Substituted (S)-Phenylpiperidines and Rigid Congeners as Preferential **Dopamine Autoreceptor Antagonists: Synthesis and Structure-Activity** Relationships

Clas Sonesson,^{*,†} Chiu-Hong Lin,[‡] Lars Hansson,[†] Nicholas Waters,[§] Kjell Svensson,[§] Arvid Carlsson,[§] Martin W. Smith,[‡] and Håkan Wikström^{||}

Medicinal Chemistry Unit, Department of Pharmacology, University of Göteborg, Medicinaregatan 7, S-413 90 Göteborg, Sweden, Upjohn Laboratories, The Upjohn Company, Kalamazoo, Michigan 49001, Department of Pharmacology, University of Göteborg, Medicinaregatan 7, S-413 90 Göteborg, Sweden, and Department of Medicinal Chemistry, University Centre for Pharmacy, University of Groningen, Antonius Deusinglaan 2, NL-9713 AW, Groningen, The Netherlands

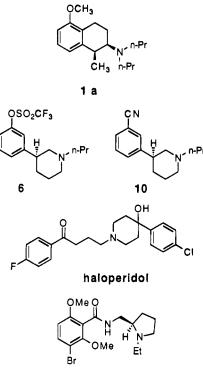
Received April 14, 1994[®]

A series of (S)-phenylpiperidines in which the substituents on the aromatic ring and nitrogen have been varied has been prepared. They have been evaluated pharmacologically to explore the importance of these substituents for the interaction with central dopamine (DA) receptors. On the basis of biochemical and behavioral data in rats, several of these compounds are characterized as centrally acting DA autoreceptor antagonists. (S)-Phenylpiperidines having an aromatic substituent with a high group dipole moment in the 3-position, i.e., meta with respect to the piperidine ring, and being N-substituted with a propyl group were found to be highly active in vivo on the synthesis and turnover of dopamine. However, they do not induce strong hypoactivity or catalepsy. Interestingly, the most active compounds in vivo were found to display only low affinity for DA D_2 and D_3 receptors in vitro. In addition, 7-triflate-substituted octahydrobenzo[f]quinolines and 6-triflate-substituted hexahydro-1H-benz[e]indoles have been prepared and pharmacologically evaluated. The trans isomers of these rigid structures were found to display a pharmacological profile similar to that of the flexible phenylpiperidines. The corresponding *cis* isomers were found to be inactive *in vivo*.

Introduction

In the search for compounds with pharmacological profiles similar to compound 1a (cis-(+)-5-methoxy-1methyl-2-(di-n-propylamino)tetralin, cis-(+)-UH232),^{1,2} we have found that compounds 6 and 10 with the S-configuration are structures with interesting pharmacodynamic and pharmacokinetic properties.³ Like classical neuroleptics (e.g., haloperidol)⁴ and "atypical antipsychotics" (e.g., remoxipride),⁵ they increase the firing of dopamine (DA) neurons as well as the synthesis and turnover of dopamine but are not cataleptogenic in rodents.^{6,7} Rather, compounds 6 and 10 act as mild behavioral stimulants. According to the prevailing hypothesis, the behavioral activation is thought to be caused by a preferential antagonism of nerve terminal DA autoreceptors leading to an increased synthesis and release of DA and subsequent increase in psychomotor activity. Still, 6 and 10 are able to antagonize the biochemical and behavioral effects of direct and indirect DA agonists such as apomorphine and d-amphetamine.⁸ However, this new class of phenylpiperidines have relatively low affinity for D₂ receptors in *in vitro* binding assays.³ It has been speculated that compounds of this structural class may be useful in the treatment of both positive and negative symptoms in schizophrenia^{9,10} and possibly also for other indications such as depression and the pharmacological treatment of drug addiction.^{11,12}

Compounds 6 and 10 emanate from the partial DA D₂ agonist 1b ((-)-3-(3-hydroxyphenyl)-1-propylpiperi-



remoxipride

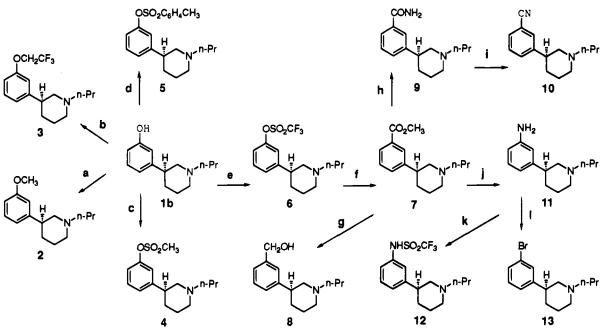
dine, (-)-3-PPP, Preclamol)¹³⁻¹⁵ and have some physicochemical properties in common. For compounds 6 and 10, it has been suggested that a lack of hydrogen bonddonor properties on the aromatic ring (e.g., OH or an isosteric hydrogen-donating functionality such as an indole N-H¹⁶) is important to diminish the intrinsic activity at the autoreceptors.³ Accordingly, removing the hydrogen bond-donating ability of 1b by O-methy-

[†] Medicinal Chemistry Unit, Department of Pharmacology, University of Göteborg. [‡] The Upjohn Co.

Department of Pharmacology, University of Göteborg. "University of Groningen.

[®] Abstract published in Advance ACS Abstracts, July 1, 1994.



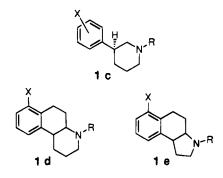


^a Reagents and conditions: (a) NEt₃, NaOH, (CH₃O)₂SO₂, THF; (b) NaH, *p*-TosOCH₂CF₃, DMF, Δ ; (c) NEt₃, CH₃SO₂Cl, CH₂Cl₂; (d) NEt₃, *p*-TosCl, CH₂Cl₂; (e) NEt₃, (CF₃SO₂)₂O, CH₂Cl₂; (f) Pd(OAc)₂/dppp, CO(g), NEt₃, MeOH, Δ ; (g) LiAlH₄/diethyl ether; (h) MeONa/MeOH, HCONH₂, DMF, Δ ; (i) POCl₃/DMF, Δ ; (j) NaN₃, concentrated H₂SO₄, CH₂Cl₂, Δ ; (k) NEt₃, (CF₃SO₂)₂O, CH₂Cl₂; (l) NaNO₂, CuBr, 48% HBr, Δ .

lation, yielding 2, results in a compound with only weak agonist activity (reserpinized rats, Table 2). In addition, 2 displays only weak, if any, antagonist-like activity in nonpretreated habituated rats. This is in sharp contrast to the results obtained with compounds 6 and 10, which are powerful antagonists at DA autoreceptors. The above results suggests that, apart from a lack of hydrogen-bonding properties, there are other structural requirements that are crucial for antagonist activity within this series of phenylpiperidines. Consequently, the importance of proper selection of the aromatic substituent, in order to yield antagonists in a series of aminotetralins, has recently been investigated.¹⁷ Substitution of the methoxy group in 1a for a triflate or a cyano group reduces the antagonist-like properties at the DA autoreceptors, shown by lower levels of released DA and metabolites than for 1a. Thus, in contrast to the phenylpiperidines, strongly electron-donating groups (e.g., -OMe) seem to be important for the antagonistic properties at presynaptic autoreceptors of 5-substituted-1-methyl-2-aminotetralins 1a.

To gain further insight about the physicochemical properties (lipophilic, electronic, and/or sterical) that are of importance for the DA receptor antagonistic properties of compounds 6 and 10, we now report on the synthesis and biological activity of a series of compounds with the generic structure 1c. All compounds have been evaluated in vitro in D₁, D₂, D₃, D₄, and 5-HT_{1A} binding assays in addition to assaying their biochemical and behavioral activity in vivo. The maximal change in tissue dihydroxyphenylalanine (DOPA accumulation) or dihydroxyphenylacetic acid (DOPAC levels) and dopamine release has been used to characterize their profile at the DA autoreceptors. Furthermore, to investigate if the aromatic substituent (e.g., OSO₂CF₃ and CN) has a different impact on the pharmacological profile in rigid as compared to flexible structures (1c), we have synthesized the conformationally restricted analogs 1d,e.

The *cis* and *trans* isomers of both generic structures have been evaluated. This evaluation may provide some information about the "antagonist conformation" of the flexible phenylpiperidine system.



Chemistry

Compounds of the general formula 1c were obtained via transformations of (-)-3-(3-hydroxyphenyl)-1-propylpiperidine (1b) or its substituted derivatives. The preparations of analogs bearing different substituents on the aromatic ring in position 3 are depicted in Schemes 1-3. Yields and analytical data are given in Table 1.

The methoxy derivative **2** was prepared from **1b** via triethylamine-catalyzed O-methylation with dimethyl sulfate in 88% yield.¹⁸ The O-trifluoroethylated analog **3** was prepared in low yield by treating the sodium phenolate of **1b** with 2,2,2-trifluoroethyl p-toluenesulfonate in DMF. The sulfonate analogs **4**-**6** were prepared according to standard procedures.¹⁹ The carboxy methyl ester **7** was prepared in high yields by palladium-catalyzed carbonylation of **6**.²⁰ A major side reaction in this step is a reductive elimination of the triflate group (10-15%) to yield **20**. This is a wellknown problem in the palladium-catalyzed reactions. It Table 1. Analytical and in Vitro Binding Data for D₂, D₃, and 5-HT_{1A} Receptors



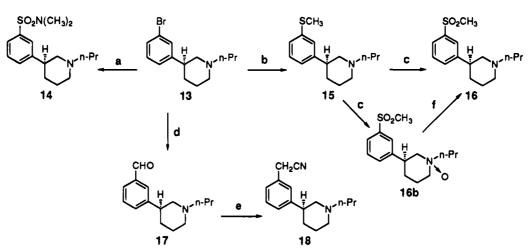
					$K_i ({ m nM})^a$			
no.	X	R	% yield	$\mathbf{formula}^b$	D ₂ c~e [³ H]spiperone	D ₂ ^f * [³ H]U-86170	D3 ^{f#} [³ H]spiperone	5-HT _{1A} c,h [³ H]-8-OH-DPAT
1 b	3-OH ⁱ	<i>n</i> -Pr			100^{h}	8.5 ± 1.1	132 ± 16	8000
2	3-OMe	<i>n</i> -Pr	88	C ₁₅ H ₂₃ NO•HCl	3440 ± 515	602 ± 138	1328 ± 57	3600
3	$3-OCH_2CF_3$	n-Pr	31	C ₁₆ H ₂₂ F ₃ NO·HCl	1670^{h}	820 ± 157	1414 ± 202	1200
4	$3-OSO_2CH_3$	<i>n</i> -Pr	88	$C_{15}H_{23}NO_3SC_4H_4O_4$	4170^{h}	192 ± 19	978 ± 34	1380 ± 44^d
5	$3-OSO_2C_6H_4CH_3$	n-Pr	78	$C_{21}H_{27}NO_3S\cdot HCl$	1320^{h}	162 ± 18	92 ± 10	1600
6	$3-OSO_2CF_3^i$	n-Pr	92	C ₁₅ H ₂₀ NF ₃ O ₃ S•HCl	408 ± 115	40 ± 2	68 ± 3	1530 ± 15^d
7	3-CO ₂ CH ₃	n-Pr	84	C ₁₆ H ₂₃ NO ₂ •HCl	2170^{h}	>1852	>1285	1600
8	3-CH₂OH	<i>n</i> -Pr	83	C ₁₅ H ₂₃ NO·HCl	410^{h}	240 ± 20	1461 ± 143	2410
9	3-CONH ₂	n-Pr	84	$C_{15}H_{22}N_2O \cdot 0.5H_2O$	>15 000	>1852	>1285	5040
10	3-CN ⁱ	<i>n</i> -Pr	92	$C_{15}H_{20}N_2$ ·HCl	1140 ± 282	119 ± 33	249 ± 39	>10 000
11	$3-NH_2$	<i>n</i> -Pr	76	$C_{14}H_{22}N_{2}^{j}$	2000^{h}	NT^{k}	NT	5360
12	3-NHSO ₂ CF ₃	n-Pr	48	$C_{15}H_{21}F_{3}N_{2}O_{2}S$	NT	>2000	>2000	>10 000
13	3-Br	n-Pr	57	C ₁₄ H ₂₀ NBr-HCl	533 ± 128	106 ± 7	171 ± 8	800
14	$3-SO_2N(CH_3)_2$	<i>n</i> -Pr	38	$C_{16}H_{26}N_2O_2SC_4H_4O_4$	3390 ^h	227 ± 15	170 ± 4	8160
1 5	3-SCH ₃	<i>n</i> -Pr	81	$C_{15}H_{23}NS C_4H_4O_4 0.25H_2O$	849 ± 133	49 ± 4	139 ± 12	887
16	3-SO ₂ CH ₃	<i>n</i> -Pr	52 + 41	C ₁₅ H ₂₃ NO ₂ S·HCl	6270 ± 580	447 ± 45	1305 ± 249	2590 ± 30^d
17	3-CHO	n-Pr	75	$C_{15}H_{21}NO \cdot HCl^{2}$	1120^{h}	20 ± 2	83 ± 5	8110
18	3-CH ₂ CN	<i>n</i> -Pr	50	$C_{16}H_{22}N_2 \cdot C_4H_4O_4$	1229 ± 11	69 ± 20	509 ± 30	3260
19	3-CCH	n-Pr	52	$C_{16}H_{21}N\cdot HCl$	709 ± 41	334 ± 18	339 ± 24	310
20	3-H	n-Pr	71	$C_{14}H_{21}N\cdot HCl$	1490 ± 86	222 ± 18	777 ± 39	2050
21	3-CH ₃	n-Pr	58	$C_{15}H_{23}N\cdot HCl$	930 ^h	430 ± 45	573 ± 37	1200
22	3-(3-thienyl)	n-Pr	70	$C_{18}H_{23}NS HCl 0.5H_2O$	760 ± 38	134 ± 22	78 ± 6	160
23	3-COCH ₃	n-Pr	74	$C_{16}H_{23}NO \cdot HCl$	6540 ± 3460	>1852	2150 ± 132	4170
25	3-CH ₂ CH ₂ CH ₃	<i>n</i> -Pr	88	$C_{17}H_{27}N$ ·HCl	392^{h}	57 ± 6	86 ± 10	135
26	3-CN	н	84	$C_{12}H_{14}N_2 \cdot C_4 H_4 O_4$	2670 ^h	314 ± 47	631 ± 45	8000
27	3-CN	Me	89	$C_{13}H_{16}N_2 HCl^m$	8330 ^h	1040 ± 159	1759 ± 92	>10 000
28	3-CN	Et	91	$C_{14}H_{18}N_2$ ·HCl	2330 ^h	235 ± 21	514 ± 21	>10 000
29	3-CN	<i>i</i> -Pr	93	$C_{15}H_{20}N_2$ ·HCl	717 ^h	106 ± 9	268 ± 37	>10 000
30	3-CN	allvl	53 64	$C_{15}H_{18}N_2$ ·HCl	1890 ± 263	$\frac{100\pm3}{254\pm38}$	208 ± 31 658 ± 48	9600
31	3-CN	propargyl	69	$C_{15}H_{16}N_2$ ·HCl	>15 000	1178 ± 257	>1362	>10 000
32	3-CN	n-Bu	76	$C_{16}H_{22}N_2 C_4H_4O_4 0.25H_2O$	617 ^h	NT	NT	4480
33	3-CN	s-Bu	70	$C_{16}H_{22}N_2 C_4H_4O_4 O_2OH_2O$ $C_{16}H_{22}N_2 C_4H_4O_4$	1050 ^h	187 ± 26	105 ± 46	8070
34	3-CN	cpm ⁿ	80	$C_{16}H_{20}N_2 C_4H_4O_4$	3100 ± 354	519 ± 77	522 ± 35	6320
35	3-CN	pheet	81	$C_{16}H_{20}N_2 C_4H_4O_4$ $C_{20}H_{22}N_2 C_4H_4O_4$	105^{h}	313 ± 77 22 ± 3	522 ± 35 59 ± 4	120
36	3-CN	pheprop ^p	72	$C_{20}H_{22}V_{2}C_{4}H_{4}O_{4}$ $C_{21}H_{24}N_{2}C_{4}H_{4}O_{4}$	80 ^h	$\frac{22 \pm 3}{29 \pm 4}$	36 ± 2	180
30 37	3-CN	thiopheneet ^q	60	$C_{21}R_{24}N_2C_4R_4O_4$ $C_{18}H_{20}N_2SC_4H_4O_4$	102 ± 27	$\begin{array}{c} 29 \pm 4 \\ 11 \pm 1 \end{array}$	$\begin{array}{c} 36 \pm 2 \\ 56 \pm 5 \end{array}$	320
37	3-CN 3-CN	$(CH_2)_3N(CH_3)_2$	38	$C_{18}H_{20}N_2SC_4H_4O_4$ $C_{17}H_{25}N_32HCl 0.5H_2O^r$	102 ± 27 2330 ^h		36 ± 5 NT	320 7120
38 39	3-OSO ₂ CF ₃		38 91	$C_{13}H_{16}F_{3}NO_{3}S^{s}$	11800^{h}	NT	NT	2240
39 40	3-OSO ₂ CF ₃ 3-OSO ₂ CF ₃	Et	91 88	$C_{13}H_{16}F_{3}NO_{3}S^{\circ}$ $C_{14}H_{18}F_{3}NO_{3}S\cdot HCl$	1180^{h}	NT	NT	2240 1040
40 41	$3-OSO_2CF_3$ $3-OSO_2CF_3$	pheet	88 84		40 ^h	11 ± 1	19 ± 3	NT
		n-Pr	84 85	$C_{20}H_{22}F_3NO_3SC_4H_4O_4$	40^{n} 92^{h}	11 ± 1 315 ± 21	19 ± 3 3113 ± 235	1840
42	$2-OSO_2CF_3^t$			$C_{15}H_{20}F_{3}NO_{3}S\cdot HCl$				
43	$4-OSO_2CF_3^t$	n-Pr	92 80	$C_{15}H_{20}F_{3}NO_{3}S\cdot HCl$	11800^{h}	1736 ± 239	1136 ± 62	>10 000
44	3,4-di-OSO ₂ CF ₃	n-Pr	80	$C_{16}H_{19}F_6NO_6S_2$ ·HCl	6250^{h}	1100 ± 99	338 ± 53	>10 000

^a K_i values for displacement of the dopamine D_2 receptor antagonist spiperone, the dopamine D_2 receptor agonist U-86170, the dopamine D_3 receptor antagonist spiperone, or the 5-HT_{1A} receptor agonist 8-OH-DPAT. ^b Analyses for C, H, and N were within $\pm 0.4\%$. ^c Data from rat striatal membrane. ^d Values were obtained with six drug concentrations $(10^{-4}-10^{-9} \text{ M})$ in which each value was determined in triplicate. ^e In addition, all compounds were also screened in D_2 cloned mammalian receptors expressed in CHO-K1 cells using [³H]spiperone as ligand. All compounds display $K_i > 217$ except for compounds 1b ($K_i = 39 \pm 7$), 6 (48 ± 8), 10 (265 ± 54), 13 (170 ± 23), 14 (91 ± 8), 17 (145 ± 20), 33 (98 ± 10), 36 (60 ± 8), 37 (134 ± 8), and 41 (73 ± 5). ^f Data from cloned mammalian receptors expressed in CHO-K1 cells. ^g Values were obtained with 11 drug concentrations in which each value was determined in duplicate. ^h Values were obtained with six drug concentrations (10⁻⁴-10⁻⁹ M) in which each value was determined from a single experiment. ⁱ Data taken from ref 3. ^j High-resolution mass spectrum calcd for C₁₄H₂₂N₂ (M⁺): 218.1782. Found: 218.1770. ^k NT means not tested. ⁱ Anal. Calcd: C, 67.3. Found: C, 66.6. ^m Anal. Calcd: C, 66.0. Found: C, 65.4. ⁿ cpm: cyclopropylmethyl. ^o pheet: 2-phenylethyl. ^p pheprop: 3-phenylpropyl. ^q thiopheneet: 3-(2-ethyl)thiophene. ^r Anal. Calcd: H, 6.9; found: H, 7.9. ^s High-resolution mass spectrum calcd for C₁₃H₁₆F₃NO₃S (M⁺): 323.0803. Found: 323.0804. ^r Racemate.

has been suggested that the hydride source may be triethylamine.^{21,22} An alternative approach to avoid this side reaction is the use of diisopropylethylamine as a base.²¹ However, when we used diisopropylethylamine, the amount of reduced triflate increased to 40-45%. Analysis of another side product from this reaction, by ¹H-NMR and GC-MS, revealed that the piperidine ring of the starting material probably acted as the hydride source.

The hydroxymethyl analog 8 was obtained from methyl ester 7 through reduction with lithum alumium hydride (LiAlH₄). The carboxamide **9** was obtained *via* an efficient one-step synthesis from the ester **7** using a formamide-sodium methoxide reagent in dimethylformamide (DMF).²³ The primary carboxamide **9** was then dehydrated with phosphorus oxychloride in dry DMF to yield the corresponding cyano analog **10**.²⁴ The preparation of this compound has been reported earlier³ in a one-step procedure starting with the triflate analog **6** *via* a palladium-mediated insertion of the cyanide group (52% yield). The overall yield of **10**, based on **6**, reported by us is 65-70%.





