Xanthines with C⁸ Chiral Substituents as Potent and Selective Adenosine A₁ Antagonists

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Several 8-substituted 1,3-dipropylxanthines were synthesized, and their receptor binding affinities at adenosine A_1 and A_2 receptors were measured. When enantiomeric pairs of compounds were examined, the *R* enantiomers were significantly more potent than the corresponding *S* enantiomers. The most potent compound at the A_1 receptor was (*R*)-3,7-dihydro-8-(1-methyl-2-phenylethyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5a; MDL 102,503), whose K_i value at the A_1 receptor was 6.9 nM. However, a more selective compound was (*R*)-3,7-dihydro-8-(1-phenylpropyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5d; MDL 102,234), which had a K_i value of 23.2 nM at the A_1 receptor and an A_2/A_1 ratio of 153.

Introduction

The recognition of purinoceptors¹ in peripheral cell membranes, specifically the A₁ and A₂ receptors,² has stimulated a surge of activity in adenosine research.³ The host of suggested therapeutic utilities for adenosine receptor ligands has prompted the design of potent and selective adenosine A₁ and A₂ antagonists and agonists. Indeed, there have been successes with three of these four targets. For example, several sets of 8-substituted 1,3dipropylxanthines with selectivity and good potency for the A₁ receptor have been described.⁴⁻⁷ Certain C⁶ N-substituted adenosines are A₁-selective agonists.⁸ And adenosines and adenosine-5'-(N-substituted carboxamides) with alkylamino and aralkylamino substituents at the 2-position have shown potency and moderate selectivity as A₂ agonists.^{9,10}

Certain substitution patterns have been found to favor A_2 -selectivity in xanthines. Very recently it has been shown that xanthines bearing an (E)-3,4-dimethoxystyryl or (E)-3,4,5-trimethoxystyryl group at the 8-position in combination with a methyl group at the 7-position were A_2 -selective antagonists.¹¹ Selectivity and potency of known adenosine receptor ligands, as well as a historical perspective on adenosine receptor research and potential areas of therapeutic intervention, are topics which have been recently and comprehensively reviewed.¹²

We have recently proposed a new binding mode for xanthines with respect to adenosine at adenosine receptors.^{13,14} This new binding mode has recently been compared to other binding models and referred to as the "N6-C8" model.¹⁵ To substantiate our model we prepared the optical isomers of 1.3-dipropyl-8-(phenylisopropyl)xanthines and showed that the R enantiomer was substantially more potent than the S enantiomer at adenosine A₁ receptors, analogous to the enhanced potency of N^6 -[(R)-1-methyl-2-phenylethyl] adenosine (R-PIA) with respect to S-PIA. This report describes additional xanthines with chiral substituents at the 8-position which also display a marked stereochemical requirement for receptor affinity, as well as related racemic 8-substituted xanthines which help define structure-activity relationships for potency at the A_1 receptor.

Chemistry

The synthetic route which we employed for the preparation of both racemic chiral 8-substituted xanthines is shown in Scheme I. Treatment of 1,3-di-*n*-propyl-5,6-diaminouracil (1)¹⁶ with the carboxylic acids gave 6-(acylamino)uracils 2. For the preparation of racemic 8-substituted xanthines 4 we cyclized 2 with ethanolic potassium hydroxide.^{16,17} The preparation of xanthines 4 bearing a chiral substituent at position 8 was achieved without the use of strong base. Thus, compounds 2 were treated with Meerwein's reagent to afford imino ethers 3, which were thermally cyclized in benzene at reflux to afford xanthines 4.¹³ This conversion proceeds very cleanly and in good yield and can easily be monitored by thin-layer chromatography. Interestingly, this cyclization is formally a 5-endo-trig, anti-Baldwin closure.^{18,19}

Table I lists the 8-substituted xanthines which were prepared using the routes shown in Scheme I. The racemic compounds were prepared directly from 3 by dehydrative cyclization with potassium hydroxide, while the chiral compounds were prepared by thermal cyclization of imino ethers 4. 13,20

Several carboxylic acids, both racemic and optically active, were required for this study. Phenylisobutyric acid 5 was prepared from propanoic acid by treating the dianion, which was made with 2 equiv of lithium diisopropylamide, with benzyl chloride. Enantiomers 5a and 5c were prepared using enzymatic resolution techniques as previously described.^{13,20} Likewise, racemic homologs 5j²¹ and $5k^{22}$ were similarly prepared in yields of 77% and 87%, respectively. Carboxylic acids 5d-i, 5l, and 5n were commercially available. Indan-2-carboxylic acid (5m) was prepared by alkylation of diethyl malonate with α, α' dibromo-o-xylene, followed by hydrolysis and decarboxylation of the resulting diester.^{23,24} cis-2-Chloro-6-phenylcyclohexanone²⁵ was prepared by treating 2-phenylcyclohexanone with sulfuryl chloride and subjected to Favorskii conditions to give trans-2-phenylcyclopentanecarboxylic acid (5g).²⁶ Scheme II describes the synthesis of a protected version of 2-benzyl-3-hydroxypropanoic acid (5p), 2-[(tert-butyldimethylsilyl)methyl]-3-phenylpropanoic acid (10), which initiated from β -propiolactone (6). Treatment of 6 with methanol and triethylamine gave 3-hydroxypropanoic acid methyl ester (7) which was alkylated with benzyl bromide to afford ester 8. The tert-

