Synthesis and Structure-Activity Relationships of Peptidyl α -Keto Heterocycles as Novel Inhibitors of Prolyl Endopeptidase

Seiji Tsutsumi,*,† Tsuneo Okonogi,† Seiji Shibahara,† Shokichi Ohuchi,† Emiko Hatsushiba,† Arthur A. Patchett,‡ and Burton G. Christensen§

Pharmaceutical Research Laboratory, Meiji Seika Kaisha, Ltd., 760 Morooka-cho, Kohoku-ku, Yokohama 222, Japan, Merck Research Laboratories, P.O. Box 2000, Rahway, New Jersey 07065, and Microcide Pharmaceuticals, Inc., 4040 Campbell Avenue, Menlo Park, California 94025

Received March 11, 19948

The preparation and in vitro prolyl endopeptidase (PEP) inhibitory activity of a series of α -keto heterocyclic compounds is described. The design is based on the introduction of α -keto heterocycles at the C-terminal end of substrate-like peptides. Many of the compounds including those substituted with thiazole, benzothiazole, benzoxazole, imidazole, and pyridine groups exhibit IC50 potencies of PEP inhibition at nanomolar levels. Structure—activity studies of the C-terminal heterocyclic groups indicate the importance of an sp² nitrogen atom at a β -position from the adjoining ketone carbonyl group. This heterocyclic nitrogen atom would provide a critical hydrogen-bond interaction with the histidine residue of the catalytic triad in PEP. Our inhibitors would extend the generality of the α -keto heterocycle design to another serine protease.

Introduction

Prolyl endopeptidase (PEP) (E.C. 3.4.21.26) is classified as a serine protease on the basis of studies with irreversible inhibitors¹ and by molecular cloning.² This enzyme degrades proline-containing neuropeptides, such as vasopressin and substance P.3 Vasopressin has been suggested to be involved in learning and memory.4 Substance P has been reported to exert a protective activity against the neurodegenerative effect of β -amyloid protein.⁵ Recently, progress in Alzheimer's disease studies has revealed that the C-terminal portion of β -amyloid may injure neurons by stimulating the cell's signaling pathway.⁶ Ishiura et al. suggest that PEP may be involved in processing the C-terminal portion of the amyloid precursor protein. On the basis of these reports, PEP inhibitors may improve memory by blocking the metabolism of endogenous neuropeptides8 and have possible potential as cognition-enhancing drugs.9 Potent PEP inhibitors which are peptidyl aldehydes¹⁰ and pyrrolidine derivatives11 have been reported to ameliorate the experimental amnesia induced by scopolamine in rats. 12

Progress in drug design has led to the development of small peptide enzyme inhibitors as therapeutic agents. ^{13,14} In the case of serine proteases, inhibitors have been developed by replacing the scissile amide bond of a protease substrate with a carbonyl group able to form a hemiketal that resembles the tetrahedral intermediate. A number of peptidyl carbonyl groups, such as aldehydes, ¹⁵ fluoro ketones, ¹⁶ keto esters, ¹⁷ and diketones, ¹⁸ have been successfully incorporated into hydrolytic enzyme inhibitors.

Recently, Edwards *et al.* determined by an X-ray crystal structure study¹⁹ that a peptidyl α -keto benzoxazole derivative was an active site-directed elastase inhibitor in which the nitrogen atom of the benzoxazole participated in stabilizing the enzyme—inhibitor com-

plex. PEP, like elastase, is a serine protease on the basis of its inactivation by both diisopropyl fluorophosphorofluoridate and the peptidyl chloromethyl ketone inhibitor Z-Gly-Pro-CH $_2$ Cl. Thus, a histidyl residue may also be located in the active site of PEP to act as a general acid—base catalyst to activate the serine residue.

In analogy with the evidence of Edwards et~al., we believe that the peptidyl α -keto heterocycle derivatives which stabilize the enzyme—inhibitor complex through formation of a hydrogen bond possess an inhibitory potency for PEP. In addition, to have oral potency and penetration into the central nervous system, we introduced the hydrophobic functionality " α -keto heterocycles" at the P_1 of peptidyl PEP inhibitors. We describe herein the design, synthesis, and biological activity of α -keto heterocyclic PEP inhibitors.

Synthesis

The compounds presented in Table 2 contain heterocycles at the P_1' position, L-proline at the P_1 position, and N-substituted L-proline derivatives in the P_2-P_3 area. These inhibitors were conveniently prepared as shown in Schemes 1-5.

Synthetic methods for N-protected α -hydroxy heterocyclic derivatives ${\bf 2a-m}$ are shown in Schemes 1–3. The first convergent approach involved the reaction of the appropriate lithiated heterocycles with N-(tert-butyloxy-carbonyl)prolinal (N-Boc-prolinal) (${\bf 4a}$). Thiazole, thiophene, and benzothiazole derivatives ${\bf 2a-c}$ were prepared via the direct preparation of lithiated heterocycles (method A). The alternative lithiation of 2-bromothiazole with LDA followed by removal of the 2-bromine atom provided the desired 5-substituted thiazole ${\bf 2e}$ (method B). 21 2-Bromopyridine was employed to direct metalation to the 2-position by treatment with n-butyllithium. The transmetalation of 6-membered heterocycles gave the desired pyridine 22 and pyrimidine 23 derivatives ${\bf 2f-h}$ (method C).

In the case of imidazole, the acidic N-H proton on this heterocycle interferes with metalation by *n*-butyllithi-

[†] Meiji Seika Kaisha, Ltd.

^{*} Merck Research Laboratories.

[§] Microcide Pharmaceuticals, Inc.

⁸ Abstract published in Advance ACS Abstracts, September 1, 1994.

Scheme 1^a

^a Reagents and conditions: (a) thiazole, n-BuLi, THF; (b) 2-bromothiazole, diisopropylamine, n-BuLi, THF; (c) 2 equiv of n-BuLi, THF; (d) 2-bromopyridine, n-BuLi, THF; (e) 1-SEM-imidazole 1l, n-BuLi, THF.

Scheme 2. Method E^a

^a Reagents: (a) acetone cyanohydrin, TEA, CH₂Cl₂; (b) (1) acetyl chloride, EtOH, CHCl₃, (2) 2-aminophenol, EtOH; (c) (1) acetyl chloride, EtOH, CHCl₃, (2) 2-amino-3-hydroxypyridine, EtOH; (d) (1) acetyl chloride, EtOH, CHCl₃, (2) 2-aminoethanethiol hydrochloride, Et₃N, EtOH.

um. Therefore, we protected this active hydrogen with the [(trimethylsilyl)ethoxy]methyl (SEM) protective group as a solution to this problem (method D),²⁴ and in this way, the derivative 2i was prepared.

Another useful synthetic method involved the formation of heterocycles from the cyanohydrin 4c which was prepared from N-protected prolinal 4b and an excess of acetone cyanohydrin and triethylamine. Treatment of **4c** with anhydrous HCl in ethanol provided the ethyl imidate hydrochloride which was cyclized with 2-aminophenol to afford the α-hydroxy benzoxazole derivative¹⁹ 2j (method E). In a similar way, the oxazolopyridine derivative 2k and the thiazoline derivative 2l were synthesized. The 4-substituted thiazole derivative 2m was synthesized by using method F outlined in Scheme 3. An α-hydroxy acid derivative, 4e, was prepared by hydrolysis of α -hydroxy ester 4d which was obtained by the reaction of NaBH₄ and α -keto ester.²⁵ The protected α -hydroxy acid **4f** was converted into an acid anhydride with ethyl chloroformate and then treated with diazomethane and hydrogen chloride to give chloromethyl ketone 4g. Cyclization of 4g with

Scheme 3. Method F^a

^a Reagents: (a) 2 N LiOH, THF; (b) (1) TBDMS-Cl, diisopropylethylamine, THF, (2) K₂CO₃, MeOH; (c) (1) ClCO₂Et, Nmethylmorpholine, CH₂Cl₂, (2) CH₂N₂; (3) 4 N HCl-dioxane; (d) HCSNH₂, MeOH.

2m

Scheme 4. Method G^a

^a Reagents: (a) pyrrole, MeMgBr, THF.

thioformamide provided the desired 4-substituted thiazole derivative 2m (method F).

Reaction of ethyl ester 4a with pyrrolylmagnesium bromide provided the corresponding 2-ketopyrrole 16a (method G).²⁶

The N-Boc and -Alloc groups were removed by treatment with trifluoroacetic acid and Pd(PPh₃)₄, respectively. Treatment of proline derivatives with phenylbutanoyl chloride or benzyloxycarbonyl chloride provided the desired N-substituted proline derivatives 5a,b. The N-substituted dipeptide inhibitors were prepared by

Scheme 5. Synthesis of α -Keto Heterocycle Derivatives^a

^a Reagents: (a) (1) TFA or (2) Pd(PPh₃)₄; (b) Z-proline (5a) or N-(4-phenylbutanoyl)proline (5b), WSCD, HOBt, CH₃CN; (c) (COCl)₂, DMSO, TEA, CH₂Cl₂; (d) 3 N HCl-MeOH, 50 °C; (e) Bu₄NF.

standard peptide coupling methods followed by oxidation as illustrated in Scheme 5. Condensation of the N-substituted proline derivatives **5a,b** with the amino alcohols **3a-m** gave the dipeptide derivatives **6aa-ma** as a mixture of diastereomers. The desired α-keto heterocycles **7a-19a** were prepared from the corresponding alcohols via Swern oxidation. In the case of the 4-substituted thiazole derivative **9a**, the O-protected TBDMS group was removed after the coupling reaction followed by oxidation. The SEM-protected intermediate was deprotected after Swern oxidation to give the desired compound **17b**.

Enzyme Inhibitory Activities

The peptidyl α -keto heterocycle derivative **7b** was a more potent inhibitor than the aldehyde **20** or the α -keto ester **21**. The epimeric isomer **7c**(R) and the alcohol analogue **6ab** were almost 1000-fold less potent than the corresponding ketone **7b** (Table 1).

Table 2 shows the inhibitory potencies of the novel peptidyl prolyl endopeptidase (PEP) inhibitors which contain a series of heterocycles at the P_1 position. On the N-terminal acyl group, most of the phenylbutanoyl derivative inhibitors (7b and 12b) were more potent than the corresponding benzyloxycarbonyl derivatives (7a and 12a).

The structure-activity relationships of a series of the substituted heterocycles at P1' illustrate several interesting requirements for PEP inhibition. Among the active analogues, the one-ring type derivatives (7b, 10a, 13a, and 17b) and the two-ring type derivatives (12b, **18a**, and **19a**) have a similar potency (IC₅₀ = 3.8-9.0nM). On the other hand, the thiophene derivatives (11a,b) had lost potency (IC₅₀ = 2020 and 1260 nM, respectively). These results suggest that peptidyl heterocycles which possess a nitrogen atom are important for inhibitory activity. We hypothesized that PEP inhibitor potency depended on the position of the nitrogen atom relative to the adjoining ketone. To examine this hypothesis, we synthesized regioisomers of the thiazole and pyridine derivatives with different nitrogen atom positions. The inhibitory potency of the thiazole derivatives followed the trend: 2-thiazole 7b

Table 1. Comparison of Electrophilic Carbonyl-Based PEP Inhibitors^a

(IC₅₀ = 5.0 nM) \geq 4-thiazole **9a** (IC₅₀ = 6.2 nM) \gg 5-thiazole **8a** (IC₅₀ = 1090 nM). Comparison of 6-membered rings showed the following trend: 2-pyridine **13a** (IC₅₀ = 6.9 nM) \geq 5-pyrimidine **15a** (IC₅₀ = 61 nM) \gg 3-pyridine **14a** (IC₅₀ = 2290 nM). Of nearly equal importance to the choice of heterocycles was the pyrrole derivative (**16a**). The 2-pyrrole ketone **16a** (IC₅₀ = 21.3 μM) with a nitrogen atom at a β-position had remarkably decreased inhibitory potency. This result suggests

 $[^]a$ See the Experimental Section for description of the method for determining the IC_{50} values of these inhibitors. b The values are the means \pm SEM for the three independent experiments. c Literature 10a $IC_{50}=8.7$ nM.

