl group, which on the basis of previous structure-activity studies of anesthetic steroids12 has been postulated to serve as a hydrogen bond donor when these compounds are bound to the receptor, is displaced 0.73 Å away from the position it occupies in three-dimensional space in steroid 43. This displacement is more readily observed in Figure 3D which shows an orthogonal ball and stick drawing of only the A-ring atoms of the steroids as fit to each other in Figure 3A. The orthogonal view has

 C^3 in front of the plane of the page and C^{10} and its respective axial-hydrogen or axial-methyl substituent behind the plane of the page.

The described difference in the relative position of the 3α -hydroxy group of steroids 4 and 43 is not large. Moreover, one needs to consider that only a slight change in alignment of steroids 4 and 43 is needed to bring about the superimposition of the oxygen atoms of the 3α -hydroxyl groups and the nitrogen atoms of the 17β -carbonitirile groups of the two steroids. Thus, it is reasonable to conclude that these rigid steroids should bind similarly to the receptor site and have similar electrophysiological activities. This is what we observed in the electrophysiological evaluation of the compounds.

Conformational analysis of the benz[e]indenes is more complex. The different methyl group substitution patterns at position C⁶ of the benz[e]indenes differentially affect the conformations available to the hydroxy group located in the flexible C7 side chain. Figure 3B shows the minimum energy conformations of benz[e]indenes 2, 35. 37, and 39 that are most similar to the modeled conformations of steroids 4 and 43. An alternate orthogonal view of partial ball and stick structures is shown in Figure 3E. The compounds are superimposed on each other in the same way that the steroids shown in Figure 3, parts A and D, are superimposed. Figure 3, parts C and F, shows the superimpositon of both the steroids and the benz-[e] indenes thereby demonstrating how similar both classes of molecules are to each other when viewed in a common alignment.

The conformation shown for benz[e]indene 2 is one of 27 minimum energy conformations that were found by our conformational search strategy. The energies of these 27 conformations differed by as much as 3.09 kcal/mol. The conformer shown has an energy that is 0.17 kcal/mol above that found for the global minimum energy conformer.31 Hence, the modeled conformation of benz[e]indene 2 will be significantly populated even in the absence of additional binding interactions from the receptor that might tend to further stabilize this conformation because of its similarity to the conformation of the active steroid analogues. As aligned in Figure 3, parts C and F, the oxygen atoms of compounds 43 and 2 are separated in space by 0.11 Å. Thus, this analysis indicates that benz[e]indene 2 is an excellent conformational mimic of steroid 43, and as reported, this benz[e]indene has a potent electrophysiological effect.

The modeled conformation of benz[e] indene 35, which contains a 6(a)-methyl group, is one of 33 minimum energy conformations that were found for this molecule. The energies of these 33 conformations differed by as much as 8.17 kcal/mol. The conformer shown has an energy that is 0.16 kcal/mol above that found for the global minimum energy conformer.³¹ In the alignment shown in Figure 3, parts C and F, the oxygen atoms of compounds 43 and 35 are separated in space by 0.40 Å. Hence, like compound 2, benz[e]indene 35 is also an excellent conformational mimic of steroid 43 and this flexible steroid analogue also has potent electrophysiological effects.

Notice also that the 1,3-diaxial interaction of the 6(a)methyl group and the axial-hydrogens at C5, C8, and C9a of benz[e]indene 35 are alleviated, as discussed earlier for steroid 4, by a leftward bending of the axial-methyl group (Figure 3B). However, since the hydroxyl group of benz-[e]indene 35 resides in a flexible side chain, whereas the 3α -hydroxyl group of steroid 4 is on a semirigid ring, the

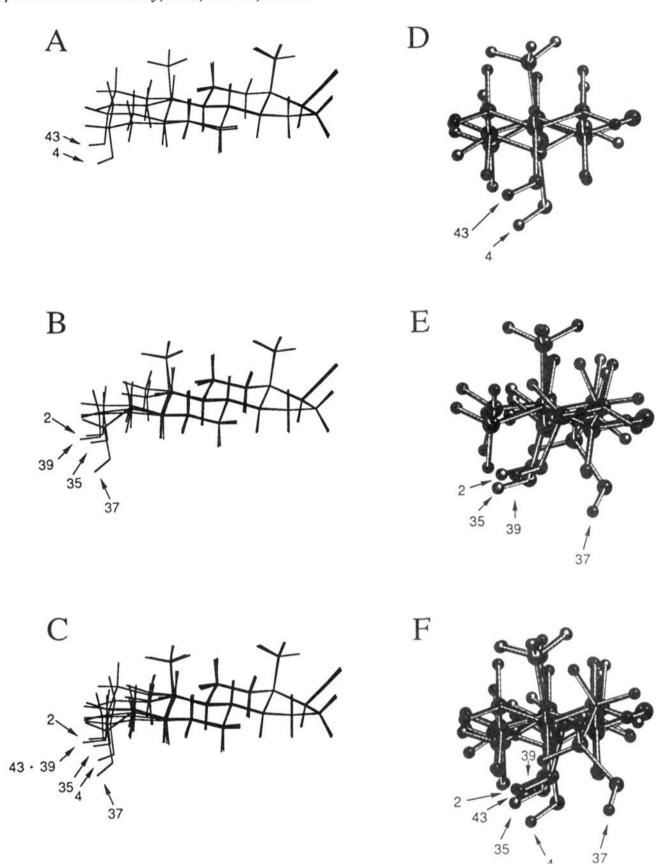


Figure 3. Molecular modeling results. Numbering of the arrows in the figure corresponds to the numbering of the compounds: (For ease of identification of the molecules, the numbered arrows point to the hydrogen atom of the hydroxyl group in each molecule.) (A) Stick drawings of minimum energy conformations of steroids 4 and 43. The molecules have been superimposed by aligning the carbon atoms of the steroid B, C, and D rings, the 18-methyl group, and the atoms in the 17β -carbonitrile group of each steroid. (B) Stick drawings of analogously superimposed minimum energy conformations of benz[e]indenes 2, 35, 37, and 39. (C) The superimposition of the molecules present in panels A and B. (D)An orthogonal ball and stick drawing of only the A-ring atoms of the steroids as fit to each other in panel A. The orthogonal view has C3 in front of the plane of the page and C10 and its respective axial-hydrogen or axial-methyl substituent behind the plane of the page. (E) An orthogonal ball and stick drawing of partial benz[e]indene structures as fit to each other in panel B. This view is analogous to that shown in panel D for the partial steroid structures. (F) The superimposition of the partial molecules present in panels D and E.

leftward bending of the axial-methyl group in compound 35 does not cause as large a downward movement of the hydroxyl group in benz[e]indene 35 as it causes for the hydroxyl group in steroid 4. Figure 3E readily demonstrates that the hydroxyl groups of compounds 2 and 35 remain close to each other in three-dimensional space even though the latter compound contains an axial-methyl group. Thus, it may be that the axial-methyl group in benz[e]indene 35 provides an additional favorable hydrophobic interaction with the receptor binding site without requiring an unfavorable movement of the hydroxyl group in the side chain. This could explain the observed increase in potency found for compound 35 relative to compound 2.

The modeled conformation of benz[e]indene 37, which contains a 6(e)-methyl group, is one of 29 minimum energy conformations that were found for this molecule. The energies of these 29 conformations differed by as much as 9.12 kcal/mol. The conformer shown in Figure 3, parts B and E, has an energy that is 2.50 kcal/mol above that found for the global minimum energy conformer,32 indicating that the modeled confomation will not be highly populated unless additional binding interactions from the receptor make this conformation more energetically favorable.