^a Reagents and conditions: (a) i, s-BuLi/THF, SO₂(g), ii, SOCl₂, 40% HNMe₂, CH₂Cl₂; (b) t-BuLi/diethyl ether, CH₃SSCH₃; (c) m-CPBA, CF₃COOH; (d) t-BuLi/diethyl ether, DMF; (e) t-BuOK/1,2-DME, TOSMIC; (f) TiCl₄/NaI, CH₃CN.

Further transformation of the ester 7, illustrated in Scheme 1, includes a Curtius rearrangement yielding the aniline 11, using excess of sodium azide in a concentrated sulfuric acid-dichloromethane mixture.²⁵ Treatment of compound 11 with trifluoromethanesulfonic anhydride afforded 12 in a modest yield (48%). Compound 13 was prepared from 11 via the Sandmeyer reaction using cuprous bromide (57%).²⁶

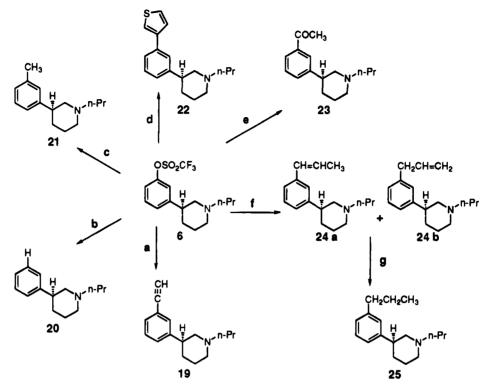
Scheme 2 shows the conditions used for the preparation of compounds 14–18. Metal-halogen exchange²⁷ of the bromo intermediate 13 with sec- or tert-butyllithium and subsequent treatment of the reaction mixture with dimethyl disulfide or DMF afforded the methyl sulfide analog 15 (81%) or the formyl analog 17 (75%), respectively. The yields were significantly improved if the bromo analog 13 was distilled prior to use. Oxidation of compound 15 with 2.3 equiv of m-chloroperoxybenzoic acid in concentrated trifluoroacetic acid afforded compound 16 in a surprisingly low yield. According to GC-MS, no starting material or sulfoxide intermediate was detectable, but after extractive workup of the reaction mixture, only 60-70% of the organic material was recovered. A chromatographic separation of the crude material afforded a 50% yield of the pure product. The aqueous phase from the extractive workup was concentrated and the residue redissolved in absolute ethanol. Analysis of the ethanolic solution by means of TLC showed a product more polar than the sulfonyl analog 16. The ¹H-NMR signals of the methylene groups around the nitrogen were shifted downward and broadened compared to those of 16. We presume this impurity may be the N-oxide 16b, although the reaction was performed in trifluoroacetic acid to protect the amino group. A survey of the literature reveals that many methods have been reported for the reduction of amine N-oxides. We found that 16 could be obtained from 16b after a mild reduction using titanium tetrachloride/sodium iodide (TiCl₄/NaI) in acetonitrile.²⁸ With this extra procedure, the combined yield of compound 16 was enhanced to 93% after the chromatographic separation.

To prepare the sulfonamide 14, the metalated compound 13 was treated with sulfur dioxide (gas), affording the corresponding lithium sulfinate which was then further treated with thionyl chloride. Subsequent treatment with a water solution of dimethylamine (40%) yielded the sulfonamide 14 (38%). Finally, the methylene cyano 18 can be prepared either from aldehyde 17, via a "reductive" cyanation procedure using tosylmethyl isocyanide²⁹/potassium *tert*-butylate in dry dimethoxyethane, or from alcohol 8, which was chlorinated with thionyl chloride in methylene chloride followed by treatment with potassium cyanide in ethanol, in acceptable yield (unpublished data).

For the synthesis of the novel analogs 19-25, we utilized an efficient palladium-catalyzed transformation of triflate derivative 6 (Scheme 3). Introduction of a ethynyl (19, 52%), methyl (21, 58%), or 3-thienyl group (22, 70%) was accomplished by palladium-catalyzed cross-coupling with the corresponding organostannanes, now commonly referred to as the Stille reaction.³⁰ Several attempts were made to obtain the allyl analog 24b from allyltributyltin. Only mixtures of isomers, 24a,b (the major product, 24a, being the result of double-bond migration into conjugation with the ring),³¹ were obtained in modest yield. Chromatographic separation of these isomers was unsuccessful. Therefore the mixture was hydrogenated to yield the propyl analog 25 using Pd/C and ammonium formate in refluxing methanol.³² The triflate substituent was conveniently removed by treatment with formic acid in the presence of a palladium(II) acetate, affording compound 20 in high yield.³³ The acetyl group was introduced by a Heck coupling of 6 with butyl vinyl ether and subsequent hydrolysis with HCl solution to yield 23.34

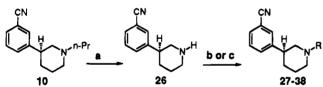
As shown in Scheme 4, compound 26 is the key intermediate in the synthesis of analogs 27-38. The first attempt to obtain 26 from 10 using the von Braun reaction with cyanogen bromide failed.³⁵ Only traces of the desired product were obtained. Selective Ndealkylation of tertiary amines using alkyl or vinyl chloroformates is probably not feasible since the nitrile is quite sensitive to both acidic and basic conditions which are required to hydrolyze the initially formed carbamate.³⁶ A recently reported reagent for selective N-dealkylation utilizes α -chloroethyl chloroformate (ACE-Cl).³⁷ The advantage of the ACE-Cl reagent compared to other chloroformates is that it only requires refluxing methanol to cleave the initially formed carbamate. Treating compound 10 with excess of ACE-Cl in refluxing 1,2-dichloroethane and then in boiling methanol gave 26 in 84% yield.

Scheme 3^a



^a Reagents and conditions: (a) Pd(PPH₃)₄, *n*-Bu₃SnCCH, LiCl, NEt₃, 1,4-dioxan, Δ ; (b) Pd(OAc)₂/PPh₃, NEt₃, HCOOH, DMF, Δ ; (c) Pd(PPh₃)₄, Me₄Sn, LiCl, NEt₃, 1,4-dioxan, Δ ; (d) Pd(PPh₃)₄, *n*-Bu₃Sn-3-thienyl, LiCl, NEt₃, 1,4-dioxan, Δ ; (e) i, butyl vinyl ether, Pd(OAc)₂/Pph₃, NEt₃, DMF, Δ , ii, 5% HCl; (f) (PPh₃)₂PdCl₂/PPh₃, *n*-Bu₃SnCH₂CH=CH₂, LiCl, NEt₃, DMF, Δ ; (g) HCONH₄, Pd/C, MeOH, Δ .

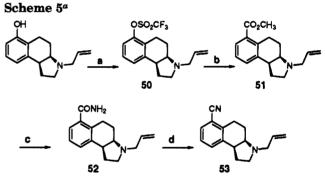
Scheme 4^a



^a Reagents and conditions: (a) i, $ClCO_2CH(Cl)CH_3$, 1,2-dichloroethane, Δ , ii, MeOH, Δ ; (b) RCHO, NaBH(OAc)₃, CH₃COOH, 1,2-dichloroethane; (c) K₂CO₃, R-X, CH₃CN.

Compounds 27-28 were obtained from 26 via reductive amination of the corresponding aldehydes, using sodium triacetoxyborohydride as the reducing agent.³⁸ The remaining tertiary amines (29-38) were prepared from the secondary amine 26 by alkylation with the corresponding alkyl halide (I and Br, etc.) using ground potassium carbonate as the base and acetonitrile as the solvent. The triflates 39-50 were prepared from the corresponding known phenols by using a standard method.³

Scheme 5 shows the synthesis of a few 6-substituted cis- and trans-hexahydro-1H-benz[e]indoles. The triflate 50 was prepared from corresponding hydroxy analog.³⁹ The triflate was converted to methyl ester 51 in a manner similar to the preparation of 7 (Scheme 1). The carboxamide 52 was obtained via hydrolysis of 51 to the carboxylic acid followed by coupling with ammonia (gas) in the presence of diethyl cyanophosphonate (DEPC). The carboxamide 52 was then converted to the nitrile 53 via a preparation similar to that of the nitrile 10 (Scheme 1). In the cis series, the enantiomers were prepared, whereas the trans analogs were tested as the racemates.



^a Reagents and conditions: (a) $(CF_3SO_2)_2O$, pyridine, CH_2Cl_2 ; (b) $Pd(OAc)_2/dppp$, CO(g), NEt_3 , MeOH, Δ ; (c) i, NaOH, $MeOH/H_2O$, ii, NH_3 , DEPC, NEt_3/DMF ; (d) $POCl_3$, DMF, Δ .

Pharmacology

The compounds were tested for their in vitro binding affinity (Tables 1 and 4) to rat striatal DA D1 receptors (utilizing [³H]SCH23390 as ligand), DA D₂ receptors (utilizing [³H]spiperone as ligand), and cloned mammalian DA receptors expressed in CHO-K1 cells (using $[^{3}H]U-86170$ and $[^{3}H]$ spiperone for D_{2} DA receptors and $[^{3}H]$ spiperone for D_{3} and D_{4} DA receptors). All compounds tested displayed less than 50% inhibition at 1 μ M for D₁ and D₄ DA receptors. They were also tested for their affinity at 5-HT_{1A} receptors using [³H]-8-OH-DPAT in either homogenized rat brain tissue or cloned CHO cells. To further establish the pharmacological profile of the compounds, attempting to gain information of their possible target(s) molecules in the CNS, we tested the compounds in an extended battery of CNS in vitro radioligand receptor binding assays according to standard methodology. All compounds tested displayed less than 50% inhibition at 1 μ M for α_1 - ([³H]prazosin), α_2 - ([³H]clonidine), and β -adrenoreceptor

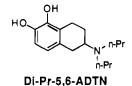
			nonpretreated habituated rats, ^a 100 μ mol/kg sc		nonhabituated rats, ^b 100 μ mol/kg sc		reserpine-pretreated rats,° 100 µmol/kg sc		brain dialysis, ^d 200 μmol/kg sc
no.	х	R	DOPAC ^e	LMA	DOPA acc ^g	LMAf	DOPA acc ^g	LMA ^h	DA release
1 b	3-OH	n-Pr	203* i	119 ± 12	141 ^{<i>j</i>}	40 ^{<i>i</i>}	25 ^j	12 ± 2^{j}	190*
2	3-OMe	n-Pr	149*	$463 \pm 28^*$	131*	$160 \pm 14^*$	66*	23 ± 9	NT^k
3	3-OCH ₂ CF ₃	n-Pr	NT	NT	96	90 ± 17	NT	NT	NT
4	3-OSO ₂ CH ₃	<i>n</i> -Pr	276*	$576 \pm 72^{*}$	249*	154 ± 38	74* [≀]	18 ± 6	234*
5	3-OSO ₂ C ₆ H ₄ CH ₃	<i>n</i> -Pr	89	152 ± 62	NT	NT	NT	NT	NT
6	3-OSO ₂ CF ₃	n-Pr	217*	$290 \pm 85^{*}$	264^{m}	129^{m}	89 ^m	26^{m}	223*
7	3-CO ₂ CH ₃	n-Pr	140	$297 \pm 84^{*}$	NT	NT	NT	NT	NT
8	3-CH ₂ OH	<i>n</i> -Pr	138*	187 ± 28	118	123 ± 17	50*	3 ± 1	175*
9	3-CONH ₂	n-Pr	114	203 ± 40	NT	NT	97^n	NT	NT
10	3-CN	n-Pr	362*	200 ± 40 $653 \pm 41^*$	268^{m}	165^{m}	105^{m}	12^{m}	252*
11	$3-NH_2$	<i>n</i> -Pr	NT	NT	128*	113 ± 17	91^{n}	72 ± 38^{n}	NT
11 12	3-NHSO ₂ CF ₃	n-Pr	91	$305 \pm 66^*$	NT	NT	NT	NT	NT
12	3-Br	<i>n</i> -Pr	145*	$470 \pm 90^{*}$	199*	140 ± 6	101	15 ± 9	363*
13 14		n-Pr	355*	$470 \pm 90^{\circ}$ 548 ± 23*	NT	140 ± 0 NT	NT	15 ± 9 NT	220* ⁿ
	$3-SO_2N(CH_3)_2$	n-Fr n-Pr	118*	$313 \pm 55^*$	NT	NT	92^{n}	5 ± 2^n	NT
15	3-SCH ₃								
16	3-SO ₂ CH ₃	n-Pr	250*	$368 \pm 43^*$	319*	135 ± 17	95	10 ± 7	293*
17	3-CHO	n-Pr	127*	208 ± 51	NT	NT	92	5 ± 3	NT
18	3-CH ₂ CN	$n-\Pr$	302*	$887 \pm 102*$	207*	147 ± 36	57*	16 ± 5	NT
19	3-CCH	<i>n</i> -Pr	138	$671 \pm 137*$	136*	$237 \pm 16^*$	NT	NT	265*
20	3-H	<i>n</i> -Pr	188*	353 ± 66	140*	126 ± 17	65* ⁿ	3 ± 1	473*
22	3-(3-thienyl)	n-Pr	88	229 ± 67	NT	NT	NT	NT	106^{n}
23	3-COCH ₃	n-Pr	183*	$824 \pm 107*$	243*	139 ± 15	73*	7 ± 1	179*
25	3-CH ₂ CH ₂ CH ₃	n-Pr	181*	$734 \pm 191*$	190* 0	$205\pm20^{*}$	62* ^p	11 ± 1	NT
26	3-CN	Н	167*	575 ± 107	271*	$193 \pm 20*$	NT	NT	NT
27	3-CN	Me	174*	$277 \pm 68^{*}$	215*	$79 \pm 2^*$	NT	NT	NT
28	3-CN	\mathbf{Et}	264*	$351 \pm 59^{*}$	318*	113 ± 7	NT	NT	212*
29	3-CN	i-Pr	335*	$366 \pm 33^{*}$	301*	102 ± 15	NT	NT	NT
30	3-CN	allyl	298*	$679 \pm 190^{*}$	257*	$143 \pm 17^*$	85	3 ± 1	522*
31	3-CN	propargyl	146*	$297 \pm 51^{*}$	NT	NT	NT	NT	NT
32	3-CN	<i>n</i> -Bu	257*	$1382 \pm 190^{*}$	NT	NT	71* ⁿ	12 ± 3	NT
33	3-CN	s-Bu	267*	$346 \pm 52^*$	NT	NT	NT	NT	256*
34	3-CN	cpm	272*	$518 \pm 153^{*}$	285*	$191 \pm 31^{*}$	NT	NT	NT
35	3-CN	pheet	231*	$510 \pm 69^*$	176*	120 ± 36	NT	NT	NT
36	3-CN		280*	$484 \pm 111^*$	NT	120 ± 30 NT	NT	NT	NT
	3-CN	pheprop	200*		NT	NT	NT	NT	
37		thiopheneet Et	200* 300*	$500 \pm 92^*$	N1 286*	103 ± 19	NT	NT	NT 120
40	$3-OSO_2CF_3$			$345 \pm 44^{*}$					
41	$3-OSO_2CF_3$	pheet	NT	NT	111^{l}	144 ± 23^{l}	66*	20 ± 20	89 N/F
42	$2-OSO_2CF_3$	n-Pr	83	$621 \pm 156^*$	102	101 ± 14	NT	NT	NT
43	4-OSO ₂ CF ₃	n-Pr	85 ^q	495*	NT	NT	NT	NT	NT
44	3,4-di-OSO ₂ CF ₃	n-Pr	96	36	NT	NT	NT	NT	NT
(R)-apomor	phine		65* /	$491 \pm 61^{* l}$	53* ^s	$277 \pm 14^{*}$		366 ± 36^t	
haloperidol			404* ^v	61 ± 20^{v}	356* ×	$2\pm1^{*}$ *	NT	NT	211* ^y
remoxipride			313*	222	411* ⁿ	60 ± 26^n	104^{n}	5 ± 1	NT

^a The animals were put into the activity meters and 60 min later injected sc with drug. The activity was measured for the subsequent 60 min. ^b Animals were injected with test drug 65 min and with NSD 1015 (100 mg/kg sc) 30 min before death. Controls received corresponding saline injections. Values are expressed as percent of saline controls. ^c Animals were injected with reserpine (5 mg/kg ip) 18 h, test drug 60 min, and NSD 1015 (100 mg/kg sc) 30 min before death. Controls received corresponding vehicle injections. Values are expressed as percent of saline controls. ^c Animals were injected with reserpine (5 mg/kg ip) 18 h, test drug 60 min, and NSD 1015 (100 mg/kg sc) 30 min before death. Controls received corresponding vehicle injections. Values are expressed as percent of saline controls. ^d Effects of tested compounds (60 min after administration) on DA release in the brain dialysis model after 200 μ mol/kg sc (striatum). Values are expressed as percent of saline controls. ^e 3,4-Dihydroxyphenylacetic acid levels in rat striatum. Values are expressed as percent of saline controls. For clarity, SEM values were omitted. However, the SEM never exceeded $\pm 22\%$. ^f Locomotor activity. Values are expressed as percent of saline controls; means \pm SEM. ^e 3,4-Dihydroxyphenylalanine accumulation in rat striatum. For clarity, SEM values were omitted. However, the SEM never exceeded $\pm 22\%$. ^f Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor activity. Values are expressed as percent of saline controls. ^h Locomotor a

 $([^{3}H]dihydroalprenolol)-, 5-HT_{1D} (\alpha + \beta, cloned cells, [^{3}H]-5-OH-tryptamin)-, ⁴⁰ 5-HT_2 ([^{3}H]ketanserin)-, acetylcholine muscarinic ([^{3}H]oxotremorine)-, benzodiazepine ([^{3}H]flunitrazepam)-, and opiate ([^{3}H]etorphine)-labeled sites.$

In addition, the compounds were tested for their ability to displace the DA receptor agonist 5,6-dihydroxy-2-(di-*n*-propylamino)tetralin (Di-Pr-5,6-ADTN) from rat striatal binding sites *in vivo* and their concomitant ability to antagonize the locomotor hyperactivity induced by Di-Pr-5,6-ADTN (Table 3).⁴¹⁻⁴³

In parallel to the *in vitro* screening, the compounds were tested for central DA and 5-HT receptor agonist and/or antagonist activity *in vivo* by the use of biochemical and behavioral methods.^{1,15,44-49} The compounds were biochemically investigated for their ability to



control the synthesis, release, and metabolism of monoamines (DA, 5-HT, and NA) in reserpine-pretreated or nonpretreated rats, with or without habituation to their environment prior to drug treatment (Tables 2 and 5). The behavioral effects of the compounds tested were also monitored in these experiments. The concept of these biochemical and behavioral screening methods has been described earlier.⁴⁹ Furthermore we employed brain microdialysis to moni-

Table 3. Interactions with Di-Pr-5,6-ADTN in Vivo

compd	Di-Pr-5,6-ADTN displacement ^a	locomotor activity, % of ctrl ^b
4	$55 \pm 2^{** c} (100)^d$	$46 \pm 19^{*}$
6	$54 \pm 3^{**}(100)$	78 ± 36
10	$66 \pm 6^{**}(100)^e$	$31 \pm 9^{**}$
16	$45 \pm 8^{**}(100)$	101 ± 17
20	$20 \pm 8(100)$	90 ± 21
23	$44 \pm 6^{**}(100)$	81 ± 6
30	$59 \pm 4^{**}(200)$	$46 \pm 13^{*}$
33	$59 \pm 4^{**}(200)$	$56 \pm 15^{*}$
37	$22 \pm 5^{*}(100)$	$61 \pm 7^{*}$
remoxipride	$48 \pm 6^{*(100)^{e}}$	91 ± 9
haloperidol	$82 \pm 6^{**}(1.3)^e$	$7 \pm 1^{**}$
Di-Pr-5,6-ADTN	0(0.25)	100

^a The animals were injected with Di-Pr-5,6-ADTN (0.25 μ mol/kg sc) 100 min and the test compounds 40 min before death. Shown is the striatal level of Di-Pr-5,6-ADTN (after subtraction of cerebellum levels) expressed as percent of Di-Pr-5,6-ADTN controls. ^b The locomotor activity was recorded 5–35 min after injection of the test compounds and is expressed as percent of Di-Pr-5,6-ADTN controls (saline, 4412 ± 1322 counts/30 min; Di-Pr-5,6-ADTN, 9054 ± 546 counts/30 min). ^c *P < 0.05, **P < 0.01 or less. ^d dose μ mol/kg. ^e Data taken from ref 57.

tor presynaptic effects on dopamine release of the tested compounds.⁵⁰ The data from experiments with haloperidol, remoxipride, cis-(+)-5-methoxy-1-methyl-2-(n-propylamino)tetralin (57), and the DA agonist apomorphine are included for comparison.

