Anti-AIDS Agents. 15. Synthesis and Anti-HIV Activity of Dihydroseselins and Related Analogs¹

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Received February 28, 1994®

Forty-two dihydroseselins based on the structure of suksdorfin (1) were synthesized in order to evaluate their anti-HIV activity. These synthetic derivatives include 3',4'-di-O-acyl- and 3'- or 4'-O-acyl-cis-dihydroseselins (8-21) and 3', 4'-trans-dihydroseselins with O-acyl and/or O-alkyl groups at the 3' and 4' positions (6, 22-43). Two 4'-azido (44, 45) and three 4'-alkylamido (46, 48, 49) derivatives were also prepared. By using optically pure reagents, three pairs of diastereoisomers were synthesized and separated as optically pure compounds (14, 15; 16, 17; 38, 39). Together with the above synthetic derivatives, seselin (3) and (\pm) -cis-(4), (+)-cis- (5), and (±)-trans-dihydroseselin-3',4'-diol (7) were also tested for their in vitro anti-HIV activity. An optically pure compound, 3',4'-di-O-(-)-camphanoyl-(+)-cis-khellactone (16), showed potent inhibitory activity and remarkable selectivity against HIV replication. The EC_{50} value and in vitro therapeutic index (TI) of 16 are $4 \times 10^{-4} \, \mu M$ and 136 719, respectively, which are better than those shown by AZT in the same assay. In addition, compound 16 is also active against HIV replication in a monocytic cell line and in peripheral blood mononuclear cells (PBMCs). Our in vitro assay indicated that, like compound 1, compound 16 is not an inhibitor of HIV-1 reverse transcriptase. Moreover, the anti-HIV activity of 16 is stereoselective as its three diastereoisomers (17, 38, 39) are at least 10 000 times less active. Since other synthetic dihydroseselin derivatives with different substituents or without any substituents are inactive or are active only at much higher concentrations, the antiviral potency of 16 could be associated with the camphanoyl moieties of its structure. Therefore, compound 16 represents a unique coumarin structure with promising anti-HIV activity.

Introduction

Acquired immunodeficiency syndrome (AIDS), the disease caused by the human immunodeficiency virus (HIV), has become a serious threat to public health due to its rapid spread, high mortality rate, and incurability. Although the first generation anti-HIV compounds, such as AZT and other HIV reverse transcriptase (RT) inhibitors, are very effective against viral replication in laboratory tests, they have limited or transient clinical benefit due to their toxicities and to the development of drug resistant virus.^{2,3} Currently, the development of new anti-HIV agents is focused on discovering compounds either with novel structures or active through new mechanism(s) of action.

Recently, in our continuing bioactivity-directed isolation and characterization of new plant anti-HIV agents, suksdorfin (1), $^{4-6}$ isolated from Lomatium suksdorfii, was found to have anti-HIV activity with an in vitro EC₅₀ value of $2.6 \pm 2.1 \,\mu\text{M}$ and a therapeutic index (TI)

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* Abstract published in Advance ACS Abstracts, September 15, 1994.

in inhibition HIV-1 replication (ddC > ddI). However, the viral inhibition mediated by combining 1 with AZT was not statistically synergistic. Furthermore, the presence of suksdorfin did not antagonize the suppression mediated by three nucleoside reverse transcriptase inhibitors. These results suggested that 1 may exert its anti-HIV activity through mechanism(s) other than inhibition of HIV RT and has potential for use in drug combination with RT inhibitors for AIDS therapy. Despite the potential clinical and research applications, 1 has a relatively low therapeutic index, which might limit its usefulness. In attempts to discover compounds with more potent and selective anti-HIV activity, our further efforts have been directed to modification of dihydroseselin analogs related to 1. The syntheses of these analogs and their bioassay results are described in this paper.

30.6 ± 22.4.7 Combinations of 1 and the anti-HIV nucleosides ddI and ddC demonstrated statiscal synergy

Chemistry

Suksdorfin $[(3'R,4'R)-3'-acetoxy-4'-(isovaleryloxy)-3',4'-dihydroseselin] (1)^{5,6}$ is a pyranocoumarin derivative, which has two *cis*-oriented acyl groups at the 3' and 4' positions (Figure 1). Our early study of coumarin derivatives, including 1 and pteryxin (2, Figure 1), suggested that changing the acyl group at the 3' and 4'

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Seselin (3)

Figure 1. Structures of suksdorfin (1), pterysin (2), seselin (3), and 3',4'-di-O-(-)-camphanoyl-(+)-cis-khellactone (16).

positions of the pyran ring affected the biological activities.⁷ This finding led us to focus on chemical modification at the 3' and 4' positions of the dihydroseselin skeleton. By introducing different chemical moieties as well as changing the configurations at the 3' and 4' positions, two groups of dihydroseselin derivatives were synthesized and classified according to their 3' and 4' configurations: one with a cis configuration as found in compound 1, and the other with a trans configuration.