The minimization of a steric interaction between the pro-R hydrogen of the hydroxymethylene group in the side chain and the 6(e)-methyl group is responsible for the observed rightward movement of the side chain shown for benz[e]indene 37 in Figure 3E. In the alignment shown in Figure 3, parts C and F, the oxygen atoms of compounds 43 and 37 are separated in space by 1.86 Å. The diminished activities of benz[e]indene 37 in the electrophysiological evaluation can be readily explained by the fact that this compound, even if stabilized in the conformation shown by its binding interactions with the receptor, is not as close a conformational mimic of the active steroids as the other more potent benz[e]indene analogues.

The modeled conformation of benz[e] indene 39 (Figure 3, parts B and E), which contains the 6,6-dimethyl groups, is one of 30 minimum energy conformations that were found for this molecule. In the alignment shown in Figure 3, parts C and F, the oxygen atoms of compounds 43 and 39 are separated in space by 0.22 Å. The energies of the 30 conformers found for benz[e]indene 39 differed by as much as 4.87 kcal/mol. The conformer shown has an energy that is 3.82 kcal/mol above that found for the global minimum energy conformer.³³ Clearly, this conformer, which is conformationally quite similar to the modeled conformation of benz[e]indene 2, will not be heavily populated in the absence of an energetically favorable binding interaction from the receptor. If, however, the hydroxyl group does indeed form a hydrogen bond with the receptor as the earlier structure-activity studies¹² of anesthetic steroids suggest, this interaction would supply the stabilization energy necessary to maintain this receptor bound conformation of compound 39. Consequently, this line of reasoning would suggest that it is not surprising that compounds 39 and 2 have similar electrophysiological activities.

Finally, some discussion of why compound 39 is better able to mimic the conformation of steroid 43 than is compound 37 is in order. As noted earlier the presence of a 6(a)-methyl group causes 1,3-diaxial interactions that are alleviated by a leftward bending of the axial-methyl group. For compound 39 this bending of the axial-methyl group also results in a downward movement of the 6(e)methyl group. Consequently, the unfavorable steric interactions that occurred between the 6(e)-methyl group and the pro-R hydrogen of the hydroxymethylene group of compound 37 are not as severe in compound 39 and a large rightward movement of the side chain is not needed to minimize this steric interaction in compound 39.

In conclusion, the studies reported provide new information regarding the conformations of 6-methyl-substituted 7-(2-hydroxyethyl)benz[e]indenes and the activites of these compounds at GABAA receptors. Additional studies are needed to explain why the dose-response curves for the two different GABAA receptor modulatory activities of 7-(2-hydroxyethyl)benz[e]indenes are further separated for these compounds than they are for the anesthetic steroids.

Experimental Section

General Methods. All melting points were determined with a capillary melting point apparatus and are uncorrected. NMR spectra were recorded at ambient temperature in CDCl₃ (unless noted otherwise) with a 5-mm probe on either a Varian Gemini-300 operating at 300 (1H) or 75 MHz (13C). For 1H and 13C NMR spectra, the internal references were TMS (δ 0.00) and CDCl₃ (δ 77.00), respectively. IR spectra were recorded as films on a NaCl plate with a Perkin-Elmer 1710 FT-IR spectrophotometer. Elemental analyses were carried out by M-H-W Laboratories, Phoenix, AZ. Solvents were used either as purchased or dried and purified by standard methodology. Flash chromatography was performed using silica gel (32-63 μm) purchased from Scientific Adsorbants, Atlanta, GA. 3α -Hydroxy- 5α -androstan17-one was purchased from Sigma Chemical Co., St. Louis, MO. 178-Hydroxyandrostan-3-one was purchased from United States Biochemical Corp., Cleveland, OH. The Econosil HPLC column was purchased from Alltech Associates, Inc., Deerfield, IL.

 $[3S-(3\alpha,3\mathbf{a}\alpha,5\mathbf{a}\beta,6\beta,7\alpha,9\mathbf{a}\alpha,9\mathbf{b}\beta)]-3-(\mathbf{Acetyloxy})-6-(\mathbf{2-hydroxy})$ ethyl)dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetic Acid (10). A solution of an approximately 2:8 mixture of $3,17\beta$ -diacetoxyandrost-3-ene and $3,17\beta$ -diacetoxyandrost-2-ene (4) (9.7 g, 26.9 mmol) in dichloromethane (400 mL) and acetic acid (30 mL) was treated with O₃ at -78 °C until a blue color persisted. Excess O₃ was discharged by an O₂ stream and then methyl sulfide (ca. $20 \mu L$) was added. The dichloromethane was removed on a rotary evaporator, and water (90 mL) and acetic acid (200 mL) were added to the remaining solution. After the mixture was stirred overnight to hydrolyze the anhydride group generated during ozonolysis, water (200 mL) and diethyl ether (500 mL) were added. The diethyl ether layer was repeatedly washed with water to remove acetic acid and then dried over MgSO₄. Solvent removal yielded a slightly yellow solid (10.7 g) that was dissolved in methanol (220 mL), cooled to 0 °C, and reacted with slowly added portions of NaBH₄ (14.3 g, 378 mmol). After 15 min, 10% aqueous HCl was added until the solution became acidic. Water (250 mL) was added, and methanol removal on a rotary evaporator was accompanied by the precipitation of the steroid product. The precipitate was filtered and dried by air to give the crude product as a white solid (8.2 g, 87% crude yield). This material was combined with crude product (4.0 g) from a similar smaller scale reaction and recrystallized from acetone to give pure product 10 (7.4 g, 61%) as white crystals: mp 182-184.5 °C; IR 3319, 1247, 1214, 1729, 1692 cm⁻¹; ¹H NMR δ 4.58 (t, J = 10.8 Hz, 1H, CHOAc), 3.72 (m, 2H, CH₂OH), 2.62 (d, J = 12.9 Hz, 1H of CH_2COOH), 2.04 (s, 3H, OCOCH₃), 0.77 (s, 6H, CH₃ on C^{3a} and CH₃ on C^6). Anal. ($C_{21}H_{34}O_5$) C, H.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-(Acetyloxy)-6-$ (2-hydroxyethyl)dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetate (12). To a solution of compound 10 (4.0 g, 10.9)mmol) in ethyl acetate (200 mL) and absolute ethanol (20 mL) was added diazomethane in diethyl ether until a yellow color persisted. The solution was allowed to stir for an additional 15 min. Excess diazomethane was destroyed by addition of several drops of formic acid. The mixture was washed with water (50 mL), saturated aqueous NaHCO₃ (50 mL), and brine (25 mL). The organic layer was dried over Na₂SO₄. The solvent was removed to give a solid, which was purified by column chromatography (silica gel, 30% ethyl acetate in hexane) to give 3.9 g (94%) of pure product 12 as white crystals: mp 115.5-117.5 °C (from diethyl ether-hexane); IR 3453, 2934, 1732, 1438, 1373, 1246 cm⁻¹; ¹H NMR δ 4.58 (t, J = 8.5 Hz, 1H, CHOAc), 3.73 (m, 1H of CH_2OH), 3.68 (s, 3H, $COOCH_3$), 3.63 (m, 1H of CH_2OH), 2.56 (dd, J = 14.4 Hz, 2.4 Hz, 1H of CH_2COOCH_3), 2.04 (8, 3H, OCOCH₃), 0.77 (s, 6H, CH₃ on C^{3a} and CH₃ on C⁶). Anal. $(C_{22}H_{36}O_5)$ C, H.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-(Acetyloxy)-6 {f [2(\it E)$-(acetyloxy)ethenyl]} {f dodecahydro-3a,6-dimethyl-1} {\it H-}$ benz[e]indene-7-acetate (E-16) and Methyl [3S- $(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)$]-3-(Acetyloxy)-6-[2(Z)-(acetyloxy) ethenyl]-dodecahydro-3a, 6-dimethyl-1H-benz[e] indene-7-acetate (\mathbb{Z} -16). A solution of compound 12 (4.7 g, 12.3 mmol) in dichloromethane (50 mL) was added rapidly to a suspension of pyridinium chlorochromate (10.8 g, 50.2 mmol) in dichloromethane and stirred at room temperature under nitrogen. After 3 h, the volume of dichloromethane was reduced to 40 mL on a rotary evaporator and then poured into diethyl ether (750 mL). The diethyl ether was passed through a small column of Florisil, and the Florisil was washed with additional diethyl ether. The solvent was removed on a rotary evaporator to give crude methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-(acetyloxy)-6-(2-oxoethyl)$ dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetate (14) as an oil (4.6 g) which was used immediately without further purification or characterization.

