Communications to the Editor

Structure-Based Design of HIV-I Protease Inhibitors: Replacement of Two Amides and a 10π -Aromatic System by a **Fused Bis-tetrahydrofuran**

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The replacement of the peptide bond with a functionality that mimics the biological mode of action continues to be a major strategy for drug design. As part of our continuing efforts in search of therapeutic agents for AIDS, we recently described a stereochemically defined tetrahydrofuran ring as a surrogate for the asparagine side chain in the design of HIV protease inhibitors.¹ We were intrigued by the possibility of further exploiting this concept with the ultimate goal of producing a polyether template to mimic the peptide region which binds to the viral enzyme.²

After visual inspection of the X-ray crystal structure of the HIV-I protease-inhibitor (Ro 31-8959) complex,³ it was tempting to speculate that a fused bicyclic tetrahydrofuran could effectively hydrogen bond to the NH of the Asp 29 and 30 residues and thereby replace the quinaldic amide-asparagine amide fragment of the Ro 31-8959 inhibitor. In the hydroxyethylamine isostere derived inhibitors of which Ro 31-8959 is prototypical, inclusion of a P_3 ligand is essential for low nanomolar activity against the HIV protease. Since there was considerable rotational freedom about the four bonds connecting the two carbonyls involved, a rigid cyclically constrained system might provide additional gains in binding energy, to offset loss of the P_3 hydrophobic binding of the quinoline ring. In this paper, we report the structure-based design of a fused bis-tetrahydrofuran that effectively replaces two amide bonds and a 10π aromatic system of the present clinical candidate 12 (Ro 31-8959).⁴

The synthetic route leading to the bis-tetrahydrofuran (bis-Thf) 4 is outlined in Scheme 1. As shown, optically pure $(3R)$ -diethyl malate 1 was alkylated utilizing the procedure reported by Seebach.⁵ The diastereomer 2 was obtained as the major (selectivity 12:1) product in 85% yield after distillation. The diastereomeric mixture was converted to the isopropylidene derivative 3 by LAH reduction in diethyl ether followed by treatment with a catalytic amount of $p\text{-}\mathrm{TsOH}$ in acetone at 23 °C for 12 h (59% isolated yield). Swern oxidation and subsequent reaction with camphorsulfonic acid (CSA) in methanol afforded the methyl acetal 4 as a mixture (ratio 4:1) in 73% yield. Methyl acetal 4 was converted to bis-Thf **5**

Scheme 1°

 a Key: (a) LDA, $CH_2=CHCH_2Br$; (b) LAH, Et₂O; (c) acetone, p -TsOH; (d) Swern oxidation; (e) CSA, MeOH; (f) ozonolysis then NaBH₄; (g) CSA, CH₂Cl₂; (h) DPC, Et₃N, CH₂Cl₂.

Scheme 2°

^a Key: (a) mixed carbonate 6 , CH_2Cl_2 ; (b) mixed carbonate 8 , CH_2Cl_2 ; (c) ref 1.

by the following reaction sequence: (1) ozonolytic cleavage of the terminal olefin, (2) NaBH₄ reduction of the resulting aldehyde in ethanol at 0° C, and (3) exposure of the corresponding alcohol with CSA in methylene chloride at 23 °C for 12 h. The desired bis-Thf ligand 5 $(\alpha^{23}$ _D -4.3°, c 0.215, CHCl₃) was obtained in 81% yield $(from 4)$ after silica gel chromatography.⁶ Similarly, bis-Thf ligand 7 (α^{24} _D +3.9°, c 0.32, CHCl₃) with a 3S,3aR,-6aS-configuration was synthesized, starting from optically pure $(3S)$ -diethyl malate following the sequence of reactions described above. The ligands 5 and 7 readily reacted with dipyridyl carbonate and triethylamine in methylene chloride to furnish the correspond-

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Figure 1. X-ray structure of the inhibitors 10 (magenta) and 12 (green) bound to the HIV-1 protease.

ing active carbonates 6 and 8 in good yields (75-80%).⁷ Reaction of the active carbonate 6 with the hydroxyethylamine^{1b} isostere 9 in methylene chloride at 23 °C provided only the inhibitor 10 (white solid, mp $98-101$ $^{\circ}$ C) by ¹H NMR and HPLC analyses (96% yield). Similarly, treatment of active carbonate 8 with amine 9 afforded the inhibitor 11 (white solid, mp $85-89$ °C) in 94% isolated yield.