Table 2. In Vitro PEP Inhibitory Activities of Peptidyl α-Keto Heterocycles^a

			0	MEI		
no.	HET	х	Y	z	Meth	nod IC ₅₀ (nM) ^b
7a	-	0	н	н	A	8.5 (±0.8)
7b	$\overset{N}{\longrightarrow}$	CH₂CH₂	Н	н	Α	5.0 (±0.3)
8a	-⟨s ^N	CH₂CH₂	н	н	В	1090 (±100)
9a	~\s\	CH₂CH₂	н	н	F	6.2 (±0.3) ·
10a	$\overset{n}{\longrightarrow}$	CH₂CH₂	н	н	E	3.8 (±0.7)
11a	-	0	н	н	A	2020 (±100)
11b	-	CH ₂ CH ₂	н	н	A	1260 (±120)
12a	\prec_s	0	Н	н	A	5.0 (±0.8)
12b	-	CH₂CH₂	н	н	A	4.0 (±1.2)
13a		CH₂CH₂	Н	н	С	6.9 (±1.0)
14a		CH₂CH₂	н	н	С	2290 (±320)
15a	N	CH ₂ CH ₂	н	н	С	61 (±13)
16a	→NH NH	CH₂CH₂	н	н	G	21300 (±1200)
17b		CH₂CH₂	н	н	D	9.0 (±2.4)
18a	$ ^{\circ}$ \bigcirc	CH₂CH₂	н	н	Ε	5.6 (±1.0)
19a	-	CH ₂ CH ₂	Н	н	E	5.5 (±1.0)
20	н	CH₂CH₂	н	н		8.7 (±0.8)
21	CO₂Et	CH₂CH₂	н	н		10.3 (±0.8)

^a See the Experimental Section for description of the method for determining the IC_{50} values of these inhibitors. ^b The values are the means \pm SEM for the three independent experiments.

that the β -attachment was a necessary, but not sufficient, condition for inhibitory potency.

Discussion

Our starting point for these studies was the peptidyl α-keto heterocycle inhibitor 7b and 9a originally reported to be a potent PEP inhibitor.²⁷ This study showed that good PEP inhibitory activity requires a nitrogen atom at a β -position to the adjoining ketone moiety. This concept of inhibition has been extended to other heterocycles in this study. Among a series of 6-membered rings, inhibitory potency was found in the compound 13a. Furthermore, PEP inhibitors are found in a wide variety of heterocyclic classes, one-ring types (10a and 13a) and two-ring types (12a, 18a, and 19a). Although Edwards et al. reported that an α-keto heterocycle derivative would not be an effective inhibitor of serine protease inhibitor, except for α-keto benzoxazole and α-keto oxazoline derivatives, 19 our result showed that variations of the heterocycles were tolerated. The second significant structure-activity relationship for PEP inhibitory activity was discovered in the series containing imidazole substituents (16a and 17b). The pyrrole derivative 16a with a nitrogen atom at a β -position did not have inhibitory potency. These results suggest that high potency requires an sp² nitrogen atom which is able to accept a hydrogen bond at a β -position from the ketone moiety.

Peptidyl trifluoromethyl ketones, α-keto esters, 17 and α-diketones¹⁸ of serine protease inhibitors were reported to be hydrated in DMSO with water from ¹³C-NMR studies. These inhibitors can serve as transition analogues and react with a nucleophilic residue (serine hydroxy) to form a hemiketal that resembles the tetrahedral intermediate. We could not detect a new peak around 100 ppm which was consistent with a hydrated ketone (gem diol) in 10% $D_2O/DMSO-d_6$ (v/v) by the ¹³C-NMR spectrum for 7b and 12b. This ¹³C-NMR study suggests that participation in hydrogen-bond interactions by the heterocycles at the P₁' position was more more important for inhibitory activity of PEP than its hydrating capability. Recently, Patel et al. also suggested that the hydrophobic and/or hydrogen-bond interactions of the adjacent moiety contributed to the inhibitory potency of α -keto ester- and α -diketone-based renin inhibitors.²⁸

In the course of this study, we found that the thiazole derivative 7a was 300 times more potent than the thiophene derivative 11a. The observation that the transition-state analogue 7a binds to the PEP 3 orders of magnitude tighter than 11a demonstrates the vital role the hydrogen bond plays in transition-state stabilization. The role of hydrogen bonding has been examined in several enzyme inhibitors.^{29,30} Bartlett et al. determined that removal of hydrogen-bond capability in an inhibitor of the zinc endopeptidase thermolysin caused the loss of inhibitory potency, reflecting the loss of 4.0 kcal mol⁻¹ in binding energy.²⁹ Applying the Gibbs-Helmholtz equation for the binding of our PEP inhibitor,³¹ we obtained a difference in the free energy of binding between 7a and 11a and between 7b and 11b of 4.60 and 4.63 kcal mol⁻¹, respectively. We believe that the general agreement of these values with those of Bartlett et al. is in accord with hydrogen-bond formation to the heterocyclic nitrogen atom and enthalpy changes are likely to be the major contributor to the observed energy differences.

Structure—activity studies of the C-terminal heterocyclic groups indicate the importance of an sp^2 nitrogen atom at a β -position from the adjoining ketone carbonyl group. In analogy with the study of Edwards $et\ al.$, this heterocyclic nitrogen atom would provide a critical hydrogen-bond interaction with the histidine residue of the catalytic triad in PEP. These design features found in our inhibitors may well be capable of extension to other serine protease or cysteine protease inhibitors.

Experimental Section

The abbreviations which were used are as follows: HOBt, 1-hydroxybenzotriazole; WSCD, 1-ethyl-3-[3'-(dimethylamino)-propyl]carbodiimide; TFA, trifluoroacetic acid; TEA, triethylamine; THF, tetrahydrofuran; Boc, tert-butyloxycarbonyl; Alloc, allyloxycarbonyl; Z-proline, N-(benzyloxycarbonyl)proline; SEM-Cl, [2-(trimethylsilyl)ethoxy]methyl chloride; TB-DMS-Cl, tert-butyldimethylsilyl chloride.

Chemistry. General Methods. Unless otherwise noted, materials were obtained from commercial suppliers and used without further purification. ¹H-NMR and ¹³C-NMR spectras were recorded at ambient temperature on a JEOL JNM-GX 400 spectrometer or a Varian Gemini-300 spectrometer. All chemical shifts are reported in δ units (ppm) relative to tetramethylsilane (assigned to 0.0 ppm). Mass spectra and high-resolution mass spectra were determined on an HITACHI mass spectrometer M-80A or M-80B. Analytical results for compounds were determined by Toray Research Center, Inc., and the Meiji College of Pharamaceutical Science. Melting points were determined with a Yanaco micromelting point apparatus. Optical rotations were obtained on a Perkin-Elmer 241 polarimeter. Thin-layer chromatography (TLC) analyses were performed on silica gel plates (0.25 mm; Merck) and visualized with UV, I2, or a ninhydrin spray reagent. Flash chromatography was performed with silica gel (200-300 mesh; Merck). Purities of final products were checked by reversephase HPLC (YMC HPLC, C-18 column, 150×60 mm, 5μ m) at flow rates of 1 mL/min, eluting with CH₃CN and water (60: 40 or 50:50), and detected by UV at 254 nm. Retention time, purity, and HPLC conditions are included in each individual experimental procedure.

Method A. Synthesis of 1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(thiazol-2-yl)methanol (2a). A solution of n-BuLi (3.3 mL of a 1.6 M solution in hexane, 5.5 mmol) was added dropwise to a solution of thiazole (470 mg, 5.5 mmol) in dry THF (10 mL) at -65 °C over 5 min under Ar. After the mixture was stirred for 30 min, a solution of N-Bocprolinal (4a)²⁰ (1.0 g, 5.0 mmol) in dry THF (10 mL) was added and the mixture was stirred at -65 °C for 4 h. The reaction mixture was poured into water (5 mL), and the product was extracted with EtOAc. The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure. Purification of the crude material by flash chromatography (1:1 hexane/EtOAc) gave 2a as a yellow solid (1.2 g, 83%): MS (EI) m/z 285 (M + 1); ¹H-NMR (CDCl₃, 300 MHz) δ 1.25–1.36 (m, 9 H), 1.70-1.90 (m, 1 H), 2.05-2.35 (m, 2 H), 2.76-2.83(m, 1 H), 3.20-3.60 (m, 2 H), 4.38-4.42 (m, 1 H), 5.12-5.20 (m, 1 H), 6.70 (d, 1 H), 7.31 (d, 1 H, J = 3.3 Hz), 7.72 (d, 1 H, J = 3.3 Hz)J = 3.3 Hz).

1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(thiophene-2-yl)methanol (2b). The title compound, 2b, was prepared from 2-thienyllithium (5.5 mL of a 1.0 M solution in THF, 5.5 mmol) and 4a (1.0 g, 5.0 mmol) using method A. The crude product was purified by chromatography (1:1 hexane/EtOAc) to afford 2b (1.07 g, 76%) as a white solid: MS (SIMS) m/z 284 (M + 1); ¹H-NMR (CDCl₃, 400 MHz) δ 1.25–1.30 (m, 9 H), 1.70–2.05 (m, 4 H), 3.25–3.55 (m, 2 H), 4.05–4.30 (m, 1 H), 4.75–5.10 (m, 1 H), 5.85–6.05 (m, 1 H), 6.85–7.27 (m, 3 H).

1-(Benzothiazol-2-yl)-1-[1-(tert-butyloxycarbonyl)-2(S)-pyrrolidinyl]methanol (2c). The title compound, 2c, was prepared from n-BuLi (33 mL of a 1.66 M solution in THF, 48 mmol), benzothiazole (6.8 g, 48 mmol), and 4a (9.2 g, 46 mmol)

using method A. The crude product was purified by chromatography (1:1 hexane/EtOAc) to afford $\bf 2c$ (10.3 g, 67%) as a yellow solid: MS (EI) $\it m/z$ 335 (M + 1); $^1\rm H\textsc{-}NMR$ (CDCl $_3$, 300 MHz) δ 1.47–1.52 (m, 9 H), 1.70–2.86 (m, 4 H), 3.26–3.47 (m, 2 H), 4.24–4.48 (m, 1 H), 4.96–5.14 (m, 1 H), 6.64–6.90 (m, 1 H), 7.35–7.39 (m, 1 H), 7.44–7.52 (m, 1 H), 7.89–7.91 (m, 1 H), 7.97–7.99 (m, 1 H).

Method B. Synthesis of 1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(thiazol-5-yl)methanol (2e). A solution of lithium diisopropylamide (LDA) was prepared as follows: to dijsopropylamine (770 µL, 5.5 mmol) in anhydrous THF (10 mL) at -60 °C was added 3.5 mL of n-BuLi (1.56 M solution in hexane, 5.5 mmol) under Ar. After stirring at 10 °C for 10 min, the reaction mixture was cooled at -70 °C. To the LDA solution was added 2-bromothiazole (900 mg, 5.5 mmol), and the mixture was stirred for 20 min. The resulting mixture was then treated with N-Boc-prolinal (4a) (1.0 g, 5.0 mmol) and stirred at -60 °C for 2 h. The reaction mixture was poured into aqueous saturated NH₄Cl solution at 0 °C. The product was extracted with EtOAc. The combined organic extracts were dried over MgSO4 and concentrated under reduced pressure. Purification of the crude material by flash chromatography (40:1 CH₂Cl₂/EtOAc) gave 2d (840 mg, 46%) as a brown oil: ${}^{1}\text{H-NMR}$ (CDCl₃, 400 MHz) δ 1.50–1.52 (m, 9 H), 1.78-2.14 (m, 3 H), 2.95-3.04 (m, 1 H), 3.27-3.49 (m, 2 H), 4.02-4.32 (m, 1 H), 4.82-5.00 (m, 1 H), 7.42 (s, 1 H). To the 2-bromothiazole derivative 2d (820 mg, 2.3 mmol) in THF (10 mL) was added 3.2 mL of n-BuLi (1.56 M solution in hexane, 5.0 mmol) at -70 °C, and the reaction mixture was stirred for 15 min under Ar. The reaction mixture was poured into aqueous saturated NH4Cl solution at 0 °C, and the product was extracted with EtOAc. The combined organic extracts were dried over MgSO4 and concentrated under reduced pressure. Purification of the crude material by flash chromatography (10:1 CH₂Cl₂/EtOAc) gave 2e (430 mg, 66%) as a brown oil: MS (EI) m/z 285 (M + 1); ¹H-NMR (CDCl₃, 400 MHz) δ 1.50–1.52 (m, 9 H), 1.77–2.13 (m, 3 H), 2.95–3.00 (m, 1 H), 3.30-3.42 (m, 2 H), 4.36-4.38 (m, 1 H), 4.94-5.09 (m, 1 H), 7.77-7.78 (m, 1 H), 8.84 (s, 1 H).

Method C. Synthesis of 1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(pyridin-2-yl)methanol (2f). A solution of n-BuLi (3.5 mL of a 1.56 M solution in hexane, 5.5 mmol) was added to a solution of 2-bromopyridine (870 mg, 5.5 mmol) in dry THF (10 mL) at -65 °C over 5 min under Ar. After the solution was stirred for 60 min, a solution of N-Boc prolinal (4a) (1.0 g, 5 mmol) in dry THF (5 mL) was added and the resulting mixture was stirred at -65 °C for 2 h. The mixture was poured into water (5 mL), and then the product was extracted with CH₂Cl₂. The combined organic extracts were dried over MgSO4 and concentrated under reduced pressure. Purification of the crude material by flash chromatography (7:1 CH₂Cl₂/EtOAc) gave 2f (970 mg, 70%) as a yellow oil: MS (FD) m/z 278 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.48-1.51 (m, 9 H), 1.70-2.05 (m, 4 H), 3.14-3.52 (m, 2 H), 4.00-4.20 (m, 1 H), 4.80-5.20 (m, 1 H), 7.22-7.51 (m, 2 H), 7.64–7.72 (m, 1 H), 8.75–8.77 (m, 1 H).