Results and Discussion

DA Agonist Properties of Substituted Phenylpiperidines. Hacksell et al.⁵¹ reported on a series of racemic phenylpiperidines with different substituents on the aromatic ring. All compounds except the hydroxy substituted (i.e., racemic 1b) were devoid of intrinsic activity (IA) at DA receptors in reserpine-pretreated rats. This assay is a model for the detection of agonist activity in vivo. Phenolic compounds with R-configuration in the N-alkyl-3-(3-hydroxyphenyl)piperidine series behave as DA receptor agonists with affinity and IA at both pre- and postsynaptic receptors.⁵² The same bifunctional profile has been shown for the S enantiomers with N-substituents bulkier than n-propyl. Likewise, the S enantiomers with ethyl or n-propyl N-substituents seem to interact with both pre- and postsynaptic receptors. However, they have IA at presynaptic receptors but appear to possess antagonist-like properties at postsynaptic receptors.⁵² More recently, developments in the area of molecular biology of DA receptors have led to the suggestion that the difference between DA autoreceptors and postsynaptic receptors is one of sensitivity rather than one of structural diversity.⁵³ This sensitivity difference is suggested to be due to greater DA receptor reserve at presynaptic versus postsynaptic sites.⁵⁴ Thus, a compound with a certain level of IA may have the ability to stimulate presynaptic DA receptors and at the same time display weak antagonistic properties at the postsynaptic receptors. Following this reasoning, compounds with higher IA may show up as postsynaptic DA D_2 agonists.

In vitro binding data on phenylpiperidines with the S-configuration (1-44, Table 1) show that they possess various affinities at the DA receptor sites defined by the dopamine agonist [³H]U-86170 or the dopamine antagonist [³H]spiperone. The affinities for these compounds are generally higher at the high-affinity agonist (HiAg)

binding site (defined by [³H]U-86170) at both rat striatal DA receptors and cloned mammalian DA D_2 and D_3 receptors than at the antagonist site, i.e., low-affinity agonist (LowAg) binding site (defined by [³H]spiperone binding). Interestingly, the ratio of the affinity constants for the HiAg site and the LowAg site has been used as a predictive model for IA at DA D₂ receptors.⁵⁵ This methodological approach has also been successful in the prediction of IA at cortical muscarinic receptors.⁵⁶ However, despite having a binding profile predicting IA. many of the novel phenylpiperidines were unable to stimulate presynaptic DA receptors in vivo as judged by measurements of DOPA accumulation (Table 2). Obviously, in contrast to some other chemical classes, the phenylpiperidines do not lend themselves to predictions of *in vivo* agonist effects by means of *in vitro* D_2 binding assays.

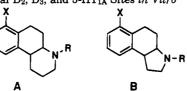
Compound 1b has a clear-cut agonist activity at the presynaptic receptors, shown by a maximal decrease of DOPA accumulation in reserpine-pretreated rats. The lack of increase in locomotor activity indicates the absence of IA at postsynaptic receptors in contrast to, for instance, apomorphine (Table 2). These data support earlier findings by Hacksell et al.⁵¹ In addition, some other (S)-(-)-phenylpiperidines seem to possess a certain, albeit weak, IA at the presynaptic receptors, shown by a slight decrease in DOPA accumulation (Table 2). The electron density in the aromatic ring appears to play an important role for the IA. This is demonstrated by compounds substituted with powerful electron-withdrawing groups (e.g., -CN and -SO₂CH₃) which lack IA in this assay. However, introduction of substituents with varying, albeit lower than -OH, electron-donating properties $(\sigma_{\rm p})$, e.g., an -OMe (2), propyl group (25), or -H (20), generally yields partial agonists (Table 2). Despite the variation of electrondonating capability among these substituents, they all show a similar level of IA. Thus, some other properties, in addition to the electron-donating capability, are implicated for the expression of IA. Interestingly, the hydroxymethyl-substituted compound 8 is the most efficacious agonist next to 1b (in reducing DOPA accumulation), despite the low electron-donating capabilitv of this substituent (Table 2). All of the above show that the IA depends on both hydrogen bond-donating properties and the electron density in the aromatic ring. In support of this, we found that compound 9 substituted with a carboxamide (hydrogen bond-donating but simultaneously strongly electron-withdrawing properties) is completely devoid of IA.

In conclusion, to obtain maximum agonist activity for phenylpiperidines with S-configuration, the requirement is an aromatic substituent simultaneously possessing hydrogen bond- and electron-donating properties (i.e., -OH).

DA Antagonist Properties of the Substituted Phenylpiperidines. Table 2 contains several compounds that are dopamine receptor antagonists, as determined in the *in vivo* biochemical assays. Like haloperidol and remoxipride, they increase the synthesis (DOPA accumulation), release (DA), and metabolism (DOPAC levels) of dopamine with various efficacies and proportions between these measures.

Some compounds, e.g., **19**, **25**, **26**, and **34**, increase the locomotor activity (LMA) in actively exploring rats

Table 4. Analytical Data and Affinities at Central D2, D3, and 5-HT1A Sites in Vitro



-						$K_{ m i} ({ m nM})^a$				
no.	struct	x	R	isomer	formula ^b	D ₂ ^{c,d} [³ H]- spiperone	D ₂ ^{c,d} [³ H]- U-86170	D ₃ ^{c,d} [³ H]- spiperone	5-HT _{1A} , ^{c,d} [³ H]- 8-OH-DPAT	
45	A	OSO ₂ CF ₃	<i>n</i> -Pr	cis-(±)	C ₁₇ H ₂₂ F ₃ NO ₃ S·HCl	>453	744 ± 117	587 ± 52	1140	
46	Α	OSO_2CF_3	<i>n-</i> Pr	trans-(±)	$C_{17}H_{23}F_3NO_3SCI-0.25H_2O$	64 ± 8	21 ± 5	22 ± 2	3 9	
47	В	OSO_2CF_3	n-Pr	$trans-(\pm)$	$C_{16}H_{18}F_3NO_3S\cdot HCl$	82 ± 10	89 ± 8	61 ± 8	72 ± 4^{e}	
48	в	OSO_2CF_3	allyl	$trans-(\pm)$	C ₁₆ H ₁₈ F ₃ NO ₃ S·HCl	>172	125 ± 16	67 ± 3	$18\pm1.4^{\prime}$	
49	В	OSO_2CF_3	allyl	cis-(+)	C ₁₆ H ₁₈ F ₃ NO ₃ S·HCl	>172	353 ± 44	173 ± 31	117 ± 15^{g}	
50	в	OSO_2CF_3	allyl	cis-(-)	C ₁₆ H ₁₈ F ₃ NO ₃ SHCl	>172	>1900	>500	>400	
52	В	$CONH_2$	allyl	cis-(-)	$C_{16}H_{20}N_2O 0.75H_2O$	>172	>1500	>1500	>1000	
53	В	CN	allyl	cis-(-)	C ₁₆ H ₁₈ N ₂ ·HCl	>172	984 ± 74	>455	>400	
54	в	OMe	allyl	cis-(+)		NT^{h}	>339 ⁱ	\mathbf{NT}	>333 ⁱ	
55	В	OMe	allyl	cis-(-)		NT	18 ± 6^i	NT	236 ± 28^i	
56	в	OMe	allyl	$trans-(\pm)$		NT	50 ± 4^i	NT	3.7 ± 2.2^i	

^a K_i values for displacement of the dopamine D₂ receptor antagonist spiperone, the dopamine D₂ receptor agonist U-86170, the dopamine D₃ receptor antagonist spiperone, and the 5-HT_{1A} receptor agonist 8-OH-DPAT. ^b Analyses for C, H, and N were within ±0.4%. ^c Data from cloned mammalian receptors expressed in CHO-K1 cells. ^d Values were obtained with 11 drug concentrations in which each value was determined in duplicate. ^e 5-HT₂ $K_i = 36 \pm 11$ (ketanserin). ^f 5-HT₂ $K_i = 108 \pm 40$ (ketanserin). ^g 5-HT₂ $K_i = 212 \pm 40$ (ketanserin). ^h NT means not tested. ⁱ Data taken from ref 39.

 Table 5. Effects on in Vivo DA Biochemistry (Striatum) and Locomotor Activity

	habituat	treated æd rats,ª .ol/kg sc	reserpine-pretreated rats, b 50 μ mol/kg sc			
n0.	DOPAC	LMA^d	DOPA acc ^e	5-HTP aco ^f	\mathbf{LMA}^{d}	
45 46 47 48 49 50 53 55 57 ⁿ	$\begin{array}{c} 94\pm 6\\ 263\pm 15^{*\ h,i}\\ 223\pm 16^{*\ k}\\ 175\pm 12^{*}\\ 108\pm 6\\ 98\pm 4\\ 172\pm 11^{*}\\ 314\pm 13^{*\ m}\\ 346\pm 29^{*\ m,o} \end{array}$	$\begin{array}{c} 266 \pm 183 \\ 200^{j} \\ 687 \pm 210^{*\;k} \\ 443 \pm 107^{*} \\ 116 \pm 34 \\ 148 \pm 70 \\ 292 \pm 28^{*\;l} \\ 182 \pm 51^{m} \\ 354 \pm 37^{*\;m.o} \end{array}$	NT ^g 85 ± 9 NT 92 ± 6 NT NT NT NT NT	$ \begin{array}{c} \mathrm{NT} \\ 62 \pm 7^* \\ \mathrm{NT} \\ 52 \pm 3^* \\ \mathrm{NT} \end{array} $	$\begin{array}{c} \text{NT} \\ 211 \pm 63^* \\ \text{NT} \\ 635 \pm 16^* \\ \text{NT} \end{array}$	

^a The animals were put into the activity meters and 60 min later injected sc with drug. The activity was measured for the subsequent 60 min.^b Animals were injected with reserpine (5 mg/kg ip) 18 h, test drug 60 min, and NSD 1015 (100 mg/kg ip) 30 min before death. Controls received corresponding saline injections. Values are expressed as percent of saline controls. c 3,4-Dihydroxyphenylacetic acid levels in rat striatum. Values are expressed as percent of saline controls; mean \pm SEM. ^d Locomotor activity. Values are expressed as percent of saline controls; means \pm SEM. ^e 3,4-Dihydroxyphenylalanine accumulation in rat striatum. Values are expressed as percent of saline controls; mean \pm SEM. f 5-Hydroxytryptophan accumulation in rat striatum. ^g NT means not tested. h * p < 0.05 or less. i Nonpretreated rats, 50 μ mol/kg, DOPA accumulation (striatum) = $235 \pm 5^*$, 5-HTP (striatum) $72 \pm 4^*$. ^j LMA, nonpretreated rats, 50 μ mol/kg: 56 ± 4^{*}. ^k 89 μ mol/kg. ¹ HVA (striatum): $255 \pm 13^*$. ^m 52 µmol/kg. ⁿ cis-(+)-5-Methoxy-1-methyl-2-(n-propylamino)tetralin. ° Data taken from ref 1.

to the same level as does a high dose of apomorphine. However, compound **32** is the most efficacious LMA stimulant in habituated rats. It has since long been accepted that locomotor activity is increased when direct or indirect DA agonists stimulate postsynaptic DA receptors in the basal ganglia. The compounds under study increase the release of DA and the levels of DOPAC, as determined by the *in vivo* dialysis experiments (Table 2). This is a general effect observed for DA receptor antagonists (ref 57 and references cited therein). In this series of (S)-phenylpiperidines, there

are a number of compounds that increase DA release to higher levels (e.g., 10, 13, 16, 19, 20, 30, and 33) than does haloperidol or raclopride.⁵⁷ Compounds 10, 19, and **30** are also some of the most behaviorally stimulating compounds within this series. Consequently, it is tempting to suggest that this strong effect on DA release underlies the pronounced increase in LMA observed (Table 2). However, there are also compounds that are relatively weak in this aspect, exemplified by compound 23, but still, it has strong stimulatory properties. Haloperidol releases DA to a similar level as compound 23 but produces hypomotility and catalepsy. Thus, DA release *per se* can not be the only variable responsible for the behavioral effects of DA antagonists in general. An additional explanation of this increase in locomotor activity may be the difference in ability of the (S)phenylpiperidines and the classical neuroleptics to block postsynaptic DA receptors. This has been assayed in the *in vivo* Di-Pr-5,6-ADTN binding model. A general trend in this assay is that the phenylpiperidines are less efficacious than haloperidol or raclopride (Table 3; see, also, Waters et al.⁴³). These data are in support of the previous suggestion that less postsynaptic antagonism favors behavioral activation. Interestingly, the noncataleptogenic antipsychotic drug remoxipride shows similar in vivo binding properties as the antagonist phenylpiperidines.

Similar to the agonist-like compounds of this chemical class, the antagonists generally show low affinity to dopamine receptors in *in vitro* binding studies (Table 1, D_1 and D_4 are not shown but the IC₅₀'s are estimated to be >1000 nM for all compounds). In addition, most antagonist-like compounds (see above) have the same peculiarity as the agonistic phenylpiperidines, i.e., they show higher affinity for the HiAg state of the dopamine D_2 receptor as compared to their affinity for the LowAg state. In addition, the few examples of compounds with high affinity for D_2 receptors *in vitro*, i.e., **17**, **37**, and **41**, have weak, or no, presynaptic effects *in vivo* (Table 2). Thus, like the agonists of this chemical class, the

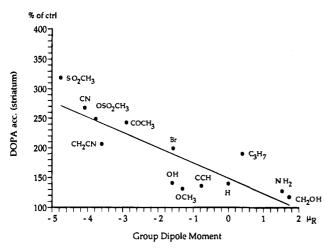


Figure 1. Relationship between DOPA accumulation (nonpretreated rats, striatum, 100 μ mol/kg) and the group dipole moment (μ_R) of the aromatic 3-substituent in (S)-(-)-3-(3-Xphenyl)-1-propylpiperidines (r = 0.86, p = 0.0002, n = 13).

antagonists do not lend themselves to predictions about *in vivo* activity based on *in vitro* binding.

On the basis of the data for compounds substituted with a triflate group, we have found that antagonist activity in the *in vivo* biochemical assays seems to be sensitive for the position on the aromatic ring (Table 2). Substitution in the meta-position leads to an active antagonist compound, while substitution in the ortho-(42) or para-position (43) yields biochemically inactive compounds. Disubstitution with triflate groups in metaand *para*-positions also leads to an inactive compound (44) in the *in vivo* biochemical assays. This further confirms that the in vivo activity is sensitive to steric bulk in the para-position. On the basis of the above results, it seems likely in the case of triflates that the substituent should be positioned in a meta-position with respect to the piperidine ring to yield compounds with antagonistic properties at DA autoreceptors in vivo.

To understand which subtituent properties determine the antagonistic activity of *meta*-substituted phenylpiperidines, substituents with various physical properties^{58,59} were introduced. The resulting series of compounds were examined for increase of striatal DOPA or DOPAC levels (compounds 1-25). It was found that introduction of a methoxy (2) or thiomethyl group (15)does not alter the *in vivo* activity relative to the unsubstituted 20 (Table 2). Compounds substituted with electron-withdrawing groups increased the levels of DOPA more than compound 20. Thus, there seems to be an increasing ability to raise DOPA accumulation paralleled with an increase of the electron-withdrawing properties of the substituent: $-H < -Br < -OSO_2CH_3 <$ $-OSO_2CF_3 \leq -CN < -SO_2CH_3$ (Table 2). However, it is not certain whether it is the lowered electron density in the aromatic ring or the substituents themselves (hydrogen bond-accepting properties, dipole-dipole interactions⁶⁰) which is the most important factor causing the DA receptor antagonist properties. Interestingly, when the maximal increase in DOPA accumulation in the striatum of nonpretreated rats was determined for 11 differently substituted cases, these levels were found to be correlated with the group dipole moment⁶⁰ of the substituent and displayed a near linear relationship (r= 0.86, P = 0.0002, n = 13, Figure 1). Likewise, a correlation between the group dipole moments and the

maximal increases in DOPAC was found (nonpretreated habituated rats, r = 0.73, P < 0.01, n = 14). In addition, no relation between Hansch's π -parameter or Hammett's σ -values and changes in striatal DOPAC was found. These results suggest that the substituent mainly interacts directly with the receptor, i.e., via one or more of several possible interactions involving the substituent dipole. Therefore the electron density or distribution in the aromatic ring may be of secondary importance. This assumption is supported by the fact that compound 18, which is substituted with a $-CH_2$ -CN group, efficiently increases the striatal DOPA levels (Table 2). This group has only weak electron-withdrawing properties ($\sigma_m = 0.16$) but has a strong group dipole moment ($\mu_{\rm R} = -3.6$, for dipole moment sign convention, see ref 60).

The low antagonistic efficacy of compounds 9 (CONH₂) and 17 (CHO) can not be explained by the above reasoning (Table 2), but the hydrogen bond-donating ability of compound 9 may perhaps cancel out the important interaction at the receptor. The lack of activity of the tosylate 5, despite its electron-withdrawing character, may be a steric hindrance effect since the molar refractivity value (used as a measure of bulk of the substituent) is very high (37).⁵⁹

The effect of replacing the N-propyl group in the piperidine ring with different alkyl groups or alkylaryl groups was also investigated (aromatic ring cyano-substituted compounds, **26-38**). The Et (**28**)- and *i*-Pr (**29**)-substituted compounds proved to be the most efficient to increase the DOPA accumulation (300-320%, Table 2). This observation was also made in the triflate series (**6** and **40**), but these compounds were less efficacious than the corresponding cyano analogs. The cyano-substituted secondary amines **26**, **10** (*n*-Pr), **30** (allyl), and **34** (cyclopropylmethyl) were the most efficacious in this series to increase the LMA (in nonpretreated rats, Table 2). A methyl N-substitution surprisingly yields a drug with slight inhibitory effects on the LMA in the same assay.

Compounds with N-substituents holding an aromatic system have the highest affinity for D₂ receptor sites in the *in vitro* binding assay (compounds **35-37** and **41**, Table 1). This may be due to a stabilizing effect *via* $\pi-\pi$ -interactions between the ligand and an accessory binding site in the receptor. However, *in vitro* highaffinity compounds of this structural class have less antagonistic efficacy *in vivo* (Tables 2 and 3).

In conclusion, to obtain pure antagonist properties within the phenylpiperidines with the S-configuration, they should be substituted in the *meta*-position with respect to the piperidine ring. The group should be small, possess a strong negative dipole moment, and also lack hydrogen bond-donating properties. With respect to biochemical effects, the N-substituent should be a *n*-propyl group for maximum *in vivo* antagonist effects at the presynaptic DA receptors.

Substituted Octahydrobenzo[f]quinolines and Hexahydro-1H-benz[e]indoles. The interesting pharmacological properties of the substituted phenylpiperidines (e.g., 6 and 10) encouraged us to investigate the effects of restricting the rotational freedom of the flexible phenylpiperidine system. This can be obtained by joining the piperidine 2-position with the aromatic ring by means of a two-carbon fragment, which can also place the pseudoplane of the piperidine ring in the same plane as the aromatic ring (see below). This resulted in the synthesis of *cis* and *trans* derivatives of the 4-npropyl-1,2,3,4,4a,5,6,10b-octahydrobenzo[f]quinolines (compounds 45 and 46). Previous data show that the trans-7-hydroxy isomer has potent central pre- and postsynaptic dopaminergic receptor agonist properties. The cis isomer was shown to have much less, if any, activity in tests designed to detect dopaminergic agonist activity.⁶¹ Of the resolved enantiomers, the 4aS10bS trans compound was found to be the most potent, while both *cis* enantiomers were inactive as agonists.⁶² The difference in dopaminergic activity between these cis and trans isomers can be rationalized on conformational grounds.^{63,64} The trans ring system is highly rigid and overall a planar molecule, like apomorphine. Molecular modeling reveals that the *cis*-fused system is not completely rigid and can adopt two interconvertible flip conformations. In one, the amino group and the aromatic ring are synclinal, and in the other, the dopamine moiety approaches the antiperiplanar disposition and the aromatic ring approaches coplanarity with the ethylamine side chain. The deviation from overall planarity was proposed to be detrimental for an interaction with the dopamine receptor(s).^{63,64}

The in vitro data (Table 4) show that the triflatesubstituted cis isomer 45 displays only low affinity for D_2 , D_3 , and 5-HT_{1A} sites while the *trans* isomer 46 has quite a high affinity for both D_2 and D_3 receptors. A comparision of the in vitro data of compounds 6 and 46 indicates that the effect of restricting the rotational freedom by forming a rigid and planar system (i.e., 46) does not increase the affinity for D_2 and D_3 receptors dramatically. The affinity for 5-HT_{1A} sites, on the other hand, increases 30-fold, as compared to 6. This increase in 5-HT_{1A} affinity is accompanied by the appearance of agonist activity in vivo, i.e., 46 decreases the 5-HTP accumulation in reserpinized rats (Table 5). In contrast to the potent DA receptor agonist 7-OH and 7-OMe analogs (7-OMe⁶⁵ ED₅₀ = 90 nmol/kg (DOPA accumulation, striatum) versus 130 nmol/kg for 7-OH⁶¹), compound 46 was inactive as an agonist, even at high doses $(50 \,\mu mol/kg, Table 5)$. Instead, presynaptic DA receptor antagonistic properties of 46 were demonstrated by the increase in DOPAC levels in nonpretreated habituated rats. In addition, 46 increases the DOPA accumulation in striatum to $235 \pm 5\%$ (nonpretreated rats, 50 μ mol/ kg, p < 0.005). Interestingly, in the same assay, 46 decreased significantly the LMA to $56 \pm 4\%$. This is in sharp contrast to compound 6 and indicates a stronger affinity for the postsynaptic receptors.