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Scheme I. Synthesis of 8-Substituted Xanthines^a



^a Reagents: (a) N-methylmorpholine, isobutyl chloroformate, THF/DMF, -20 °C; (b) KOH, H₂O-EtOH, reflux, 2 h; (c) triethyloxonium tetrafluoroborate, benzene, 50 °C, 15 h; (d) silica gel chromatography; (e) dry benzene, reflux, 2 h.

butyldimethylsilyl ether of 8 (9) was treated with aqueous potassium hydroxide to hydrolyze the ester. The resulting ester 10 was used in the xanthine synthesis of Scheme I, during which the *tert*-butyldimethylsilyl group was removed.

Structure-Activity Relationships

The (R)-phenylisopropyl group at the 8-position of the xanthine nucleus (compound 5a) confers the best potency at the A_1 adenosine receptor of the list of compounds in Table I. However, this chiral recognition unit does not provide much A_1 selectivity, with an A_2/A_1 ratio of ca. 23. Compound 5d, which is isomeric with 5a, is about 3 times less potent at the A_1 receptor than 5a but is significantly more selective, with an A_2/A_1 ratio of 153. Compound 5g, a close analog of 5d whose carbon skeleton in the chiral recognition unit is the same as 5d with the substitution of methyl for ethyl, has a binding profile very similar to that of 5d. Compounds 5h and 5i, the analogous racemate and S enantiomer, are also strictly analogous to their counterparts 5e and 5f. The significantly greater potencies of R enantiomers 5a, 5d, and 5g over their corresponding S enantiomers 5c, 5f, and 5i demonstrate the marked stereochemical requirement for receptor affinity of the substituent at position 8 on the xanthine antagonist. With all three pairs of enantiomers in Table I, the affinity and selectivity values for the racemates fall between those values registered for the enantiomers, which adds a checkpoint to the validity of the data.

Extension of methyl to ethyl and propyl in the phenylisopropyl group of 5b gave homologs 5j and 5k, respectively, which were less potent at the A1 receptor and less A_1 selective. Addition of a hydroxy group to the methyl group of 5b gave compound 5p, which was markedly less potent at both A_1 and A_2 receptors. This addition of hydrophilicity suggests an unfavorable interaction with the receptors. In a model of the G-protein-coupled adenosine A1 receptor which we recently described,27 the proposed ligand binding site contains several hydrophobic amino acid residues in the pocket which accepts the xanthine substituent at the 8-position. Favorable interactions of a hydrophobic C⁸ substituent with these residues, which are located on helices III, IV and VI, may be necessary for good binding affinity. Moving the methyl group of **5b** to the β -position of the side chain (compound



<u></u>		stereo-	Ki, nM		
compd	R	chem	A ₁ receptor ^a	A_2 receptor ^b	A_2/A_1
5a °	H CH	R	6.9 ± 1.6^{d}	157 ± 27^{d}	22.8
5b	H CH	racemic	32.6 ± 4.6^{d}	644 ± 209 ^d	19.8
5c		S	60.7 ± 5.3^{d}	848 ± 99 ^d	14.0
5d°		R	23.2 ± 3.5	3510 ± 250	153
5e		racemic	33.5 ± 1.6	3210 ± 1000	95
5f		S	136.1 ± 21.6	7500 ± 1600	55
5 g	<u>н</u> , сн _а	R	25.3 ± 1.45	4220 ± 590	160
5 h	H₃C H	racemic	49.4 ± 5.3	4900 ± 2000	99
5 i		S	174.7 ± 15.7	12900 ± 1500	74
5j		racemic	161.2 ± 27.2	1230 ± 330	8
5 k	CH CH	racemic	73.8 ± 8.7	610 ± 150	8
51		racemic	94.3 ± 6.4	1740 ± 240	18
5m			64.3 ± 3.6	8350 ± 1500	129
5 n	-	racemic	94.6 ± 17.2	10300 ± 2700	108
5p		racemic	1294 ± 10.7	12600 ± 1790	10
5q	\Rightarrow	racemic	164.3 ± 8.2	2720 ± 160	10
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^a Binding of [³H]CHA in whole rat brain membranes was measured at 25 °C. Values are geometric means \pm standard error, n = 3 separate determinations. See: Goodman, R.; Cooper, M.; Gavish, M.; Snyder, S. Mol. Pharmacol. 1982, 21, 329. ^b Binding of [³H]NECA was measured in rat brain striatum at 25 °C. Values are geometric means \pm standard error, n = 3 separate determinations. See: Bruns, R. R.; Lu, G. H.; Pugaley, T. A. Mol. Pharmacol. 1986, 29, 331. ^c MDL 102,503. ^d Reference 13.^e MDL 102,234.