The binding properties of the inhibitors 10 and 11 were determined by enzyme inhibition assays.⁸ As is evident, the ring stereochemistry and conformational rigidity associated with bis-Thf ligands have a significant effect on their in vitro potencies. The inhibitor 10 derived from $(3R,3aS,6aR-bis$ -Thf has shown an enzyme inhibitory activity (IC_{50}) of 1.8 ± 0.2 nM $(n = 6)$. In comparison, inhibitor 12 (Ro 31-8959), containing asparagine in the P_2 -position and quinaldic amide in the P₃-position, exhibited an IC₅₀ value of 0.23 ± 0.10 nM $(n = 3)$. The inhibitor with $(3S,3aR,6aS)$ -bis-Thf as the P₂-ligand 11, IC₅₀ = 6.4 nM ($n = 2$) was less potent than 10. More strikingly, the enhanced inhibitory potency of 10 relative to the $(3S$ -tetrahydrofuranylurethane $(IC_{50}$ 132 nM; CIC_{95} >800 nM) or the BOC derivative (IC₅₀ $>3 \mu M$ ⁹ was also reflected in its antiviral potency. Inhibitor 10 has prevented the spread of HIV-I in MT4 human T-lymphoid cells infected with IIIb isolate¹⁰ at an average concentration $(n = 4)$ of 46 ± 4 nM (CIC₉₅). In head to head comparison, inhibitor 10 was equipotent to present clinical candidate 12 (Ro 31-8959), CIC_{95} = 23 ± 7 nM.¹¹ In contrast, inhibitor 11 has shown an antiviral potency of 200 nM.

In an effort to gain insight into the ligand binding site interactions, a single crystal of the inhibitor 10 complexed with HIV-I protease was generated, and the three-dimensional structure was determined by X-ray diffraction to 2.10-Å resolution.¹² A stereoview of the bound conformation of inhibitors 10 (magenta) and 12

(green) as determined by X-ray crystallographic analysis of the enzyme—inhibitor complex is shown in Figure I.¹³ The (R) -hydroxyl group of inhibitor 10 is positioned symmetrically between the two aspartates of the enzyme. Both the asparagine of 12 and the bis-Thf of 10 are located in the S_2 subsite.¹⁴ As shown, bis-Thf oxygen-1 of 10 and the P_2 asparagine carbonyl of 12 are within hydrogen bonding distance (3.5 and 3.2 A, respectively) to the Asp 30 NH present in the S_2 binding domain of the HIV-I protease. Also, the bis-Thf oxygen-6 and the P_3 quinoline amide carbonyl of 12 interact with the Asp 29 NH (bonding distance 3.0 and 3.3 A, respectively) positioned in the region. Like other reported protein–ligand complex structures, the P_2 bis-Thf urethane carbonyl and the *tert*-butyl amide carbonyl of 10 hydrogen bond to the carbon bond to the carbon bond that the carbon bond that the carbon bond that the carbon bond that of **IU** hydrogen bond to the critical water inc

Thus, incorporation of a conformationally constrained bis-Thf as the P_2 ligand provided an inhibitor 10, with comparable *in vitro* antiviral activities to inhibitors in the hydroxyethylamine class with both P_2 and P_3 ligands. Design of such a high-affinity ligand led to improved aqueous solubility,¹⁵ decreased log *P* value,¹⁶ and reduction in molecular weight due to exclusion of the P_3 ligand. The molecular weight of the bis-Thf is essentially half the combined molecular weight of the P_2 asparagine and P_3 quinoline ligands of inhibitor 12 (Ro 31-8959). The present studies offer many important aspects of the molecular design that could facilitate the design of other novel protease inhibitors with improved $\frac{1}{2}$ and $\frac{1}{2}$ actions.