The *cis* isomer **45** is inactive in this assay (Table 5). Furthermore, **42**, a compound where a bulky triflate group in the *ortho*-position with respect to the piperidine ring will force this ring to adopt a conformation where its plane is nearly perpendicular to the plane of the aromatic ring, was found to be inactive. Taken together, these results suggest that the flexible phenylpiperidine **6** interacts with the DA receptor(s) in an almost coplanar conformation.

Recently, SAR on a series of 2,3,3a,4,5,9b-hexahydro-1*H*-benz[*e*]indoles **1e** (BI) was reported.³⁹ The authors found that analogs with 6-methoxy or 6-hydroxy substituents on the aromatic ring act as dopamine receptor antagonists. The *cis* analogs were found to be more potent and selective, versus serotonergic activity, than the corresponding *trans* analogs. The methoxy substitution was found to enhance the antagonistic efficacy relative to hydroxy substitution. It was also shown that the antagonistic activity resides in the 3aR (-)-enantiomers. These results are in line with what has been reported for *cis*- and *trans*-2-amino-5-methoxy-1-methyltetralins, here exemplified by $1a.^2$

Introduction of a triflate group on the aromatic ring in the series of BI's with a *trans* junction (47 and 48) leads to slightly lower affinity for D₂ and 5-HT_{1A} receptors as compared to the corresponding methoxy analog 56. These compounds were also found to display some preference for the DA D₃ receptor *in vitro*. Compound 48 was found to be inactive as a DA agonist, while a partial decrease in 5-HTP accumulation in reserpinized rats was observed at 50 μ mol/kg (sc). This indicates intrinsic activity at the 5-HT_{1A} receptor (Table 5). In nonpretreated habituated rats, 47 and 48 increased the DOPAC levels as well as the LMA (Table 5). Thus, 47 and 48 appear to be antagonists at the presynaptic DA receptors.

Triflate substitution of compounds with the cis junction (49 and 50) leads to loss of *in vitro* affinity for D_2 , D_{3} , and 5-HT_{1A} receptors. This is accompanied by a loss in *in vivo* activity both behaviorally and biochemically. The cyano analog 53 weakly increased the DOPAC levels in nonpretreated habituated rats. These results are intriguing since the corresponding methoxy analog 55 is the most efficient in that sense. It appears that the presence of an electron-attracting group diminishes the in vivo activity for the cis-BI analogs (e.g., OMe versus CN) and an increase in the bulkiness abolished all activity (e.g., OSO_2CF_3). The difference in activity between the methoxy and cyano analogs may be attributed to the difference in lipophilic properties rather than electronic properties since the methoxy group is more lipophilic than the cyano group.⁵⁸ It is of particular interest to note that the trend for the antagonistlike activity in the biochemical assays *in vivo* is OMe > $CN \gg OTf$ for the *cis*-(-)-2,3,3a,4,5,9b-hexahydro-1*H*benz[e]indoles, which has also been found to be valid for the 5-substituted cis-(+)-2-amino-1-methyltetralins.¹⁷

In conclusion, for triflate-substituted octahydrobenzo-[f]quinolines and benz[e]indoles, the trans isomers displayed similar pharmacological properties as the corresponding phenylpiperidines whereas the cis isomers of the former are inactive. Both of the above trans structures have the basic N atom in, or near, the plane of the aromatic ring.⁶⁶ This suggests that the active antagonist conformation of the flexible phenylpiperidines is the one where the two rings are coplanar, or nearly so. The dependence on the aromatic substituent for the presynaptic antagonism of the cis-benz[e]indoles shows the same trend as for the 5-substituted cis-(1S)methyl-(2R)-aminotetralins. This suggests that these latter structural classes interact similarly with the DA autoreceptors but differently from the interaction exerted by the phenylpiperidines.

Experimental Section

General. ¹H- and ¹³C-NMR spectra were recorded in $CDCl_3$ at 300 and 75.4 MHz, respectively, using a Varian XL 300 spectrometer. The spectra for the amines were recorded on

the free bases. Chemical shifts are reported as δ values (ppm) relative to an internal standard (tetramethylsilane). Lowresolution mass spectra were recorded on a HP 5970A instrument operating at an ionization potential of 70 eV. The mass detector was interfaced with a HP5700 gas chromatograph equipped with a fused silica column (11 m 0.22 mm i.d.) coated with cross-linked SE-54 (film thickness 0.3 mm, He gas, flow 40 cm/s). Elemental analyses were performed by Mikro Kemi AB, Uppsala, Sweden. Melting points were determined with a melting point microscope (Reichert Thermovar) and are uncorrected. For flash chromatography, silica gel 60 (0.040-0.063 mm, E. Merck, No. 9385) was used. The amine products were converted into their corresponding HCl salts by dissolving the free base in an ethanolic HCl solution. The solvent was removed and azeotroped with absolute ethanol or toluene in vacuo followed by recrystallization from an appropriate solvent. The fumarate salts were prepared using a similar procedure, substituting HCl for fumaric acid.

Materials. All chemicals used are commercially available and were used without further purification. (S)-(-)-3-(3-Hydroxyphenyl)-1-propylpiperidine (1b) was synthesized by Astra Södertälje using the literature method.⁶⁷ The 3-(tributylstannyl)thiophene was synthesized according to the literature method.⁶⁸

 $(S) \cdot (-) \cdot 3 \cdot (3 \cdot Methoxyphenyl) \cdot 1$ -propylpiperidine (2) (Scheme 1). A solution of 1b (220 mg, 1 mmol), triethylamine (3 mg, 0.03 mmol), and ground sodium hydroxide (100 mg, 2.58 mmol) in 10 mL of THF was stirred at room temperature for 30 min.¹⁸ The temperature was then increased to 30 °C and dimethyl sulfate (129 mg, 1.02 mmol; Warning! Dimethyl sulfate is acutely toxic and also carcinogenic!) was added over a period of about 1.5 h, maintaining the reaction temperature at 25-30 °C by external cooling. The reaction mass was subsequently digested at about 60 °C for 2 h. Most of the conversion takes place prior to digestion and digestion is mainly intended to destroy unreacted dimethyl sulfate. Water (20 mL) was then added and the mixture stirred overnight at room temperature. The aqueous phase was extracted with diethyl ether (three portions), the combined organic phases were washed with brine $(2 \times 5 \text{ mL})$ and dried (MgSO₄), and the solvent was evaporated under reduced pressure. The residue was chromatographed on a silica column using MeOH: CH_2Cl_2 (1:9 (v/v)) as eluent. Concentration of the pooled fractions afforded 205 mg (88%) of pure 2. The amine was converted to the hydrochloride salt using HCl-saturated ethanol and then recrystallized from ethanol/isopropyl ether. This product has been characterized elsewhere:⁵² mp 202-206 °C (HCl, lit mp 200.5-202 °C); MS m/z (rel intensity, 70 eV) 233.15 (M⁺, 5.2), 204.15 (100), 121.05 (15.5), 90.95 (15.3), 70.05 (20.1)

(S)-(-)-1-Propyl-3-[3-(2,2,2-trifluoroethoxy)phenyl]piperidine (3) (Scheme 1). A three-necked round-bottomed flask equipped with a dropping funnel and a magnetic stirring bar was charged with sodium hydride dispersion (60% active, 332 mg, 8.66 mmol in mineral oil). The hydride was washed with *n*-hexane $(2 \times 5 \text{ mL})$ and suspended in 10 mL of anhydrous DMF. Compound 1b (1.86 g, 8.49 mmol), dissolved in anhydrous DMF (30 mL), was slowly added over a period of 45 min. The mixture was then stirred at 40 °C for 1 h, under a nitrogen atmosphere, followed by dropwise addition of 2,2,2trifluoroethyl p-toluenesulfonate (2.27 g, 8.91 mmol) dissolved in 20 mL of DMF.⁶⁹ The mixture was then stirred at 80 °C for 20 h. The reaction mixture was then cooled and poured into an ice/water mixture and the aqueous solution extracted with diethyl ether (4×30 mL). The combined etheral extracts were washed with a 5% aqueous sodium hydroxide solution and brine, dried $(MgSO_4)$, and evaporated. The residue was purified by flash chromatography (petroleum ether:ethyl acetate: Et_3N , 85:10:5 (v/v)), which, after concentration of the pooled fractions, afforded 790 mg (31%) of the title compound as a colorless oil. The amine was converted into the hydrochloride salt and recrystallized from ethanol/diethyl ether: mp 156-160 °C (HCl); ¹H-NMR (300 MHz, D_2O) δ 1.0 (t, 3H), 1.7–2.2 (m, 6H), 2.85-3.2 (m, 5H), 3.6 (m, 2H), 4.6 (q, J = 8.7 Hz, 2H), 7.05 (m, 3H), 7.4 (t, J = 7.6 Hz, 1H); MS m/z (rel intensity,

70 eV) 301.15 (M⁺, 4.2), 273.05 (15.8), 272.15 (100), 189.05 (12.5), 86.10 (9.7), 70.20 (10.8); $[\alpha]^{20}{}_{\rm D}$ –6.7° (c 1.0, MeOH).

(S)-(-)-3-Methanesulfonic Acid 3-(1-Propylpiperidin-3-yl)phenyl Ester (4) (Scheme 1). A solution of 1b (200 mg, 0.91 mmol) and triethylamine (101 mg, 1 mmol) in 15 mL of CH₂Cl₂ was cooled to 0 °C. Then mesyl chloride (136 mg, 1.19 mmol) dissolved in 5 mL of CH₂Cl₂ was added dropwise. The reaction mixture was allowed to reach room temperature and then stirred for 2 h at 25 °C. The reaction was finally quenched with water. The organic layer was separated and washed with 10% HCl and then 10% Na₂CO₃. After drying (MgSO₄), the solvent was removed under reduced pressure. The residue was chromatographed on a silica column using MeOH: $CH_2Cl_2(1:12 (v/v))$ as eluent. The fractions containing pure 4 were collected, and the solvent was removed in vacuo, affording 238 mg (88%) of an oil. The amine was then converted into the fumarate salt and recrystallized from ethanol: mp 164-165 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.35–2.0 (m, 8H), 2.33 (m, 2H), 2.82–3.0 (m, 3H), 3.14 (s, 3H), 7.1-7.4 (m, 4H); MS m/z (rel intensity, 70 eV) 297.05 (M⁺, 2.1), 269.05 (14.6), 268.05 (100), 189.1 (34.5), 188.1 (10.2), 120.0 (8.6); $[\alpha]^{20}D^{-11.6^{\circ}}$ (c 1.0, MeOH).

(S)-(-)-Toluene-4-sulfonic Acid 3-(1-Propylpiperidin-3-yl)phenyl Ester (5) (Scheme 1). This compound was prepared as described for 4, using 1b (150 mg, 0.68 mmol), p-toluenesulfonyl chloride (196 mg, 1.02 mmol), and pyridine (107 mg, 1.36 mmol).¹⁹ Purification of the crude product by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)) afforded 199 mg (78%) of pure 5. The amine was converted into the HCl salt and recrystallized from ethanol/diethyl ether: mp 154 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.25 (qd, J =12.2, 4.5 Hz, 1H), 1.4-2.0 (m, 7H), 2.3 (m, 2H), 2.45 (s, 3H), 2.7 (tt, J = 11.7, 3.6 Hz, 1H), 2.9 (m, 2H), 6.7 (s, 1H), 6.9 (dd, J = 8, 2.5 Hz, 1H), 7.1 (d, J = 7.6 Hz, 1H), 7.2 (t, J = 7.6 Hz, 1H), 7.3 (d, J = 8.7 Hz, 2H), 7.7 (d, J = 9.2 Hz, 2H); MS m/z (rel intensity, 70 eV) 373.15 (M⁺, 2.0), 345.15 (21.1), 344.15 (100), 218.1 (12.2), 189.1 (35.7), 188.1 (11.8); $[\alpha]^{20}_{\rm D} - 4.3^{\circ}$ (c 1.0, MeOH).

(S)-(-)-Trifluoromethanesulfonic Acid 3-(1-Propylpiperidin-3-yl)phenyl Ester (6) (Scheme 1). A solution of 1b (3.3 g, 15.07 mmol) and triethylamine (1.68 g, 16.58 mmol) in 300 mL of CH_2Cl_2 was cooled to -30 °C. Then triflic anhydride (4.68 g, 16.58 mmol) in 30 mL of CH_2Cl_2 was added dropwise. The reaction mixture was allowed to reach room temperature and stirred for 2 h at 25 °C. The reaction was quenched with cold water and the organic layer separated and washed with two portions of cold 5% HCl solution. Following a wash of the organic portion with brine and drying $(MgSO_4)$, the solvent was removed under reduced pressure and the residue was chromatographed on a silica column with MeOH: CH_2Cl_2 (1:19 (v/v)) as eluent. The solvents from the collected fractions containing pure 6 were evaporated yielding a pale oil 4.87 g (92%). Addition of ethanolic HCl to an etheral solution afforded 6 HCl. This product has been characterized elsewhere:³ mp 156-159 °C (HCl).

(S)-(-)-3-(1-Propylpiperidin-3-yl)benzoic Acid Methyl Ester (7) (Scheme 1). A mixture of compound 6 (5.5 g, 15.67 mmol), triethylamine (3.17 g, 31.34 mmol), MeOH (20 g, 626.8 mmol), $Pd(OAc)_2$ (0.105 g, 0.47 mmol), and 1,3-bis(diphenylphosphino)propane (0.194 g, 0.47 mmol) in 60 mL of DMSO was stirred at room temperature for 15 min or until all particles were dissolved.²⁰ A stream of CO(g) (Caution! Highly toxic!) was passed through the solution for 4-5 min, and then the reaction vessel was placed under a slightly positive pressure of CO(g) (1 atm). The temperature was increased to 70 °C (oil bath). After 6 h, GC analysis revealed the complete absence of any 6 and indicated 7 as the major product (see the Discussion). The reaction mixture was allowed to cool to room temperature. Water (200 mL) was then added, and the aqueous solution was extracted with five portions of diethyl ether. The combined organic phases were washed with water until neutral, dried $(MgSO_4)$, and evaporated. The residue was chromatographed on a silica column using MeOH:CH₂Cl₂ (1: 12 (v/v) as eluent. The solvents from the collected fractions containing pure 7 were evaporated yielding 3.43 g (84%). The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: m.p 166–67 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.4–2.1 (m, 8H), 2.3 (m, 2H), 2.9 (tt, 1H), **3.**0 (m, 2H), 3.9 (s, 3H), 7.36 (t, J = 7.62 Hz, 1H), 7.41 (dt, J = 7.8, 1.5 Hz, 1H), 7.88 (dt, J = 7.5, 1.6 Hz, 1H), 7.9 (t, J = 1.3 Hz, 1H); MS m/z (rel intensity, 70 eV) 261.15 (M⁺, 3.0), 233.05 (16.2), 232.05 (100), 100.55 (12.8), 86.05 (7.3); [α]²⁰_D –6.6° (c 1.0, MeOH).

(S)-(-)-[3-(1-Propylpiperidin-3-yl)phenyl]methanol(8) (Scheme 1). A solution of 7 (396 mg, 1.51 mmol) in 30 mL of dry diethyl ether was cooled to 0 °C. Solid LiAlH₄ (400 mg, 10.5 mmol) was then added in small portions.⁷⁰ The mixture was stirred for 1 h at 0 °C. The reaction was then terminated by the addition of 1 mL of water, 1 mL of 15% sodium hydroxide solution, and finally 3 mL of water. The resulting mixture was filtered through a pad of Celite, dried (MgSO₄), and evaporated to dryness. The residue was chromatographed on a silica column with MeOH: CH_2Cl_2 (1:3 (v/ v)) as eluent. The solvents from the collected fractions containing pure 8 were evaporated yielding 293 mg (83%) as an oil. The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: m.p 144-148 °C (HCl); ¹H-NMR (300 MHz, $CDCl_3$) δ 0.9 (t, 3H), 1.4–2.0 (m, 8H), 2.25 (m, 2H), 2.7 (br s, 1H), 2.85 (tt, 1H), 2.95 (m, 2H), 4.7 (s, 2H), 7.1 (d, 1H), 7.2-7.3 (m, 3H); MS m/z (rel intensity, 70 eV) 233.15 (M⁺, 3.7), 205.20 (14.3), 204.1 (100), 131.0 (6.0), 86.0 (6.9); $[\alpha]^{20}{}_{\rm D}$ –7.9° (c 1.0, MeOH).

(S) - (-) - 3 - (1 - Propylpiperidin-3-yl) benzamide (9) (Sceme1). A solution of 7 (12.0 g, 46 mmol) and freshly distilled formamide (7.23 g, 161 mmol) in DMF (150 mL) was heated to 100 °C under a blanket of argon. Sodium methoxide (14.6 g, 82.8 mmol) in methanol (30%) was added dropwise over 30 min under continuous stirring.²³ After 1 h, GC analysis revealed the complete absence of 7 and indicated 9 as the sole product. The reaction mixture was then allowed to reach room temperature. To the reaction mixture was then added 2-propanol (700 mL), and the resulting solution was filtered through a pad of Celite and evaporated to dryness. The residue was chromatographed on a silica column using CH₂Cl₂:MeOH (9:1 + 1% NEt₃ (v/v)) as eluent. Collection of the fractions containing pure product followed by evaporation of the solvent afforded 9.6 g of 9 as an oil (84%) which crystallized on standing: mp 130 °C (free base); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.45–1.6 (m, 2H), 1.7–1.88 (m, 2H), 1.92–2.1 (m, 4H), 2.35 (m, 2H), 2.85-3.1 (m, 3H), 5.8 (br s, 1H), 6.1 (br s, 1H), 7.35-7.46 (m, 2H), 7.65 (dt, J = 7.5, 1.5 Hz, 1H), 7.76(s, 1H); MS m/z (rel intensity, 70 eV) 246.15 (M⁺, 2.7), 217.15 (100), 131.05 (23.1), 100.55 (18.8), 70.05 (19.5); $[\alpha]^{20}$ _D -7.4° (*c* 1.0, CH₂Cl₂).

(S)-(-)-3-(1-Propylpiperidin-3-yl)benzonitrile (10) (Scheme 1). A solution of 9 (6.5 g, 26.4 mmol) and freshly distilled POCl₃ (6 mL, 66 mmol) in dry DMF (50 mL) was heated at 80 °C for 3 h under an argon atmosphere.²⁴ Evaporation of the solvent yielded a dark, oily residue, which was dissolved in water. The solution was then basified with a saturated sodium carbonate solution and extracted several times with CH₂Cl₂. The combined organic layers were evaporated under reduced pressure, and the residue was purified by flash chromatography (acetone:MeOH, 20:1 (v/v)), affording 5.5 g (92%) of pure 10 as an oil. The amine was converted to the hydrochloride salt using HCl-saturated ethanol and then recrystallized from ethanol/isopropyl ether affording the pure salt. This compound has been characterized elsewhere:³ mp 197-199 °C (HCl).

(S)-(-)-3-(1-Propylpiperidin-3-yl)phenylamine (11) (Scheme 1). To a solution of 7 (10 g, 38.31 mmol) in concentrated sulfuric acid (240 mL) and CH₂Cl₂ (400 mL) was added carefully sodium azide (15 g, 231 mmol). After the addition was completed, the mixture was brought to reflux (50 °C). Over a period of 6 h, small portions of additional sodium azide (3 \times 2 g) were added to the reaction mixture.²⁵ After refluxing for a further 20 h, the reaction mixture was cooled to room temperature and the reaction quenched with ice water. The aqueous solution was basified with 20% NaOH, and the organic layer was separated. The aqueous phase was extracted with CH₂Cl₂ (3 \times 200 mL), and the combined organic phases were dried over MgSO₄, filtered, and concentrated *in* vacuo to give 7.12 g (85%) of crude 11 (95% pure according to GLC), which was used in subsequent steps without further purification. A small sample (200 mg) was purified by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)), affording 178 mg (76%) of pure 11 as an oil (the HCl salt is highly hygroscopic): ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.45–2.2 (m, 8H), 2.45 (m, 2H), 2.9 (tt, 1H), 3.1 (m, 2H), 3.75 (br s, 2H), 6.5–6.65 (m, 3H), 7.1 (t, J = 8.3 Hz, 1H); MS m/z (rel intensity, 70 eV) 218.45 (M⁺, 13.2), 190.4 (13.4), 189.4 (100), 120.25 (19.1), 70.15 (19.0); [α]²⁰_D -7.0° (c 1.0, CH₂Cl₂, base).

(S)-(+)-C,C,C-Trifluoro-N-[3-(1-propylpiperidin-3-yl)phenyl]methanesulfonamide (12) (Scheme 1). This compound was prepared as described for 6, using 11 (230 mg, 1.05 mmol) and triflic anhydride (326 mg, 1.16 mmol). Purification of the crude product by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)) afforded 176 mg (48%) of pure 12, which crystallized on standing: mp 124–128 °C (free base); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.4–2.3 (m, 5H), 2.6 (m, 2H), 2.8–3.1 (m, 3H), 3.3 (m, 2H), 3.75 (m, 1H), 6.7 (d, 1H), 7.0 (m, 1H), 7.2– 7.4 (m, 2H); MS *m/z* (rel intensity, 70 eV) 350.2 (M⁺, 2.6), 322.15 (16.2), 321.15 (100), 188.25 (12.3), 187.25 (20.0); $[\alpha]^{20}_{D}$ +123.8° (c 1.0, CH₂Cl₂, free base).