Since the syntheses began with a corresponding racemic material (4, 6, 7), most of the *cis* and *trans* products are (\pm) -mixtures except for 21, which was synthesized from (+)-*cis*-khellactone (5). Optically pure compounds 14-17, 38, and 39 were obtained by introducing an optically active acyl group, which generally gave separable diastereoisomers.

Scheme 1 shows the syntheses of the compounds with a cis configuration. The (\pm) -3',4'-di-O-acyl-cis-khellactone derivatives with two identical acyl groups at the 3' and 4' positions (8-17) were synthesized from (\pm) -cis-khellactone (4) using standard procedures. When using (S)-(+)- α -methylbutyric anhydride, monoesterified compounds (18, 19) were produced. Subsequent acetylation of 18 with Ac₂O/pyridine gave compound 20 with different ester groups at C3' and C4'. Although 18-20 are mixtures of diastereoisomers, separation by column chromatography was not successful. Compound 21, one optically pure component of racemic 12, was prepared from (+)-cis-khellactone (5), which was obtained by hydrolysis of 1.

For syntheses of compounds with a *trans* configuration at the 3' and 4' carbons (which is different from the configuration of 1), we introduced various esters at the 3' and/or 4' positions of the *trans*-khellactone molecule. In addition to the usual ester derivatives, compounds with azido (44, 45), amide (46, 48, 49), and ether (24–34, 43) substituent(s) were also prepared.

Scheme 2 summarizes the general synthetic procedures for the *trans* compounds. Compounds 37-42 with two identical ester groups were synthesized directly from 7, while 43 was produced by treatment of 6 with NaH and MeI. Compounds 22-27, which have two different substituents at C3' and C4', were obtained by

Scheme 1. Syntheses of cis-Khellactone Derivatives

* Optically pure compounds

Scheme 2. Syntheses of trans-Khellactone Derivatives

* Optically pure compounds

treatment of the monoester 6 with acylating or alkylating reagents. By using 3,4-dihydro-2*H*-pyran, two isomers (26, 27) were generated as a result of equal

addition of the hydroxyl oxygen to the pyran double bond from two opposite directions. These isomers with either R or S configuration at the acetal carbon were separated as their own 3',4'-trans racemic mixtures. Subsequent treatment of the 4'-O-alkyl compounds 24—26 with base gave the hydrolysis products 28—30. Acylation of 28—30 yielded a series of 3'-O-alkyl-4'-O-acylkhellactones (31-34). Finally, compound 36, which is the trans isomer of 1, was synthesized by acidic removal of the tetrahydropyranyl moiety of 33 to give 35, followed by acetylation of 35 with Ac_2O /pyridine.

Two 4'-azido- (44, 45) and three 4'-(alkylamido)dihydroseselin derivatives (46, 48, 49) were synthesized from cis-khellactone (4) (Scheme 3). Treatment of 4 with sodium azide and trifluoroacetic acid gave selective substitution at the 4'-position and produced a mixture of cis and trans azido isomers (44). After acetylation of 44 with Ac₂O/pyridine, the product (45) was further reduced by hydrogenation on palladium/carbon. The expected amine derivative (47) was obtained togther with an acetamido derivative (46). This amide derivative (46) was a product of intramolecular migration of the 3'-acetyl group to the 4'-amine. This migration probably occurred in the cis isomer, since a similar migration of an acetyl group has been observed in a cis dihydroseselin.8 The amine (47) was acylated to give 48. Acetylation of 48 yielded compound 49, which is a trans amido analog of 1.

Stereochemistry of Compounds 16, 17, 38, 39. With the optically pure reagent (-)-camphanic chloride, compounds 16 and 17, 38 and 39 were obtained as a mixture of diastereoisomers, which were separated by repeated column chromatography. On the other hand, hydrolysis with suksudorfin (1) with KOH in dioxane yielded (+)-cis-kehllactone [3'(R),4'(R)-configurations] as well as (-)-trans-kehllactone [3'(R),4'(S)-confugurations]. 6,9 Treatment of (+)-cis-kehllactone with (-)camphanoyl chloride gave 3',4'-di-O-(-)-camphanoyl-(+)-cis-kehllactone, which was shown to be identical with 16. Therefore, the configurations at C-3' and C-4' in compound 17, the diastereoisomer of 16, were assigned as S and S, respectively. In contrast, (-)-transkehllactone was treated with (-)-camphanoyl chloride, furnishing 3',4'-di-O-(-)-camphanoyl-(-)-trans-kehllactone, which was identical with 38, thus confirming its stereostructure. Accordingly, the structure of 39 was assigned as 3',4'-di-O-(-)-camphanoyl-(+)-trans-kehllactone.