1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(pyridin-3-yl)methanol (2g). The title compound, 2g, was prepared from n-BuLi (1.3 mL of a 1.56 M solution in THF, 2.0 mmol), 3-bromopyridine (316 mg, 2.0 mmol), and 4a (400 mg, 2.0 mmol) using method C. The crude product was purified by chromatography (1:1 hexane/EtOAc) to afford 2g (123 mg, 22%) as a yellow oil: MS (EI) m/z 279 (M + 1); 1 H-NMR (CDCl₃, 400 MHz) δ 1.51–1.52 (m, 9 H), 1.68–2.85 (m, 4 H), 3.28–3.55 (m, 2 H), 4.05–4.37 (m, 1 H), 4.59–4.88 (m, 1 H), 7.30–7.36 (m, 1 H), 7.68–7.81 (m, 1 H), 8.51–8.54 (m, 2 H)

1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(pyrimidin-5-yl)methanol (2h). The title compound, 2h, was prepared from n-BuLi (1.3 mL of a 1.56 M solution in THF, 2.0 mmol), 5-bromopyrimidine (318 mg, 2.0 mmol), and 4a (400 mg, 2.0 mmol) using method C. The crude product was purified by chromatography (1:1 hexane/EtOAc) to afford 2h (153 mg, 28%) as a brown oil: MS (SIMS) m/z 280 (M + 1); 1 H-NMR (CDCl₃, 400 MHz) δ 1.51–1.52 (m, 9 H), 1.64–2.10

(m, 3 H), 2.78-2.90 (m, 1 H), 3.36-3.53 (m, 2 H), 4.10-4.38 (m, 1 H), 4.64-4.86 (m, 1 H), 8.69-8.76 (m, 2 H), 9.16-9.17 (m, 1 H).

Method D. Synthesis of 1-[1-(tert-Butyloxycarbonyl)-2(S)-pyrrolidinyl]-1-[1-[[2-(trimethylsilyl)ethoxy]methyl]imidazol-2-yl]methanol (2i). A solution of n-BuLi (6.56 mL of a 1.6 M solution in hexane, 10.5 mmol) was added to a solution of N-SEM-imidazole²⁴ (1.9 g, 9.6 mmol) in dry THF (10 mL) at -40 °C over 5 min under Ar. After the mixture was stirred for 60 min, a solution of N-Boc-prolinal (4a) (1.9)g, 9.6 mmol) in dry THF (10 mL) was added and the resulting mixture was stirred at -40 °C for 1 h and at 0 °C for 3 h. The mixture was poured into aqueous saturated NH₄Cl solution. The product was extracted with CH2Cl2. The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure. Purification of the crude material by flash chromatography (3:1 hexane/EtOAc) gave 2i (3.80 g, 99%) as a yellow oil: \dot{MS} (EI) m/z 397 (M); 1H -NMR (CDCl₃, 400 MHz) δ 0.00 (s, 9 H), 0.92 (t, 2 H), 1.48-1.51 (m, 9 H), 1.75-2.10 (m, 4 H), 3.35-3.57 (m, 4 H), 4.14-4.38 (m, 1 H), 5.29-5.49 (m, 3 H), 6.99-7.06 (m, 2 H).

Method E. Synthesis of 1-[1-(Allyloxycarbonyl)-2(S)pyrrolidinyl]-1-(benzoxazol-2-yl)methanol (2j). A solution of N-Alloc-prolinal (4b) (5.5 g, 30 mmol) in CH₂Cl₂ (70 mL) was treated with acetone cyanohydrin (7.2 g, 85 mmol) followed by TEA (2.5 mL, 18 mmol) and stirred for 24 h at room temperature under Ar. The reaction mixture was concentrated under reduced pressure, and the residue was taken up in Et₂O. The Et₂O solution was washed with brine, dried over MgSO₄, and evaporated. The crude product was purified by chromatography (50:1 CH₂Cl₂/EtOAc) to afford cyanohydrin 4c (6.2 g, 97%) as a colorless oil: MS (SIMS) m/z211 (M + 1); ¹H-NMR (CDCl₃, 400 MHz) δ 1.80-2.28 (m, 4 H), 3.52-3.56 (m, 1 H), 3.68-3.74 (m, 1 H), 4.14-4.18 (m, 1 H), 4.54-4.70 (m, 3 H), 5.26-5.37 (m, 2 H), 5.91-6.01 (m, 1 H). Acetyl chloride (4.3 mL, 60 mmol) was added to a solution of CHCl₃ (4 mL) and EtOH (3 mL) at 0 °C over 15 min under Ar. Cyanohydrin 4c (420 mg, 2 mmol) in CHCl₃ (4 mL) was added; the reaction mixture was stirred at 0 °C and at room temperature overnight. After evaporation, the crude imidate in EtOH (10 mL) was treated with 2-aminophenol (240 mg, 2.2 mmol) and heated at reflux for 6 h. The reaction mixture was concentrated under reduced pressure, and then the residue was taken up in EtOAc. The organic extract was washed with water and aqueous saturated NaHCO₃ solution, dried over MgSO4, and evaporated. The crude product was purified by chromatography (25:1 CH₂Cl₂/EtOAc) to afford 2j (560 mg, 93%) as a yellow oil: MS (SIMS) m/z 303 (M + 1); ¹H-NMR (CDCl₃, 400 MHz) δ 1.80–1.88 (m, 4 H), 3.49–3.61 (m, 2 H), 4.20-4.93 (m, 4 H), 5.22-5.37 (m, 2 H), 5.94-5.98 (m, 1 H), 7.34-7.39 (m, 2 H), 7.53-7.57 (m, 1 H), 7.71-7.74 (m, 1 H)

1-[1-(Allyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(oxazolo-[4,5-b]pyridin-2-yl)methanol (2k). The title compound, 2k, was prepared from 4c (406 mg, 2.0 mmol) and 2-amino-3-hydroxypyridine (240 mg, 2.2 mmol) using method E. The crude product was purified by chromatography (10:1 CH₂Cl₂/EtOAc) to afford 2k (90 mg, 15%) as a yellow oil: MS (SIMS) m/z 304 (M + 1); ¹H-NMR (CDCl₃, 400 MHz) δ 1.77-2.24 (m, 4 H), 3.14-3.55 (m, 2 H), 4.45-4.47 (m, 1 H), 4.61-4.64 (m, 2 H), 5.20-5.30 (m, 3 H), 5.81-5.90 (m, 1 H), 5.91-5.97 (m, 1 H), 7.31 (dd, 1 H, J = 4.7, 8.1 Hz), 7.84 (d, 1 H, J = 8.1 Hz), 8.57 (d, 1 H, J = 4.7 Hz).

1-[1-(Allyloxycarbonyl)-2(S)-pyrrolidinyl]-1-(thiazolin-2-yl)methanol (21). The title compound, 21, was prepared from 4c (406 mg, 2.0 mmol), TEA (610 μ L, 4.4 mmol), and aminoethanethiol hydrochloride (250 mg, 2.2 mmol) using method E. The crude product was purified by chromatography (1:2 toluene/EtOAc) to afford 21 (276 mg, 51%) as a yellow oil. MS (FD) m/z 270 (M); ¹H-NMR (CDCl₃, 400 MHz) δ 1.72–2.10 (m, 4 H), 3.28–3.65 (m, 4 H), 4.15–4.34 (m, 3 H), 4.60–4.72 (m, 2 H), 4.85–4.95 (m, 1 H), 5.20–5.40 (m, 2 H), 5.90–6.04 (m, 1 H).

Method F. Synthesis of 1-[1-(Allyloxycarbonyl)-2(S)-pyrrolidinyl]-1-[(tert-butyldimethylsilyl)oxy]-1-(thiazol-4-yl)methane (2m). Ethyl 2-[1-(allyloxycarbonyl)-2(S)-pyr-

rolidinyl]-2-oxoacetate²⁵ (6.2 g, 25.8 mmol) in EtOH (80 mL) was reduced with NaBH₄ (380 mg, 10 mmol) at 0 °C for 30 min. After evaporation, the reaction mixture was diluted with EtOAc, washed with water and brine, and dried over MgSO₄. The organic layer was concentrated and the residue purified on silica gel to give the α -hydroxy ester derivative **4d** (4.7 g, 75.0%). A solution of the α -hydroxy ester 4d (1.0 g, 3.89 mmol) in THF (10 mL) was treated with 2 N LiOH (2.05 mL) at room temperature for 2 h. The reaction mixture was then acidified with 2 N HCl and extracted with EtOAc to afford 4e (715 mg, 80%) which was used without further purification. A solution of 4e (710 mg, 3.1 mmol) in dry DMF (20 mL) was treated with diisopropylethylamine (1.7 mL, 9.9 mmol) and TBDMS-Cl (1.49 g, 9.9 mmol). The resulting mixture was stirred at room temperature for 15 h. The reaction mixture was poured into brine and extracted with EtOAc. The combined organic extracts were concentrated under reduced pressure. After the residue was dissolved in MeOH (55 mL) and THF (20 mL), 0.5 M K₂CO₃ solution (20 mL) was added and the solution was stirred at room temperature for 1 h. The reaction mixture was concentrated and neutralized to pH 4 by 1 N HCl. The aqueous layer was extracted with EtOAc. The combined organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by chromatography (EtOAc) to afford 4f (870 mg, 82%) as a yellow oil: ${}^{1}\text{H-NMR}$ (CDCl₃, 300 MHz) δ 0.00-0.32 (m, 6 H), 0.94 (s, 9 H), 1.78-2.12 (m, 4 H), 3.36-3.55 (m, 2 H), 4.06-4.14 (m, 1 H), 4.24-4.79 (m, 3 H), 5.20-5.38 (m, 2 H), 5.79-6.02 (m, 1 H). A solution of 4f~(870~mg,~2.65~mmol) in $CH_2Cl_2~(15~mL)$ was treated with ClCO₂Et ($304 \mu L$, 3.2 mmol) and N-methylmorpholine (378 μ L, 3.4 mmol) at -10 °C for 30 min under Ar. To the reaction mixture was added diazomethane solution in Et₂O (10 mL, 5.0 mmol) at -10 °C. The reaction mixture was stirred at -10 °C for 30 min and treated with 1.3 mL of 4 N HCl/dioxane solution, and the mixture was stirred at 0 °C for 30 min. Aqueous NaHCO3 solution was added, and the separated aqueous layer was extracted with CH₂Cl₂. The combined organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product was purified by chromatography (10:1 toluene/EtOAc) to afford 4g (530 mg, 54%, two steps) as a yellow oil: ¹H-NMR (CDCl₃, 300 MHz) δ 0.02-0.15 (m, 6 H), 0.90 (s, 9 H), 1.78-2.10 (m, 4 H), 3.35-3.60 (m, 2 H), 4.08-4.70 (m, 6 H), 5.20-5.36 (m, 2 H), 5.58-6.02 (m, 1 H). A solution of 4g (400 mg, 1.06 mmol) in MeOH (4 mL) was treated with thioformamide (130 mg, 2.2 mmol) and stirred at 45 °C for 18 h. The reaction mixture was diluted with EtOAc and washed with water. The organic layer was concentrated under reduced pressure. The crude product was purified by chromatography (10:1 toluene/EtOAc) to afford 2m (160 mg, 39%) as a yellow oil: $^{1}\text{H-NMR}$ (CDCl₃, 300 MHz) δ 0.00-0.20 (m, 6 H), 0.96 (s, 9 H), 1.56-2.12 (m, 4 H), 3.00-3.11 (m, 1 H), 3.28-3.46 (m, 1 H), 4.17-4.26 (m, 1 H), 4.58-4.72 (m, 2 H), 5.21-5.69 (m, 3 H), 5.88-6.04 (m, 1 H), 7.30 (d, 1.72 (m, 2 H))1 H), 8.85 (d, 1 H).

Method G. Synthesis of 2-[[1-[[1-(4-Phenylbutanoyl)- $2(S) \hbox{-pyrrolidinyl}] \hbox{-} arbonyl] \hbox{-} 2(S) \hbox{-pyrrolidinyl}] \hbox{-} carbonyl] \hbox{-}$ pyrrole (16a). A solution of methylmagnesium bromide (1.78 mL of a 3 M solution in Et₂O, 5.34 mmol) in toluene (7 mL) was treated with pyrrole (557 mL, 8.0 mmol) at -40 °C, and the resulting mixture was then stirred at -10 °C for 10 min under Ar. A solution of this pyrrolylmagnesium bromide (4.5 mL) was added to a solution of the proline ethyl ester derivative 4h (250 mg, 0.67 mmol) in toluene (1 mL) at -65 °C over 5 min. The mixture was stirred at -65 °C for 3 h and then at 0 °C for 3 h and at room temperature for 15 h. The reaction was quenched with aqueous saturated NH₄Cl. The product was extracted with EtOAc and dried over MgSO4, and the solvent was removed under reduced pressure. Purification of the crude product by column chromatography (1:1 hexane/ EtOAc) gave 16a (150 mg, 55%) as a white foam: $[\alpha]_D - 95.5^{\circ}$ (c 1.1, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.89-2.20 (m, 9 H), 2.22–2.34 (m, 3 H), 2.64–2.68 (m, 2 H), 3.37–3.43 (m, 1 H), 3.54-3.60 (m, 1 H), 3.65-3.71 (m, 1 H), 3.93-3.99 (m, 1 H), 4.71–4.74 (m, 1 H), 5.27–5.31 (m, 1 H), 6.25–6.27 (m, 1 H), 6.98-7.01 (m, 2 H), 7.14-7.28 (m, 5 H), 9.50 (brs, 1 H); HPLC retention time 4.35 min (96.6%), eluting with 60% CH₃-

CN in water, 1.0 mL/min; MS (EI) m/z 407 (M); HRMS for $C_{24}H_{29}N_3O_3$ calcd 407.2207, found 407.2180. Anal. ($C_{24}H_{29}N_3O_3$) C. H. N.