Scheme II. Synthesis of Acid 10^a



^a Reagents: (a) Et₃N, CH₃OH, 3 days; (b) LDA, -50 °C, C₆H₅CH₂Br, -20 °C; (c) TBDMSCl, imidazole, DMF, 1.5 h; (d) 30% KOH, 0 °C, 5 h.

51) also reduced affinity, approximately 3-fold at both A_1 and A_2 receptors, without affecting selectivity.

Three conformationally restricted versions of the phenylisopropyl compound **5b** were prepared, all of which were less potent than **5b**. The indanyl-substituted xanthine **5m**, in which the methyl group in the side chain of **5b** is connected directly to the aromatic ring, was ca. 2-fold less potent than **5b** at the A₁ receptor. However, **5m** was more A₁ selective than **5b** by a factor of 6. The tetrahydronaphthyl-substituted xanthine **5n**, in which the methyl group of **5b** is tethered to the aromatic group via a methylene spacer, was ca. 3-fold less potent at the A₁ receptor and similarly more selective. *trans*-2-Phenylcyclopentyl compound **5q**, in which the methyl group in the side chain of **5b** is tethered to the side chain methylene group via an ethylylene spacer, was ca. 5-fold less potent than **5b** at the A₁ receptor, with the same A₁ selectivity.

The restrictions that were imposed with compounds 5m, 5n, and 5q demonstrate that affinity at the A₁ receptor is better for the unrestricted 5b than for compounds wherein the methyl group is tethered to the substituent in three different ways. Certainly, additional conformational restrictions could be applied to 5b in an attempt to freeze a side-chain conformation which is precisely accommodated by the A₁ receptor pocket.

Conclusion

Several 8-substituted 1,3-dipropylxanthines were prepared, and their binding affinities to adenosine A_1 and A_2 receptors were measured as shown in Table I. Three pairs of compounds were prepared where the first carbon of the substituent was chiral. In all three cases there was a marked stereochemical requirement for affinity, i.e., the R enantiomers were significantly more potent than the Senantiomers. The most potent compound of the series at the adenosine A_1 receptor was (R)-3,7-dihydro-8-(1-methyl-2-phenylethyl)-1,3-dipropyl-1H-purine-2,6-dione (5a; MDL 102,503), having a K_i value of 6.9 nM. However, the A_2/A_1 ratio for MDL 102,503 was only 23. An isomer of 5a, (R)-3,7-dihydro-8-(1-phenylpropyl)-1,3-dipropyl-1H-purine-2,6-dione (MDL 102,234, 5d), was less potent but more A_1 selective, with a K_i value of 23.2 nM at the A₁ receptor and an A₁/A₁ ratio of 153. Compound 5d (MDL 102,234) is currently undergoing evaluation in cognition enhancement studies.28

Experimental Section

Melting points were determined with a Thomas-Hoover capillary melting point apparatus and are uncorrected. IR spectra were recorded with a Perkin-Elmer Model 727B spectrophotometer and NMR spectra with Varian Gemini 300 and Varian VXR-300 spectrometers; MS data were collected at 70 eV with a Finnigan TCQ GC/MS/MS instrument, and HRMS data were collected at 70 eV with a VG ZABZ-SE spectrometer, using computerized peak matching with perfluorokerosene as the reference and a resolution of 10 000. Chemical shifts for ¹H NMR signals are reported in ppm downfield from TMS (δ). TLC analyses were performed with Merck DC-F254 on Analtech GHLF silica gel plates, with visualization by alkaline permanganate and UV irradiation. Gas chromatography was performed on an HP 5890A fitted with a 15-m × 0.321-mm DB-5 column. Optical rotations were determined on a JASCO DIP-360. Flash chromatography was performed using Merck silica gel, 230-400 mesh. A Model 7924T Chromatotron from Harrison Research was used for radial chromatography. All reactions were run under an atmosphere of nitrogen using commercially dried solvents.

Carboxylic acids used for the preparation of 8-substituted xanthines using the route shown in Scheme I were (i) prepared as previously described in the literature and referenced accordingly; (ii) prepared using new routes, as shown in Scheme II for 10 and as in refs 19 and 29 for 2a-c; or (iii) obtained commercially: (R)-, (S)-, and (\pm) -2-phenylbutanoic acids (2d, 2f, and 2e, respectively); (R)-, (S)-, and (\pm) -2-phenylputanoic acids (2g, 2i, and 2h, respectively); 3-phenylbutanoic acid (21); and 1,2,3,4-tetrahydro-2-naphthoic acid (2n).