 $(S) \cdot (-) \cdot 3 \cdot (3 \cdot Bromophenyl) \cdot 1 \cdot propylpiperidine$ (13) (Scheme 1). To a solution of 11. HCl (16.28 g, 55.96 mmol) in 100 mL of 48% aqueous HBr at 0 °C was added dropwise with stirring a solution of sodium nitrite (4.2 g, 60.96 mmol) in 4 mL of water.²⁶ The reaction mixture was stirred for 1 h at 0 °C under an argon atmosphere. Cuprous bromide (8.2 g, 57.16 mmol) dissolved in 20 mL of 48% HBr was than added and the solution heated at 80 °C for 40 min. After cooling, 100 mL of water was added and the reaction mixture was made alkaline using concentrated aqueous ammonia. The aqueous solution was extracted with CH_2Cl_2 (3 × 60 mL). The combined organic phases were dried (MgSO₄) and filtered, and the solvent was evaporated in vacuo to give 13.6 g (85%) of crude 13. The residue was purified by flash chromatography using CH₂Cl₂:MeOH (9:1 (v/v)) as eluent, affording pure 13 (9.05 g, 57.3%). The amine was converted to the hydrochloride salt with ethanolic HCl and recrystallized from ethanol/ isopropyl ether: mp 209-211 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.33–2.0 (m, 8H), 2.3 (m, 2H), 2.81 (tt, J = 11.4, 3.3 Hz, 1H), 2.97 (m, 2H), 7.15 (d, J = 4.8 Hz, 2H), 7.32 (dt, J = 4.4, 2.0 Hz, 1H), 7.37 (br, s, 1H); MS m/z (rel intensity, 70 eV) 283.05 (M^+ + 1, 2.8), 282.05 (M^+ , 1.9), 281.05 $(M^+ - 1, 3.4), 253.95 (94.2), 251.95 (100), 129.95 (30.8), 128.95$ (31.7); $[\alpha]^{20}_{D} - 7.9^{\circ}$ (c 1.0, MeOH).

(S)-(-)-N,N-Dimethyl-3-(1-propylpiperidin-3-yl)benzenesulfonamide (14) (Scheme 2). To a solution of 13 (700 mg, 2.49 mmol) in dry THF (20 mL), at -78 °C, was added a solution of sec-butyllitium in hexane (1.4 M, 2.66 mL, 3.73 mmol). The mixture was stirred at -78 °C for 15 min and then allowed to reach 0 °C. After an additional 30 min at 0 $^{\circ}$ C, the solution was brought to -78 $^{\circ}$ C. Dry sulfur dioxide gas was passed into the reaction vessel for 20 min via a needle positioned just above the surface of the solution and gave a copious precipitate.⁷¹ The reaction mixture was then allowed to reach room temperature and stirred for 1 h under a sulfur dioxide atmosphere. The reaction mixture was concentrated in vacuo and pretreated with CH₂Cl₂ (25 mL). The suspension was cooled to 0 °C and SOCl₂ (3 mL) was added dropwise. After 2 h, the mixture was concentrated in vacuo to remove excess of SOCl₂. The oily residue was redissolved in CH₂Cl₂ (35 mL) and added dropwise to a water solution of dimethylamine (25 mL, 40% water solution).⁷² The mixture was then brought to reflux for 3 h under vigorous stirring. The phases were separated, and the water phase was extracted several times with CH₂Cl₂. The combined organic phases were washed with brine, dried (MgSO₄), filtered, and concentrated in vacuo. The residue was purified by flash chromatography using CH₂-Cl₂:MeOH (9:1 (v/v)) as eluent, affording pure 14 (603 mg, 38%). The amine was converted to the fumarate salt and recrystallized from ethanol/isopropyl ether: mp 162-164 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.45-1.6 (m, 3H), 1.7-1.85 (m, 2H), 1.92-2.1 (m, 3H), 2.35 (m, 2H), 2.7 (s, 6H), 2.85–3.1 (m, 3H), 7.45–7.65 (m, 4H); MS m/z (rel intensity, 70 eV) 310.10 (M⁺, 2.9), 282.10 (17.0), 281.10 (100), 173.10 (12.3), 129.0 (10.5); $[\alpha]^{20}_{D} - 6.8^{\circ} (c \ 1.0, CH_2Cl_2).$

(S)-(-)-3-[3-(Methylsulfanyl)phenyl]-1-propylpiperidine (15) (Scheme 2). Compound 13 (5 g) was distilled at 145 °C (0.2 mbar, Kugelrohr distillation) yielding 4.6 g (16.5 mmol) of pure 13 which then was dissolved in dry diethyl ether (150 mL). The solution was cooled to -78 °C, and tertbutyllitium in hexane (1.7 M, 12.63 mL, 21.47 mmol) was added dropwise. The resulting mixture was stirred at -78°C for 15 min, allowed to warm to 0 °C, stirred for additional 30 min at 0 °C, and brought to -78 °C and then treated with freshly distilled dimethyl disulfide (2.5 mL, 26.5 mmol). The reaction mixture was then allowed to reach room temperature and stirred for 1 h. The reaction mixture was then diluted with 10% Na₂CO₃, and the phases were separated. The aqueous phase was extracted with diethyl ether $(3 \times 50 \text{ mL})$, and the combined organic phases were washed with brine, dried (MgSO₄), filtered, and concentrated in vacuo to give 3.87 g (94%) of crude 15. The residue was purified by flash chromatography using CH₂Cl₂:MeOH (12:1 (v/v)) as eluent, affording pure 15 (2.61 g, 81%). The amine was converted to the fumarate salt and recrystallized in ethanol/isopropyl ether: mp 184 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 1.0 (t, 3H), 1.6-2.1 (m, 5H), 2.5 (s, 3H), 2.6-2.7 (m, 3H), 2.9 (m, 2H), 3.55 (m, 3H), 7.0 (d, 1H), 7.1-7.3 (m, 3H); MS m/z (rel intensity, 70 eV) 249.25 (M⁺, 7.9), 221.15 (15.3), 220.15 (100), 150.15 (5.2), 129.15 (7.3); $[\alpha]^{20}D^{-9.4^{\circ}}$ (c 1.0, MeOH).

(S)-(-)-3-[3-(Methylsulfonyl)phenyl]-1-propylpiperidine (16) (Scheme 2). To a solution of 15 (1.7 g, 6.83 mmol) in trifluoroacetic acid (20 mL) was added a solution of m-chloroperoxybenzoic acid (2.71 g, 15.7 mmol) in trifluoroacetic acid (20 mL).73 The mixture was stirred at room temperature for 3 h and poured into ice water. The resulting mixture was made alkaline with 15% NaOH and extracted with CH_2Cl_2 (3 × 25 mL). The combined organic phases were dried (MgSO₄), filtered, and concentrated in vacuo (1.35 g, 71%). The oily residue was purified by flash chromatography using CH_2Cl_2 : MeOH (9:1 (v/v)) as eluent, affording pure 16 (1.0 g, 52%). The amine was converted into the HCl salt and recrystallized in ethano/isopropyl ether: mp 181 °C (HCl); 1 H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.45–1.6 (m, 3H), 1.7– 1.85 (m, 2H), 1.9-2.0 (m, 3H), 2.3 (m, 2H), 2.9-3.1 (m, 3H), 3.1 (s, 3H), 7.5 (m, 2H), 7.8 (m, 2H); MS m/z (rel intensity, 70 eV) 281.25 (M⁺, 2.9), 253.15 (16.1), 252.15 (100), 129.15 (9.6), 70.15 (6.4); $[\alpha]^{20}_{D} -6.1^{\circ}$ (c 1.0, MeOH).

The water phase above was evaporated to dryness, and the resulting salt was redissolved in absolute ethanol (50 mL). Nonsoluble particles were filtered off, and the solution was concentrated *in vacuo* yielding a residue (800 mg, one spot on TLC) which was redissolved in CH₃CN (20 mL). To the *N*-oxide solution was added TiCl₄ (507 mg, 2.68 mmol) and NaI (1.2 g, 8.05 mmol).²⁸ The reaction mixture turned dark brown almost immediately, and TLC, after 40 min, confirmed complete reaction. The reaction was interrupted by adding 10% KOH (30 mL). The water phase was extracted with diethyl ether (3 × 30 mL). The combined organic phases were dried (MgSO₄), filtered, and concentrated *in vacuo*. The residue was further purified as above yielding 310 mg (41%) of pure 1**6**.

(S)-(-)-3-(1-Propylpiperidin-3-yl)benzaldehyde (17) (Scheme 2). To a solution of 13 (3.45 g, 12.3 mmol) in dry diethyl ether (40 mL) at -78 °C was added a solution of tertbutyllitium in hexane (1.7 M, 8 mL, 13.5 mmol). The mixture was stirred at -78 °C under an argon atmosphere for 15 min and allowed to warm to 0 °C. After an additional 30 min at 0 °C, the solution was brought to -78 °C and dry dimethylformamide (1.1 mL, 15.9 mmol) was added. The reaction mixture was allowed to reach room temperature and stirred for 1 h. The reaction mixture was then diluted with 10% Na₂CO₃, and the phases were separated. The aqueous phase was extracted with diethyl ether (3 \times 30 mL), and the combined organic phases were washed with brine, dried (MgSO₄), filtered, and concentrated in vacuo to give 2.72 g (96%) of crude 17. The residue was purified by flash chromatography using CH₂Cl₂: MeOH (9:1 (v/v)) as eluent, affording pure 17 (2.12 g, 75%). The amine was converted to the hydrochloride salt with etheral HCl. The 17·HCl was recrystallized in ethanol/isopropyl ether: mp 166–68 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.45–1.6 (m, 3H), 1.7–1.85 (m, 2H), 1.9–2.0 (m, 3H), 2.3 (m, 2H), 2.85–3.1 (m, 3H), 7.35–7.5 (m, 2H), 7.7–7.9 (m, 2H), 10.0 (s, 1H); MS *m/z* (rel intensity, 70 eV) 231.15 (M⁺, 3.6), 203.2 (14.3), 202.2 (100), 131.10 (17.5), 91 (12.4); [α]²⁰_D –6.0° (c 1.0, MeOH).

 $(S) {\boldsymbol{\cdot}} (-) {\boldsymbol{\cdot}} [3 {\boldsymbol{\cdot}} (1 {\boldsymbol{\cdot}} Propyl piperidin {\boldsymbol{\cdot}} 3 {\boldsymbol{\cdot}} y l) phenyl] acetoni$ trile (18) (Scheme 2). A solution of tosylmethyl isocyanide²⁹ (446 mg, 2.29 mmol) in 1,2-dimethoxyethane (distilled from LiAlH₄, 10 mL) was added dropwise to a stirred suspension of potassium tert-butylate (500 mg, 4.45 mmol) in 10 mL of 1,2dimethoxyethane. The reaction mixture was kept below -30°C under nitrogen. Then a solution of 17 (500 mg, 2.16 mmol) in 15 mL of 1,2-dimethoxyethane was added dropwise to the mixture at -50 to -60 °C. After 60 min, methanol (6 mL) was added to the cold solution, which was then heated to reflux for 20 min. The solution was evaporated to dryness, and the residue was taken up in 10% HCl solution (30 mL). The water phase was extracted with CH₂Cl₂, and the combined organic phases were washed with a saturated solution of NaHCO₃, dried (MgSO₄), filtered, and concentrated in vacuo to give 850 mg (160%) of crude 18. The residue was purified by flash chromatography using CH₂Cl₂:MeOH (9:1 (v/v)) as eluent, affording pure 18 (261 mg, 50%). The amine was converted to the fumarate salt and crystallized in ethanol/isopropyl ether: mp 112-119 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.85 (t, 3H), 1.3-1.5 (m, 3H), 1.6-1.8 (m, 2H), 1.9-2.0 (m, 3H), 2.3 (m, 2H), 2.82 (tt, J = 11.5, 3.7 Hz, 1H), 2.95 (m, 2H), 3.7 (s, 2H), 7.1-7.35 (m, 4H); MS m/z (rel intensity, 70 eV) $242.05\,(M^+,2.3),214.1\,(15.4),213.1\,(100),130.00\,(12.6),115.0$ $(9.7); [\alpha]^{20}_{D} - 12.2^{\circ} (c \ 1.0, \text{ MeOH}).$

(S)-(-)-3-(3-Ethynylphenyl)-1-propylpiperidine (19) (Scheme 3). To a solution of 6 (1.2 g, 3.41 mmol) in 20 mL of 1,4-dioxane were added tri-*n*-butylethenylstannane (1.13 g,3.58 mmol), LiCl (446 mg, 10.2 mmol), Pd(PPh₃)₄ (78.7 mg, 0.068 mmol), and a few crystals of 2,6-di-tert-butyl-4-methylphenol. $^{31}\,$ The resulting mixture was refluxed under nitrogen for 6 h, cooled to room temperature, and treated with 1 mL of pyridine and 2 mL of pyridinium fluoride. The resulting mixture was stirred at room temperature for 16 h. The mixture was diluted with diethyl ether, filtered through a small pad of Celite, and washed with water, 10% HCl, and concentrated sodium chloride solution. The organic phase was dried (MgSO₄) and concentrated, and the residue was purified by flash chromatography using acetone: MeOH (25:1 (v/v)) as eluent, affording pure 19 (400 mg, 52%). The amine was converted into the hydrochloride and recrystallized from ethanol/isopropyl ether: mp 172-174 °C (HCl); ¹H-NMR (300 MHz, $CDCl_3$) δ 0.92 (t, 3H), 1.4–2.0 (m, 8H), 2.31 (m, 2H), 2.81 (tt, 1H), 2.98 (m, 2H), 3.05 (s, 1H), 7.2-7.45 (m, 4H); MS m/z (rel intensity, 70 eV) 227.1 (M⁺, 5.2), 199.1 (15.4), 198.1 (100), 128.05 (15.7), 15.05 (20.7), 70.05 (9.2); $[\alpha]^{20}{}_{D} - 9.7^{\circ} (c$ 1.0, MeOH).

(S)-(-)-3-Phenyl-1-propylpiperidine (20) (Scheme 3). To a stirred solution of 6 (500 mg, 1.42 mmol) in DMF (20 mL) under an argon atmosphere at room temperature were sequentially added triethylamine (575 mg, 5.68 mmol), formic acid (261 mg, 5.68 mmol), PPh₃ (74.4 mg, 0.28 mmol), and Pd-(OAc)₂ (47.8 mg, 0.21 mmol).³³ The reaction temperature was raised to 60 °C. After 6 h, the reaction was complete (GLC) and the reaction mixture was cooled to room temperature; 5% HCl (30 mL) was added, and after another 0.5 h of stirring, the mixture was poured into CH_2Cl_2 (75 mL). The aqueous layer was extracted with CH_2Cl_2 (3 \times 15 mL), and the combined organic layers were washed with water until neutrality, dried (MgSO₄), filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (CH₂Cl₂: MeOH, 9:1 (v/v)), affording 204 mg (71%) of pure 20 as an oil: mp 195-200 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.92 (t, 3H), 1.4-2.0 (m, 8H), 2.35 (m, 2H), 2.82 (tt, 1H), 3.05 (m, 2H), $7.2-7.35 \text{ (m, 5H)}; \text{MS } m/z \text{ (rel intensity, 70 eV) } 203.2 \text{ (M}^+, 5.0),$ 175.1 (12.6), 174.1 (100), 91.05 (16.1), 70.05 (7.9); $[\alpha]^{20}{}_{D}$ -4.7° $(c \ 1.0, MeOH).$

(S)-(-)-1-Propyl-3-*m*-tolylpiperidine (21) (Scheme 3). This compound was prepared as described for 19 from 6 (1.06

g, 3.02 mmol) and tetramethylstannane (0.57 g, 3.18 mmol). Purification of the crude reaction mixture by flash chromatography (acetone:MeOH, 20:1 (v/v)) afforded 380 mg (58%) of pure **21**. The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: mp 193–196 °C (HCl); ¹H-NMR (300 MHz, D₂O, HCl salt) δ 0.95 (t, 3H), 1.7–2.15 (m, 6H), 2.35 (s, 3H), 2.9–3.15 (m, 5H), 3.6 (m, 2H), 7.1–7.25 (m, 3H), 7.32 (t, J = 7.8 Hz, 1H); MS m/z (rel intensity, 70 eV) 217.15 (5.1, M⁺), 189.15 (14.4), 188.15 (100), 105.05 (18.2), 70.05 (14.5); [α]²⁰_D – 5.8° (c 1.0, MeOH).

(S)-(-)-1-Propyl-3-(3:thiophene-3:ylphenyl) piperidine (22) (Scheme 3). This compound was prepared as described for 19 from 6 (1.22 g, 3.47 mmol), 3-(tributylstannyl)-thiophene⁶⁸ (1.55 g, 4.16 mmol), (PPh₃)₂PdCl₂ (195 mg, 0.28 mmol), and PPh₃ (146 mg, 0.56 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂: MeOH, 12:1 (v/v)) afforded 690 mg (70%) of pure 22 as an oil. The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: mp 155-160 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.4-2.1 (m, 8H), 2.3 (m, 2H), 2.85 (tt, 1H), 3.0 (m, 2H), 7.07 (dd, J = 3.6 Hz, 1H), 7.15 (dt, J = 7.7 Hz, 1H), 7.24-7.33 (m, 3H), 7.43 (t, J = 1.8 Hz, 1H), 7.47 (d, J = 1.2 Hz, 1H); MS *m/z* (rel intensity, 70 eV) 285.1 (M⁺, 7.2), 257.1 (20.1), 256.1 (100), 186.00 (15.8), 173.0 (14.8), 128.0 (20.5); [α]²⁰_D -3.0° (c 1.0, MeOH).

 $(S) {\boldsymbol{\cdot}} (-) {\boldsymbol{\cdot}} 1 {\boldsymbol{\cdot}} [3 {\boldsymbol{\cdot}} (1 {\boldsymbol{\cdot}} Propyl piperidin {\boldsymbol{\cdot}} 3 {\boldsymbol{\cdot}} y l) phenyl] ethanone$ (23) (Scheme 3). To a stirred solution of 6 (1.87 g, 5.34 mmol) in DMF (18 mL) under an argon atmosphere at room temperature was sequentially added triethylamine (1.63 g, 16 mmol), butyl vinyl ether (4.01 g, 40 mmol), 1,3-bis(diphe-nylphosphino)propane (309 mg, 0.749 mmol), and Pd(OAc)₂ (129 mg, 0.575 mmol).³⁴ The reaction flask was heated to 80 °C. After 0.5 h, the conversion was complete (GLC) and the reaction mixture was cooled to room temperature; 5% HCl (30 mL) was added, and after another 0.5 h of stirring, the mixture was poured into CH2Cl2 (60 mL). The aqueous layer was extracted with CH_2Cl_2 (3 \times 30 mL), and the combined organic layers were washed with water until neutrality, dried (anhydrous MgSO₄), filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)), affording pure 23 (964 mg, 74%). The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: mp 151-156 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.4–2.1 (m, 8H), 2.2 (m, 2H), 2.25 (s, 3H), 2.95 (tt, 1H), 3.0 (m, 2H), 7.2-7.4 (m, 2H), 7.6-7.8 (m, 2H); MS m/z (rel intensity, 70 eV) 245.15 (M⁺, 3.3), 217.05 (15.8), 216.05 (100), 100.55 (6.1), 86.05 (6.2); $[\alpha]^{20}D$ -5.1° (c 1.0, MeOH).

 $(S) \cdot (-) \cdot 3 \cdot (3 \cdot Allylphenyl) \cdot 1 \cdot propylpiperidine$ (24)(Scheme 3). This compound was prepared as described for 19 from 6 (910 mg, 2.59 mmol) and allyltributyltin (900 mg, 2.72 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 333 mg (53%) of 24 as two double-bond isomers according to GC-MS (24a + 24b). MS *m/z* (rel intensity, 70 eV) 243.15 (M⁺, 5.7), 215.1 (17.5), 214.1 (100), 149.0 (12.3), 129.0 (15.4), 86 (15.0).

(S)-(-)-1-Propyl-3-(3-propylphenyl)piperidine 25 (Scheme 3). Compound 24 (459 mg, 1.88 mmol) was dissolved in 25 mL of methanol. Solid ammonium formate (225 mg, 3.57 mmol) and Pd/C (30 mg) were added.³² The resulting mixture was refluxed under a nitrogen atmosphere for 4 h. The mixture was filtered through a pad of Celite, and the solvent was evaporated in vacuo. The residue was redissolved in 15 mL of 10% Na₂CO₃. The water phase was extracted with CH₂-Cl₂ (4 \times 20 mL). The combined organic phases were dried (MgSO₄), filtered, and evaporated to dryness. The crude product was purified by flash chromatography (CH2Cl2:MeOH, 9:1 (v/v)), affording 25 (407 mg, 88%). The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: mp 176-179 °C (HCl); ¹H-NMR (300 MHz, $CDCl_3$) δ 0.9 (m, 6H), 1.4–2.2 (m, 10H), 2.4 (m, 2H), 2.6 (t, 2H), 2,95 (tt, 1H), 3.2 (m, 2H), 7.0-7.3 (m, 4H); MS m/z (rel intensity, 70 eV) $245.2 (M^+, 6.7), 217.1 (17.4), 216.1 (100),$ 117.0 (8.5), 70.1 (13.1); $[\alpha]^{20}$ _D -4.4° (*c* 1.0, MeOH).