Method H. General Deprotection Procedure. (1) The N-Boc-proline derivatives $2\mathbf{a}-\mathbf{i}$ (0.2-0.4 mM) were dissolved in 90% trifluoroacetic acid in CH_2Cl_2 and stirred at 0 °C for 1 h. After removal of solvent, the pH of the residue was adjusted to 10 with 1 N NaOH and the mixture was extracted with CH_2 - Cl_2 . The organic layers were dried over MgSO₄ to give the desired amines $3\mathbf{a}-\mathbf{i}$ which were used without further purification.

(2) To a solution of the N-Alloc-proline derivatives 2j-m (0.2–0.4 mM) in 50% EtOAc in CH_2Cl_2 was added dimedone (2 equiv) or pyrrolidine (5 equiv) at room temperature followed by tetrakis(triphenylphosphine)palladium (0.05 equiv). The mixture was stirred at room temperature for 2–4 h and then evaporated in vacuo. The crude product was purified by chromatography ($CH_2Cl_2/MeOH$) to afford the desired amines 3j-m as a brown oil.

Method I. General Procedure for Peptide-Bond Formation. To a solution containing an N-protected proline derivative, **5a** or **5b** (1 equiv, 0.02–0.05 mM), in CH₃CN were added a pyrrolidine methanol derivative (0.9–1.1 equiv), HOBt (1.0–1.2 equiv), and WSCD (1.0–1.2 equiv). In the case of a pyrrolidine derivative as an HCl salt, TEA was further added to the reaction mixture. The mixture was stirred overnight at 25 °C. After evaporation in vacuo, the residue was dissolved in EtOAc, washed with brine, dried over MgSO₄, and then concentrated. The product was purified by flash chromatography to give the desired carbinols **6aa**–**mb**.

Method J. General Procedure for the Preparation of α-Keto Heterocycles. DMSO (3.0-4.0 equiv) was added to a solution of $(COCl)_2$ (1.5-2.0 equiv) in dry CH_2Cl_2 at -60 °C, and the mixture was stirred at this temperature for 10 min under Ar. To the solution were added dropwise pyrrolidine methanol derivatives 6aa-mb (0.1-0.2 mmol) in dry CH_2Cl_2 , and the reaction mixture was stirred at -60 °C for 30 min. TEA (6.0-8.0 equiv) was added, and the mixture was allowed to warm to 0 °C and stirred for 1-6 h. The mixture was diluted with EtOAc, washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The product was purified by flash chromatography to afford the desired α-keto heterocycles 7a-19a.

1-[1-[[1-(Benzyloxycarbonyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiazol-2-yl)methanol (6aa). Compound 3a was prepared by method H (1) starting from 2a. Reaction of Z-proline (5a) (750 mg, 3.0 mmol) with the alcohol 3a (560 mg, 3.0 mmol), HOBt (410 mg, 3.0 mmol), and WSCD (470 mg, 3.0 mmol) using method I gave 6aa as an oil (1.0 g, 80%) which was purified by column chromatography (1.0 g, 80%) which was purified by column chromatography (3.0 MHz) δ 1.75-3.05 (m, 8 H), 3.38-3.80 (m, 4 H), 4.30-4.70 (m, 2 H), 4.90-5.25 (m, 3 H), 7.10-7.40 (m, 6 H), 7.66-7.75 (m, 1 H).

2-[[1-[[1-(Benzyloxycarbonyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]thiazole (7a). Reaction of the alcohol 6aa (1.0 g, 2.40 mmol) with (COCl)₂ (320 μ L, 3.65 mmol), DMSO (490 μ L, 6.9 mmol), and TEA (2.1 mL, 15.2 mmol) using method J gave 7a as a colorless foam (850 mg, 86%) which was purified by column chromatography (2:1 CH₂Cl₂/EtOAc): $[\alpha]_D$ -144.3° (c 1.0, CHCl₃); two rotamers [maj/min (3:2)]; ¹H-NMR (CDCl₃, 400 MHz) δ 1.86-2.46 (m, 8 H), 3.46-3.93 (m, 4 H), 4.48 (dd, J = 3.9, 8.5 Hz, min), 4.61(dd, J = 3.9, 8.5 Hz, maj), 5.00-5.19 (m, 2H), 5.61 (dd, J = 3.9)4.6, 9.0 Hz, min, 5.78 (dd, J = 4.6, 9.0 Hz, maj), 7.28-7.39(m, 5 H), 7.66 (d, J = 3.1 Hz, min), 7.68 (d, J = 3.1 Hz, maj)8.00 (d, J = 3.1 Hz, maj), 8.02 (d, J = 3.1 Hz, min); HPLCretention time 5.65 min (93.5%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 413 (M); HRMS for $C_{21}H_{23}N_3O_4S_1$ calcd 413.1408, found 413.1450. $(C_{21}H_{23}N_3O_4S_1\cdot 0.2H_2O)$ C, H, N.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiazol-2-yl)methanol (6ab). Reaction of N-(4-phenylbutanoyl)proline (5b) (50 mg, 0.2 mmol) with the alcohol 3a (36 mg, 0.2 mmol), HOBt (30 mg, 0.22 mmol), and WSCD (35 mg, 0.23 mmol) using method I gave

6ab (64 mg, 75%) purified by column chromatography (15:1 CH₂Cl₂/MeOH): MS (EI) m/z 427 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.75–2.05 (m, 9 H), 2.10–2.50 (m, 3 H), 2.60–2.70 (m, 2 H), 3.40–3.70 (m, 4 H), 4.20–4.62 (m, 2 H), 4.90–5.20 (m, 1 H), 7.15–7.35 (m, 6 H), 7.66–7.75 (m, 1 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]thiazole (7b). Reaction of the alcohol 6ab (51 mg, 0.12 mmol) with $(COCl)_2$ (16 μ L, 0.18 mmol), DMSO (24 μ L, 0.34 mmol), and TEA (160 μ L, 1.15 mmol) using method J gave 7b as a colorless oil (39 mg, 76%) which was purified by column chromatography (30:1 $CH_2Cl_2/MeOH$): [α]_D -121.9° (c 0.8, $CHCl_3$); ¹H-NMR ($CDCl_3$, 400 MHz) δ 1.90–2.21 (m, 9 H), 2.25–2.40 (m, 2 H), 2.45– 2.55 (m, 1 H), 2.65-2.75 (m, 2 H), 3.40-3.50 (m, 1 H), 3.55-3.65 (m, 1 H), 3.72-3.80 (m, 1 H), 4.00-4.05 (m, 1 H), 4.73 (dd, 1 H, J = 3.6, 8.0 Hz), 5.75 (dd, 1 H, J = 4.7, 9.0 Hz), 7.15 -7.30 (m, 5 H), 7.65 (d, 1 H, J = 3.1 Hz), 8.00 (d, 1 H, J = 3.1 Hz)Hz); ¹³C-NMR (CDCl₃, 400 MHz) δ 24.6, 25.2, 26.0, 28.6, 29.0, 33.6, 35.3, 47.2, 47.3, 57.6, 61.8, 125.9, 126.3, 128.4, 128.6, 141.9, 144.9, 165.1, 170.6, 171.6, 191.1; HPLC retention time 5.47 min (93.9%), eluting with 60% CH₃CN in water, 1.0 mL/ min; MS (EI) m/z 425 (M); HRMS for C23H27N3O3S1 calcd 425.1772, found 425.1790.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(R)-pyrrolidinyl]-1-(thiazol-2-yl)methanol (6ac). Reaction of N-(4-phenylbutanoyl)proline (5b) (75 mg, 3.0 mmol) with the alcohol 3a(R) (56 mg, 3.0 mmol), HOBt (41 mg, 3.0 mmol), and WSCD (47 mg, 3.0 mmol) using method I gave 6ac as an oil (105 mg, 82%) which was purified by column chromatography (20:1 CH₂Cl₂/MeOH): MS (EI) m/z 427 (M); H-NMR (CDCl₃, 300 MHz) δ 1.85–2.21 (m, 8 H), 2.28–2.38 (m, 4 H), 2.63–2.70 (m, 2 H), 3.46–3.77 (m, 4 H), 4.50–4.63 (m, 2 H), 5.34 (d, 1 H), 5.82–5.85 (m, 1 H), 7.15–7.30 (m, 6 H), 7.73 (m, 1 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(R)-pyrrolidinyl]carbonyl]thiazole (7c(R)). Reaction of the alcohol 6ac (80 mg, 0.19 mmol) with (COCl)₂ (22 μ L, 0.25 mmol), DMSO (34 μ L, 0.48 mmol), and TEA (170 μ L, 1.2 mmol) using method J gave 7c(R) as a white foam (65 mg, 81%) which was purified by column chromatography (30:1 CH₂Cl₂/MeOH): [α]_D + 35.0° (c 1.2, CHCl₃); ¹H-NMR (CDCl₃, 300 MHz) δ 1.80–2.09 (m, 8 H), 2.14–2.38 (m, 4 H), 2.62–2.79 (m, 2 H), 3.33–3.42 (m, 1 H), 3.45–3.77 (m, 3 H), 4.20–4.44 (m, 1 H), 5.64–6.20 (m, 1 H), 7.14–7.28 (m, 5 H), 7.62–7.76 (m, 1 H), 7.97–8.07 (m, 1 H); HPLC retention time 5.22 min (94.7%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 425 (M); HRMS for C₂₃H₂₇N₃O₃S₁ calcd 425.1772, found 425.1735. Anal. (C₂₃H₂₇N₃O₃S₁) C, H, N.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiazol-5-yl)methanol (6e). Compound 3e was prepared by method H (1) starting from 2e. Reaction of N-(4-phenylbutanoyl)proline (5b) (52 mg, 0.2 mmol) with the alcohol 3e (40 mg, 0.22 mmol), HOBt (30 mg, 0.22 mmol), and WSCD (35 mg, 0.23 mmol) using method I gave 6e as a yellow oil (63 mg, 74%) which was purified by column chromatography (50:1 CH₂Cl₂/MeOH): ¹H-NMR (CDCl₃, 300 MHz) δ 1.84–2.08 (m, 8 H), 2.23–2.41 (m, 4 H), 2.64–2.71 (m, 2 H), 3.09–3.13 (m, 1 H), 3.42–3.67 (m, 3 H), 4.60–4.71 (m, 2 H), 5.11–5.13 (m, 1 H), 7.14–7.30 (m, 5 H), 7.75–7.77 (m, 1 H), 8.76–8.80 (m, 1 H).