(S)-N-(6-Amino-1,2,3,4-tetrahydro-2,4-dioxo-1,3-dipropyl-5-pyrimidinyl)- α -methylbenzenepropanamide (3a). (R)-2-(Phenylmethyl)propanoic acid (2a)^{20,29} (0.69 g, 4.6 mmol) was dissolved in 15 mL of THF and cooled to -20 °C. The solution was treated with N-methylmorpholine (0.46 mL, 4.6 mmol) followed by dropwise addition via syringe of isobutyl chloroformate (0.60 mL, 4,6 mmol). After the mixture was stirred at -20 °C for 30 min, 1,3-dipropyl-5,6-diaminouracil¹⁶ (0.84 g, 3.8 mmol) in 5 mL of DMF was added. After being stirred for 4 h at -20 °C, the reaction mixture was allowed to warm to room temperature overnight. The solvent was removed in vacuo, and the residue was purified by flash chromatography (5% to 10% to 20% IPA/ hexane) to give amide 3a (0.87 g, 64%) as a foam: $[\alpha]^{20} - 42.7^{\circ}$ (c 0.82, CHCl₃); ¹H NMR (CDCl₃) δ 7.40-7.18 (m, 6 H), 4.98 (s, 2 H), 3.83 (m, 4 H), 3.00 (m, 1 H), 2.80 (m, 2 H), 1.75-1.55 (m, 4 H), 1.28 (d, J = 6.7 Hz, 3 H), 0.96 (t, J = 8.6 Hz, 3 H), 0.90 (t, J = 3 H); MS (70 eV, CI, CH₄) m/z 373 (M⁺ + 1), 401 (M⁺ + 29), 413 (M^+ + 41); exact mass calcd for C₂₀H₂₆N₄O₃ 372.2161, found 372.21**6**3.

(R)-3,7-Dihydro-8-(1-methyl-2-phenylethyl)-1,3-dipropyl-1H-purine-2,6-dione (5a). Amide 3a (0.85 g, 2.28 mmol) was dissolved in 100 mL of benzene and treated with 1 M triethyloxonium tetrafluoroborate in CH₂Cl₂ (14.8 mL, 14.8 mmol) with stirring. The solution was heated to 50 °C for 15 h. After cooling, the solution was poured into 500 mL of Et₂O, rinsed with 300 mL of 0.1 M phosphate buffer, and 200 mL of saturated NaCl, and dried over MgSO4. After filtering, the solvent was removed in vacuo and the residue purified by radial chromatography (2% to 5% methanol/CHCl₃) to yield the unstable imino ether 4a (0.36 g. 39%), which was used immediately in the next step. Imino ether 4a (0.36 g, 0.9 mmol) was dissolved in 100 mL of benzene and heated at reflux for 3 h. The solvent was removed in vacuo, and the residue was purified by radial chromatography (50% ethyl acetate/hexane, 2-mm plate) to yield 5a (0.23g). The solid was recrystallized from 20% Et₂O/hexane and dried under high vacuum at 30 °C to yield pure 5a (187 mg, 59%) as a white solid: $[\alpha]_{D}^{20} - 42^{\circ}$ (c 0.75, CHCl₃); mp 141-142 °C; ¹H NMR (CDCl, δ 12.29 (s, 1 H), 7.22–7.09 (m, 5 H), 4.11 (t, J = 7.3 Hz, 2 H), 4.03 (t, J = 7.8 Hz, 2 H), 3.39-3.20 (m, 2 H), 2.95 (dd, J =13.7 Hz, J = 7.3 Hz, 1 H), 1.85–1.65 (m, 4 H), 1.41 (d, J = 7.0 Hz, 3 H), 0.98 (m, 6 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 $(M^+ + 29)$, 395 $(M^+ 41)$. Anal. $(C_{20}H_{28}N_4O_2)$ C, H, N.

(±)-3,7-Dihydro-8-(1-phenylpropyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5e). 2-Phenylbutanoic acid (2e; 1.1 g, 6.4 mmol) was dissolved in 15 mL of THF and treated with *N*-methylmorpholine (0.58 mL, 5.3 mmol). The solution was cooled to -20°C, and isobutyl chloroformate (0.69 mL, 5.3 mmol) was added dropwise via syringe with stirring. After 20 min, 1,3-dipropyl-5,6-diaminouracil (1.2 g, 5.3 mmol) in 5 mL of DMF was added. After stirring at -20 °C for 4 h, the reaction mixture was allowed to warm to room temperature overnight. The mixture was then diluted with 500 mL of CHCl₃, rinsed with 300 mL of saturated NaHCO₃, dried over MgSO₄, filtered, and concentrated under vacuum to yield crude 3a (1.97 g). This intermediate was used immediately in the next step without further purification. The crude amide 3e (1.97 g, 5.3 mmol) was dissolved in 40 mL of ethanol and 100 mL of 30% KOH, and the solution was heated at reflux for 2 h. The solution was then cooled to 0 °C and cautiously acidified with dilute HCl (42 mL of concentrated HCl in 300 mL of H_2O). The mixture was extracted with CHCl₃ (3 \times 200 mL), and the combined organic extracts were dried over MgSO₄, filtered, and concentrated under vacuum. The residue was purified by radial chromatography (25% to 50% ethyl)acetate/hexane, 4-mm plate) and triturated wth 5% Et₂O/hexane. The white solid was dried under high vacuum at 39 °C to yield 5e (454 mg, 24%): mp 137-138 °C; ¹H NMR (CDCl₃) δ 12.45 (s, 1 H), 7.42 (d, J = 8.6 Hz, 2 H), 7.38–7.20 (m, 3 H), 4.20–4.00 (m, 5 H), 2.38 (m, 1 H), 2.19 (m, 1 H), 1.90-1.70 (m, 4 H), 0.98 (m, 9 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 (M⁺ + 29), 395 $(M^+ + 41)$. Anal. $(C_{20}H_{26}N_4O_2)$ C, H, N.