A solution of crude compound 14 (4.6 g) and p-toluenesulfonic acid (0.5 g) in isopropenyl acetate (60 mL) was refluxed for 16 h. The solution was gently distilled for 1 h until about 10 mL of distillate was collected and then cooled to room temperature. The solution was poured into dichloromethane and washed with water (50 mL), 5% aqueous NaHCO₃ (50 mL), water (3 \times 100 mL), and dried over MgSO₄. The solvent was removed on a

rotary evaporator to give slightly yellow crystals (5.6 g), which were purified by chromatography (silicagel eluted with 4% ethyl acetate in dichloromethane) to give 3.7 g of product. Recrystallization from ethyl acetate—hexane gave 1.83 g (36%) of a mixture of the E and Z enol acetates. The enol acetate isomers were separated by HPLC (Econosil 5 μ m, 250 mm × 4.6 mm, eluted at 2.0 mL/min with 10% ethyl acetate in hexane).

The major 6-[2(E)-(acetyloxy)ethenyl] isomer E-16 had experimental data as follows: mp 191–192 °C; IR 2918, 1756, 1733, 1666, 1451, 1373, 1227 cm⁻¹; ¹H NMR δ 6.98 (d, J = 12.7 Hz, 1H, AcOCH=CH), 5.15 (d, J = 12.7 Hz, 1H, AcOCH=CH), 4.56 (t, J = 8.5 Hz, 1H, CHOAc), 3.63 (s, 3H, COOCH₃), 2.41 (dd, J = 15.2 Hz, J = 2.8 Hz, 1H of CH₂COOCH₃), 2.03 (s, 3H, OCOCH₃), 2.11 (s, 3H, OCOCH₃), 0.87 (s, 3H, CH₃ on C⁶), 0.76 (s, 3H, CH₃ on C^{3a}). Anal. (C₂₄H₃₆O₆) C, H.

The minor 6-[2(Z)-(acetyloxy)ethenyl] isomer Z-16 had experimental data as follows: mp 109.5–112 °C; IR 2936, 1759, 1735, 1668, 1437, 1371, 1246, 1217 cm⁻¹; ¹H NMR δ 6.70 (d, J = 7.6 Hz, 1H, AcOCH=CH), 4.58 (t, J = 8.5 Hz, 1H, CHOAc), 4.39 (d, J = 7.5 Hz, 1H, AcOCH=CH), 3.64 (s, 3H, COOCH₃), 2.44 (d, J = 14.3 Hz, 1H of CH₂COOCH₃), 2.11 (s, 3H, OCOCH₃), 2.03 (s, 3H, OCOCH₃), 1.06 (s, 3H, CH₃ on C⁵), 0.79 (s, 3H, CH₃ on C^{3a}). Anal. (C₂₄H₃₆O₆) C, H.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\alpha,7\alpha,9a\alpha,9b\beta)]$ -3-(Acetyloxy)-dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetate (20). A solution of compound 16 (200 mg, 0.48 mmol) in dichloromethane (25 mL) and HOAc (0.5 mL) was treated with O₃ at -78 °C until a blue color persisted. The excess O₃ was removed with a stream of O₂ and methyl sulfide (ca. 20 μ L) was added. Additional dichloromethane (75 mL) was added and the solution was washed with water, 5% aqueous NaHCO₃, and water and dried over Na₂SO₄. The solvent was removed to give the crude methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]$ -3-(acetyloxy)-6-formyl-dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetate (18) as a yellow oil (158 mg), which was used immediately without purification or characterization.

Crude aldehyde 18 and Wilkinson's catalyst (3×500 mg, added in equal portions, initially, and after 72 and 120 h) in benzonitrile (25 mL) was heated to 150–180 °C for 144 h under nitrogen. After cooling, ethyl acetate (50 mL) was added and the mixture was filtered. Removal of the solvents from the filtrate yielded a brown sludge (2.0 g) which was chromatographed (silica gel eluted successively with 2.5%, 5.0%, and 7.5% ethyl acetate in hexane) to give 25.4 mg (17.5%) of pure product 20 as a white solid: mp 61-63 °C; IR 2922, 1737, 1437, 1373, 1248 cm⁻¹; ¹H NMR δ 4.61 (t, J=7.8 Hz, 1H, CHOAc), 3.67 (s, 3H, COOCH₃), 2.56 (s, 3H, OCOCH₃), 0.78 (s, 3H, CH₃ on C^{3a}). Anal. (C₂₀H₃₂O₄) C, H.

 17β -(Acetyloxy)-3-oxa-A-homo- 5α -androstan-4-one (21). A mixture of compound 12 (378 mg, 1.0 mmol) and Raney nickel (1.2 g) in toluene (10 mL) was refluxed for 3.5 h under nitrogen. The mixture was cooled and filtered, and the Raney nickle was washed with EtOH (10 mL). The filtrates were combined and evaporated to give a solid which was recrystallized from EtOH to give pure compound 21 as white crystals (335 mg, 96.8%): mp 230-233 °C (lit.84 mp 218-223 °C); IR 2924, 2846, 1728, 1452, 1374, 1259 cm⁻¹; ¹H NMR δ 4.57 (t, J = 8.5 Hz, 1H, CHOAc), 4.27 $(t, J = 13 \text{ Hz}, 1 \text{H of COOCH}_2), 4.16-4.18 \text{ (m, 1 H of COOCH}_2),$ $2.85 \text{ (dd, } J = 11.4 \text{ Hz, } J = 2.9 \text{ Hz, } 1\text{H of } \text{CH}_2\text{COO), } 2.02 \text{ (s, } 3\text{H, } 1\text{Hz)}$ OCOCH₃), 0.93 (s, 3H, CH₃ on C¹⁹), 0.77 (s, 3H, CH₃ on C¹⁸); ¹³C NMR: δ 175.80 (CH₂COO), 171.10 (CH₃COO), 82.54 (C³), 64.55 $(COOCH_2)$, 12.06 $(CH_3 \text{ on } C^{18} \text{ or } C^{19})$, 12.03 $(CH_3 \text{ on } C^{18} \text{ or } C^{19})$, 53.62, 50.47, 43.26, 42.34, 41.61, 37.89, 37.46, 36.70, 34.33, 31.29, 30.30, 27.45, 23.37, 21.13, 20.73 ppm. Anal. $(C_{21}H_{32}O_4)$ C, H.