Table 1. Anti-HIV Activities of Dihydroseselin Analogs in Acutely Infected H9 Lymphocytes

	- Industry Infocuted 110 Lymphoty ton							
compd	$EC_{50}(\mu M)$	TI^a	compd	$EC_{50}(\mu M)$	TI^a			
3	3.5	5	24	241	0.34			
4	_b	_	25	3.1	5			
5	_	_	26	_	_			
6	4.2	2.4	27	8.3	10			
7	_	_	28	43	2.7			
8	69.4	0.4	29	_	_			
9	7	2	30	_	_			
10	7.0	14.4	3 1	38	2.3			
11	4.7	3.5	32	_	-			
12	_	_	33	_	_			
13	<1.4	>2.2	34	-	_			
14	_	_	35	_	_			
15	11.4	2.9	36	_	_			
16	4×10^{-4}	136,719	37	43	3			
17	144.7	1.1	38	24.1	3			
18	_	_	39	32	>5			
19	_	_	40	_	_			
20	_	_	41	_	-			
2 1	21.8	2	42	20.9	3			
22	_	_	43	_	-			
23	9.3	1.4	44	115.5	2.9			
1	2.6 ± 2.1	30.6 ± 22.4	45	115.9	1			
2	4 .6	4.5	46	_	-			
			48	_	-			
AZT	0.15	12,500	49	_	_			

 a In vitro therapeutic index, ratio of IC₅₀:EC₅₀. b -: no suppression.

Compounds 14 and 15 are another pair of diastereoisomers for which the absolute configurations still remain to be established.

Results and Discussion

The anti-HIV assay indicates that compound 16, 3',4'-di-O-(-)-camphanoyl-(+)-cis-khellactone (Figure 1) has potent anti-HIV activity in acutely infected H9 lymphocytes (EC₅₀ = 4 × 10⁻⁴ μ M) and a remarkable therapeutic index (136 719), which are better than those of AZT and compound 1 in the same assay (Table 1).

Two cis compounds (10 and 11) also show moderate anti-HIV activities with EC₅₀ values of 7.0 and 4.7 μ M, and TIs 14.4 and 3.5, respectively. Three trans compounds with a 4'-m-chlorobenzoyl group (23, 25, and 27) are slightly active with EC₅₀ values ranging from 3.1 to 8.3 μ M, and TIs from 1.4 to 10. However, none of them has biological profiles better than compound 1. Other dihydroseselin derivatives in Table 1, either with various 3' and 4' substituents or without any substituents, such as seselin (3, Figure 1) or khellactones (4, 5, and 7), are inactive or toxic in the assay.

Moreover, our study indicates that the anti-HIV activity of compound 16, the (+)-cis-khellactone derivative, is highly stereospecific as the (-)-cis (17), (-)-trans (38), and (+)-trans (39) isomers of 16 are at least ten thousand times less active. In addition, the data suggest that the stereochemistry of 1 may also play a role in its anti-HIV activity, as its (\pm) -trans isomer (36) and amido (\pm) -trans analog (49) are both inactive in the same assay. Since the stereochemistry in compounds 1 and 16 is identical at the 3' and 4' positions, the 3' and 4' configurations of the khellactone derivatives might be very important for the anti-HIV activity in this type of compound.

Therefore, a possible explanation for the low anti-HIV activity observed in most of the synthetic dihydroseselin analogs is that they are racemic mixtures. Since the anti-HIV activity of this type of compound is stereospe-

Table 2. Anti-HIV Activity of 16 in Different in Vitro Assays

		H9 lymphocytes	PBMCs	U937
16	EC ₅₀ (μM)	4×10^{-4}	2.9×10^{-2}	2.1×10^{-3}
	TI^a	136 719	472	11,719

a In vitro therapeutic index, ratio of IC50:EC50.

cific as evidenced in 1 and 16, an optically pure compound with the preferred configuration would display a lower EC_{50} value than its corresponding isomeric mixture.