The following racemic compounds were obtained using a method similar to the preparation of 5e.

3,7-Dihydro-8-(1-methyl-2-phenylethyl)-1,3-dipropyl-1*H***-purine-2,6-dione (5b).** Compound **5b** (570 mg, 77%) was prepared from 1 and 2-methyl-3-phenylpropanoic acid and isolated as a white solid: mp 140–142 °C; ¹H NMR (CDCl₃) δ 12.61 (s, 1 H), 7.22–7.09 (m, 5 H), 4.13 (t, J = 7.4 Hz, 2 H), 4.05 (t, J = 8.1 Hz, 2 H), 3.38 (dd, J = 14 Hz, J = 7.1 Hz, 1 H), 3.25 (dd, J = 7.6 Hz, 1 H), 2.96 (dd, J = 13.4 Hz, J = 7.4 Hz, 1 H), 1.85–1.65 (m, 4 H), 1.43 (d, J = 6.7 Hz, 3 H), 0.98 (m, 6 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 (M⁺ + 29), 395 (M⁺ + 41). Anal. (C₂₀H₂₈N₄O₂) C, H, N.

3,7-Dihydro-8-(1-phenylethyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5h). Compound 5h (584 mg, 65%) was prepared from 1 and 2-phenylpropanoic acid and isolated as a white solid: mp 148-150 °C; ¹H NMR (CDCl₃) δ 11.81 (s, 1 H), 7.40-7.20 (m, 5 H), 4.35 (q, J = 7.0 Hz, 2 H), 4.11 (t, J = 7.7 Hz, 2 H), 4.00 (t, J = 7.8 Hz, 2 H), 1.90-1.60 (m, 4 H), 1.78 (d, J = 7.6 Hz, 3 H), 0.99 (t, J = 7.4 Hz, 3 H), 0.95 (t, J = 7.4 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 341 (M⁺ + 1), 369 (M⁺ + 29), 381 (M⁺ + 41). Anal. (C₁₉H₂₄N₄O₂) C, H, N.

3,7-Dihydro-8-[1-(phenylmethyl)propyl]-1,3-dipropyl-1*H***-purine-2,6-dione (5j).** Compound **5j** (407 mg, 35%) was prepared from 1 and 1-(phenylmethyl)butanoic acid²¹ and isolated as a white solid: mp 186–188 °C; ¹H NMR (CDCl₃) δ 12.41 (s, 1 H), 7.20–7.03 (m, 5 H), 4.12 (t, J = 7.5 Hz, 2 H), 4.02 (t, J = 7.6 Hz, 2 H), 3.20–3.00 (m, 3 H), 1.95–1.65 (m, 6 H), 0.98 (m, 3 H), 0.96 (t, J = 7.6 Hz, 3 H), 0.89 (t, J = 7.5 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 369 (M⁺ + 1), 397 (M⁺ + 29), 401 (M⁺ + 41); exact mass calcd for C₂₁H₂₆N₄O₂ 368.2212, found 368.2197.

3,7-Dihydro-8-[1-(phenylmethyl)butyl]-1,3-dipropyl-1*H***-purine-2,6-dione (5k).** Compound **5k** (217 mg, 41%) was prepared from 1 and 2-(phenylmethyl)pentanoic acid²² and was isolated as a white solid: mp 158–160 °C; ¹H NMR (CDCl₃) δ 12.15 (s, 1 H), 7.20–7.00 (m, 5 H), 4.12 (t, J = 7.6 Hz, 2 H), 4.01 (t, J = 7.6 Hz, 2 H), 3.30–2.97 (m, 3 H), 1.98–1.65 (m, 6 H), 1.25 (m, 2 H), 0.97 (t, J = 7.3 Hz, 6 H), 0.88 (t, J = 7.1 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 383 (M⁺ + 1), 411 (M⁺ + 29), 423 (M⁺ + 41). Anal. (C₂₂H₃₀N₄O₂) C, H, N.

3,7-Dihydro-8-(2-phenylpropyl)-1,3-dipropyl-1*H*-purine-**2,6-dione (51).** Compound **51** (180 mg, 8%) was prepared from 1 and 3-phenylbutanoic acid and isolated as a white solid: mp 136-137 °C; ¹H NMR (CDCl₃) δ 12.59 (s, 1 H), 7.25-7.10 (m, 5 H), 4.11 (t, J = 7.3 Hz, 2 H), 4.03 (t, J = 7.5 Hz, 2 H), 3.40 (m, 1 H), 3.20-3.00 (m, 2 H), 1.88-1.65 (m, 4 H), 1.34 (d, J = 7.2 Hz, 3 H), 0.98 (t, J = 7.3 Hz, 3 H), 0.95 (t, J = 7.3 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 (M⁺ + 29), 395 (M⁺ + 41). Anal. (C₂₀H₂₆N₄O₂) C, H, N.