Methyl [3S-(3 α ,3a α ,5a β ,6 β ,7 α ,9a α ,9b β)]-3-(Acetyloxy)-dodecahydro-6-(1,3-dithiolan-2-yl)-3a-methyl-1H-benz[e]indene-7-acetate (22). A solution of compound 17 (1.7 g, 5.0 mmol) and ethanedithiol (690 mg, 7.5 mmol) in dry dichloromethane (20 mL) was treated with boron trifluoride etherate (1.0 mL). After the mixture was stirred for 2 h at room temperature, additional dichloromethane (50 mL) was added and the mixture was washed with cold 10% aqueous NaOH (50 mL) and brine (50 mL) and dried over Na₂SO₄. Evaporation of the solvent gave an oil, which was purified by chromatography (silica gel eluted with 15% EtOAc in hexane) to yield compound 22 as a pure colorless oil (1.75 g, 85%): IR 2923, 2851, 1734, 1435, 1373, 1245, 1036 cm⁻¹, ¹H NMR δ 5.00 (s, 1H, SCHS), 4.54 (t, J = 8.4 Hz, 1H,

CHOAc), 3.62 (s, 3H, COOCH₃), 2.00 (s, 3H, OCOCH₃), 0.76 (s, 3H, CH₃); 13 C NMR δ 173.4 (COOCH₃), 170.98 (CH₃COO), 82.58 (C³), 55.77 (SCHS), 37.58 (SCH₂CH₂S), 11.92 (C³a), 51.22, 49.81, 47.24, 45.91, 42.22, 40.53, 39.96, 37.64, 36.68, 31.92, 29.22, 27.37, 26.64, 23.23.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-(Acetyloxy)$ dodecahydro-6-(1,3-dithiolan-2-yl)-3a,6-dimethyl-1*H*-benz-[e]indene-7-acetate (23). To a solution of crude aldehyde 18 prepared by ozonolysis (see preparation of compound 20 for details) of compound 16 (1.2 g, 2.7 mmol) was added ethanedithiol (2.0 mL) and boron trifluoride etherate (1.0 mL). After the mixture was stirred for 1 h at room temperature, dichloromethane (50 mL) was added and the solution was washed with cold 10% aqueous NaOH (100 mL) and brine (100 mL), and dried over Na₂SO₄. Evaporation of the solvent give an oil, which was purified by chromatography (silica gel eluted with 25% EtOAc in hexane) to yield compound 23 as a pure colorless oil (1.10 g, 88%): IR 2927, 2851, 1734, 1435, 1373, 1293, 1039 cm⁻¹; ¹H NMR δ 4.92 (s. 1H, SCHS), 4.43 (t, J = 8.4 Hz, 1H, CHOAc), 3.51 (s, 3H, COOCH₃), 1.89 (s, 3H, OCOCH₃), 0.93 (s, 3H, CH₃ on C⁶), 0.76 (s, 3H, CH₃ on C^{3a}); ¹³C NMR δ 173.67 (COOCH₈), 170.41 (CH₃COO), 82.13 (C³), 62.87 (SCHS), 37.61 (SCH₂CH₂S), 15.03 (CH₃ on C⁶), 11.92 (CH₃ on C^{3a}), 50.98, 50.57, 50.18, 41.62, 39.61, 36.91, 36.49, 35.48, 30.26, 28.54, 27.12, 23.17, 21.28, 20.71.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-(Acetyloxy)$ dodecahydro-3a,6-dimethyl-1H-benz[e]indene-7-acetate (24). A suspension of compound 22 (1.75 g, 4.1 mmol), Raney nickle (20 g, thoroughly washed with 3×30 mL of distillated water and 3 × 50 mL of absolute EtOH) in absolute ethanol (100 mL) was refluxed for 3 h under-nitrogen. Then the mixture was filtered, and the Raney nickle was washed with EtOH (2 × 30 mL). The combined solution was evaporated to give an oil, which was purified by chromatography (silica gel eluted with 1:1:8 EtOAc/ CH₂Cl₂/hexane) to yield pure product 24 (1.1 g, 80%) as white crystals which had: mp 83-85 °C (from EtOEt-hexane); IR 2922, 2862, 1739, 1437, 1372, 1246, 1041 cm⁻¹; ¹H NMR δ 4.59 (t. J = 8.5 Hz, 1H, CHOAc), 3.66 (s, 3H, COOCH₃), 2.03 (s, 3H, OCOCH₃), 0.79 (s, 3H, CH₃ on C^{3a}); 13 C NMR δ 174.10 (COOCH₃), 171.17 (CH₃COO), 82.83 (C³), 16.42 (CH₃ on C⁶), 12.01 (CH₃ on C^{3a}), 51.36, 49.84, 49.33, 42.49, 41.46, 41.06, 40.52, 39.70, 36.92, 32.24, 30.15, 27.50, 26.07, 23.25, 21.16. Anal. (C₂₀H₃₂O₄) C, H.

Methyl $[3S-(3\alpha,3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]$ -3-(Acetyloxy)-dodecahydro-3a,6,6-trimethyl-1H-benz[e]indene-7-acetate (25). By using the same desulfurization procedure as above, a solid was obtained from compound 23 (1.1 g, 2.5 mmol) which was purified by chromatography (silica gel eluted with 4% EtOAc in hexane) to yield pure product 25 (675 mg, 77%) as white crystals: mp 98.5-99.5 °C (from EtOEt-hexane); IR 2943, 2877, 2850, 1733, 1437, 1371, 1246, 1036 cm⁻¹; ¹H NMR δ 4.58 (t, J = 7.9 Hz, 1H, CHOAc), 3.66 (s, 3H, COCH₃), 2.03 (s, 3H, OCOCH₃), 0.91 (s, 3H, ax-CH₃ on C⁶), 0.77 (s, 3H, CH₃ on C^{3a}), 0.72 (s, 3H, eq-CH₃ on C⁶); ¹³C NMR δ 174.49 (COCCH₃), 171.01 (CH₃COO), 82.68 (C³), 21.27 (eq-CH₃ on C⁶), 14.67 (ax-CH₃ on C⁶), 11.93 (CH₃ on C^{3a}), 52.89, 51.37, 50.65, 44.60, 42.21, 36.90, 36.20, 35.81, 35.21, 31.00, 27.95, 27.47, 26.39, 23.38, 21.06. Anal. (C₂₁H₃₄O₄) C, H.