5-[[1-([1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]- 2(S)-pyrrolidinyl]carbonyl]thiazole (**8a**). Reaction of the alcohol **6e** (64 mg, 0.15 mmol) with (COCl)₂ (26 μ L, 0.3 mmol), DMSO (41 μ L, 0.6 mmol), and TEA (125 μ L, 0.9 mmol) using method J gave **8a** as a yellow solid (48 mg, 75%) which was purified by column chromatography (EtOAc): [α]_D -85.0° (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.91-2.06 (m, 4 H), 2.09-2.24 (m, 4 H), 2.27-2.34 (m, 3 H), 2.62-2.68 (m, 3 H), 3.40-3.42 (m, 1 H), 3.54-3.56 (m, 1 H), 3.68-3.73 (m, 1 H), 3.94-4.00 (m, 1 H), 4.70 (dd, 1 H, J = 3.6, 8.0 Hz), 5.33 (dd, 1 H, J = 4.6, 8.7 Hz), 7.15-7.27 (m, 5 H), 8.54 (s, 1 H), 9.00 (s, 1 H); ¹³C-NMR (CDCl₃, 400 MHz) δ 24.6, 25.1, 25.8, 28.4, 29.0, 33.4, 35.1, 46.9, 47.1, 57.4, 62.6, 125.7, 128.2, 128.4, 128.6, 137.4, 141.7, 147.4, 159.0, 170.7, 171.5, 191.0; HPLC retention time 4.34 min (97.0%), eluting with 60% CH₃CN in water, 1.0

mL/min; MS (EI) m/z 425 (M); HRMS for $C_{23}H_{27}N_3O_3S_1$ calcd 425.1772, found 425.1717. Anal. ($C_{23}H_{27}N_3O_3S_1\cdot 1.0H_2O$) C, H N

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiazolin-2-yl)methanol (61). Compound 31 was prepared by method H (2) starting from 21. Reaction of N-(4-phenylbutanoyl)proline (5b) (136 mg, 0.5 mmol) with the alcohol 31 (88 mg, 0.47 mmol), HOBt (70 mg, 0.52 mmol), TEA (70 μ L, 0.5 mmol), and WSCD-HCl (119 mg, 0.62 mmol) using method I gave 61 as a yellow oil (120 mg, 60%) which was purified by column chromatography (1:1 acetone/EtOAc); ¹H-NMR (CDCl₃, 300 MHz) δ 1.85–2.34 (m, 12 H), 2.65–2.72 (m, 2 H), 3.20–3.30 (m, 2 H), 3.38–3.65 (m, 3 H), 3.90–3.96 (m, 1 H), 4.15–4.30 (m, 2 H), 4.45–4.80 (m, 3 H), 7.17–7.32 (m, 5 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]thiazoline (10a). Reaction of the alcohol 61 (120 mg, 0.28 mmol) with $(COCl)_2$ (36 μ L. 0.41 mmol), DMSO (59 μ L, 0.83 mmol), and TEA (266 μ L, 1.92 mmol) using method J gave 10a as a colorless oil (105 mg, 88%) which was purified by column chromatography (1:2 acetone/EtOAc): $[\alpha]_D -90.4^{\circ}$ (c 1.1, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.89-2.18 (m, 10 H), 2.21-2.33 (m, 2 H), 2.64-2.68 (m, 2 H), 3.27-3.32 (m, 2 H), 3.36-3.42 (m, 1 H), 3.55-3.66 (m, 2 H), 3.88-3.94 (m, 1 H), 4.43-4.59 (m, 2 H), 4.69 (dd, 1 H, J = 3.6, 8.0 Hz), 5.47 (dd, 1 H, J = 5.0, 8.8 Hz), 7.15-7.28 (m, 5 H); 13 C-NMR (CDCl₃, 400 MHz) δ 24.6, 25.0, 25.9, 28.4, 28.5, 32.4, 33.5, 35.2, 46.9, 47.2, 57.4, 61.6, 66.3, 125.8, 128.3, 128.5, 141.8, 168.7, 170.4, 171.5, 192.9; HPLC retention time 5.70 min (90.1%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 427 (M); HRMS for $C_{23}H_{29}N_3O_3S_1$ calcd 427.1928, found 427.1936. Anal. ($C_{23}H_{29}N_3O_3S_1\cdot 0.3H_2O$) C, H, N.

1-[1-[[1-(Benzyloxycarbonyl)-2(S)-pyrrolidinyl]carbonyl-2(S)-pyrrolidinyl]-1-(thiophene-2-yl)methanol (6ba). Compound 3b was prepared by method H (1) starting from 2b. Reaction of Z-proline (5a) (620 mg, 2.5 mmol) with the alcohol 3b (460 mg, 2.5 mmol), HOBt (365 mg, 2.7 mmol), and WSCD (420 mg, 2.7 mmol) using method I gave 6ba as a colorless oil (1.0 g, 97%) which was purified by column chromatography (4:1 CH₂Cl₂/EtOAc): MS (EI) m/z 414 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.74–2.28 (m, 8 H), 3.30–3.90 (m, 4 H), 4.20–4.60 (m, 2 H), 4.70–5.23 (m, 3 H), 6.92–7.00 (m, 2 H), 7.24–7.30 (m, 6 H).

2-[[1-[[1-(Benzyloxycarbonyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]thiophene (11a). Reaction of the alcohol **6b**a (1.0 g, 2.4 mmol) with (COCl)₂ (420 μ L, 4.8 mmol), DMSO (660 μ L, 9.3 mmol), and TEA (2.1 mL, 15.2 mmol) using method J gave 11a as a white solid (640 mg, 64%) which was purified by column chromatography (10:1 CH₂Cl₂/EtOAc): [α]_D -112.4° (c 1.0, CHCl₃); two rotamers [maj/min (3:2)]; ¹H-NMR (CDCl₃, 400 MHz) δ 1.90-2.30 (m, 8 H), 3.47-3.89 (m, 4 H), 4.48 (dd, J = 3.6, 8.3 Hz, min), 4.59 (dd, J = 3.6, 8.3 Hz, min), 4.99-5.19 (m, 2 H), 5.44 (dd, 1 H, J = 4.2, 8.9 Hz), 7.12-7.14 (m, 1 H), 7.14-7.30 (m, 5 H), 7.63-7.65 (m, 1 H), 7.77-7.82 (m, 1 H); HPLC retention time 5.97 min (98.5%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 412 (M); HRMS for C₂₂H₂₄N₂O₄S₁ calcd 412.1455, found 412.1502. Anal. (C₂₂H₂₄N₂O₄S₁) C, H, N.

1-[1-[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiophene-2-yl)methanol (6bb). Reaction of N-(4-phenylbutanoyl)proline (5b) (260 mg, 1.0 mmol) with the alcohol 3b (183 mg, 1.0 mmol), HOBt (150 mg, 1.1 mmol), and WSCD (180 mg, 1.2 mmol) using method I gave 6bb as a yellow oil (310 mg, 73%) which was purified by column chromatography (1:1 CH₂Cl₂/EtOAc): MS (EI) m/z 426 (M); 1 H-NMR (CDCl₃, 400 MHz) δ 1.90–2.08 (m, 9 H), 2.10–2.30 (m, 3 H), 2.65–2.75 (m, 2 H), 3.40–4.00 (m, 4 H), 4.45–5.24 (m, 3 H), 6.90–7.05 (m, 2 H), 7.21–7.30 (m, 6 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl-2(S)-pyrrolidinyl]carbonyl]thiophene (11b). Reaction of the alcohol **6bb** (300 mg, 0.72 mmol) with (COCl)₂ (126 μ L, 1.44 mmol), DMSO (196 μ L, 2.76 mmol), and TEA (650 μ L, 4.7 mmol) using method J gave 11b as a colorless oil (190 mg, 62%) which was purified by column chromatography (40:1 CH₂Cl₂/MeOH): [α]_D -84.7° (c 0.9, CHCl₃); ¹H-NMR (CDCl₃,

400 MHz) δ 1.90–2.13 (m, 9 H), 2.20–2.32 (m, 3 H), 2.64–2.68 (m, 2 H), 3.39–3.56 (m, 2 H), 3.67–3.73 (m, 1 H), 3.91–3.97 (m, 1 H), 4.72 (dd, 1 H, J = 3.6, 7.8 Hz), 5.41 (dd, 1 H, J = 4.2, 8.8 Hz), 7.12 (dd, 1 H, J = 3.9, 5.0 Hz), 7.23–7.27 (m, 5 H), 7.63 (dd, 1 H, J = 1.1, 5.0 Hz), 7.80 (dd, 1 H, J = 1.1, 3.9 Hz); $^{13}\text{C-NMR}$ (CDCl₃, 400 MHz) δ 24.6, 25.0, 25.9, 28.4, 29.3, 33.5, 35.2, 46.9, 47.2, 57.5, 61.8, 125.8, 128.2, 128.5, 132.4, 133.9, 141.7, 141.8, 170.6, 171.5, 191.1; HPLC retention time 6.05 min (94.7%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 424 (M); HRMS for $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_3\text{S}_1$ calcd 424.1819, found 424.1850. Anal. $(\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_3\text{S}_1)$ C, H, N.

1-(Benzothiazol-2-yl)-1-[1-[[1-(benzyloxycarbonyl)-2-(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]methanol (6ca). Compound 3c was prepared by method H (1) starting from 2c. Reaction of Z-proline (5a) (2.24 g, 9.0 mmol) with the alcohol 3c (1.90 g, 8.1 mmol), HOBt (1.21 g, 9.0 mmol), and WSCD (1.39 g, 9.0 mmol) using method I gave 6ca as a white foam (3.36 g, 90%) which was purified by column chromatography (1:2 hexane/EtOAc): MS (EI) m/z 465 (M); H-NMR (CDCl₃, 300 MHz) δ 1.83-2.10 (m, 5 H), 2.20-2.60 (m, 2 H), 2.90-3.12 (m, 1 H), 3.37-3.91 (m, 4 H), 4.27-4.72 (m, 2 H), 4.91-5.21 (m, 3 H), 7.27-7.38 (m, 7 H), 7.87-7.98 (m, 2 H).

2-[[1-[[1-(Benzyloxycarbonyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]benzothiazole (12a). Reaction of the alcohol **6ca** (3.87 g, 8.3 mmol) with (COCl)₂ (1.57 mL, 18 mmol), DMSO (2.45 mL, 35 mmol), and TEA (6.9 mL, 50 mmol) using method J gave **12a** as colorless crystals (2.42 g, 63%) which were purified by column chromatography (1:1 hexane/EtOAc): mp 142–144 °C; $[\alpha]_D$ –122.0° (c 1.0, CHCl₃); two rotamers $[maj/min\ (3:2)]$; ¹H-NMR (CDCl₃, 400 MHz) δ 1.83–2.48 (m, 8 H), 3.43–3.98 (m, 4 H), 4.48–4.63 (m, 1 H), 5.01–5.18 (m, 2 H), 5.68 (dd, J = 4.7, 8.9 Hz, min), 5.89 (dd, J = 4.7, 8.9 Hz, maj), 7.27–7.34 (m, 5 H), 7.49–7.59 (m, 2 H), 7.96–7.98 (m, 1 H), 8.16–8.20 (m, 1 H); HPLC retention time 10.7 min (99.6%), eluting with 50% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 463 (M); HRMS for $C_{25}H_{25}N_3O_4S_1$ calcd 463.1564, found 463.1746. Anal. ($C_{25}H_{25}N_3O_4S_1$) C, H, N.

1-(Benzothiazol-2-yl)-1-[1-[[1-(4-phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]methanol (6cb). Reaction of N-(4-phenylbutanoyl)proline (5b) (1.3 g, 4.0 mmol) with the alcohol 3c (940 mg, 4.0 mmol), HOBt (600 mg, 4.4 mmol), and WSCD (685 mg, 4.4 mmol) using method I gave 6cb as a yellow foam (1.7 g, 89%) which was purified by column chromatography (4:1 CH₂Cl₂/EtOAc): MS (EI) m/z 477 (M); ¹H-NMR (CDCl₃, 400 MHz) δ 1.80–2.20 (m, 8 H), 2.25–2.38 (m, 3 H), 2.51–2.56 (m, 1 H), 2.65–2.69 (m, 2 H), 3.06–3.86 (m, 4 H), 4.52–4.73 (m, 2 H), 5.18–5.21 (m, 1 H), 7.15–7.26 (m, 5 H), 7.34–7.47 (m, 2 H), 7.87–7.91 (m, 1 H), 7.94–7.98 (m, 1 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]benzothiazole (12b). Reaction of the alcohol **6cb** (200 mg, 0.42 mmol) with (COCl)₂ (88 μ L, 1.0 mmol), DMSO (140 μ L, 2.0 mmol), and TEA (0.82 mL, 5.9 mmol) using method J gave 12b as a yellow solid (125 mg, 63%) which was purified by column chromatography (2:1 hexane/EtOAc): $[\alpha]_D - 122.0^\circ$ (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.84-2.26 (m, 9 H), 2.15-2.25 (m, 2 H), 2.37-2.43 (m, 1 H), 2.56-2.60 (m, 2 H), 3.31-3.33 (m, 1 H), 3.45-3.49 (m, 1 H), 3.64-3.70 (m, 1 H), 3.94-3.98 (m, 1 H), 4.67-4.69 (m, 1 H), 5.80 (dd, 1 H, J = 5.0, 8.9 Hz), 7.10-7.24 (m, 5)H), 7.43-7.49 (m, 2 H), 7.87-7.91 (m, 1 H), 8.08-8.12 (m, 1 H); ${}^{13}\text{C-NMR}$ (CDCl₃, 400 MHz) δ 24.6, 25.3, 25.9, 28.4, 29.0, 33.5, 35.2, 47.1, 57.5, 61.7, 122.3, 125.6, 125.8, 126.9, 128.3, 128.5, 137.2, 141.8, 153.6, 164.3, 170.5, 171.4, 192.7; HPLC retention time 10.5 min (93.4%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 475 (M); HRMS for $C_{27}H_{29}N_3O_3S_1$ calcd 475.1928, found 475.1960. $(C_{27}H_{29}N_3O_3S_1)$ C, H, N.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(pyridin-2-yl)methanol (6f). Compound 3f was prepared by method H (1) starting from 2f. Reaction of N-(4-phenylbutanoyl)proline (5b) (144 mg, 0.55 mmol) with the alcohol 3f (88 mg, 0.53 mmol), HOBt (81 mg, 0.6 mmol), and WSCD (93 mg, 0.6 mmol) using method I gave 6f as a yellow oil (165 mg, 74%) which was purified by column