3,7-Dihydro-8-(2-indanyl)-1,3-dipropyl-1*H*-**purine-2,6-dione (5m).** Compound **5m** (1.10 g, 53%) was prepared from 2-indancarboxylic acid^{23,24} and isolated as a white solid: mp 223-224 °C; ¹H NMR (CDCl₃) δ 12.82 (s, 1 H), 7.30–7.18 (m, 4 H), 4.10 (t, *J* = 8.0 Hz, 2 H), 3.95 (quin, *J* = 9.0 Hz, 1 H), 3.81 (t, *J* = 7.5 Hz, 2 H), 3.50–3.38 (m, 4 H), 1.81 (m, 2 H), 1.58 (m, 2 H), 0.97 (t, *J* = 7.7 Hz, 3 H), 0.79 (t, *J* = 7.1 Hz, 3 H); MS (70 eV, CI, CH₄) *m/z* 353 (M⁺ + 1), 381 (M⁺ + 29), 393 (M⁺ + 41). Anal. (C₂₀H₂₄N₄O₂) C, H, N.

3,7-Dihydro-1,3-dipropyl-8-(1,2,3,4-tetrahydro-2-naphthalenyl)-1*H***-purine-2,6-dione (5n). Compound 5n (1.04 g, 55%)** was prepared from 1 and 1,2,3,4-tetrahydro-2-naphthoic acid and isolated as a white solid: mp 202-204 °C; ¹H NMR (CDCl₃) δ 12.63 (s, 1 H), 7.15 (m, 4 H), 4.11 (t, J = 7.8 Hz, 2 H), 3.87 (t, J = 7.7 Hz, 2 H), 3.40-3.10 (m, 3 H), 2.95 (m, 2 H), 2.40-2.10 (m, 2 H), 1.82 (m, 2 H), 1.55 (m, 2 H), 0.98 (t, J = 7.7 Hz, 3 H), 0.68 (t, J = 7.7 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 367 (M⁺ + 1), 395 (M⁺ + 29), 407 (M⁺ + 41). Anal. (C₂₁H₂₆N₄O₂) C, H, N.

3,7-Dihydro-8-[1-(hydroxymethyl)-2-phenylethyl]-1,3dipropyl-1*H***-purine-2,6-dione (5p). Compound 5p (820 mg, 36%) was prepared from 1 and acid 10 and isolated as a white solid: mp 145-146 °C; ¹H NMR (CDCl₃) \delta 12.38 (s, 1 H), 7.25-7.10 (m, 5 H), 4.11 (t, J = 7.6 Hz, 2 H), 4.05-3.85 (m, 4 H), 3.58 (m, 1 H), 3.38 (m, 1 H), 3.22 (dd, J = 13.8 Hz, J = 7.4 Hz, 1 H), 3.10 (dd, J = 13.6 Hz, J = 8.8 Hz, 1 H), 1.82 (m, 2 H), 1.70 (m, 2 H), 0.99 (t, J = 7.6 Hz, 3 H), 0.93 (t, J = 7.8 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 371 (M⁺ + 1), 399 (M⁺ + 29), 411 (M⁺ + 41). Anal. (C₂₀H₂₈N₄O₃) C, H, N.**

trans-3,7-Dihydro-8-(2-phenylcyclopentyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5q). Compound 5q (63 mg, 45%) was prepared from 1 and trans-2-phenylcyclopentanecarboxylic acid (2q)^{25,26} and isolated as a white solid: mp 152-153 °C; ¹H NMR (CDCl₃) δ 11.95 (s, 1 H), 7.18-7.08 (m, 5 H), 4.09 (t, J = 7.4 Hz, 2 H), 3.98 (t, 7.6 Hz, 2 H), 3.54 (dd, J = 18.6 Hz, J = 9.3 Hz, 1 H), 3.33 (dd, J = 17.7 Hz, J = 10.2 Hz, 1 H), 2.30 (m, 2 H), 2.20-1.60 (m, 7 H), 0.98 (t, J = 7.6 Hz, 6 H); MS (70 eV, CI, CH₄) m/z 381 (M⁺ + 1), 409 (M⁺ + 29), 421 (M⁺ + 41). Anal. (C₂₂H₂₈N₄O₂) C, H, N.

The following chiral compounds were obtained using a method similar to the preparation of 5a.