 $[3S-(3\alpha,3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]-3-Hydroxydodecahydro-3a$ methyl-1H-benz[e]indene-7-ethanol (26). To a stirred solution of compound 19 (0.65 g, 2.0 mmol) in dry dichloromethane (50 mL) was added diisobutyl aluminum hydride (1.0 M solution in toluene, 12 mL, 12 mmol) at 0 °C. After 2.0 h, dichloromethanemethanol (1:1, 4 mL) and then 10% aqueous HCl (10 mL) were added. The mixture was washed with water (50 mL), saturated aqueous NaHCO₃ (50 mL), and brine (50 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a solid, which was recrystallized from ethyl alcohol to yield pure product 26 (0.47g, 93%) as white crystals: mp 145– 147 °C; IR 3279, 2914, 2870, 2858, 2841, 1469, 1443, 1381, 1348, 1067, 1056 cm⁻¹; ¹H NMR (CD₃OD) δ 3.73-3.65 (m, 3H, CH₂OH and CHOH), 0.76 (s, 3H, CH₃); ¹³C NMR (CD₃OD) δ 82.63 (C³), 60.78 (CH₂OH), 11.71 (C^{3a}), 51.36, 45.35, 44.84, 42.91, 41.29, 41.23, 37.98, 35.64, 34.28, 31.72, 30.74, 30.56, 23.93. Anal. (C₁₈H₂₈O₂) C, H.

 $[3S-(3\alpha,3a\alpha,5a\beta,6\alpha,7\alpha,9a\alpha,9b\beta)]$ -3-Hydroxydodecahydro-3a,6-dimethyl-1*H*-benz[e]indene-7-ethanol (27). By using the procedure reported for the preparation of compound 26, a solid was obtained from compound 20 (294 mg, 1.0 mmol) which was

purified by chromatography (silica gel eluted with 2:2:1 EtOAc/ hexane/CH₂Cl₂) to yield pure product 23 (253 mg, 95%) as white crystals: mp 126-128 °C (from CH₂Cl₂-hexane); IR 3343, 2920, 2872, 1445, 1386, 1055, 1028 cm⁻¹; ¹H NMR δ 3.69-3.61 (m, 3H, CH2OH and CHOH), 0.76 (s, 3H, CH3 on C6), 0.73 (s, 3H, CH3 on C^{3a}); ¹³C NMR δ 81.87 (C³), 61.10 (CH₂OH), 11.06 (CH₃ on C3a), 7.41 (CH₃ on C6), 50.84, 48.19, 43.38, 38.58, 37.29, 36.73, 36.11, 33.83, 31.50, 30.46, 26.48, 26.19, 23.23. Anal. (C₁₇H₃₀O₂)

 $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-3-Hydroxydodecahydro-$ 3a,6-dimethyl-1H-benz[e]indene-7-ethanol (28). By using the procedure reported for the preparation of compound 26, a solid was obtained from compound 24 (1.0 g, 3.0 mmol), which was purified by chromatography (silica gel eluted with 50% EtOAc in hexane) to yield pure product 24 (756 mg, 96%) as white crystals: mp 133-134.5 °C (from EtOH-EtOEt); IR 3339, 2918, 2865, 1445, 1379, 1066, 1029 cm⁻¹; ¹H NMR δ 3.73–3.61 (m, 3H, CH₂OH and CHOH), 0.75 (s, 3H, CH₃ on C^{3a}); ¹³C NMR (CD₃OD) δ 82.54 (C3), 60.99 (CH2OH), 17.01 (CH3 on C6), 11.73 (CH3 on C^{3a}), 51.58, 51.44, 49.56, 44.05, 43.02, 42.34, 42.01, 38.18, 38.11, 33.08, 31.72, 30.67, 27.59, 24.16. Anal. $(C_{17}H_{30}O_2)$ C, H.

 $[3S-(3\alpha,3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]-3-Hydroxydodecahydro-3a,6,6$ trimethyl-1H-benz[e]indene-7-ethanol (29). By using the procedure reported for the preparation of compound 26, a solid was obtained from 25 (600 mg, 1.7 mmol), which was purified by chromatography (silica gel eluted with 50% EtOAc in hexane) to yield pure product 29 (470 mg, 98%) as white crystals: mp 175-177 °C (from EtOH-EtOEt); IR 3358, 2938, 2869, 1471, $1367, 1054 \,\mathrm{cm^{-1}}; {}^{1}\text{H NMR } \delta 3.71 - 3.55 \,\mathrm{(m, 3H, C}H_{2}\text{OH and C}H\text{OH)},$ 0.91 (s, 3H, eq-CH₃ on C⁶), 0.73 (s, 3H, ax-CH₃ on C⁶), 0.72 (s, 3H, CH₃ on C^{3a}); ¹³C NMR (CD₃OD) δ 82.49 (C³), 62.31 (CH₂OH), 22.73 (eq-CH₃ on C^6), 15.24 (ax-CH₃ on C^6), 11.66 (CH₃ on C^{3a}), 55.08, 52.47, 45.57, 43.84, 38.23, 37.06, 34.90, 32.70, 30.70, 29.03, 27.02, 24.32. Anal. (C₁₈H₃₂O₂) C, H.

 $[3aS-(3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)dodecahy$ dro-3a-methyl-3H-benz[e]inden-3-one (30). To a stirred solution of compound 26 (254 mg, 1.0 mmol) in glacial acetic acid (5 mL) was added dropwise at room temperature over ~ 10 min a 5.25% aqueous solution of sodium hypochlorite (1.5 mL, 1.05 mmol). After 1.0 h, 2-propanol (2.0 mL) was added to quench any excess oxidant and water (5.0 mL) was added. The mixture was extracted with EtOAc ($2 \times 25 \text{ mL}$). The combined organic layer was washed with water (25 mL), saturated aqueous NaHCO. (25 mL), water (25 mL), and brine (25 mL) and dried over Na_2SO_4 . The solvent was removed to give an oil, which was purified by column chromatography (silica gel eluted with 1:1 ethyl acetatehexane) to give pure product 30 (161 mg, 64%) as white crystals: mp 38-40 °C (from EtOEt-hexane); IR: 3435, 2917, 1739, 1452, 1406, 1373, 1258, 1097, 1046 cm⁻¹; ¹H NMR δ 3.71 (t, J = 6.5 Hz, 2H, CH₂OH), 0.87 (s, 3H, CH₃); ¹³C NMR δ 222.05 (C=O), 60.07 (CH_2OH) , 13.44 (C^{3a}) , 50.15, 48.15, 43.55, 10.10, 39.80, 39.34, 35.46, 33.88, 32.46, 31.13, 29.38, 28.63, 21.06. Anal. (C₁₆H₂₈O₂) C, H.

 $[3aS-(3a\alpha,5a\beta,6\alpha,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)dodecahy$ dro-3a,6-dimethyl-3H-benz[e]inden-3-one (31). By using the procedure reported for the preparation of compound 30, an oil was obtained from 27 (1.6 g, 6.0 mmol) which was purified by column chromatography (silicagel eluted with 50% ethyl acetate in hexane) to give pure product 31 (1.2 g, 76%) as a colorless oil: IR 3435, 2922, 2860, 1740, 1455, 1407, 1387, 1053, 1004 cm⁻¹; ¹H NMR δ 3.58 (t, J = 6.8 Hz, 2H, CH₂OH), 0.80 (s, 3H, CH₃ on C^{3a}), $0.71 (s, 3H, CH_3 \text{ on } C^6); {}^{13}C NMR \delta 221.55 (C=O), 60.53 (CH_2OH),$ 13.55 (CH₃ on C^{3a}), 7.19 (CH₃ on C⁶), 51.16, 48.01, 38.32, 36.92, 35.81, 35.68, 33.13, 31.40, 30.61, 26.07, 25.58, 21.47. Anal.