chromatography (100:1 CH₂Cl₂/MeOH): MS (EI) m/z 421 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.83–2.17 (m, 10 H), 2.24–2.36 (m, 2 H), 2.65–2.72 (m, 2 H), 3.34–3.47 (m, 2 H), 3.59–3.63 (m, 1 H), 3.81–3.85 (m, 1 H), 4.40–4.66 (m, 2 H), 4.78–5.22 (m, 1 H), 7.16–7.31 (m, 6 H), 7.48–7.50 (m, 1 H), 7.71–7.74 (m, 1 H), 8.49–8.53 (m, 1 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]pyridine (13a). Reaction of the alcohol 6f (160 mg, 0.38 mmol) with (COCl)₂ (67 μ L, 0.76 mmol), DMSO (105 μ L, 1.5 mmol), and TEA (346 μ L, 2.5 mmol) using method J gave 13a as a white oil (90 mg, 57%) which was purified by column chromatography (1:2 CH₂Cl₂/EtOAc): $[\alpha]_D$ = 93.9° (c 1.3, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.93= 2.21 (m, 9 H), 2.23-2.37 (m, 2 H), 2.40-2.48 (m, 1 H), 2.63-2.67 (m, 2 H), 3.37 - 3.43 (m, 1 H), 3.53 - 3.57 (m, 1 H), 3.68 -3.74 (m, 1 H), 3.97-4.02 (m, 1 H), 4.75 (dd, 1 H, J = 3.3, 7.4 (m, 1 H)Hz), 5.98 (dd, 1 H, J = 4.9, 9.0 Hz), 7.14-7.29 (m, 5 H), 7.43(ddd, 1 H, J = 0.8, 4.9, 7.7 Hz), 7.78 (ddd, 1 H, J = 1.56, 7.7,7.7 Hz), 8.02 (d, 1 H, J = 7.7 Hz), 8.65 (d, 1 H, J = 4.9 Hz); ¹³C-NMR (CDCl₃, 400 MHz) δ 24.6, 25.1, 25.9, 28.5, 29.0, 33.5, 35.2, 47.2, 57.5, 60.9, 122.5, 125.7, 127.1, 128.2, 128.5, 136.8, 141.8, 148.8, 152.1, 170.3, 171.4, 198.6; HPLC retention time 5.42~min~(93.3%), eluting with $60\%~CH_3CN$ in water, 1.0~mL/min; MS (EI) m/z 419 (M); HRMS for $C_{25}H_{29}N_3O_3$ calcd 419.2207, found 419.2234. Anal. (C₂₅H₂₉N₃O₃·0.9H₂O) C, H,

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(pyridin-3-yl)methanol (6g). Compound 3g was prepared by method H (1) starting from 2g (123 mg, 0.44 mmol). To a solution of the resulting TFA salt were added TEA (150 μ L, 1.1 mmol), N-(4-phenylbutanoyl)proline (5b) (130 mg, 0.5 mmol), HOBt (68 mg, 0.5 mmol), and WSCD (75 mg, 0.5 mmol). The reaction using method I gave 6g as a yellow oil (156 mg, 84%) which was purified by column chromatography (10:1 CH₂Cl₂/MeOH): MS (EI) m/z 421 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.76–2.10 (m, 10 H), 2.17–2.39 (m, 2 H), 2.63–2.70 (m, 2 H), 3.00–3.82 (m, 4 H), 4.46–4.68 (m, 2 H), 4.79–5.21 (m, 1 H), 7.15–7.46 (m, 6 H), 7.75–7.91 (m, 1 H), 8.50–8.67 (m, 2 H).

3-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]pyridine (14a). Reaction of the alcohol **6g** (42 mg, 0.1 mmol) with $(COCl)_2$ (17.5 μ L, 0.2 mmol), DMSO (27 μ L, 0.4 mmol), and TEA (83 μ L, 0.6 mmol) using method J gave 14a as a colorless oil (24.3 mg, 58%) which was purified by column chromatography (25:1 CH₂Cl₂/MeOH): [α]_D -91.3° (c 1.2, CHCl₃); 1 H-NMR (CDCl₃, 400 MHz) δ 1.87-2.20 (m, 9 H), 2.25-2.35 (m, 3 H), 2.65-2.68 (m, 2 H), 3.41-3.55 (m, 1 H), 3.54-3.59 (m, 1 H), 3.70-3.76 (m, 1 H), 3.96-4.02 (m, 1 H), 4.70 (dd, 1 H, J = 3.6, 7.7 Hz), 5.49 (dd, 1 H, J = 4.9,9.0 Hz), 7.14-7.29 (m, 5 H), 7.46-7.48 (m, 1 H), 8.34 (m, 1 H), 8.78 (m, 1 H), 9.19 (brs, 1 H); ¹³C-NMR (CDCl₃, 400 MHz) δ 24.7, 25.1, 25.9, 28.4, 28.5, 33.5, 35.1, 46.9, 47.2, 57.5, 61.0, 123.7, 125.8, 128.2, 128.5, 130.8, 136.0, 141.7, 149.5, 153.4, 170.6, 171.5, 197.4; HPLC retention time 4.25 min (95.1%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 419 (M); HRMS for $C_{25}H_{29}N_3O_3$ calcd 419.2207, found 419.2226. Anal. $(C_{25}H_{29}N_3O_3\cdot 0.3H_2O)$ C, H, N.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(pyrimidin-5-yl)methanol (6h). Compound 3h was prepared by method H (1) starting from 2h (153 mg, 0.55 mmol). To a solution of the resulting TFA salt were added TEA (150 μ L, 1.1 mmol), N-(4-phenylbutanoyl)proline (5b) (157 mg, 0.6 mmol), HOBt (81 mg, 0.6 mmol), and WSCD (93 mg, 0.6 mmol). The reaction using method I gave 6h as a yellow oil (196 mg, 84%) which was purified by column chromatography (10:1 CH₂Cl₂/MeOH): MS (EI) m/z 422 (M); H-NMR (CDCl₃, 300 MHz) δ 1.81–2.00 (m, 7 H), 2.10–2.37 (m, 4 H), 2.62–2.70 (m, 3 H), 2.94–3.00 (m, 1 H), 3.43–3.50 (m, 1 H), 3.60–3.67 (m, 1 H), 3.80–3.86 (m, 1 H), 4.56–4.66 (m, 2 H), 4.90–4.94 (m, 1 H), 7.16–7.85 (m, 5 H), 8.65–8.84 (m, 2 H), 9.13–9.16 (m, 1 H).

5-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]pyrimidine (15a). Reaction of the alcohol 6h (42 mg, 0.1 mmol) with (COCl)₂ (17.5 μ L, 0.2 mmol), DMSO (27 μ L, 0.4 mmol), and TEA (83 μ L, 0.6 mmol) using method J gave 15a as a white solid (27.4 mg, 65%) which

was purified by column chromatography (25:1 CH₂Cl₂/MeOH): $[\alpha]_D$ –76.3° (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.87–2.21 (m, 9 H), 2.25–2.37 (m, 3 H), 2.64–2.68 (m, 2 H), 3.39–3.44 (m, 1 H), 3.53–3.59 (m, 1 H), 3.70–3.76 (m, 1 H), 3.98–4.04 (m, 1 H), 4.68–4.70 (m, 1 H), 5.43 (dd, 1H, J = 4.9, 8.7 Hz), 7.15–7.28 (m, 5 H), 9.26 (s, 2 H), 9.35 (s, 1 H); ¹³C-NMR (CDCl₃, 400 MHz) δ 24.7, 25.3, 25.9, 28.4, 28.5, 33.5, 35.2, 47.0, 47.2, 57.5, 61.1, 125.8, 128.3, 128.5, 141.7, 156.8, 161.4, 170.8, 171.6, 196.4; MS (EI) m/z 420 (M); HPLC retention time 3.98 min (96.2%), eluting with 60% CH₃CN in water, 1.0 mL/min; HRMS for C₂₄H₂₈N₄O₃ calcd 420.2160, found 420.2263. Anal. (C₂₄H₂₈N₄O₃) C, H, N.

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-[1-[[2-(trimethylsilyl)ethoxy]methyl])imidazol-2-yl]methanol (6i). Compound 3i was prepared by method H (1) starting from 2i. Reaction of N-(4-phenylbutanoyl)proline (5b) (780 mg, 3.0 mmol) with the alcohol 3i (900 mg, 3.0 mmol), HOBt (480 mg, 3.5 mmol), and WSCD (543 mg, 3.5 mmol) using method I gave 6i as a brown oil (960 mg, 59%) which was purified by column chromatography (30:1 CH₂Cl₂/MeOH): MS (EI) m/z 540 (M); ¹H-NMR (CDCl₃, 400 MHz) δ -0.03 (s, 9 H), 0.86-0.95 (m, 2 H), 1.71-2.06 (m, 10 H), 2.20-2.67 (m, 2 H), 2.65-2.69 (m, 2 H), 3.37-3.45 (m, 1 H), 3.53-3.68 (m, 4 H), 3.86-3.91 (m, 1 H), 4.47-4.66 (m, 2 H), 5.00-5.18 (m, 1 H), 5.54-5.89 (m, 2 H), 7.17-7.29 (m, 6 H), 7.81-7.86 (m, 1 H).

1-[[2-(Trimethylsilyl)ethoxy]methyl]-2-[[1-[[1-(4-phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]imidazole (17a). Reaction of the alcohol 6i (1.0 g, 1.85 mmol) with (COCl)₂ (324 μ L, 3.7 mmol), DMSO (505 μ L, 7.1 mmol), and TEA (1.66 mL, 12.0 mmol) using method J gave 17a as a yellow foam (720 mg, 72%) which was purified by column chromatography (50:1 CH₂Cl₂/MeOH): MS (EI) m/z 538 (M); ¹H-NMR (CDCl₃, 400 MHz) δ -0.03 (s, 9 H), 0.91 (t, 2 H, J = 8.2 Hz), 1.93-2.13 (m, 9 H), 2.33-2.30 (m, 2 H), 2.38-2.45 (m, 1 H), 2.63-2.67 (m, 2 H), 3.36-3.40 (m, 1 H), 3.50-3.58 (m, 3 H), 3.65-3.71 (m, 1 H), 3.95-3.99 (m, 1 H), 4.74-4.76 (m, 1 H), 5.61 (d, 1 H, J = 10.5 Hz), 5.64 (dd, 1 H, J = 4.9, 9.0 Hz), 5.76 (d, 1 H, J = 10.5 Hz), 7.14-7.28 (m, 7 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]imidazole (17b). The SEMprotected derivative 17a (720 mg, 1.34 mmol) was dissolved in 3 N HCl (30 mL) in EtOH (30 mL) and stirred at 50 °C until the reaction was judged complete by TLC analysis. The pH of the solution was adjusted to 8 with NaHCO₃, and the product was extracted with CH2Cl2. The combined organic extracts were dried over MgSO₄ and then concentrated. The product was purified by silica gel chromtography (10:1 CH₂-Cl₂/MeOH) to give the title compound, 17b, as a white foam (413 mg, 75%): $[\alpha]_D = 117.0^\circ$ (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.92-2.10 (m, 8 H), 2.14-2.33 (m, 3 H), 2.42-2.45 (m, 1 H), 2.66-2.70 (m, 2 H), 3.39-3.45 (m, 1 H), 3.56-3.61 (m, 1 H), 3.70-3.74 (m, 1 H), 3.98-4.02 (m, 1 H), 4.86 (d, 1 H, J = 3.9 Hz), 5.64 (d, 1 H, J = 4.7 Hz), 7.07 (s, 1 H), 7.12(s, 1 H), 7.15-7.32 (m, 5 H); $^{13}\text{C-NMR}$ (CDCl₃, 400 MHz) δ 24.5, 25.1, 25.9, 28.4, 29.1, 33.5, 35.1, 47.2, 57.6, 61.4, 120.6, 125.7, 128.2, 128.5, 131.0, 141.8, 143.3, 170.8, 171.5, 188.2; HPLC retention time 3.52 min (96.1%), eluting with 60% CH₃-CN in water, 1.0 mL/min; MS (EI) m/z 408 (M); HRMS for calcd 408.2160, found 408.2175. $C_{23}H_{28}N_4O_3$ (C₂₃H₂₈N₄O₃•0.1H₂O) C, H, N.

1-(Benzoxazol-2-yl)-1-[1-[[1-(4-phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]methanol (6j). Compound 3j was prepared by method H (2) starting from 2j. Reaction of N-(4-phenylbutanoyl)proline (5b) (80 mg, 0.3 mmol) with the alcohol 3j (65 mg, 0.3 mmol), HOBt (45 mg, 0.33 mmol), and WSCD (52 mg, 0.33 mmol) using method I gave 6j as a yellow oil (130 mg, 94%) which was purified by column chromatography (70:1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$): MS (EI) m/z 461 (M); ¹H-NMR (CDCl₃, 300 MHz) δ 1.88-2.09 (m, 7 H), 2.11-2.36 (m, 4 H), 2.65-2.70 (m, 3 H), 3.41-3.99 (m, 4 H), 4.66-5.30 (m, 3 H), 7.15-7.29 (m, 5 H), 7.34-7.47 (m, 2 H), 7.53-7.58 (m, 1 H), 7.70-7.76 (m, 1 H).