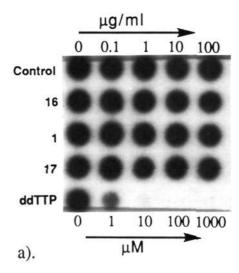
Further comparison of the 4' acyl groups of 1 and 16 suggested a structural similarity between the isovaleryl group in 1 and the (-)-camphanoyl group in 16. Compound 16 also contains an isovaleryl domain. However, the isovaleryl domain in 16 is conformationally more rigid, since it is within the frame of 4'-camphanoyl group neighboring another bulky camphanoyl substituent. On the other hand, the isovaleryl in 1 is more conformationally flexible, in which its terminal carbon atoms are disordered over two orientations in a X-ray crystallographic analysis (data not shown). Therefore, it seems the conformational feature of the isovaleryl domains at C4' could play a role in the anti-HIV activity of 1 and 16. However, a general comparison of the substitution pattern between 16 and 1, as well as other related compounds (9-12, 21), indicated that the dramatically increased potency in 16 should be related to the presence of the unique camphanoyl groups, which appear to be structurally required for high anti-HIV activity and remarkable selectivity in the dihydroseselin type

Compound 16 also demonstrated potent inhibition of viral replication in further assays using HIV-1 infected PHA-stimulated PBMCs and a promonocytic cell line, U937 (Table 2). In an attempt to determine the mechanism of anti-HIV activity, compounds 16 and 1 were tested for anti-HIV-RT activity. However, in an in vitro RT assay using the poly(A) as the template, both compounds showed no activity up to 100 µg/mL, while dideoxythymidine triphosphate (ddTTP) displayed a dose-dependent inhibition of RT activity (Figure 2).

Recently, four coumarin derivatives, calanolides A and B and inophyllums B and P, isolated from Calophyllum lanigerum and Calophyllum inophyllum Linn, respectively, were reported as potent anti-HIV agents. 10,11 Their anti-HIV activity is due to inhibition of the HIV-1 reverse transcriptase. Structurally, these coumarin derivatives possess the same tricyclic pyranocoumarin subunit found in compounds 1 and 16; however, they have an additional 5,6-ring system and a different substituent pattern at the C-4, C-2', C-3', and C-4' positions (Figure 3). Moreover, the anti-HIV mechanisms of these tetracyclic coumarins and of the tricyclic compounds 1 and 16 are different. The anti-HIV activity of compound 16 is also about 2 orders of magnitude greater than that reported for the tetracyclic coumarins. Consequently, compounds 16 and 1, especially 16, are functionally and structurally unique anti-HIV coumarins that might exert their antiviral activities by a mechanism other than inhibition of HIV-RT. Their mechanisms of action are currently under investigation.

Experimental Section

Chemistry. Melting points were measured with a Fisher-Johns melting point apparatus without correction. Optical



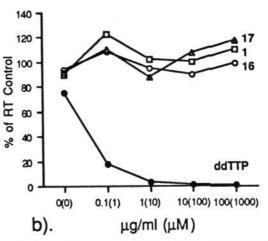


Figure 2. Assay of compounds 1, 16, and 17 on HIV-1 RT. ^{15,16} (a) Autoradiography of RT assay products. The intensity of the dots is proportional to the amount of catalytic product of HIV-1 reverse transcriptase. Control: samples are the reaction mixtures in the absence of drug. The concentrations of compounds 1, 16, and 17 are indicated on the top of the figure, while the concentrations of ddTTP are at the bottom. (b) Effect of compounds on RT activity as percent of control. For 1, 16, and 17, each point represents the mean of two experiments.

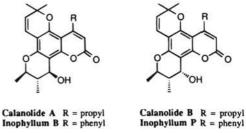


Figure 3. Structures of calanolides and inophyllums.

rotations were determined with a Rudolph Research Autopol III polarimeter. IR spectra were recorded on a Perkin-Elmer Model 1320 spectrometer. The proton nuclear magnetic resonance (¹H NMR) spectra were measured on a Bruker AC-300 spectrometer with Me₄Si (TMS) as the internal reference with CDCl₃ as solvent. Elemental analyses were determined

by Atlantic Microlab, Inc., Norcross, GA. HR-FAB MS and positive FAB MS were recorded on a HX-110 JEOL spectrometer. Thin-layer chromatography (TLC) silica gel plates were purchased from Analtech, Inc. Silica gel (230–400 mesh) from Aldrich, Inc., was used for column chromatography.