 $[3aS-(3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)dodecahy$ dro-3a,6-dimethyl-3H-benz[e]inden-3-one (32). By using the procedure reported for the preparation of compound 30, an oil was obtained from 28 (700 mg, 2.63 mmol) which was purified by column chromatography (silica gel eluted with 50% EtOAc in hexane) to give pure product 32 (489 mg, 70%) as a colorless oil: IR 3433, 2921, 2863, 1740, 1454, 1407, 1376, 1050 cm⁻¹; ¹H NMR δ 3.73-3.64 (m, 2H, CH₂OH), 0.93 (s, 3H, CH₃ on C⁶), 0.88 (s, 3H, CH₃ on C^{3a}); 13 C NMR δ 221.61 (C=O), 60.92 (CH₂OH), 16.46 (CH₃ on C⁶), 13.74 (CH₃ on C^{3a}), 50.58, 49.84, 47.73, 41.55, 40.56, 40.25, 37.09, 35.85, 31.69, 31.60, 29.67, 25.89, 21.56. Anal. $(C_{17}H_{28}O_2)$ C, H.

 $[3aS-(3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)dodecahy$ dro-3a,6,6-trimethyl-3H-benz[e]inden-3-one (33). By using the procedure reported for the preparation of compound 30, a solid was obtained from 29 (450 mg, 1.6 mmol) which was purified by column chromatography (silica gel eluted with 40% EtOAc in hexane) to give pure product 33 (335 mg, 75%) as white crystals: mp 150-151 °C; IR 3430, 2942, 1739, 1472, 1454, 1373, 1056 cm⁻¹; ¹H NMR δ 3.72–3.57 (m, 2H, CH₂OH), 0.92 (s, 3H, eq-CH₃ on C⁶), 0.86 (s, 3H, CH₃ on C^{3a}), 0.74 (s, 3H, ax-CH₃ on C⁶); 13 C NMR δ 221.58 (C=O), 61.97 (CH₂OH), 21.10 (eq-CH₃ on C^6), 14.58 (ax-CH₃ on C^6), 13.59 (CH₃ on C^{3a}), 53.36, 51.35, 47.47, 44.06, 36.07, 35.80, 34.98, 33.79, 31.55, 30.55, 27.56, 26.30, 21.62. Anal. $(C_{18}H_{30}O_2)$ C, H.

 $[3S-(3\alpha,3a\alpha,5a\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)dodecahy$ dro-3a-methyl-1H-benz[e]indene-3-carbonitrile (2) and [3R- $(3\alpha,3a\beta,5a\alpha,7\beta,9a\beta,9b\alpha)$]-7-(2- Hydroxyethyl)dodecahydro-3a-methyl-1*H*-benz[e]indene-3-carbonitrile (34). To a stirred solution of compound 30 (320 mg 1.28 mmol) in dimethoxyethane (32 mL) and ethanol (2.0 mL) at room temperature was added a 1.0 M solution of t-BuOK in dimethoxyethane (12.8 mL, 12.8 mmol). A solution of tosylmethyl isocyanide (500 mg, 2.56 mmol) in dimethoxyethane (6.5 mL) was slowly (ca. 20 min) added from a syringe. After 3.0 h, the mixture was quenched with water (50 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layer was washed with water $(2 \times 50 \text{ mL})$ and brine (50 mL)mL) and dried over Na₂SO₄. The solvent was removed to yield an oil which was purified by column chromatography (silica gel eluted with 10% acetonitrile in dichloromethane) to give a 6:4 mixture (200 mg, 60%) of products 2 and 34, respectively, as a colorless oil. The isomers were separated by HPLC (Ultrasphere-Si, $5 \mu m$, 250-mm \times 10-mm column eluted with 30% ethyl acetate in hexane at 3.0 mL/min).

The 3S isomer, compound 2, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (90 mg, 27%): mp 82-83 °C; IR 3294, 2917, 2853, 2233, 1470, 1384, 1338, 1056, 1021 cm⁻¹; ¹H NMR δ 3.64–3.61 (m, 2H, C H_2 OH), 2.25 (t, J = 9.5 Hz, 1H, CHCN), 0.88 (s, 3H, CH₃); ¹³C NMR δ 121.43 (CN), 60.30 (CH₂OH), 14.05 (CH₃), 53.12, 44.71, 43.22, 41.34, 39.95, 39.87, 39.29, 36.76, 33.88, 32.62, 30.54, 29.03, 26.17, 23.79. Anal. $(C_{17}H_{27}NO)$ C, H, N.

The 3R isomer, compound 34, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (55 mg, 17%): mp 44-46 °C; IR 3361, 2920, 2858, 2233, 1474, 1450, 1384, 1211, 1168, 1095 cm⁻¹; ¹H NMR δ 3.66 (t, J = 6.7 Hz, 2H, CH_2OH), 2.55 (dd, J = 6.8 Hz, J = 2.0 Hz, 1H, CHCN), 0.81 (s, 3H, CH₃); 13 C NMR δ 122.48 (CN), 60.55 (CH₂OH), 17.87 (CH₃), 50.84, 44.71, 42.83, 41.43, 40.04, 39.82, 39.49, 34.85, 33.95, 32.76, 30.76, 29.17, 26.98, 24.20. Anal. (C₁₇H₂₇NO) C, H, N.

 $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydroxyethyl)$ dodecahydro-3a,6-dimethyl-1H-benz[e]indene-3-carbonitrile (35). By using the procedure reported for the preparation of compounds 2 and 34, a yellowish gum was obtained from 31 (900 mg, 3.4 mmol) which was purified by column chromatography (silica gel eluted with dichloromethane) to yield a solid. The ¹³C NMR spectra showed only the 3S isomer, compound 35, was present. The compound was purified further by HPLC (Ultrasphere-Si, $5 \mu m$, 250-mm $\times 10$ -mm column eluted with 30% ethyl acetate in hexane at 3.0 mL/min) and recrystallized from EtOEt and hexane to yield product 32 (338 mg, 36.0%) as white crystals: mp 96-98 °C; IR 3402, 2922, 2877, 1450, 1389, 1056 cm⁻¹; ¹H NMR δ 3.61 (t, J = 6.7 Hz, 2H, CH₂OH), 2.67 (s, 1H, OH), 2.28 (t, J = 9.6 Hz, 1H, CHCN), 0.90 (s, 3H, CH₃ on C^{3a}), 0.76 (d, J = 39.9 Hz, 3H, CH₃ on C⁶); ¹³C NMR δ 121.11 (CN), 60.29 (CH₂OH), 13.99 (CH₃ on C^{3a}), 7.10 (CH₃ on C⁶), 53.90, 47.48, 44.48, 39.81, 38.12, 36.85, 36.77, 35.57, 33.86, 31.56, 26.21, 26.05, 25.84, 24.11. Anal. (C₁₈H₂₉NO) C, H, N.

 $[3R-(3\alpha,3a\beta,5a\alpha,6\alpha,7\beta,9a\beta,9b\alpha)]-7-(2-Hydroxyethyl)$ dodecahydro-3a,6-dimethyl-1H-benz[e]indene-3-carbonitrile (36) and $[3S-(3\alpha,3a\alpha,5a\beta,6\beta,7\alpha,9a\alpha,9b\beta)]-7-(2-Hydrox$ yethyl)dodecahydro-3a,6-dimethyl-1H-benz[e]indene-3carbonitrile (37). By using the procedure reported for the preparation of compounds 2 and 34, a yellowish gum was obtained from 32 (450 mg, 1.7 mmol) which was purified by column chromatography (silica gel eluted with 10% acetonitrile in dichloromethane) to give a 1:1 mixture (125 mg, 27%) of products 36 and 37 as a colorless oil. The mixture was separated by HPLC (Ultrasphere-Si, 5 μ m, 250-mm × 10-mm column eluted with 30% EtOAc in hexane at 3.0 mL/min).