2-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]benzoxazole (18a). Reaction

of the alcohol 6j (130 mg, 0.28 mmol) with (COCl)₂ (49 μ L, 0.56 mmol), DMSO (77 μ L, 1.1 mmol), and TEA (250 μ L, 1.8 mmol) using method J gave 18a as a colorless oil (63.3 mg, 49%) which was purified by column chromatography (50:1 $CH_2Cl_2/MeOH$): [α]_D -86.7° (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.83–2.17 (m, 9 H), 2.21–2.34 (m, 2 H), 2.42– 2.49 (m, 1 H), 2.63-2.67 (m, 2 H), 3.35-3.40 (m, 1 H), 3.51-3.56 (m, 1 H), 3.75-3.79 (m, 1 H), 4.02-4.06 (m, 1 H), 4.72-4.75 (m, 1 H), 5.73 (dd, 1 H, J = 5.4, 8.5 Hz), 7.14-7.31 (m, 5)H), 7.42-7.46 (m, 1 H), 7.49-7.54 (m, 1 H), 7.62-7.64 (m, 1 H), 7.86-7.88 (m, 1 H); ${}^{13}\text{C-NMR}$ (CDCl₃, 400 MHz) δ 24.5, 25.4, 25.9, 28.4, 28.8, 33.5, 35.2, 47.1, 57.4, 62.1, 111.8, 122.4, 125.8, 128.2, 128.5, 140.5, 141.8, 150.7, 156.0, 170.6, 171.5, 188.1; HPLC retention time 7.71 min (95.6%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 459 (M); HRMS for C27H29N3O4 calcd 459.2156, found 459.2102. Anal. $(C_{27}H_{29}N_3O_4{\cdot}0.3H_2O)~C,~H,~N.$

1-(Oxazolo[4,5-b]pyridin-2-yl)-1-[1-[[1-(4-phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]methanol (6k). Compound 3k was prepared by method H (2) starting from 2k. Reaction of N-(4-phenylbutanoyl)proline (5b) (78 mg, 0.3 mmol) with the alcohol 3k (55 mg, 0.25 mmol), HOBt (48 mg, 0.35 mmol), and WSCD (55 mg, 0.35 mmol) using method I gave 6k as a yellow oil (100 mg, 87%) which was purified by column chromatography (50:1 CH2Cl2/ MeOH): MS (EI) m/z 462 (M); ¹H-NMR (CDCl₃, 400 MHz) δ 1.83-2.07 (m, 6 H), 2.17-2.36 (m, 5 H), 2.64-2.69 (m, 3 H), 3.12-3.60 (m, 3 H), 3.84-3.90 (m, 1 H), 4.59-4.71 (m, 2 H), 4.87-5.15 (m, 1 H), 7.18-7.33 (m, 6 H), 7.80-7.84 (m, 1 H), 8.55-8.58 (m, 1 H).

2-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]carbonyl]oxazolo[4,5-b]pyridine (19a). Reaction of the alcohol **6k** (100 mg, 0.22 mmol) with (COCl)₂ $(38.5 \,\mu\text{L}, 0.44 \,\text{mmol}), \, DMSO \, (60 \,\mu\text{L}, \, 0.85 \,\text{mmol}), \, \text{and TEA} \, (194 \,\mu\text{L}, \, 0.85 \,\mu\text{L})$ μ L, 1.4 mmol) using method J gave 19a as a white foam (34 mg, 34%) which was purified by column chromatography (30:1 $CH_2Cl_2/MeOH$); [α]_D -92.5° (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.93-2.32 (m, 11 H), 2.46-2.67 (m, 3 H), 3.35-3.40 (m, 1 H), 3.51-3.56 (m, 1 H), 3.75-3.79 (m, 1 H), 4.02-4.06 (m, 1 H), 4.72 (dd, 1 H, J = 3.0, 8.4 Hz), 5.61 (dd, 1 H, J= 5.9, 8.2 Hz), 7.14-7.31 (m, 5 H), 7.48 (dd, 1 H, J = 4.9, 8.5Hz), $7.98 \, (dd, 1 \, H, J = 1.5, 8.5 \, Hz), 8.74 \, (dd, 1 \, H, J = 1.5, 4.9)$ Hz); 13 C-NMR (CDCl₃, 400 MHz) δ 24.5, 25.6, 25.9, 28.2, 28.9, 33.5, 35.1, 47.1, 57.4, 62.9, 120.2, 123.1, 125.8, 128.2, 128.5, 141.8, 143.4, 148.7, 154.0, 157.4, 170.8, 171.5, 188.0; HPLC retention time 4.92 min (94.7%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 460 (M); HRMS for $C_{26}H_{28}N_4O_4$ calcd 406.2109, found 460.2119.

1-[(tert-Butyldimethylsilyl)oxy]-1-[1-[[1-(4-phenylbu $tanoyl) \hbox{-} 2(S) \hbox{-} pyrrolidinyl] \hbox{-} 2(S) \hbox{-} pyrrolidinyl] \hbox{-} 1 \hbox{-}$ (thiazol-4-yl)methane (6ma). Compound 3m was prepared by method H (2) starting from 2m. Reaction of N-(4-phenylbutanoyl)proline (5b) (86 mg, 0.33 mmol) with the alcohol 3m (90 mg, 0.3 mmol), HOBt (49 mg, 0.36 mmol), and WSCD·HCl (68 mg, 0.36 mmol) using method I gave 6ma as a yellow oil (150 mg, 92%) which was purified by column chromatography (1:1 toluene/EtOAc); ¹H-NMR (CDCl₃, 300 MHz) δ 0.00 (s, 3 H), 0.18 (s, 3 H), 0.92 (m, 9 H), 1.88-2.10 (m, 8 H), 2.23-2.38 (m, 4 H), 2.65-2.73 (m, 2 H), 2.95-3.00 (m, 1 H), 3.38-3.45 (m, 1 H), 3.60-3.67 (m, 2 H), 4.42-4.48 (m, 1 H), 4.62 (dd, 1 H), 5.64 (d, 1 H), 7.14–7.30 (m, 5 H), 7.20 (s, 1 H), 8.68 (s, 1 H).

1-[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-1-(thiazol-4-yl)methanol (6mb). To a solution of 6ma (150 mg, 0.28 mmol) in THF (2 mL) was added tetrabutylammonium fluoride (0.41 mL of a 1 M solution in THF, 0.41 mmol) at room temperature. After 1 h, the solution was evaporated and the residue was partitioned between EtOAc and water. The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by chromatography (1:1 acetone/EtOAc) to give 6mb as a yellow oil (93 mg, 79%); ¹H-NMR (CDCl₃, 300 MHz) δ 1.85–2.36 (m, 12 H), 2.65–2.72 (m, 2 H), 3.46–4.00 (m, 4 H), 4.38–5.25 (m, 3 H), 5.82–6.05 (m, 1 H), 7.14–7.30 (m, 5 H), 7.40 (m, 1 H), 8.72–8.80 (m, 1 H).

4-[[1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-

2(S)-pyrrolidinyl]carbonyl]thiazole (9a). Reaction of the alcohol 6mb (93 mg, 0.22 mmol) with $(COCl)_2$ (28 μ L, 0.32 mmol), DMSO (46 μ L, 0.65 mmol), and TEA (206 μ L, 1.49 mmol) using method J gave 9a as a colorless foam (50 mg, 54%) which was purified by column chromatography (1:2 acetone/EtOAc): $[\alpha]_D$ -99.8° (c 1.0, CHCl₃); ¹H-NMR (CDCl₃, 400 MHz) δ 1.90-2.24 (m, 9 H), 2.21-2.44 (m, 3 H), 2.64-2.68 (m, 2 H), 3.37-3.43 (m, 1 H), 3.54-3.59 (m, 1 H), 3.70-3.75 (m, 1 H), 3.89-4.01 (m, 1 H), 4.75 (dd, 1 H, J = 3.5, 7.8)Hz), 5.71 (dd, 1 H, J = 4.7, 9.0 Hz), 7.16-7.27 (m, 5 H), 8.25 $({\rm d}, 1~{\rm H}, J = 2.0~{\rm Hz}), 8.88~({\rm d}, 1~{\rm H}, J = 2.0~{\rm Hz}); {\rm ^{13}C\text{-}NMR}~({\rm CDCl_3},$ 400 MHz) δ 24.6, 25.1, 25.9, 28.6, 33.5, 35.2, 47.1, 57.5, 62.7, 125.7, 126.2, 128.2, 128.5, 141.8, 152.8, 153.8, 170.4, 171.4, 192.4; HPLC retention time 5.29 min (94.2%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 425 (M); HRMS for $C_{23}H_{27}N_3O_3S_1$ calcd 425.1772, found 425.1751. Anal. $(C_{23}H_{27}N_3O_3S_1\hbox{-}0.1H_2O)\ C,\ H,\ N.$

1-[[1-(4-Phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]- $\mathbf{2}(\mathbf{S})\text{-pyrrolidine}$ carboxaldehyde (20) was prepared by the method of Henning et al.25 and purified by column chromatography (2:1 CH₂Cl₂/EtOAc) to give **20** as a white solid: ¹H-NMR (CDCl₃, 400 MHz) δ 1.85-2.48 (m, 12 H), 2.57-2.69 (m, 2 H), 3.24-3.85 (m, 4 H), 4.59-4.78 (m, 2 H), 7.16-7.29 (m, 5 H), 9.50 (d, 1 H, J = 1.1 Hz); HPLC retention time 4.17 min (93.7%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS (EI) m/z 342 (M); HRMS for $C_{20}H_{26}N_2O_3$ calcd 342.1942, found 342.1947.

Ethyl [1-[[1-(4-phenylbutanoyl)-2(S)-pyrrolidinyl]carbonyl]-2(S)-pyrrolidinyl]-2-oxoacetate (21) was prepared by the method of Henning et al.25 and purified by column chromatography (2:1 toluene/EtOAc) to give 21 as a colorless oil: ¹H-NMR (CDCl₃, 400 MHz) δ 1.35 (t, 3 H, J = 7.2 Hz), 1.90-2.15 (m, 9 H), 2.22-2.36 (m, 3 H), 2.64-2.68 (m, 2 H), 3.38-3.42 (m, 1 H), 3.55-3.66 (m, 2 H), 3.91-3.97 (m, 1 H), 4.31 (q, 2 H, J = 7.2 Hz), 4.65 (dd, 1 H, J = 3.6, 8.2 Hz), 5.17(dd, 1 H, J = 5.9, 8.8 Hz), 7.16-7.29 (m, 5 H); HPLC retentiontime 5.60 min (91.8%), eluting with 60% CH₃CN in water, 1.0 mL/min; MS m/z 414 (M); HRMS for C₂₃H₃₀N₂O₅ calcd 414.2153, found 414.2121.

Enzyme Assays for Prolyl Endopeptidase (PEP). The pig kidney prolyl endopeptidase used in assays of inhibitor potency was a partially purified preparation following the method of Yoshimoto et al. 32 Prolyl endopeptidase is present in all tissues, and the enzyme from different tissues is biologically and immunologically identical.³³ Inhibition assays of the text compounds (7a-21) were carried out with Z-Gly-Pro-p-nitroanilide as substrate. The inhibitor solution (3 μ L of a DMSO solution of varying concentrations) was added to the substrate solution (0.26 mM in dioxane and 0.2 M phosphate buffer solution). To this solution was added 20 μ L of the enzyme solution (0.05 units/mL), and this reaction mixture was incubated for 20 min at 37 °C. p-Nitroaniline formation was monitored spectrophotometrically at 405 nm. The IC₅₀ value (n = 3-6) was estimated from the inhibitor concentration vs the activity curve.

For K_i determinations, PEP from Flavobacterium meningosepticus was purchased from Seikagaku Kougyo Co., Ltd., Tokyo. The substrate concentration was varied between 0.46 and 0.058 mM, the inhibitor concentration was varied between 0 and 0.75 μ g, and the PEP concentration was 25 nM. K_i values were determined by Dixon plots. The average of triplicate assays, plotting 1/v vs inhibitor concentration, gave intersecting lines with a correlation coefficient ≥ 0.97 . The **7a,b**, and 11a,b PEP inhibitor K_i values are 0.25, 0.19, 440, and 350 nM, respectively. The calculated differences in binding energies were obtained from these measured K_i values for thiazole (7a,b) and thiophene (11a,b) derivatives.

Acknowledgment. We wish to acknowledge the contributions of Mr. Y. Arai for the preparation of some of the compounds. We also thank Ms. S. Miki for obtaining the high-resolution mass spectra and Ms. N. Ichihara and Dr. S. Gomi for analyzing the ¹H-NMR and ¹³C-NMR spectra.