(S)-(+)-3-Piperidin-3-ylbenzonitrile (26) (Scheme 4). A

solution of 10 (3.93 g, 17.2 mmol) in 1,2-dichloroethane (50 mL) was cooled to 0 °C. Then α -chloroethyl chloroformate (3.69 g, 25.8 mmol) in 1,2-dichloroethane (30 mL) was added dropwise at 0 °C.³⁷ The reaction mixture was then brought to reflux and followed by GLC. Five portions (1 mL) of a-chloroethyl chloroformate were added during a period of 2 days, and the conversion was complete (GLC) after 3 days. The heating was interrupted, and the volatiles were evaporated in vacuo. The residue was triturated with methanol (150 mL) and refluxed for 2.5 h. The solvent was evaporated to afford 26 HCl as light-brown crystals. The product was chromatographed on a silica column with CH₂Cl₂:MeOH (25:1 (v/v), the methanol was saturated with $NH_3(g)$) as eluent. Collection of the fractions containing pure product and evaporation of the solvent afforded pure 26 (2.7 g, 84%). A small sample was converted to the fumarate salt and recrystallized in ethanol/isopropyl ether: m.p 120-124 °C (fumarate); ¹H-NMR (300 MHz, $CDCl_3$) δ 1.69 (dq, J = 12.7, 3.2 Hz, 1H), 2.13 (m, 3H), 2.94 (m, 2H), 3.37 (m, 1H), 3.58 (m, 2H), 7.43-7.6 (m, 4H); MS m/z (rel intensity, 70 eV) 186.05 (M⁺, 48.6), 185.05 (28.1), 129.00 (60.8), 128.00 (27.1), 57.00 (100), 56.00 (55.8); $[\alpha]^{20}_{D}$ +1.7° (c 1.0, MeOH).

(S)·(+)·3·(1·Methylpiperidin·3·yl)benzonitrile (27) (Schemes 4). Compound 26 (500 mg, 2.69 mmol), glacial acetic acid (161 mg, 2.69 mmol), and paraformaldehyde (88.7 mg, 2.96 mmol) were mixed in 1,2-dichloroethane (30 mL). Sodium triacetoxyborohydride (854 mg, 4.03 mmol) was added to the solution, and the reaction mixture was stirred at room temperature under a nitrogen atmosphere for 5 h (GLC analysis indicated a complete reaction).³⁸ The reaction was quenched with saturated aqueous NaHCO3, and the product was extracted with CH₂Cl₂. The combined organic phases were dried $(MgSO_4)$ and filtered, and the solvent was evaporated to afford 27 as a oily residue. The product was chromatographed on a silica column with CH₂Cl₂:MeOH (9:1 (v/v)) as eluent. Collection of the fractions containing pure product and evaporation of the solvent afforded pure 27 (481 mg, 89%). The amine was converted into the hydrochloride salt and recrystallized from ethanol/isopropyl ether: mp 210-212 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.68 (qd, $J_1 = 12.6, 2.8$ Hz, 1H), 2.10 (t, J = 16.7 Hz, 2H), 2.46 (m, 1H), 2.73–2.9 (m, 5H), 3.5-3.72 (m, 3H), 7.28-7.61 (m, 4H); MS m/z (rel intensity, 70 eV) 200.05 (M+, 33.1), 199.05 (22.0), 128.95 (16.4), 71.05 (31.4), 57.95 (100); $[\alpha]^{20}_{D}$ +2.5° (c 1.0, methanol).

(S)-(-)-3-(1-Ethylpiperidin-3-yl)benzonitrile (28) (Schemes 4). This compound was prepared as described for 27 from 26 (363 mg, 1.95 mmol) and acetaldehyde (98.7 mg, 2.15 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 12:1 (v/v)) afforded 380 mg (91%) of pure 28 as an oil. The amine was converted to the hydrochloride salt with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 192-194 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.50 (t, J = 7.3 Hz, 3H), 1.68 (qd, J = 12.9, 3.4 Hz, 1H), 2.10 (qt, J = 13.0 Hz, 2H), 2.49 (m, 1H), 2.70 (q, J = 11.5 Hz, 2H), 3.13 (q, J = 7.3 Hz, 2H), 3.54-3.73 (m, 3H), 7.45-7.60 (m, 4H); MS *m/z* (rel intensity, 70 eV) 214.15 (M⁺, 16.3), 199.15 (100), 115.95 (13.7), 72.05 (43.8), 57.05 (47.4); [α]²⁰_D -10.3° (c 1.0, methanol).

(S)-(-)-3-(1-Isopropylpiperidin-3-yl)benzonitrile (29) (Scheme 4). A suspension of 26 (500 mg, 2.69 mmol) and ground K₂CO₃ (400 mg) was stirred in CH₃CN (30 mL) at room temperature. A solution of 2-bromopropane (331 mg, 2.69 mmol) in CH_3CN (5 mL) was added dropwise. The mixture was stirred at 50 °C for 5 days, and 0.5 equiv of 2-bromopropane was added each day. The reaction mixture was filtered, and the volatiles were evaporated in vacuo. The oily residue was chromatographed on a silica column with MeOH:CH₂Cl₂ (1:25 (v/v)) as eluent. Collection of the fractions containing pure product and evaporation of the solvent afforded pure 29 (570 mg, 93%). The amine was converted into the hydrochloride and recrystallized from ethanol/isopropyl ether: mp 185-186 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.1 (dd, J = 6.6 Hz, 6H), 1.46 (qd, J = 12.3, 4.3 Hz, 1H), 1.7–2.0 (m, 3H), 2.2 (m, 2H), 2.8-3.0 (m, 4H), 7.4-7.6 (m, 4H); MS m/z (rel intensity, 70 eV) 228.1 (M⁺, 4.8), 214.1 (15.2), 213.1 (100), 129.05 (5.4), 116.05 (9.8), 70.95 (4.5); $[\alpha]^{20}_{D}$ –14.6° (c 1.0, MeOH).

(S)-(+)-3-(1-Allylpiperidin-3-yl)benzonitrile (30)(Scheme 4). This compound was prepared as described for 29 from 26 (173 mg, 0.93 mmol) and allyl bromide (118 mg, 0.98 mmol, added dropwise over a period of 3 h). The reaction mixture was stirred at room temperature overnight. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 112 mg (64%) of pure 30 as an oil. The amine was converted to the hydrochloride salt with HCl-saturated ethanol and recrystallized from ethanol/ isopropyl ether: mp 183-185 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.67 (qd, J = 12.8, 2.7 Hz, 1H), 2.09 (t, J = 16.7 Hz, 2H), 2.46 (m, 1H), 2.73 (m, 2H), 3.5-3.73 (m, 5H), 5.48-5.57 (m, 2H), 6.21 (m, 1H), 7.44-7.6 (m, 4H); MS m/z (rel intensity, 70 eV) 226.05 (M⁺, 39.8), 225.05 (34.2), 199.05 (37.9), 84.00 (100), 82.00 (42.1); $[\alpha]^{20}_{D}$ +0.2° (c 1.0, MeOH).

(S)-(-)-3-(1-Prop-2-ynylpiperidin-3-yl)benzonitrile (31) (Scheme 4). This compound was prepared as described for 29 from 26 (363 mg, 1.95 mmol) and propargyl bromide (237 mg, 1.99 mmol). The reaction mixture was stirred at room temperature for 1 h. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 25:1 (v/v)) afforded 302 mg (69%) of pure 31 as an oil. The amine was converted to the hydrochloride salt with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 195-196 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 1.45 (qd, J = 12.2, 4.1 Hz, 1H), 1.65-2.0 (m, 3H), 2.3 (m, 3H), 2.9 (m, 3H), 3.35 (d, J= 2.5 Hz, 2H), 7.44-7.6 (m, 4H); MS m/z (rel intensity, 70 eV) 224.05 (M⁺, 82.4), 223.05 (95.3), 129.05 (79.6), 116.00 (54.9), 94.00 (100); [α]²⁰D -7.6° (c 1.0, MeOH).

(S)-(-)-3-(1-Butylpiperidin-3-yl)benzonitrile (32) (Scheme 4). This compound was prepared as described for 29 from 26 (250 mg, 1.34 mmol) and 1-bromobutane (193 mg, 1.41 mmol). The reaction mixture was refluxed for 2 days. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 25:1 (v/v)) afforded 246 mg (76%) of pure 32 as an oil. The amine was converted to the fumarate salt and crystallization from ethanol/isopropyl ether yielded a gum: mp 117-119 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) ∂ 0.92 (t, 3H), 1.2-1.6 (m, 5H), 1.65-2.0 (m, 5H), 2.35 (m, 2H), 2.87 (tt, 1H), 2.96 (br, d, 2H), 7.37-7.53 (m, 4H); MS *m/z* (rel intensity, 70 eV) 242.15 (M⁺, 2.6), 200.1 (14.8), 199.1 (100), 156.0 (5.7), 129.0 (5.5), 116.0 (7.5); [α]²⁰_D -9.6° (c 1.0, MeOH).

(S)-(-)-3-(1-sec-Butylpiperidin-3-yl)benzonitrile (33) (Scheme 4). This compound was prepared as described for 29 from 26 (0.7 g, 3.76 mmol) and 2-iodobutane (0.7 g, 3.8 mmol). The mixture was stirred at 40 °C for 30 h. Purification of the crude reaction mixture by flash chromatography (CH₂-Cl₂:MeOH, 19:1 (v/v)) afforded 700 mg (77%) of pure 33 as an oil. The amine was converted into the fumarate salt and recrystallized from ethanol/isopropyl ether: mp 153-157 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 0.92 (t, 3H), 1.0 (dd, 3H), 1.2-2.0 (m, 6H), 2.1-2.4 (m, 2H), 2.5 (m, 1H), 2.8 (m, 3H), 7.37-7.53 (m, 4H); MS *m/z* (rel intensity, 70 eV) 242.25 (M⁺, 1.1), 227.25 (8.2), 214.25 (15.9), 213.25 (100), 129.1 (5.0), 116.1 (10.6); [α]²⁰_D -19.9° (c 1.0, MeOH).

 $(S) \hbox{-} (-) \hbox{-} 3 \hbox{-} [1 \hbox{-} (Cyclopropylmethyl) piperidin \hbox{-} 3 \hbox{-} yl] ben$ zonitrile (34) (Scheme 4). This compound was prepared as described for 29 from 26 (500 mg, 2.69 mmol) and (bromomethyl)cyclopropane (399 mg, 2.95 mmol). The reaction mixture was stirred at 40 °C for 8 h. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 25:1 (v/v)) afforded 516 mg (80%) of pure 34 as an oil. The amine was converted to the fumarate salt and recrystallized from ethanol/ isopropyl ether: mp 139-142 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 0.12 (m, 2H), 0.54 (m, 2H), 0.90 (m, 1H), 1.48 (dq, J = 12.2 Hz, 4.6 Hz, 1H), 1.75-1.85 (m, 2H), 1.92-2.09(m, 3H), 2.32 (d, J = 6.6 Hz, 2H), 2.92 (tt, J = 11.4, 3.6 Hz, 1H), 3.14 (br, d, 2H), 7.39-7.56 (m, 4H); MS m/z (rel intensity, 70 eV) 240.15 (M⁺, 20.1), 199.05 (94.7), 129.00 (66.8), 116.0 $(34.7), 98.0 (49.5), 96.0 (49.5), 57.0 (74.4), 55.0 (100); [\alpha]^{20}$ 19.1° (c 1.0, MeOH).

(S)-(-)-3-(1-Phenethylpiperidin-3-yl)benzonitrile (35) (Scheme 4). This compound was prepared as described for 29 from 26 (167 mg, 0.9 mmol) and 2-phenylethyl bromide (182 mg, 0.99 mmol). The reaction mixture was stirred at 50 °C overnight. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 211 mg (81%) of pure **35** as an oil. The amine was converted to the fumarate salt and recrystallized from ethanol/isopropyl ether: mp 185–187 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.44 (dq, J = 12.5, 4.45 Hz, 1H), 1.71–2.12 (m, 5H), 2.66 (t, 2H), 2.8–3.07 (m, 5H), 7.17–7.54 (m, 9H); MS m/z (rel intensity, 70 eV) 290.12 (M⁺, 0.1), 200.03 (17.47), 199.03 (100), 156.02 (4.37), 129.07 (3.22); [α]²⁰_D –6.6° (c 1.0, MeOH).

(S)-(-)-3-[1-(3-Phenylpropyl)piperidin-3-yl]benzonitrile (36) (Scheme 4). This compound was prepared as described for 29 from 26 (350 mg, 1.88 mmol) and 1-bromo-3-phenylpropane (237 mg, 1.99 mmol). The reaction mixture was refluxed for 4 h. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 30:1 (v/v)) afforded 410 mg (72%) of pure 36 as an oil. The amine was converted into the fumarate salt and recrystallized from 2-propanol/ isopropyl ether: mp 158-159 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 1.44 (dq, J = 12.0, 4.41 Hz, 1H), 1.71-2.1 (m, 7H), 2.4 (m, 2H), 2.66 (m, 2H), 2.8-3.07 (m, 5H), 7.17-7.54 (m, 9H); MS m/z (rel intensity, 70 eV) 304.15 (M⁺, 3.3), 200.1 (14.3), 199.1 (100), 116.0 (5.7), 91.0 (8.3); $[\alpha]^{20}_{D}$ -18.6° (c 1.0, MeOH).

(S)-(-)-3-[1-(2-Thiophene-3-ylethyl)piperidin-3-yl]benzonitrile (37) (Scheme 4). This compound was prepared as described for 29 from 26 (183 mg, 0.98 mmol) and 2-(thiene-2-yl)ethyl methanesulfonate (222 mg, 1.08 mmol). The reaction mixture was refluxed for 4 h. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 144 mg (60%) of pure 37 as an oil. The amine was converted to the fumarate salt and recrystallized from ethanol/isopropyl ether: mp 195-199 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 1.51 (dq, 1H), 1.73-2.19 (5H), 2.74 (m, 2H), 2.95 (tt, 1H), 3.05-3.11 (m, 4H), 6.86 (dd, 1H), 6.98 (dd, 1H), 7.17 (dd, 1H), 7.3-7.59 (m, 4H); MS *m*/z (rel intensity, 70 eV) 295.95 (M⁺, 0), 200.05 (14.5), 199.05 (100), 156.0 (7.0), 129.0 (5.4), 116.0 (10.5); [α]²⁰D -10.1° (c 1.0, MeOH).

(S)-(-)-3-[1-[3-(Dimethylamino)propyl]piperidin-3-yl]benzonitrile (38) (Scheme 4). This compound was prepared as described for 29 from 26 (416 mg, 2.24 mmol) and 3dimethylaminopropyl chloride hydrochloride (371 mg, 2.35 mmol). The reaction mixture was refluxed for 18 h. Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 3:1 (v/v)) afforded 230 mg (38%) of pure 38 as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol and recrystallized from methanol/ isopropyl ether: mp 264-266 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 1.3 (dq, 1H), 1.5-1.9 (m, 7H), 2.1 (s, 6H), 2.15-2.35 (m, 4H), 2.75 (tt, 1H), 2.85 (m, 2H), 7.1-7.5 (m, 4H); MS m/z (rel intensity, 70 eV) 271.25 (M⁺, 4.8), 226.15 (60.8), 211.15 (69.0), 199.15 (47.4), 197.15 (25.4), 110.05 (28.7), 86.05 (100); [α]²⁰_D -21.6° (c 1.0, MeOH).

(S)-(+)-Trifluoromethanesulfonic Acid 3-(1-Methylpiperidin-3-yl)phenyl Ester (39). This compound was prepared as described for 6 from (S)-3-(3-hydroxyphenyl)-N-methylpiperidine⁵² (200 mg, 1.05 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂: MeOH, 9:1 (v/v)) afforded 307 mg (91%) of pure **39** as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol, and crystallization from methanolisopropyl ether yielded a gum: ¹H-NMR (300 MHz, CDCl₃) δ 1.65 (qd, 1H), 2.05 (dt, 1H), 2.15 (m, 1H), 2.45 (m, 1H), 2.6-2.75 (m, 2H), 2.8 (s, 3H), 3.5-3.7 (m, 3H), 7.1-7.5 (m, 4H); MS *m/z* (rel intensity, 70 eV) 324.0 (M⁺ + 1, 2.5), 323.0 (M⁺, 14.0), 190.05 (42.0), 189.15 (11.5), 118.95 (5.8), 90.95 (14.6), 83.95 (10.3), 70.95 (36.4), 68.95 (16.9), 57.95 (100); $[\alpha]^{20}_{D}$ +1.3° (c 1.0, MeOH).

(S)-(-)-Trifluoromethanesulfonic Acid 3-(1-Ethylpiperidin-3-yl)phenyl Ester (40). This compound was prepared as described for 6 from (S)-3-(3-hydroxyphenyl)-N-ethylpiperidine⁵² (200 mg, 0.97 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)) afforded 289 mg (88%) of pure 40 as an oil. The amine was converted to the hydrochloride salt with HCl-saturated ethanol, and crystallization from ethanol/isopropyl ether yielded a gum: ¹H-NMR (300 MHz, CDCl₃) δ 1.45 (t, 3H), 1.6 (qd, 1H), 2.0 (dt, 1H), 2.15 (m, 1H), 2.4-2.65 (m, 3H), 3.05 (q, 2H), 3.5-

3.75 (m, 3H), 7.1–7.3 (m, 2H), 7.35 (d, 1H), 7.45 (t, 1H); MS m/z (rel intensity, 70 eV) 338.0 (M⁺ + 1, 2.8), 337.0 (M⁺, 15.7), 323.0 (15.5), 322.0 (100), 204.05 (36.4), 203.05 (16.7), 189.05 (50.6), 90.95 (24.7); $[\alpha]^{20}_{D} - 8.0^{\circ}$ (c 1.0, MeOH).

(S)-(-)-Trifluoromethanesulfonic Acid 3-[1-(Phenylethyl)piperidin-3-yl]phenyl Ester (41). This compound was prepared as described for 6 from (S)-3-(3-hydroxyphenyl)-N-(2-phenylethyl)piperidine⁵² (200 mg, 0.71 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 25:1 (v/v)) afforded 246 mg (84%) of pure 41 as an oil. The amine was converted to the fumarate salt and recrystallized in ethanol/isopropyl ether: mp 202-205 °C (fumarate); ¹H-NMR (300 MHz, CDCl₃) δ 1.46 (dq, 1H), 1.7-2.0 (m, 3H), 2.1 (t, 2H), 2.66 (m, 2H), 2.8-3.2 (m, 5H), 7.17-7.54 (m, 9H); MS *m*/z (rel intensity, 70 eV) 323.05 (16.0), 322.05 (100, M⁺ - 91), 189.10 (50.2), 188.20 (11.3), 105.00 (11.1), 91.00 (12.0); [α]²⁰D - 3.0° (c 1.0, base, MeOH).

Trifluoromethanesulfonic Acid 2-(1-**Propylpiperidin-3-yl)phenyl Ester (42).** This compound was prepared as described for **6** from 3-(2-hydroxyphenyl)-1-propylpiperidine⁵¹ (200 mg, 0.91 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 25:1 (v/v)) afforded 271 mg (85%) of pure **42** as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 202–204 $^{\circ}$ C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.4–2.4 (m, 10H), 2.9 (m, 2H), 3.2 (m, 1H), 7.25–7.5 (m, 4H); MS *m/z* (rel intensity, 70 eV) 351.05 (M⁺, 4.1), 322.05 (100), 218.05 (21.6), 189.10 (48.8), 147.00 (31.3), 107.00 (17.1).

Trifluoromethanesulfonic Acid 4-(1-**Propylpiperidin-3-yl)phenyl Ester (43).** This compound was prepared as described for **6** from 3-(4-hydroxyphenyl)-1-propylpiperidine⁵¹ (210 mg, 0.96 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 9:1 (v/v)) afforded 309 mg (92%) of pure **43** as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 166–170 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.65 (qd, 1H), 1.8–3.0 (m, 9H), 3.5 (m, 3H), 7.2 (d, 2H), 7.3 (d, 2H); MS *m/z* (rel intensity, 70 eV) 351.15 (M⁺, 4.1), 323.1 (18.0), 322.05 (100), 189.15 (19.6), 86.15 (33.1), 69.00 (12.6).