The 3R isomer, compound 36, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (62 mg, 13%): mp 131–133 °C; IR 3433, 2922, 2863, 2233, 1473, 1451, 1383, 1058, 1035 cm⁻¹; ¹H NMR δ 3.66–3.57 (m, 2H, CH₂OH), 2.54 (dd, J = 7.0, J = 2.0 Hz, 1H, CHCN), 0.79 (s, 3H, CH₃ on C^{3a}); ¹³C NMR δ 122.19 (CN), 60.71 (CH₂OH), 17.94 (CH₃ on C^{3a}) 16.38 (CH₃ on C⁶), 51.22, 48.98, 44.11, 41.41, 41.05, 40.40, 39.96, 37.03, 35.10, 31.79, 30.92, 27.18, 26.26, 24.56. Anal. (C₁₈H₂₉NO) C, H, N.

The 3S isomer, compound 37, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (62 mg, 13%): mp 123–125 °C; IR 3481, 2920, 2867, 2236, 1462, 1450, 1386, 1336, 1055 cm⁻¹; ¹H NMR δ 3.71–3.62 (m, 2H, CH₂OH), 2.27 (t, J = 9.5 Hz, 1H, CHCN), 0.92 (s, 3H, CH₃ on C^{3a}); ¹³C NMR δ 121.33 (CN), 60.73 (CH₂OH), 16.39 (CH₃ on C⁶), 14.21 (CH, on C^{3a}), 53.52, 49.36, 41.37, 41.04, 40.38, 40.18, 37.11, 36.95, 31.69, 30.75, 26.47, 26.19, 24.23. Anal. (C₁₈H₂₉NO) C, H, N.

[3R-(3 α ,3a β ,5a α ,7 β ,9a β ,9b α)]-7-(2-Hydroxyethyl)dodecahydro-3a,6,6-trimethyl-1*H*-benz[e]indene-3-carbonitrile (38) and [3S-(3 α ,3a α ,5a β ,7 α ,9a α ,9b β)]-7-(2-Hydroxyethyl)dodecahydro-3a,6,6-trimethyl-1*H*-benz[e]indene-3-carbonitrile (39). By using the procedure reported for the preparation of compounds 2 and 34, a yellowish solid was obtained from 33 (332 mg, 1.7 mmol) which was purified by column chromatography (silica gel eluted with 10% acetonitrile in dichloromethane) to give a 3:7 mixture (220 mg, 63%) of products 38 and 39, respectively, as a solid. The mixture was separated by HPLC (Ultrasphere-Si, 5 μ m, 250-mm × 10-mm column eluted with 30% EtOAc in hexane at 3.0 mL/min).

The 3R isomer, compound 38, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (70 mg, 20%): mp 157–159 °C; IR 3469, 2924, 2876, 2235, 1475, 1451, 1387, 1367, 1055 cm⁻¹; ¹H NMR δ 3.67–3.54 (m, 2H, CH₂OH), 2.54 (dd, J = 7.0 Hz, J = 2.0 Hz, 1H, CHCN), 0.90 (s, 3H, eq-CH₃ on C⁶), 0.78 (s, 3H, CH₃ on C^{3a}), 0.69 (s, 3H, ax-CH₃ on C⁶); ¹³C NMR δ 122.20 (CN), 61.95 (CH₂OH), 21.51 (eq-CH₃ on C⁶), 17.82 (CH₃ on C^{3a}), 14.54 (ax-CH₃ on C⁶), 52.48, 52.04, 43.89, 39.81, 35.89, 35.77, 35.09, 33.78, 31.77, 27.68, 27.16, 26.30, 24.77. Anal. (C₁₈H₃₁NO) C, H, N.

The 3S isomer, compound 39, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (150 mg, 43%): mp 175.5–177.5 °C; IR 3508, 2942, 2904, 2845, 2233, 1474, 1448, 1436, 1389, 1368, 1066 cm⁻¹; ¹H NMR δ 3.67–3.52 (m, 2H, CH₂OH), 2.42 (s, 1H, OH), 2.27 (t, J=9.6 Hz, 1H, CHCN), 0.91 (s, 3H, eq-CH₃ on C⁶), 0.88 (s, 3H, CH₃ on C^{3a}), 0.72 (s, 3H, ax-CH₃ on C⁶); ¹³C NMR δ 121.16 (CN), 61.65 (CH₂OH), 21.38 (eq-CH₃ on C⁶), 14.47 (ax-CH₃ on C⁶), 14.02 (CH₃ on C^{3a}), 54.19, 52.82, 43.96, 43.80, 39.92, 37.01, 35.81, 35.69, 33.60, 31.53, 27.49, 26.34, 26.18, 24.31. Anal. (C₁₇H₃₁NO) C, H, N.

 3α -Hydroxy- 5α -estran-17-one (41). To a cooled (-78 °C), stirred solution of 5α -estrane-3,17-dione (40, 274 mg, 1.0 mmol) in dry THF (20 mL) under nitrogen was added a solution of K-Selectride (1.0 M solution in THF, 2.0 mL, 2.0 mmol). The reaction was monitored by TLC (silica gel plate eluted with 30% EtOAc in hexane). After 1.5 h, the reaction was stopped by the addition of 10% aqueous NaOH (10 mL) followed by 30% H₂O₂ (10 mL). The reaction mixture was allowed to warm to room temperature and stirring was continuned for another half hour. The mixture was extracted with EtOAc (3 \times 50 mL). combined organic layer was washed with brine (2 × 50 mL) and dried over Na₂SO₄. Evaporation of solvent gave a white solid which was recrystallized from EtOAc to give compound 38 (237 mg, 86%) as white crystals: mp 167-168.5 °C; IR 3309, 2916, 2859, 1724 cm⁻¹; ¹H NMR δ 4.06 (t, J = 2.8 Hz, 1H, CHOH), 0.84 (s, 3H, CH₈); ¹³C NMR δ 221.63 (C=O), 66.19 (CHOH), 13.73 (CH₈), 50.55, 48.12, 47.83, 46.89, 40.63, 40.40, 35.87, 35.78, 33.31, 32.83, 31.44, 29.72, 24.80, 23.60, 21.54. Anal. (C₁₈H₂₈O₂) C, H.

 3α -Hydroxy- 5α -estrane- 17β -carbonitrile (43) and 3α -Hydroxy- 5α -estrane- 17α -carbonitrile (44). By using the procedure reported for the preparation of compounds 2 and 34, a yellowish solid was obtained from compound 41 (415 mg, 1.5 mmol) which was purified by column chromatography (silica gel eluted with 30% EtOAc in hexane) to give a 1.2:1 mixture (228 mg, 52%) of products 43 and 44, respectively, as a solid. The

mixture was separated by HPLC (Ultrasphere-Si, $5 \mu m$, 250-mm \times 10-mm column eluted with 30% EtOAc in hexane at 3.0 mL/min).