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Supplementary Material Available: Experimental procedures and spectral data for compounds $2-12$ and log P determination protocol for **10** and **12** (9 pages). Ordering information is given on any current masthead page.

References

- (1) (a) Thompson, W. J.; Ghosh, A. K.; Holloway, M. K.; Lee, H. Y.; Munson, P. M.; Schwering, J. E.; Wai, J. M.; Darke, P. L.; Zugay, J. A.; Emini, E. A.; Schleif, W. A.; Huff, J. R.; Anderson, P. S. 3'-Tetrahydrofuranylglycine as a Novel, Unnatural Amino Acid Surrogate for Asparagine in the Design of Inhibitors of the HIV Protease. J. Am. Chem. Soc. 1993, 115, 801–03. (b) Ghosh, A.
K.; Thompson, W. J.; Holloway, M. K.; McKee, S. P.; Duong, T.
T.; Lee, H. Y.; Munson, P. M.; Smith, A. M.; Wai, J. M.; Darke,
P. L.; Zugay, J. A.; Emini, E. A.; ment of Tetrahydrofuranylglycine as Novel P₂-Ligands and Pyrazine Amides as P3-Ligands. *J. Med. Chem.* **1993,***36,*2300- 10.
- (2) The polyether molecule with antiprotease activity has been reported recently. See: Corey, E. J.; Rao, K. S. Enantioselective Total Synthesis of Ginkgolide Derivatives Lacking the iert-Butyl Group, an Essential Subunit for Antagonism of Platelet Activating Factor. *Tetrahedron Lett.* **1993,** *32,* 4623-26.
- (3) A complex between **12** (Ro 31-8959, L-697, 803) and HIV-I protease was crystallized in the space group $P6_1$, $a = b = 63.33$ and c = 83.31 A. A model for the complex was partially refined against data extending from 8.0 to 2.2 A in resolution; the *R* value for the current model is 0.196, and deviations from ideal bond distances are 0.018 A. Full details of the crystallographic analysis will be published elsewhere.
- (4) Roberts, N. A.; Martin, J. A.; Kinchington, D.; Broadhurst, A. V.; Craig, J. C; Duncan, I. B.; Galpin, S. A.; Handa, B. K.; Kay, J.; Krohn, A.; Lambert, R. W.; Merrett, J. H.; Mills, J. S.; Parkes, K. E. B.; Redshaw, S.; Ritchie, A. J.; Taylor, D. L.; Thomas, G. J.; Machin, P. J. Rational Design of Peptide-Based HIV Pro-teinase Inhibitors. *Science* **1990,** *248,* 358-61.
- (5) Seebach, D.; Aebi, J.; Wasmuth, D. Diastereoselective a-Alkylation of /3-Hydroxycarboxylic Esters Through Alkoxide Enolates: (+)-Diethyl (2S,3R)-3-Allyl-2-Hydroxysuccinate from (-)-Diethyl S-Malate. *Org. Synth.* **1983,** *65,* 109-120.
- (6) Proton NMR and infrared spectra are consistent with assigned structures. Satisfactory $(\pm 0.4\%)$ elemental analysis were obtained for all new compounds, and all melting points were determined on a Thomas-Hoover capillary melting apparatus and are uncorrected.
- (7) (a) Ghosh, A. K.; Duong, T. T.; McKee, S. P. Di(2-Pyridyl) Carbonate Promoted Alkoxycarbonylation of Amines: A Convenient Synthesis of Functionalized Carbamates. *Tetrahedron Lett.* **1991,** *32,* 4251-54. (b) Ghosh, A. K; Duong, T. T.; McKee, S. P.; Thompson, W. J. N,N'-Disuccinimidyl Carbonate: A Useful

Reagent for Akoxycarbonylation of Amines. *Tetrahedron Lett.* **1992,** *33,* 2781-84. (a) Ghosh, A. K; Duong, T. T.; McKee, S. P. *Tetrahedron Lett.* **1991,** *32,* 4251. (b) Ghosh, A. K.; Duong, T. T; McKee, S. P. *Tetrahedron Lett.* **1992,** *33,* 2781.