(S)-(-)-Trifluoromethanesulfonic Acid 5-(1-Propylpiperidin-3-yl)-2-[[(trifluoromethyl)sulfonyl]oxy]phenyl Ester (44). This compound was prepared as described for 6 from (S)-(-)-3-(3,4-dihydroxyphenyl)-1-propylpiperidine⁷⁴ (340 mg, 1.08 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 12:1 (v/v)) afforded 430 mg (80%) of pure 44 as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 138-140 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.3-2.1 (m, 8H), 2.4 (m, 2H), 2.9 (m, 3H), 7.4 (m, 3H); MS *m/z* (rel intensity, 70 eV) 499.1 (M⁺, 2.8), 472.05 (11.2), 471.1 (17.5), 470.1 (100), 204.2 (20.5), 176.15 (10.4); [α]²⁰D -6.9° (c 1.0, MeOH).

cis-(±)-Trifluoromethanesulfonic Acid 4-Propyl-1,2,3,4,-4a,5,6,10b-octahydrobenzo[f]quinolin-7-yl Ester (45). This compound was prepared as described for 6 from cis-(±)-7hydroxy-4-n-propyl-1,2,3,4,4a,5,6,10b-octahydrobenzo[f]quinoline⁶¹ (235 mg, 0.91 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 300 mg (87%) of pure 45 as an oil. The amine was converted into the hydrochloride with HCl-saturated ethanol and recrystallized from ethanol/isopropyl ether: mp 198–204 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.5–2.0 (m, 8H), 2.4–2.7 (m, 5H), 3.0–3.2 (m, 3H), 7.05 (dd, J₁ = 7.6 Hz, J₂ = 1.6 Hz, 1H), 7.1–7.2 (m, 2H); MS *m*/z (rel intensity, 70 eV) 377.2 (M⁺, 7.2), 349.05 (18.2), 348.05 (100), 244.2 (18.4), 215.1 (20.3).

trans-(\pm)-Trifluoromethanesulfonic Acid 4-Propyl-1,2,3,4,4a,5,6,10b-octahydrobenzo[f]quinolin-7-yl Ester (46). This compound was prepared as described for 6 from trans-(\pm)-7-hydroxy-4-n-propyl-1,2,3,4,4a,5,6,10b-octahydrobenzo[f]quinoline⁶¹ (473 mg, 1.93 mmol). Purification of the crude reaction mixture by flash chromatography (CH₂Cl₂:MeOH, 19:1 (v/v)) afforded 660 mg (91%) of pure 46 as an oil. The amine was converted into the hydrochloride salt with HCl-saturated Sonesson et al.

ethanol and recrystallized from ethanol/diethyl ether: mp 239–241 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 0.9 (t, 3H), 1.25 (m, 1H), 1.4–1.9 (m, 5H), 2.15 (dt, J = 10.8, 2.4 Hz, 1H), 2.25–3.1 (m, 9H), 7.08 (d, J = 8.0 Hz, 1H), 7.2 (t, J = 8.0 Hz, 1H), 7.3 (d, J = 8.0 Hz, 1H); MS m/z (rel intensity, 70 eV) 377.2 (M⁺, 5.7), 349.05 (18.5), 348.05 (100), 244.2 (8.6), 215.1 (25.7).

 $trans.(\pm)$ -Trifluoromethanesulfonic Acid 3-Propyl-2,3,-3a,4,5,9b-hexahydro-1H-benz[e]indol-6-yl Ester (47). A solution of trans-(±)-3-propyl-2,3,3a,4,5,9b-hexahydro-1H-benz[e]indol-6-ol³⁹ (134 mg, 0.5 mmol) and pyridine (10 mL) in 45 mL of methylene chloride was cooled to 0-5 °C, and trifluoromethanesulfonic anhydride was added slowly over a period of 5 min. The resulting brown solution was stirred at room temperature for 2 h. The reaction was quenched with aqueous saturated sodium bicarbonate and extracted with ethyl acetate $(2 \times 1 L)$. The organic layer was washed with water and brine, dried $(MgSO_4)$, filtered, and concentrated in vacuo to give a brown oil. The oil was purified on 400 g of silica gel, eluting with hexane: acetone (4:1 (v/v)) and collecting 40 mL fractions. Fractions homogeneous by TLC were collected and concentrated in vacuo to give pure 47 as a light-yellow oil (160 mg, 88.9%). The amine was converted into the hydrochloride salt with HCl-saturated methanol and recrystallized from EtOAc/hexane: mp 196-197 °C (HCl); ¹H-NMR (300 MHz, $CDCl_3$) δ 1.06 (t, J = 7.3 Hz, 3H), 1.70–4.24 (m, 14 H), 7.10– 7.30 (m, 3H).

trans-(±)-Trifluoromethanesulfonic Acid 3-Allyl-2,3,-3a,4,5,9b-hexahydro-1*H*-benz[e]indol-6-yl Ester (48). This compound was prepared as described for 47 from trans-(±)-3-allyl-2,3,3a,4,5,9b-hexahydro-1*H*-benz[e]indol-6-ol³⁹ (425 mg, 1.6 mmol), and the reaction was run in neat pyridine. Purification of the crude reaction mixture by liquid chromatography (hexane:acetone, 4:1 (v/v)) afforded 420 mg (72%) of pure 48 as an oil. The amine was converted into the hydrochloride with HCl-saturated methanol and recrystallized from EtOAc/hexane: mp 174-175 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 1.88-4.14 (m, 12H), 5.5-6.24 (m, 3H), 7.03-7.34 (m, 3H); MS m/z (rel intensity, 70 eV) 361 (M⁺), other ions at m/z 334, 228, 200.

cis-(+)-Trifluoromethanesulfonic Acid 3-Allyl-2,3-(3aS),4,5(9bR)-hexahydro-1H-benz[e]indol-6-yl Ester (49). This compound was prepared as described for 47 from cis-(+)-3-allyl-2,3,3a,4,5,9b-hexahydro-1H-benz[e]indol-6-ol³⁹ (1.85 g, 8.0 mmol). Purification of the crude reaction mixture by liquid chromatography (hexane:acetone, 9:1 (v/v)) afforded 2.5 g (87%) of pure 49 as an oil. The amine was converted into the hydrochloride with HCl-saturated methanol and recrystallized from EtOAc/hexane: mp 144–145 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.6–3.54 (m, 12H), 5.1–6.05 (m, 3 H), 7.0–7.2 (m, 3H); $[\alpha]_D^{25}$ +6° (c 0.72, MeOH).

cis-(-)-Trifluoromethanesulfonic Acid 3-Allyl-2,3-(3aR),4,5(9bS)-hexahydro-1H-benz[e]indol-6-yl Ester (50) (Scheme 5). This compound was prepared as described for 47 from cis-(-)-3-allyl-2,3,3a,4,5,9b-hexahydro-1H-benz[e]indol-6-ol³⁹ (1.72 g, 7.5 mmol). Purification of the crude reaction mixture by flash chromatography (hexane:acetone, 9:1 (v/v)) afforded 1.83 g (68%) of pure 50 as an oil. The amine was converted into the hydrochloride with HCl-saturated methanol and recrystallized from EtOAc/hexane: mp 144-145 °C; $[\alpha]_D^{25}$ -9° (c 0.72, MeOH).

cis-(-)-3-Allyl-2,3(3aR),4,5(9bS)-hexahydro-1H-benz[e]indole-6-carboxylic Acid Methyl Ester (51) (Scheme 5). This compound was prepared as described for 7 from compound 50 (1.8 g, 5 mmol). Purification of the crude reaction mixture by flash chromatography (hexane:acetone, 4:1 (v/v)) afforded 0.7 g (52%) of pure 51 as an oil: ¹H-NMR (300 MHz, CDCl₃) δ 1.58-3.52 (m, 12H), 3.88 (s, 3H), 5.05-6.03 (m, 3 H), 7.17 (t, J = 8 Hz, 1H), 7.27, (d, J = 8 Hz, 1H), 7.63, (d, J = 8 Hz, 1H).

cis-(-)-3-Allyl-2,3(3aR),4,5(9bS)-hexahydro-1H-benz[e]indole-6-carboxylic Acid Amide (52) (Scheme 5). A solution of methyl ester 51 (0.7 g, 2.6 mmol), 3 N NaOH (1.3 mL), and methanol (5.2 mL) was refluxed overnight. TLC analysis showed no starting material remaining. The mixture was neutralized with 6 N HCl (to pH 5) and concentrated to dryness using methanol and toluene. A light yellow solid as a free acid was recovered. A solution of this solid in DMF (20 mL) and triethylamine (0.54 mL, 3.9 mmol) was flushed with ammonia gas for 10 min at 0 °C and treated with DEPC (0.6 mL, 3.9 mmol). Ammonia gas was bubbled through the solution overnight at room temperature. The reaction mixture was concentrated *in vacuo* to a solid and purified by liquid chromatography on 200 g of silica gel, eluting first with 500 mL of CH₂Cl₂ followed by CH₂Cl₂:MeOH (9:1, v/v) and collecting 40 mL fractions. Homogeneous fractions were combined and concentrated to yield a white solid (0.2 g, 30%). The solid was recrystallized from hexane/acetone: mp 170–172 °C; ¹H-NMR (300 MHz, CDCl₃) δ 1.6–3.6 (m, 12H), 5.1–6.05 (m, 3 H), 7.16–7.27 (m, 3H); $[\alpha]^{25}_{\rm D}$ –56° (c 0.41, MeOH).

cis-(-)-3-Allyl-2,3(3aR),4,5(9bS)-hexahydro-1H-benz[e]indole-6-carbonitrile (53) (Scheme 5). This compound was prepared as described for 10 from compound 52 (109 mg, 0.42 mmol) and POCl₃ (0.2 mL, 2.1 mmol). Purification of the crude reaction mixture by liquid chromatography (hexane:acetone, 4:1 (v/v)) afforded 93 mg (92%) of pure 53 as an oil. The amine was converted into the hydrochloride salt with HCl-saturated methanol and recrystallized from EtOAc/methanol: mp 242– 243 °C (HCl); ¹H-NMR (300 MHz, CDCl₃) δ 1.62–4.14 (m, 12H), 5.5–6.42 (m, 3H), 7.26–7.37 (m, 2H), 7.55 (d, J = 7.5Hz, 1H); [α]²⁵_D –30.8° (c 0.5, MeOH).

Pharmacology. Animals. Animals used in the biochemical and motor activity experiments were male rats of the Sprague–Dawley strain (Bekay, Sollentuna, Sweden), weighing 200–300 g. The rats were kept 5/cage with free access to water and food at least 1 week from arrival until used in the experiments. The animals treated orally with drug were starved 18 h before the experiment.

Drugs. All substances to be tested were dissolved in physiological (0.9%) saline immediately before use, occasionally with the addition of a few drops of glacial acetic acid and/or moderate heating in order to obtain complete dissolution. Reserpine was dissolved in a few drops of glacial acetic acid and made up to volume with 5.5% glucose. Injection volumes were 5 or 10 mL/kg, and all solutions had neutral pH values (except for the solutions of reserpine).

Biochemistry (Biochemically Monitored DA and 5-HT Receptor Agonist or Antagonist Activity). The concept of the biochemical screening method is that a DA or 5-HT receptor agonist stimulates the corresponding receptor and through regulatory feedback systems induces a decrease in tyrosine or tryptophan hydroxylase activity, respectively, and a subsequent reduction in the synthesis rate of DA and 5-HT. DOPA or 5-HTP formation, as determined after in vivo inhibition of the aromatic L-amino acid decarboxylase with NSD 1015 ((3-hydroxybenzyl)hydrazine hydrochloride), is taken as indirect measures of DA and 5-HT synthesis rates, respectively.^{48,75} The biochemical experiments and the determinations of DOPA and 5-HTP by means of HPLC with electrochemical detection were performed according to a modification of a previously described method.^{76,77} Receptor antagonist effects are seen as increases in the synthesis rate of neurotransmitter. This is a result of inhibition of the feedback down regulation of transmitter synthesis. The effects on DOPA accumulation are expressed as percent of controls, which is DOPA striatum = 1350 ± 25 ng/g, mean \pm SEM, n =77 (Tables 2 and 5). In the experiments with habituated rats, no NSD 1015 was administered and the animals were killed 1 h after drug administration. The brains were dissected, and the levels of DOPAC (control levels: striatum 1022 ± 38 ng/g, mean \pm SEM, n = 24) were assayed by means of HPLC with electrochemical detection.⁷⁸ The levels are expressed as percent of controls (DOPAC, Tables 2 and 5).

Motor Activity. Reserpine-Pretreated Animals. The motor activity was measured by means of photocell recordings (M/P 40 Fc electronic motility meter, Motron Products, Stockholm) as previously described.⁷⁵ Eighteen hours prior to the motility testing (carried out between 9 a.m. and 1 p.m.), the rats were injected with reserpine (5 mg/kg sc) in the neck region. On the day of experiment, the different test compounds were administered subcutaneously in the neck region (n = 4). Immediately after drug administration, the rats were put into the motility meters (1 rat/cage). Motor activity was then

followed and recorded for the subsequent 30 min (reserpine control values 3 ± 1 counts/30 min, mean \pm SEM, n = 13, Tables 2 and 5). Observations of gross behavior were made throughout the activity sessions through semitransparent mirrors.

Nonpretreated Animals. The motor activity was measured as described for reserpine-pretreated animals. The different test compounds were administered subcutaneously in the flank. Immediately after drug administration, the rats were placed in the test cages (1 rat/cage) and put into the motility meters. Motor activity was then followed and recorded for the subsequent 30 min (Table 2). Observations of gross behavior were made throughout the activity sessions through semitransparent mirrors. Control levels were 229 \pm 24 counts/30 min (means \pm SEM, n = 4).

Nonpretreated Habituated Animals. These experiments were performed as described above, but the animals were habituated in the test cages 1 h before the injection of the test compound or saline (controls). The habituation resulted in a locomotor activity of about 10% of that seen in nonpretreated animals. The locomotor activity after the test compound was then recorded for 60 min (Tables 2 and 5). Control levels were 44 ± 15 counts/60 min (means \pm SEM, n = 4).

Microdialysis. Following the placement of a small burr hole in the appropriate location on the cranium over the corpus striatum (A 1, L 2.6, and D 6), male Sprague-Dawely rats were stereotaxically implanted with a flexible plastic dialysis probe.⁵⁷ The rats were then allowed to recover for 48 h before the experiment started. After connection to a perfusion pump, delivering a Ringers solution containing in mmol/L: NaCl 140, CaCl₂ 1.2, KCl 3.0, MgCl₂ 1.0, and ascorbic acid 0.04, the rats were placed in an open cage and allowed to move freely within its domains. The dialysate contents of DA and its metabolites were analyzed on a HPLC-EC system, allowing 5 min runs for each sample using a sample splitting technique.⁷⁹ Drugs were dissolved in physiological saline and injected sc in the flank. The effect of the drugs was studied during 3 h. After the experiment, the rats were decapitated and the brains taken out and frozen on a block of dry ice. The location of the probes was controlled by means of a Leitz freezing microtome (Table 3)

Receptor Binding. IC_{50} values were estimated from a nonlinear single-site fit to data obtained from competition binding experiments run in single, duplicate, or triplicate. Radio receptor binding studies with [³H]-8-OH-DPAT (5-HT_{1A} agonist, 143–158 Ci/mmol, New England Nuclear, Boston, MA), and [³H]spiperone (D₂ antagonist, 21–24 Ci/mmol, New England Nuclear, Boston, MA) were performed using rat striatal membrane preparations as previously described.^{80,81}

Radioligands used in cloned mammalian receptors expressed in CHO-K1 cells^{82,83} were [³H]U-86170 (62 Ci/mmol, 2 nM)⁸⁴ and [³H]spiperone (107 Ci/mmol, 0.5 nM) for D₂ dopamine and [³H]spiperone (107 Ci/mmol, 0.6 nM) for D₃ dopamine receptors. The buffer used was 20 mM HEPES and 10 mM MgSO₄, pH 7.4, for D₂ dopamine receptors. The buffer used for D₃ dopamine receptors was 20 mM HEPES, 10 mM MgCl₂, 150 mM NaCl, and 1 mM EDTA, pH 7.4. Incubation of the 0.9 mL binding mixtures was for 1 h at room temperature. Reactions were stopped by vacuum filtration. Counting was with a 1205 betaplate (Wallac) using MeltiLex B/HS (Wallac) as scintillant. Dissociation constants (K_i) were calculated with the Cheng and Prushoff equation.⁸⁵ The data in Tables 1 and 4 are in nM ± SEM.

In Vivo Di-Pr-5,6-ADTN Displacement and Motor Activity Measurements. The procedures applied in these experiments are essentially the same as described by Feenstra et al.⁴¹ but altered according to Carlsson and Löfberg.⁴² The rats were injected with Di-Pr-5,6-ADTN (0.25 μ mol/kg sc); 60 min later, the test drug was injected. After 5 min, the rats were placed in the motility meter boxes. The locomotor activity was recorded for 30 min. After an additional 5 min, the rats were killed by decapitation and their brains rapidly taken out. The striatum and cerebellum were dissected for further analysis of their content of Di-Pr-5,6-ADTN and DOPAC. The "specific" binding to striatal binding sites was calculated by subtracting the Di-Pr-5,6-ADTN content of the cerebellum from that of the striatum. The motility measurements were carried out using a set of eight photocell animal motility meters (digiscan activity monitor RXYZM(16)TAO, Omnitech Electronics, Inc., Columbus, OH). The motility meters were kept in sound- and light-proof boxes equipped with semitransparent mirrors allowing observation of the animals during the course of the experiments. The data are presented in Table 3.

Acknowledgment. The financial support provided by The Upjohn Co. is gratefully aknowledged. We also thank AB Astra Södertälje for the gift of (-)-3PPP. We thank Jonas Lidborg and Maria Boije for the synthetic work they performed. We thank Charles F. Lawson, Sinsaidh K. Schlachter, and Agneta Ekman assisted by Marianne Thorngren and Lucia Gaete for in vitro binding studies as well as Kirsten Sönniksen, Lena Wollter, Mari-Charlotte Ericsson, Elisabeth Ljung, Arja Schedwin, and Lena Löfberg for their work with behavioral, biochemical, and microdialysis experiments and HPLC analyses. We also gratefully acknowledge Stellan Ahl for the construction and building of the motility meter boxes and Torben Kling-Petersen for the computerization of the same.

Supplementary Material Available: List of ¹³C-NMR data and C, H, and N analyses (7 pages). Ordering information is given on any current masthead page.

References

- Svensson, K.; Johansson, A. M.; Magnusson, T.; Carlsson, A. (+)-AJ 76 and (+)-UH 232: Central Stimulants Acting as Prefer-
- AJ 76 and (+)-OH 232: Central Stimulants Acting as Freier-ential Dopamine Autoreceptor Antagonists. Naunyn-Schmiede-berg's Arch. Pharmacol. 1986, 334, 234-245. Johansson, A. M.; Arvidsson, L.-E.; Hacksell, U.; Nilsson, L. G.; Svensson, K.; Carlsson, A. Resolved cis- and trans-2-Amino-5-methoxy-1-methyltetralins: Central Dopamine Recep-tor Agonists and Antagonists. J. Med. Chem. 1987, 30, 602-611.
- (3) Sonesson, C.; Waters, N.; Svensson, K.; Carlsson, A.; Smith, M. W.; Piercey, M. F.; Meier, E.; Wikström, H. Substituted 3-Phenylpiperidines: New Centrally Acting Dopamine Autoreceptor Antagonists. J. Med. Chem. 1993, 36, 3188-3196. Carlsson, A.; Lindqvist, M. Effect of Chlorpromazine or Halo-
- peridol on Formation of 3-Methoxytyramine and Normethanephrine in Mouse Brain. Acta Pharmacol. Toxicol. 1963, 20, 140-144
- (5) Högberg, T. Novel Substituted Salicylamides and Benzamides as Selective Dopamine D2-Receptor Antagonists. Drugs Future **1991**, *16*, 333–357.
- Waters, N.; Sonesson, C.; Wikström, H.; Piercey, M.; Smith, M.; Carlsson, A.; Svensson, K. (-)-DS121 A Novel Dopamine D3and Autoreceptor Preferring Antagonist: Biochemical and Elec-trophysiological Effects. Soc. Neurosci. Abstr. 1993, Part 1: 39.15, 80.
- Svensson, K.; Waters, N.; Sonesson, C.; Wikström, H.; Nichols, (7)N. F.; Carlsson, A. (-)-DS121, A Novel Dopamine D3- and Autoreceptor Preferring Antagonist: Effects on Locomotor Activity in the Rat. Soc. Neurosci. Abstr. 1993, Part 1: 39.16, 80.
- Waters, N.; et al. In preparation. Sedvall, G. Monoamines and Schizophrenia. Acta Psychiatr. Scand. 1990, 82 (Suppl. 358), 7-13.
- (10) Seeman, P. Dopamine Receptors and the Dopamine Hypothesis
- Seeman, P. Dopamine Receptors and the Dopamine Hypothesis of Schizophrenia. Synapse 1987, 1, 133-152.
 Piercey, M. F.; Lum, J. T.; Hoffmann, W. E.; Carlsson, A.; Ljung, E.; Svensson, K. Antagonism of Cocaine's Pharmacological Effects by the Stimulant Dopaminergic Antagonists, (+)-AJ76 and (+)-UH232. Brain Res. 1992, 588, 217-222.
 Clark, D.; Exner, M.; Sonesson, C.; Svensson, K.; Furmidge, L. J. Effects of the Putative Dopamine Autoreceptor Antagonist (-)-DS121 on the Discriminative Stimulus Properties of d-Amphet-amic and Coraine Eur. J. Pharmacol. 1994 in press
- amine and Cocaine. Eur. J. Pharmacol. 1994, in press.
 (13) Clark, D.; Hjorth, S.; Carlsson, A. Dopamine Receptor Agonists: Mechanism underlying Autoreceptor Selectivity. I Review of the Evidence. J. Neural Transm. 1985, 62, 1-52.
- (14) Clark, D.; Hjorth, S.; Carlsson, A. Dopamine Receptor Agonists: Mechanism Underlying Autoreceptor Selectivity. II Theo-
- retical Considerations. J. Neural Transm. 1985, 62, 171-207.
 (15) Hjorth, S.; Carlsson, A.; Clark, D.; Svensson, K.; Wikström, H.; Sanchez, D.; Lindberg, P.; Hacksell, U.; Arvidsson, L.-E.; Johansson, A.; Nilsson, J. L. G. Central Dopamine Receptor Agonist and Antagonist Actions of the Enantiomers of 3-PPP. Device the beam of the Party 1000 01. Psychopharmacology (Berlin) 1983, 81, 89-99.