The 17β -carbonitrile, product 43, was obtained as a solid which was recrystallized from EtOAc as white crystals (118 mg, 27%): mp 193–195 °C; IR 3503, 2917, 2237, 1446, 1386 cm⁻¹; ¹H NMR δ 4.06 (t, J = 2.7 Hz, 1H, CHOH), 2.26 (t, J = 9.6 Hz, 1H, CHCN), 0.89 (s, 3H, CH₉); ¹³C NMR δ 121.34 (CN), 66.15 (CHOH), 14.23 (CH₉), 47.68, 46.74, 44.42, 41.46, 40.34, 40.22, 36.98, 35.75, 33.35, 32.79, 30.83, 26.42, 25.13, 24.25, 23.58. Anal. (C₁₉H₂₉NO) C, H, N

The 17α -carbonitrile, product 44, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (98 mg, 23%): mp 138–139 °C; IR 3468, 2919, 2862, 2233, 1446, 1384 cm⁻¹; ¹H NMR δ 3.98 (t, J=2.7 Hz, 1H, CHOH), 2.50 (dd, J=7.0 Hz, J=2.0 Hz, 1H, CHCN), 0.74 (s, 3H, CH₃); ¹³C NMR δ 122.05 (CN), 65.76 (CHOH), 17.79 (CH₃), 51.03, 47.07, 46.65, 44.05, 41.28, 40.22, 39.77, 35.46, 34.81, 33.27, 32.61, 30.83, 26.95, 25.02, 24.43, 23.42. Anal. (C₁₉H₂₉NO) C, H, N.

 3α -Hydroxy- 5α -androstane- 17β -carbonitrile (4) and 3α -Hydroxy- 5α -androstane- 17α -carbonitrile (45). By using the procedure reported for the preparation of compounds 2 and 34, a yellowish solid was obtained from compound 42 (436 mg, 1.5 mmol) which was purified by column chromatography (silica gel eluted with 30% EtOAc in hexane) to give a 1.5:1 mixture (267 mg, 59%) of products 4 and 45, respectively, as a solid. The mixture was separated by HPLC (Ultrasphere-Si, 5 μ m, 250-mm \times 10-mm column eluted with 30% ethyl acetate in hexane at 3.0 mL/min).

The 17β -carbonitrile, product 4, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (160 mg, 35%): mp 159–160.5 °C (lit. 14 mp 156–158 °C); IR 3435, 2923, 2236, 1449, 1387, 1267, 1166 cm⁻¹; 14 NMR δ 3.99 (t, J = 2.5 Hz, 1H, CHOH), 2.23 (t, J = 9.6 Hz, 1H, CHCN), 0.86 (s, 3H, 18-CH₃), 0.74 (s, 3H, 19-CH₃); 13C NMR δ 121.27 (CN), 66.09 (CHOH), 14.20 (18-CH₃), 11.03 (19-CH₃), 44.26, 40.37, 38.80, 36.97, 35.93, 35.70, 35.56, 32.00, 31.77, 28.76, 28.15, 26.37, 24.37, 20.26.

The 17α -carbonitrile, product 45, was obtained as a solid which was recrystallized from EtOEt and hexane as white crystals (107 mg, 24%): mp 183–185 °C (lit. 14 mp 183–185 °C); IR 3480, 2926, 2234, 1450, 1386, 1072 cm⁻¹; 14 NMR δ 4.02 (t, J=2.5 Hz, 1H, CHOH), 2.54 (dd, J=7.0 Hz, J=2.0 Hz, 1H, CHCN), 0.78 (s, 3H, 18-CH₃), 0.76 (s, 3H, 19-CH₃); 13 C NMR δ 122.24 (CN), 66.24 (CHOH), 17.98 (18-CH₃), 11.06 (19-CH₃), 53.41, 52.04, 44.14, 39.88, 38.77, 35.97, 35.73, 35.02, 32.03, 31.96, 28.82, 28.24, 27.15, 24.70, 20.35.

Electrophysiology. Hippocampal cultures were prepared from 1-2 day old albino rat pups and maintained as described previously.35 Experiments were carried out at room temperature (~22 °C) using cultures that had been maintained in vitro for 3-10 days. At the time of an experiment the growth media was exchanged for a solution containing (in mM): 140 NaCl, 5 KCl, 2 CaCl₂, 2 MgCl₂, 10 glucose, 10 N-(2-hydroxyethyl)piperazine-N'-2-ethanesulfonic acid (HEPES), and 0.001 tetrodotoxin (TTX) with pH adjusted to 7.3. TTX was included to block voltagegated Na+ currents and to diminish spontaneous synaptic currents. Voltage clamp recordings were obtained using wholecell patch clamp methods.30 Recording electrodes were fashioned from 1.2-mm borosilicate glass capillaries (World Precision Instruments) using a Flaming-Brown P-87 horizontal pipet puller (Sutter Instruments) and had resistances of 5–8 M Ω when firepolished and filled with a solution containing (in mM): 140 CsCl, 4 NaCl, 4 MgCl₂, 0.5 CaCl₂, 10 HEPES, and 5 ethylene glycol bis(β-aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA) with pH adjusted to 7.3 using CsOH. Currents were filtered at 1.5 kHz and were digitized at 0.25 kHz using pCLAMP V 5.5 (Axon Instruments). Data were analyzed using pCLAMP V 5.5, Sigmaplot V 4.0 and routines written in Axobasic. The data in this paper represents mean \pm SEM.

GABA stock solutions were prepared in the extracellular solution. Test compound stock solutions were prepared in DMSO and were diluted with the extracellular solution at the time of an experiment. The final DMSO concentration was <0.2%, a concentration that does not alter GABA currents in hippocampal neurons. Compounds were applied by pressure ejection from pipets positioned within 5 μ m of the recorded neuron using a 100–500-ms jet of compressed air at 10–20 psi. This system allows

no discernable drug leakage between applications and affords reliable repeated drug delivery. The concentrations of drugs reported are those in the pipet. The actual concentration at the cell is likely to be less due to diffusion and the fact that the entire cell is not uniformly exposed to the pipet contents.

Molecular Modeling. Molecular modeling was performed on a SiliconGraphics Iris Indigo Elan 4000 computer using the Sybyl Molecular Modeling Software, Version 6.0 from Tripos Associates, Inc., St. Louis, MO. Conformational searches were carried out on steroids starting from an initial minimum energy conformation with rotations in 120° increments for the carbonoxygen bond of the 3α -hydroxyl group. Conformations generated in this way were then minimized using the Powell minimization option (without charges) in the Sybyl program. Conformational searches for the benz[e] indenes were similar except that rotations about the C7-C1'bond (i.e., the bond connecting the side chain to the ring) were carried out in $\sim 60^{\circ}$ increments, whereas the $C^{1'}$ - $C^{2'}$ and $C^{2'}$ -Obonds in the side chain were rotated in $\sim 120^{\circ}$ increments. Conformational searches for the benz[e]indenes started with minimum energy conformations that most closely mimicked a steroid conformation.

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- (31) For benz[elindenes 2 and 35, the global minimum energy conformations differ from the modeled conformations only by an ~120° counterclockwise rotation of the hydrogen atom of the hydroxyl
- (32) The global minimum energy conformation has the hydroxymethylene group oriented in space at a position similar to where it would be located if it were a 4-ax-substituent on a saturated steroid A-ring.
- The global minimum energy conformation has the hydroxymeth ylene group oriented in space at a position similar to where it would be located if it were a 4-eq-substituent on a saturated steroid A
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