References

- Yoshimoto, T.; Orlowski, R. C.; Walter, R. Postproline Cleaving Enzyme: Identification as Serine Protease Using Active Site Specific Inhibitors. *Biochemistry* 1977, 16, 2942-2948.
- (2) Rennex, D.; Hemmings, B. A.; Hofsteenge, J.; Stone, S. R. cDNA Cloning of Porcine Brain Prolyl Endopeptidase and Identification of the Active-Site Seryl Residue. *Biochemistry* 1991, 30, 2195– 2203.
- (3) Welches, W. R.; Brosnihan, K. B.; Ferrario, C. M. A Comparison of the Properties and Enzymatic Activities of Three Angiotensin Processing Enzymes: Angiotensin Converting Enzyme, Prolyl Endopeptidase and Neutral Endopeptidase 24.11. *Life Sci.* 1993, 52, 1461-1480.
- (4) (a) Burbach, J. P. H.; Kovacs, G. L.; De Wied, D.; Van Nispen, J. W.; Greven, H. M. A Major Metabolite of Arginine Vasopressin in the Brain is a Highly Potent Neuropeptide. Science 1983, 221, 1310-1312. (b) De Wied, D.; Gaffori, O.; Van Ree, J. M.; De Jong, W. Central Target for the Behavioural Effects of Vasopressin Neuropeptides. Nature 1984, 308, 276-278.
- (5) Kowall, N. W.; Beal, M. F.; Busciglio, J.; Duffy, L. K.; Yankner, B. A. An in vivo Model for the Neurodegenerative Effects of β Amyloid and Protection by Substance P. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 7247-7251.
- (6) Nishimoto, I.; Okamoto, T.; Matsuura, Y.; Takahashi, S.; Okamoto, T.; Murayama, Y.; Ogata, E. Alzheimer Amyloid Protein Precursor Complexes with Brain GTP-Binding Protein G₀. Nature 1993, 362, 75-79.
- (7) Ishiura, S.; Tsukahara, T.; Tabira, T.; Shimizu, T.; Arahata, K.; Sugita, H. Identification of a Putative Amyloid A4-Generating Enzyme as a Prolyl Endopeptidase. FEBS Lett. 1990, 260, 131– 134
- (8) Angelucci, L.; Calvisi, P.; Catini, R.; Cosentino, U.; Cozzolino, R.; De Witt, P.; Ghirardi, O.; Giannessi, F.; Giuliani, A.; Guaraldi, D.; Misiti, D.; Ramacci, M. T.; Scolastico, C.; Tinti, M. O. Synthesis and Amnesia-Reversal Activity of a Series of 7-and 5-membered 3-Acylamino Lactams. J. Med. Chem. 1993, 36, 1511-1519.
- (9) Moos, W. H.; Davis, R. E.; Schwarz, R. D.; Gamzu, E. R. Cognition Activators. Med. Res. Rev. 1988, 8, 353-391.
- (10) (a) Saito, M.; Hashimoto, M.; Kawaguchi, N.; Fukami, H.; Tanaka, T.; Higuchi, N. Synthesis and Inhibitory Activity of Acyl Peptidyl-Prolinal Derivatives toward Post-Proline Cleaving Enzyme as Nootropic Agents. J. Enzyme Inhib. 1990, 3, 163-178. (b) Kolosko, N.; Bakker, A. V.; Faraci, W. S.; Nagel, A. A. Novel Inhibitors of Prolyl Endopeptidase: Effects of Stereochemistry. Drug Des. Discovery 1994, 11, 61-71.
- chemistry. Drug Des. Discovery 1994, 11, 61-71.

 (11) (a) Tsuru, D.; Yoshimoto, T.; Koriyama, N.; Furukawa, S. Thiazolidine Derivatives as Potent Inhibitors Specific for Prolyl Endopeptidase. J. Biochem. 1988, 104, 580-586. (b) Saito, M.; Hashimoto, M.; Kawaguchi, N.; Shibata, H.; Fukami, H.; Tanaka, T.; Higuchi, N. Synthesis and Inhibitory Activity of Acyl Peptidyl-Pyrrolidine Derivatives toward Post-Proline Cleaving Enzyme; A Study of Subsite Specificity. J. Enzyme Inhib. 1991, 5, 51-75.
- (12) (a) Yoshimoto, T.; Kado, K.; Matsubara, F.; Koriyama, N.; Kaneto, H.; Tsuru, D. Specific Inhibitors for Prolyl endopeptidase and Their Anti-Amnesic Effect. J. Pharmacobio-Dyn. 1987, 10, 730-735. (b) Atack, J. R.; Suman-Chauhan, N.; Dawson, G.; Kulagowski, J. J. In vitro and in vivo Inhibition of Prolyl Endopeptidase. Eur. J. Pharmacol. 1991, 205, 157-163.
- (13) (a) Hirschmann, R. Medicinal Chemistry in the Golden Age of Biology: Lessons from Steroid and Peptide Research. Angew. Chem., Int. Ed. Engl. 1991, 30, 1278-1307. (b) Giannis, A.; Kolter, T. Peptidomimetics for Receptor Ligands-Discovery, Development, and Medicinal Perspectives. Angew. Chem., Int. Ed. Engl. 1993, 32, 1244-1267.
- (14) (a) Rich, D. H. Peptidase Inhibitors. In Comprehensive Medicinal Chemistry; Hansch, C., Sammes, P. G., Taylor, J. B., Eds.; Pergamon Press: Oxford, 1990; Vol. 2, pp 391-441. (b) Wiley, R. A.; Rich, D. H. Peptidomimetics Derived from Natural Products. Med. Res. Rev. 1993, 13, 327-384.
- (15) Andersson, L.; Isley, T. C.; Wolfenden, R. a-Aminoaldehyde: Transition State Analogue Inhibitors of Leucine Aminopeptidase. Biochemistry 1982, 21, 4177-4180.
- (16) (a) Gelb, M. H.; Svaren, J. P.; Abeles, R. H. Fluoro Ketone Inhibitions of Hydrolytic Enzymes. Biochemistry 1985, 24, 1813-1817. (b) Riendeau, D.; Guay, J.; Weech, P. K.; Laliberté, F.; Yergey, J.; Li, C.; Desmarais, S.; Perrier, H.; Liu, S.; Nicoll-Griffith, D.; Street, I. P. Arachidonyl Trifluoromethyl Ketone, a Potent Inhibitor of 85-kDa Phospholipase A2, Block Production of Arachidonate and 12-Hydroxyeicosatetraenoic Acid by Calcium Ionophore-challenged Platelets. J. Biol. Chem. 1994, 269, 15619-15624. (c) Angliker, H.; Anagli, J.; Shaw, E. Inactivation of Calpain by Peptidyl Fluoromethyl Ketones. J. Med. Chem. 1992, 35, 216-220.

- (17) Peet, N. P.; Burkhart, J. P.; Angelastro, M. R.; Giroux, E. L.; Mehdi, S.; Bey, P.; Kolb, M.; Neises, B.; Schirlin, D. Synthesis of Peptidyl Fluoromethyl Ketones and Peptidyl α-Keto Esters as Inhibitors of Porcine Pancreatic Elastase, Human Neutrophil Elastase, and Rat and Human Neutrophil Cathepsin G. J. Med. Chem. 1990, 33, 394-407.
- (18) Angelastro, M. R.; Mehdi, S.; Burkhart, J. P.; Peet, N. P.; Bey, P. α-Diketone and α-Keto Ester Derivatives of N-Protected Amino Acids and Peptides as Novel Inhibitors of Cysteine and Serine Proteinases. J. Med. Chem. 1990, 33, 11-13.
- Serine Proteinases. J. Med. Chem. 1990, 33, 11-13.
 (19) Edwards, P. D.; Meyer, E. F., Jr.; Vijayalakshmi, J.; Tuthill, P. A.; Andisik, D. A.; Gomes, B.; Strimpler, A. Design, Synthesis, and Kinetic, Evaluation of a Unique Class of Elastase Inhibitors, the Peptidyl α-Ketobenzoxazoles, and the X-ray Crystal Structure of the Covalent Complex between Porcine Pancreatic Elastase and Ac-Ala-Pro-Val-2-Benzoxazole. J. Am. Chem. Soc. 1992, 114, 1854-1863.
- (20) Hamada, Y.; Shioiri, T. New Methods and Reagents in Organic Synthesis. 29.¹. A Practical Method for the Preparation of Optically Active N-Protected α-Amino Aldehydes and Peptide Aldehydes. Chem. Pharm. Bull. 1982, 30, 1921-1924.
- (21) Katritzky, A. R.; Laurenzo, K. S.; Relyea, D. I. The Preparation and Fungicidal Activity of a Series of Thiazolyl- and Isothiazolyl-Diarylcarbinols. Can. J. Chem. 1988, 66, 1617–1624.
- (22) Saari, W. S.; Halczenko, W.; Huff, J. R.; Guare, J. P., Jr.; Hunt, C. A.; Randall, W. C.; Lotti, V. J.; Yarbrough, G. G. Adrenoceptor and Tetrabenazine Antagonism Activities of Some Pyridinyltetrahydropyridines. J. Med. Chem. 1984, 27, 1182-1185.
- (23) Jones, C. D.; Winter, M. A.; Hirsch, K. S.; Stamm, N.; Taylor, H. M.; Holden, H. E.; Davenport, J. D.; Krumkalns, E. V.; Suhr, R. G. Estrogen Synthetase Inhibitors. 2.¹ Comparison of the in vitro Aromatase Inhibitory Activity for a Variety of Nitrogen Heterocycles Substituted with Diarylmethane or Diarylmethanol Groups. J. Med. Chem. 1990, 33, 416-429.
- (24) (a) Whitten, J. P.; Mattews, D. P.; McCarthy, J. R. [2-(Trimethylsilyl)ethoxy]methyl (SEM) as a Novel and Effective Imidazole and Fused Aromatic Imidazole Protecting Group. J. Org. Chem. 1985, 51, 1891–1894. (b) Lipshutz, B. H.; Vaccaro, W.; Huff, B. Protection of Imidazoles as Their β-Trimethylsilylethoxymethyl (SEM) Derivatives. Tetrahedron Lett. 1986, 27, 4095–4098.
- (25) Henning, R.; Urbach, H.; Hock, F. Derivatives of Cyclic Amino Acids, Agents Containing Them, and the Use Thereof. U.S. Patent 4983623, 1991.
- (26) Nicolaou, K. C.; Claremon, D. A.; Papahatjis, D. P. A. Mild Method for the Synthesis of 2-Ketopyrroles from Carboxylic Acids. *Tetrahedron Lett.* 1981, 22, 4647-4650.
- (27) Tsutsumi, S.; Okonogi, T.; Shibahara, S.; Patchett, A. A.; Christensen, B. G. α-Ketothiazole Inhibitors of Prolyl Endopeptidase. *Bioorg. Med. Chem. Lett.* 1994, 4, 831-834.
- (28) Patel, D. V.; Rielly-Gauvin, K.; Ryono, D. E.; Free, C. A.; Smith, S. A.; Petrill, E. W., Jr. Activated Ketone Based Inhibitors of Human Renin. J. Med. Chem. 1993, 36, 2431-2447.
- (29) (a) Bartlett, P. A.; Marlowe, C. K. Evaluation of Intrinsic Binding Energy from a Hydrogen Bonding Group in an Enzyme Inhibitor. Science 1987, 235, 569-571. (b) Tronrud, D. E.; Holden, H. M.; Matthews, B. W. Structures of Two Thermolysin-Inhibitor Complexes That Differ by a Single Hydrogen Bond. Science 1987, 235, 571-574. (c) Bash, P. A.; Singh, U. C.; Brown, F. K.; Langridge, R.; Kollman, P. A. Calculation of the Relative Change in Binding Free Energy of a Protein-Inhibitor Complex. Science 1987, 235, 574-576.
- (30) Kati, W. M.; Wolfenden, R. Major Enhancement of the Affinity of an Enzyme for a Transition-State Analog by a Single Hydroxy Group. Science 1989, 243, 1591-1593.
- (31) Reddy, M. R.; Viswanadhan, V. N.; Weinstein, J. N. Relative Differences in the Binding Free Energies of Human Immunodeficiency Virus Inhibitors: A Thermodynamic Cycle-Perturbation approach. *Proc. Natl. Acad. Sci. U.S.A.* 1991, 88, 10287– 10291.
- (32) Yoshimoto, T.; Ogita, K.; Walter, R.; Koida, M.; Tsuru, D. Post-Proline Cleaving Enzyme. Synthesis of a New Fluorogenic Substrate and Distribution of the Endopeptidase in Rat Tissues and Body Fluids of Man. *Biochim. Biophys. Acta* 1979, 569, 184-192.
- (33) (a) Andrews, P. C.; Minth, C. D.; Dixon, J. E. Immunochemical Characterization of a Proline Endopeptidase from Rat Brain. J. Biol. Chem. 1982, 257, 5861-5865. (b) Yoshimoto, T.; Oyama, H.; Koriyama, N.; Tsuru, D. Prolyl Endopeptidase from Bovine Testis: Purification, Characterization and Comparison with the Enzymes from Other Tissues. Chem. Pharm. Bull. 1988, 36, 1456-1462.