- (16) Boissier, J. R.; Nedelec, L.; Oberlander, C. Simplified Ergolines as New Dopamine Agonists. Symposium on Dopamine Receptor Agonists; Swedish Pharmaceutical Press: Stockholm, 1983; pp 120-131.
- (17) Haadsma-Svensson, S. R.; Smith, M. W.; Lin, C. H.; Duncan, N. J.; Sonesson, C.; Wikström, H.; Waters, N.; Carlsson, A.; Svensson, K. Synthesis and Biological Activity of C-5 Modified Derivatives of (+)-AJ76 and (+)-UH232: Increased Dopamine D3 Receptor Preference and Improved Pharmacokinetic Properties. Bioorg. Med. Chem. Lett. 1994, 4, 689-694
- (18) Fafadia, N. J.; Wakankar, D. M.; Nambudiry, M. E. Triethy-lamine-Catalyzed O-Methylation of Alcohols With Dimethyl Sulfate. Chem. Ind. (London) 1988, 6, 203-204.
- (19) Kabalka, G. W.; Varma, M.; Varma, R. S.; Srivastava, P. C.; Knapp, F. F. Tosylation of Alcohols. J. Org. Chem. 1986, 51, 2386 - 2388
- (20) Dolle, R. E.; Schmidt, S. J.; Kruse, L. I. Palladium Catalyzed Alkoxycarbonylation of Phenols to Benzoate Esters. J. Chem. Soc., Chem. Commun. 1987, 904–905.
- (21) Saa, J. M.; Dopico, M.; Martorell, G.; Garcia-Raso, A. Deoxygenation of Highly Hindered Phenols. J. Org. Chem. 1990, 55, 991-995
- (22) Cabri, W.; Candiani, I.; DeBernardinis, S.; Francalanci, F.; Penco, S.; Santi, R. Heck Reaction on Anthraquinone Derivatives: Ligand, Solvent, and Salt Effects. J. Org. Chem. 1991, 56, 5796-5800.
- (23) Jagdmann, E. G.; Munson, R. H.; Gero, T. W. A Mild Efficient Procedure for the Conversion of Carbocylic Acid Esters to Primary Amides Using Formamide/Methanolic Sodium Methoxide. Šynth. Commun. 1990, 20, 1203-1208.
- (24) Surrey, A. R. Malononitrile. Organic Syntheses; Wiley: New York, 1955; Collect. Vol. No. III, pp 535-536.
- Werner, N. W.; Casanova, J. Cyclobutylamine. Organic Syntheses; (25)Wiley: New York, 1973; Collect. Vol. No. V, p 273.
- (26) Hartwell, J. L. o-Chlorobrombenzene. Organic Syntheses; Wiley: New York, 1955; Collect. Vol. No. III, p 185. Wakefield, B. J. Preparation of Metal-Halogen Exchange. The
- Chemistry of Organolithium Compounds; Pergamon Press Ltd: Tokyo, 1974; pp 51-62. (28) Balicki, R. TiCl4/NaI – A Novel, Efficient Reagent for Mild
- Reduction of The N-O Bond in Amine N-Oxides and Nitrones. Chem. Ber. 1990, 123, 647-648
- van Leusen, A. M.; Oomkes, P. G. One-Step Conversion of (29)Aldehydes to Nitriles. Introduction of a One-Carbon Unit. Synth. Commun. 1980, 10, 399-403.
- (30) Stille, J. K. The Palladium-Catalyzed Cross-Coupling Reactions of Organotin Reagents with Organic Electrophiles. Angew. Chem., Int. Ed. Engl. 1986, 25, 508-524. (31) Echavarren, A.; Stille, J. K. Palladium-Catalyzed Coupling of
- Aryl Triflates with Organostannanes. J. Am. Chem. Soc. 1987, 109, 5478-5486.
- (32) Cortese, N. A.; Heck, R. F. Palladium-Catalyzed Reductions of α,β-Unsaturated Carbonyl Compounds, Conjugated Dienes, and Acetylenes with Trialkylammonium Formates. J. Org. Chem. **1978**, 43, 3985–3987. (33) Cacchi, S.; Ciattini, P. G.; Morera, E.; Ortar, G. Palladium-
- Catalyzed Triethylammonium Formate Reduction of Aryl Triflates. A Selective Method for the Deoxygenation of Phenols. Tetrahedron Lett. 1986, 27, 5541-5544.
- (34) Cabri, W.; Candiani, I.; Bedeschi, A. Ligand-Controlled α-Regioselectivity in Palladium-Catalyzed Arylation of Butyl Vinyl Ether. J. Org. Chem. 1990, 55, 3654-3655.
- (35)von Braun, J. Die Aufspaltung Cyclisher Basen Durch Bromcyan. Chem. Ber. 1909, 42, 2035–2057.
- (36) Olofson, R. A.; Schnur, R. C.; Bunes, L.; Pepe, J. P. Selective N-Dealkylation of Tertiary Amines with Vinyl Chloroformate: An Improved Synthesis Of Naloxone. Tetrahedron Lett. 1977, 1567 - 1570.
- (37) Olofson, R. A.; Martz, J. T.; Senet, J.-P.; Piteau, M.; Malfroot, T. A New Reagent for the Selective, High-Yield N-Dealkylation of Tertiary Amines: Improved Syntheses of Naltrexone and Nalbuphine. J. Org. Chem. 1984, 49, 2081–2082.
 (38) Abdel-Magid, A. F.; Maryanoff, C. A.; Carson, K. G. Reductive
- Amination of Aldehydes and Ketones by Using Sodium Triac-etoxyborohydride. Tetrahedron Lett. 1990, 31, 5595-5598.
- EURYDUTODYGTIGE. 1etrahedron Lett. 1990, 31, 5595-5598.
 Lin, C.-H.; Haadsma-Svensson, S. R.; Lahti, R. A.; McCall, R. B.; Piercey, M. F.; Schreur, P. J. K. D.; von Voigtlander, P. F.; Smith, M. W.; Chidester, C. G. Centrally Acting Serotonergic and Dopaminergic Agents. 1. Synthesis and Structure-Activity Relationships of 2,3,3a,4,5,9b-Hexahydro-1H-benz[e]indole Derivatives. J. Med. Chem. 1993, 36, 1053-1068.
 Veldmar, S. A. Bischewick, M. J. Claster, J. F. (39)
- (40) Veldman, S. A.; Bienkowski, M. J. Cloning and Pharmacological Characterization of a Novel Human 5-Hydroxytryptamine-1D Receptor Subtype. *Mol. Pharmacol.* 1992, 42, 439-444.
 (41) Feenstra, M. G. P.; Rollema, H.; Mulder, T. B. A.; Westerink, B.
- H. C.; Horn, A. S. In Vivo Dopamine Receptor Binding with a Non-Radioactively Labeled Agonist, dipropyl-5,6-ADTN. Life Sci. 1983, 32, 1313-1323.

- (42) Carlsson, A.; Löfberg, L. In Vivo Displacement by 3-PPP Enan-tiomers of N,N-Dipropyl-5,6-ADTN from Dopamine Receptor-Binding Sites in Rat Striatum. J. Neural Transm. 1985, 64, 173 - 185
- (43) Waters, N.; Löfberg, L.; Haadsma-Svensson, S.; Svensson, K.; Sonesson, C.; Carlsson, A. Differential effects of dopamine D2 and D3 receptor antagonists in regard to dopamine release, in vivo receptor displacement and behaviour. J. Neural. Transm. Submitted for publication.
- Carlsson, A.; Davis, J. N.; Kehr, W.; Lindqvist, M.; Atack, C. V. (44)Simultaneous measurement of tyrosine and tryptophan hydroxylase activities in brain in vivo using an inhibitor of the aromatic amino acid decarboxylase. Naunyn-Schmiedeberg's Arch. Pharmacol. 1972, 275, 153-168.
 (45) Aghajanian, G. K.; Bunney, B. S.; Kuhar, M. J. New Concepts
- Neurotransm. Regul., Proc. Symp. Drug Abuse Metab. Regul. Neurotransm., 1972, 115–134.
- (46) Neckers, L. M.; Neff, N. H.; Wyatt, R. J. Increased serotonin turnover in corpus striatum following an injection of kainic acid: evidence for neuronal feedback regulation of synthesis. Naunyn-Schmiedeberg's Arch. Pharmacol. 1979, 306, 173–177.
- (47) Hjorth, S.; Carlsson, A.; Lindberg, P.; Sanchez, D.; Wikström, H.; Arvidsson, L. E.; Hacksell, U.; Nilsson, L. G. J. 8-Hydroxy-2-di-n-propylaminotetralin, 8-OH-DPAT: A Potent and Selective Simplified Ergot Congener with Central 5-HT Receptor Stimu-lating Activity. J. Neural Transm. 1982, 55, 169–188.
- (48) Wikström, H.; Lindberg, P.; Martinson, P.; Hjorth, S.; Carlsson, A.; Hacksell, U.; Svensson, U.; Nilsson, J. L. G. Pivaloyl Esters of N,N-Dialkylated Dopamine Congeners. Central Dopamine-Receptor Stimulating Activity. J. Med. Chem. 1978, 21, 864.
- (49) Andén, N.-E ; Carlsson, A ; Häggendal, J. Adrenergic mechanisms. Annu. Rev. Pharmacol. **1969**, 9, 119–134.
- (50) Zetterström, T.; Sharp, T.; Marsden, C.A.; Ungerstedt, U. In Vivo Measurement of Dopamine and Its Metabolites by Intracerebral Dialysis: Changes After d-Amphetamine. J. Neurochem. 1983, 41, 1769-1773.
- (51) Hacksell, U.; Arvidsson, L.-E.; Svensson, U.; Nilsson, L. G.; Sanchez, D.; Wikström, H.; Lindberg, P.; Hjorth, S.; Carlsson, A. 3-Phenylpiperidines. Central Dopamine-Autoreceptor Stimulating Activity. J. Med. Chem. 1981, 24, 1475-1482.
 Wikström, H.; Sanchez, D.; Lindberg, P.; Hacksell, U.; Arvidsson,
- L.-E.; Johansson, A.; Thorberg, S.-O.; Nilsson, J. L. G.; Svensson, K.; Hjorth, S.; Clark, D.; Carlsson, A. Resolved 3-(3-hydroxyphenyl)-N-n-propylpiperidine, 3-PPP and its Analogues: Central Dopamine Receptor Activity. J. Med. Chem. 1984, 27, 1030-1036.
- (53)Carlsson, A. Dopamine Receptor Agonists: Intrinsic Activity vs. State of Receptor. J. Neural Transm. 1983, 57, 309-315.
- Meller, E.; Bohmaker, K.; Namba, Y.; Friedhoff, A.; Goldstein, (54)M. Relationship Between Receptor Occupancy and Response at Striatal Dopamine Autoreceptors. Mol. Pharmacol. 1987, 31, 592 - 598
- (55) Lahti, R. A.; Figur, L. M.; Piercey, M. F.; Ruppel, P. L., Evans, D. L. Intrinsic Activity Determinations at the Dopamine D2 Guanine Nucleotide-Binding Protein-Coupled Receptor: Utilization of Receptor State Binding Affinities. Mol. Pharmacol. 1992, 42, 432–438.
- (56) Freedman, S. B.; Harley, E. A.; Iversen, L. L. Relative Affinities of Drugs Acting at Cholinoceptors in Displacing Agonist and Antagonist Radioligands: the NMS/Oxo-M ratio as an Index of Efficacy at Cortical Muscarinic Receptors. Br. J. Pharmacol. 1988, 93, 437-445.
- (57) Waters, N.; Lagerkvist, S.; Löfberg, L.; Piercey, M.; Carlsson, A. The Dopamine D3 Receptor and Autoreceptor Preferring Antagonists (+)-AJ76 and (+)-UH232; a Microdialysis Study. Eur. J. Pharmacol. 1993, 242, 151–163. (58) Hansch, C.; Leo, A.; Taft, R. W. A Survey of Hammett Substituent
- Constants and Resonance and Field Parameters. Chem. Rev. 1991, 91, 165-195.
- (59) Hansch, C.; Leo, A.; Unger, S. H.; Kim, K. H.; Nikaitani, D.; Lien, E. J. Aromatic Substituent Constants for Structure-Activity Correlations. J. Med. Chem. 1973, 16, 1207-1216.
- (60) Lien, E. J.; Guo, Z.-R.; Li, R.-L.; Su, C.-T. Use of Dipole Moment as a Parameter in Drug-Receptor Interaction and Quantitative Structure-Activity Relationship Studies. J. Pharm. Sci. 1982, 71, 641-655.
- (61) Wikström, H.; Sanchez, D.; Lindberg, P.; Arvidsson, L.-E.; Hacksell, U.; Johansson, A.; Nilsson, L. G.; Hjorth, S.; Carlsson, A. Monophenolic Octahydrobenzo[f]quinolines: Central Dopamine- and Serotonin-Receptor Stimulating Activity. J. Med. Chem. 1982, 25, 925-931.
- (62) Wikström, H.; Andersson, B.; Sanchez, D.; Lindberg, P.; Arvidsson, L.-E.; Johansson, A. M.; Nilsson, L. G.; Svensson, K.; Hjorth, S.; Carlsson, A. Resolved Monophenolic 2-Aminotetralines and ,2,3,4,4a,5,6,10b-Octahydrobenzo[/]quinolines: Structural and Stereochemical Considerations for Centrally Acting Pre- and Postsynaptic Dopamine-Receptor Agonists. J. Med. Chem. 1985, 28, 215-225.

- (63) Cannon, J. G.; Suarez-Gutierrez, C.; Lee, T.; Long, J. P.; Costall, B.; Fortune, D. H.; Naylor, R. J. Rigid Congeners of Dopamine Based on Octahydrobenzo[/]quinoline: Peripheral and Central Effects. J. Med. Chem. 1979, 22, 341-347.
 (64) Liljefors, T.; Bøgesø, K. P.; Hyttel, J.; Wikström, H.; Svensson, K. Congace, A. Pac, and Pacture et al. Dearwing and Activities.
- K.; Carlsson, A. Pre- and Postsynaptic Dopaminergic Activities of Indolizidine and Quinolizidine Derivatives of 3-(3-Hydrox-yphenyl)-N-(n-propyl)piperidine (3-PPP). Further Developments of a Dopamine Receptor Model. J. Med. Chem. 1990, 33, 1015-1022
- (65) Unpublished results from our group.
 (66) Chidester, C. G.; Lin, C.-H.; Lahti, R. A.; Haadsma-Svensson, S. R.; Smith, M. W. Comparision of 5-HT1A and Dopamine D2 Pharmacophores. X-ray Structures and Affinities of Conformationally Constrained Ligands. J. Med. Chem. 1993, 36, 1301-1315.
- (67) Thorberg, S.-O.; Gawell, L.; Csöregh, I.; Nilsson, J. L. G. Large Scale Synthesis and Absolute Configuration of (-)-3-PPP, a Selective Dopamine Autoreceptor Agonist. Tetrahedron 1985, 41, 129 - 139.
- (68) Gronowitz, S.; Peters, D. Convenient Synthesis of Various Terheterocyclic Compounds by Palladium(0)-Catalyzed Coupling
- Reactions. Heterocycles 1990, 30, 645–658. Camps, F.; Coll, J.; Messeguer, A.; Pericas, M. A. A simple Method for Preparation of Aryl 2,2,2-Trifluoroethyl Ethers. Synthesis 1980, 727–728. (69)
- (70) Hajos, A. Reduction of Carboxylic acids and their derivatives. Complex Hydrides; Elsevier Scientific Publishing Co.: New York, 1979; pp 89-97. (71) Kitching, W.; Fong, C. W. Insertion of Sulfur Dioxide and Sulfur
- Trioxide Into Metal-Carbon Bonds. Organomet. Chem. Rev., Sect. A 1**970**, *5*, 281–321.
- (72) de Boer, T. J.; Backer, H. J. p-Tolylsulfonylmethylnitrosamide. Organic Syntheses; Wiley: New York, 1963, Collect. Vol. No. IV, 943
- (73) Russel, G. A.; Ochrymowycz, L. A. β -Keto sulfoxides. IX. Conversion into Acetylenic Sulfoxides and Sulfones. J. Org. Chem. 1970, **35, 2**106–2108
- (74) Rollema, H.; Mastebroek, D.; Wikström, H.; Svensson, K.; Carlsson, A.; Sundell, S. Enantiomers of 3-(3,4-Dihydroxyphenyl)- and 3-(3-Hydroxyphenyl)-N-n-propylpiperidine: Central Pre- and Postsynaptic Dopaminergic Effects and Pharmacoki-netics. J. Med. Chem. 1986, 29, 1889-1895.
- (75) Hacksell, U.; Svensson, U.; Nilsson, J. L. G.; Hjorth, S.; Carlsson, A.; Wikström, H.; Lindberg, P.; Sanchez, D. N-Alkylated-2-Aminotetralins: Central dopamine-receptor stimulating activity. J. Med. Chem. 1979, 22, 1469–1475.
- (76) Svensson, K. Dopamine autoreceptor antagonists. A new class of central stimulants. University of Göteborg, Sweden, 1986, ISBN 91-7900-078-9.
- (77) Shum, A.; Sole, M. J.; van Loon, G. R. Simultaneous measurement of 5-hydroxytryptophan and L-dihydroxyphenylalanine by high performance liquid chromatography with electrochemical detection. Measurement of serotonin and chatecholamine turnover in discrete brain regions. J. Chromatogr. 1982, 228, 123-130.
- (78) Magnusson, O.; Nilsson, L. B.; Westerlund, D. Simultaneous Determination of Dopamine, Dopac, and Homovanillic acid. Direct Incection of Supernatants from Brain Tissue Homogenates in a Liquid Chromatography-Electrochemical Detection System. J. Chromatogr. 1980, 221, 237-247.
- (79) Lagerkvist, S. Sample splitting provides a fast and selective method for determining brain dialysate dopamine and its metabolites. *Monitoring Molecules in Neuroscience*; Krips Repro, Meppel: Groningen, The Netherlands, 1991; pp 136-138.
 (80) Svensson, K.; Ekman, A.; Piercey, M. F.; Hoffmann, W. E.; Lum, C. P. (2000)
- J. T.; Carlsson, A. Effects of the Partial Dopamine Agonists SDZ 208-911, SDZ 208-912 and terguride on Central Monoamine Receptors. Naunyn-Schmiedeberg's Arch. Pharmacol. 1991, 344, 263-274.
- (81) Sonesson, C.; Boije, M.; Svensson, K.; Ekman, A.; Carlsson, A.; Romero, A. G.; Martin, I. J.; Duncan, J. N.; King, L. J.; Wikström, H. Orally Active Central Dopamine and Serotonin Receptor Ligands: 5-, 6-, 7- and 8-[(Trifluoromethyl)sulfonyl]-oxy-2-(di-n-propylamino)tetralins and the Formation of Active
- Metabolites in Vivo. J. Med. Chem. 1993, 36, 3409-3416.
 (82) Chio, C. L.; Hess, G. F.; Graham, R. S.; Huff, R. M. A second molecular form of D2 dopamine receptor in rat and bovine
- (83) Chio, C. L.; Lajiness, M. E.; Huff, R. M. Activation of Heterologously Expressed D3 Dopamine Receptors: Comparision with D2 Dopamine Receptors. Mol. Pharmacol. 1994, 45, 51-60.
 (84) Lahti, R. A.; Evans, D. L.; Figur, L. M.; Carrigan, K. J.; Moon, M. W.; Hsi, R. S. Dopamine D2 receptor binding properties of [³H] U-86170, a dopamine agonist. Eur. J. Pharmacol. 1991, 202, 286-291 289-291.
- (85) Cheng, Y. C.; Prushoff, W. H. Relationship Between the Inhibition Constant and the Concentration of Inhibitor which Causes 50 Percent Inhibition of Enzymatic Reaction. Biochem. Pharmacol. 1973, 22, 3099-3108.