(S)-3,7-Dihydro-8-(1-methyl-2-phenylethyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5c). Compound 5c (87 mg, 82%) was prepared from 1 and (S)-2-(phenylmethyl)propanoic acid (2c) and isolated as a white solid: $[\alpha]^{20}_D + 38.5^\circ$ (c 0.69, CHCl₃); mp 141-142 °C; ¹H NMR (CDCl₃) δ 12.38 (s, 1 H), 7.22-7.09 (m, 5 H), 4.13 (t, J = 7 Hz, 2 H), 4.03 (t, J = 8 Hz, 2 H), 3.38 (m, 1 H), 3.23 (dd, J = 13 Hz, J = 8 Hz, 1 H), 2.98 (dd, J = 13 Hz, J = 7Hz, 1 H), 1.85-1.65 (m, 4 H), 1.41 (d, J = 7 Hz, 3 H), 0.98 (m, 6 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 (M⁺ + 29), 395 (M⁺ + 41). Anal. (C₂₀H₂₆N₄O₂) C, H, N.

(*R*)-3,7-Dihydro-8-(1-phenylpropyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5d). Compound 5d (190 mg, 9%) was prepared from 1 and (*R*)-2-phenylbutanoic acid (2d) and isolated as a white solid: $[\alpha]^{20}_{D}$ +4.4° (*c* 1.00, CHCl₃); mp 128–130 °C; ¹H NMR (CDCl₃) δ 12.41 (s, 1 H), 7.42 (d, *J* = 8.6 Hz, 2 H), 7.38–7.20 (m, 3 H), 4.20–4.00 (m, 5 H), 2.38 (m, 1 H), 2.19 (m, 1 H), 1.90–1.70 (m, 4 H), 0.98 (m, 9 H); MS (70 eV, CI, CH₄) *m/z* 355 (M⁺ + 1), 383 (M⁺ + 29), 395 (M⁺ + 41). Anal. (C₂₀H₂₈N₄O₂) C, H, N.

(S)-3,7-Dihydro-8-(1-phenylpropyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5f). Compound 5f (547 mg, 28%) was prepared from 1 and (S)-2-phenylbutanoic acid (2f) and isolated as a white solid: $[\alpha]^{20}$ _D -4.0° (c 1.07, CHCl₃); mp 128-131 °C; ¹H NMR (CDCl₃) δ 12.52 (s, 1 H), 7.42 (d, J = 8.6 Hz, 2 H), 7.38-7.20 (m, 3 H), 4.20-4.00 (m, 5 H), 2.38 (m, 1 H), 2.19 (m, 1 H), 1.90-1.70 (m, 4 H), 0.98 (m, 9 H); MS (70 eV, CI, CH₄) m/z 355 (M⁺ + 1), 383 (M⁺ + 29), 395 (M⁺ + 41). Anal. (C₂₀H₂₈N₄O₂) C, H, N.

(*R*)-3,7-Dihydro-8-(1-phenylethyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5g). Compound 5g (374 mg, 65%) was prepared from 1 and (*R*)-2-phenylpropanoic acid (2g) and isolated as a white solid: $[\alpha]^{20}_{\rm D}$ -8.5° (c 100, CHCl₃); mp 136-137 °C; ¹H NMR (CDCl₃) δ 11.18 (s, 1 H), 7.40-7.20 (m, 5 H), 4.35 (q, *J* = 7.1 Hz, 1 H), 4.11 (t, *J* = 7.4 Hz, 2 H), 4.00 (t, *J* = 7.9 Hz, 2 H), 1.90-1.60 (m, 4 H), 1.78 (d, *J* = 7.3 Hz, 3 H), 0.98 (m, 6 H); MS (70 eV, CI, CH₄) m/z 341 (M⁺ + 1), 369 (M⁺ + 29), 381 (M⁺ + 41). Anal. (C₁₉H₂₄N₄O₂) C, H, N.

(S)-3,7-Dihydro-8-(1-phenylethyl)-1,3-dipropyl-1*H*-purine-2,6-dione (5i). Compound 5i (252 mg, 41%) was prepared from 1 and (S)-2-phenylpropanoic acid (2i) and isolated as a white solid: $[\alpha]^{20}_{D} + 8.5^{\circ}$ (c 1.04, CHCl₃); mp 134.5-136°C; ¹H NMR (CDCl₃) δ 12.05 (s, 1 H), 7.40-7.20 (m, 5 H), 4.37 (q, J = 7.1 Hz, 1 H), 4.10 (t, J = 7.0 Hz, 2 H), 4.01 (t, J = 7.8 Hz, 2 H), 1.90-1.60 (m, 4 H), 1.80 (d, J = 7.5 Hz, 3 H), 0.99 (t, J = 7.4 Hz, 3 H), 0.95 (t, J = 7.4 Hz, 3 H); MS (70 eV, CI, CH₄) m/z 341 (M⁺ + 1), 369 (M⁺ + 29), 381 (M⁺ + 41). Anal. (C₁₉H₂₄N₄O₂) C, H, N.