- (8) Heimbach, J. C; Garsky, V M.; Michelson, S. R.; Dixon, R. A.; Sigal, I. S.; Darke, P. L. Affinity Purification of the HIV-I Protease. *Biochem. Biophys. Res. Commun.* **1989,** *164,* 955- 60.
- (9) Ghosh, A. K; Thompson, W. J.; McKee, S. P.; Duong, T. T.; LyIe, T. A.; Chen, J. C; Darke, P. L.; Zugay, J. A.; Emini, E. A.; Schleif, W. A.: Huff, J. R.; Anderson, P. S. 3-Tetrahydrofuran and Pyran Urethanes as High Affinity P2-Ligands for HIV-I Protease Inhibitors. *J. Med. Chem.* **1993,** *36,* 292-94.
- (10) For assay protocol, see: Thompson, W. J.; Fitzgerald, P. M. D.; Holloway, M. K.; Emini, E. A.; Darke, P. L.; McKeever, B. M.; Schleif, W. A.; Quintero, J. C.; Zugay, J. A.; T. J.; Scher, T. J.; Schwering, J. E.; Homni Huff, J. R. Synthesis and Antiviral Activity of a Series of HIV-I Protease Inhibitors with Functionality Tethered to the P_1 or P_1' Phenyl Substituents: X-ray Crystal Structure Assisted Design. *J. Med. Chem.* **1992,** *35,* 1685-01 and references cited therein.
- (11) Craig, J. C; Duncan, I. B.; Hockley, D.; Grief, C; Roberts, N. A.; Mills, J. S. *Antiviral Res.* **1991,***16,* 295-05 and references cited therein. These authors report IC_{90} values of 6-30 nM in cell culture. However, the assay protocol differs widely in that syneytia formation rather tha p24 production was monitored as endpoint, and cell types other than MT4 were employed.
- (12) A complex between **10** (L-739, 594) and HIV-I protease was crystallized in the space group $P6_1$, $a = b = 63.40$ and $c = 83.48$ A. A model for the complex was refined to completion against data extending from 8.0 to 2.1 A in resolution; the *R* value for the final model is 0.166 and deviations from ideal bond distances are 0.018 A. Full details of the crystallographic analysis will be published elsewhere.
- (13) Figure 1 was generated by superposition of the X-ray crystal structure of **12** on the protein-inhibitor complex of **10,** in the same frame of references.
- (14) For X-ray analysis of the compound **12** (Ro 31-8959) and HIV protease complex, see: (a) Martin, J. A. Recent Advances in the Design of HIV Protease Inhibitors. *Antiviral Res.* **1992,***1*7, 265- 278. (b) Krohn, A.; Redshaw, S.; Ritchie, J.; Graves, B. J.; Hatada, M. H. Novel Binding Mode of Highly Potent HIV-Proteinase Inhibitors Incorporating the (R)-Hydroxyethylene Isostere. *J. Med. Chem.* **1991,** *34,* 3340-42.
- (15) Aqueous solubility of inhibitor **10** was determined to be 0.235 mg/mL in phosphate buffer $(pH = 7.4)$. Under these same conditions, compound **12** was less than 0.01 mg/mL. The (3S) tetrahydrofuranylurethane (ref 8) was also found to be less soluble (0.15 mg/mL) in this buffer.
- (16) The log *P* value of inhibitors 10 and **12** were determined to be 3.5 and 5.7 respectively. The protocol for log *P* determination is provided in the supplementary material.