3-Hydroxypropanoic Acid Methyl Ester (7). β -Propiolactone (6) (5.5 g, 76 mmol) was dissolved in 100 mL of methanol. Triethylamine (10.8 mL, 76 mmol) was added with stirring at room temperature.³¹ After 3 days, GC (40 °C/1.5 min \rightarrow 40 °C/

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min $\rightarrow 60$ °C/3 min, $t_{\rm R} = 1.92$ min) indicated completion of reaction. The solvent was removed under vacuum, and the residue was purified by flash chromatography (10% to 20% 2-propanol/hexane) to yield 7 (3.30 g, 42%) as an oil: ¹H NMR (300 MHz, CDCl₃) § 3.88 (dt, 2 H), 3.72 (s, 3 H), 2.60 (broad triplet, J = 6.1 Hz, 3 H).

2-(Hydroxymethyl)-3-phenylpropanoic Acid Methyl Ester (8).³¹ Compound 7 (3.23 g, 31 mmol) was dissolved in 100 mL of THF and cooled to -50 °C. Lithium diisopropylamide [prepared from 2.5 M n-BuLi (26.1 mL, 65 mmol) and diisopropylamine (9.1 mL, 65 minol) in 100 mL of THF] was added slowly to produce the dianion. After 20 min at -50 °C, the reaction mixture was treated with benzyl bromide (3.68 mL, 31 mmol). The mixture was then warmed to -20 °C over 1 h and quenched with 500 mL of saturated NH₄Cl. The aqueous mixture was extracted with diethyl ether $(2 \times 500 \text{ mL})$, and the combined organic extracts were dried over MgSO4, filtered, and concentrated under vacuum. The residue was purified by flash chromatography (10% to 20% 2-propanol/hexane) to yield 8 (2.65 g, 44%) as an oil (GC conditions, 150 °C isotherm, $t_{\rm R} = 2.08$ min): ¹H NMR (CDCl₃) § 7.35-7.19 (m, 5 H), 3.80-3.65 (m, 2 H), 3.70 (s, 3 H), 3.03 (dd, J = 16.9 Hz, J = 9.2 Hz, 1 H), 2.88 (m, 2 H), 2.18 (t, 2 H), 2.18 (t,J = 7.7 Hz, 1 H); MS (70 eV, CI, CH₄) m/z 195 (M⁺ + 1), 223 (M⁺ + 29); exact mass calcd for $C_{11}H_{15}O_3$ 195.1021, found 195.1019.

2-[(tert-Butyldimethylsilyl)methyl]-3-phenylpropanoic Acid Methyl Ester (9). Compound 8 (2.6 g, 13.4 mmol) was dissolved in 75 mL of DMF and treated with tert-butyldimethylsilyl chloride (2.2 g, 14.7 mmol) and imidazole (2.0 g, 29.4 mmol) with stirring. After 1.5 h, the reaction mixture was diluted with 500 mL of diethyl ether. The mixture was rinsed with 50%aqueous NaCl (3 × 200 mL), saturated NaCl (300 mL), dried over MgSO₄, filtered, and concentrated under vacuum. The residue was purified by flash chromatography (5% to 10% 2-propanol/hexane) to yield 9 (3.49 g, 85%) (GC conditions, 200 °C isotherm, $t_{\rm R} = 1.61$ min): ¹H NMR (CDCl₃) δ 7.31-7.15 (m, 5 H), 3.75 (m, 2 H), 3.62 (s, 3 H), 2.89 (m, 3 H), 0.89 (s, 9 H), 0.04 (s, 6 H); MS (70 eV, CI, CH₄) m/z 309 (M⁺ + 1), 337 (M⁺ + 29), 349 (M⁺ + 41); exact mass calcd for $C_{17}H_{29}O_3Si$ 309.1885, found 309.1882.

2-[(tert-Butyldimethylsilyl)methyl]-3-phenylpropanoic Acid (10). Compound 9 (3.3 g, 10.7 mmol) was dissolved in 100 mL of methanol, cooled to 0 °C and treated with 50 mL of 30% KOH with vigorous stirring. The solution was allowed to warm to room temperature over 5 h, diluted with 200 mL of H_2O , and rinsed with 200 mL of diethyl ether. The aqueous phase was cooled to 0-5 °C, and CH₂Cl₂ (100 mL) was added. HCl (1 N; 260 mL) was added slowly with stirring. The layers were separated, and the aqueous phase was extracted with CH₂Cl₂ (3 \times 200 mL). The combined organic extracts were dried over MgSO₄, filtered, and concentrated under vacuum. The residue was purified by radial chromatography (2% to 4% methanol/ chloroform, 4-mm plate) to yield 10 (2.14 g, 68%): ¹H NMR (CDCl₃) § 7.35-7.18 (m, 5 H), 3.75 (m, 2 H), 3.00 (m, 1 H), 2.85 (m, 2 H), 0.90 (m, 9 H), 0.05 (s, 6 H); MS (70 eV, CI, CH₄) m/z295 (M⁺ + 1), 277 (M⁺ + 1 - H_2O), 237 (M⁺ + 1 - C_4H_{10}), 323 $(M^+ + 29)$, 335 $(M^+ + 41)$; exact mass calcd for $C_{16}H_{26}O_3Si$ 295.1729, found 295.1734.

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