Seselin (3). Seselin (3) was prepared according to the procedure reported by Hlubuek.¹² A mixture of 7-hydroxycoumarin (Aldrich Chemical Co.) (8 g, 49 mmol), K2CO3 (8 g, 58 mmol), and KI (1.2 g, 7.2 mmol) in 98% aqueous acetone (200 mL) was stirred at room temperature for 1 h, and then 3-chloro-3-methylbut-1-yne (8 g, 49 mmol) was added. The whole mixture was refluxed for 24 h with stirring. After addition of additional 3-chloro-3-methylbut-1-yne (8 g, 49 mmol) and K2CO3 (8 g, 58 mmol), the mixture was further refluxed for 24 h. After cooling, the mixture was filtered and concentrated to dryness. The residue was dissolved in Et₂O, washed successively with H₂O and brine, dried over Na₂SO₄, and concentrated. The residue was then dissolved in N.Ndiethylaniline (100 mL), and the solution was refluxed for 2 h. After cooling, the solution was diluted with Et₂O, washed successively with 2 N H₂SO₄, water, and brine, dried over Na₂-SO₄, and concentrated. The product was purified by silica gel chromatography [hexane-EtOAc $(4:1 \rightarrow 3:1)$] to yield 3 (7.11 g, 63.6% yield) as pale yellow prisms (from MeOH): mp 119-120 °C; IR (KBr) 1715 (CO), 1620 (C=C) cm⁻¹; ¹H NMR δ 1.47 $(s, 6H, 2'-(CH_3)_2), 5.73 (d, 1H, J = 10 Hz, H-3'), 6.23 (d, 1H, J)$ = 9.5 Hz, H-3, 6.72 (d, 1H, J = 8.5 Hz, H-6), 6.88 (d, 1H, J = 8.5 Hz, H-6)10 Hz, H-4'), 7.21 (d, 1H, J = 8.5 Hz, H-5), 7.60 (d, 1H, J =

(\pm)-cis-Kehllactone (4). To a solution of seselin (3) (500 mg, 2.2 mmol) and N-methylmorpholine N-oxide (505 mg, 4.4 mmol) in 80% aqueous acetone (11 mL) was added 3% OsO₄ aqueous solution (1 mL), and the whole mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with H_2O and concentrated to give an aqueous solution, which was extracted with EtOAc. The EtOAc layer was dried over Na₂SO₄ and concentrated to give a residue, which was crystallized from benzene, yielding 4 (497 mg, 86% yield) as colorless needles; mp 157-158 °C; IR (KBr) 3400 (OH), 1730 (CO), 1690, 1610 (C=C) cm⁻¹; ¹H NMR δ 1.41, 1.47 (each s, 3H, 2′-(CH₃)₂), 3.26 (d, 1H, J = 5 Hz, 3′-OH), 3.87 (t, 1H, J = 5 Hz, H-3', 4.08 (br s, 1H, 4'-OH), 5.21 (d, 1H, J = 5 Hz, H-4'),6.25 (d, 1H, J = 9.5 Hz, H-3), 6.80 (d, 1H, J = 8.5 Hz, H-6), 7.32 (d, 1H, J = 8.5 Hz, H-5), 7.65 (d, 1H, J = 9.5 Hz, H-4), which were identified as (\pm) -cis-kehllactone by comparison of the physical and spectral data with those described in the

4'-O-(m-Chlorobenzoyl)-(\pm)-trans-kehllactone (6). A mixture of seselin (3) (850 mg, 3.7 mmol) and m-chloroperbenzoic acid (800 mg, 5.6 mmol) in CHCl₃ (25 mL) was kept at room temperature for 2 days. The reaction mixture was diluted with ether, washed successively with saturated NaH-CO₃ aqueous solution, water, and brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed over silica gel column [hexane-EtOAc (3: 1)] to give 6 as colorless prisms (MeOH) (770 mg, 51% yield): mp 223-224 °C; IR (KBr) 3440 (OH), 1720 (CO), 1605 (C=C) cm⁻¹; ¹H NMR δ 1.48, 1.52 (each s, 3H, 2'-(CH₃)₂), 4.06 (d, 1H, J = 4.5 Hz, H-3', 6.21 (d, 1H, J = 9.5 Hz, H-3), 6.39 (d, 1H, J = 9.5 Hz, H-3', 6.39 (d, 1H, J = 9.5 (d, 1H, J = 9.5 Hz, H-3', 6.39 (d, 1H, J = 9.5 (d, 1H, J = 9.5J = 8.5 Hz, H-6), 7.36 (t, 1H, J = 8 Hz, H-5''), 7.41 (d, 1H, J= 8.5 Hz, H--5), 7.52 (dd, 1H, J = 2, 8 Hz, H-4"), 7.62 (d, 1H, J = 2, 8 Hz, H--4")J = 9.5 Hz, H-4), 7.87 - 7.98 (m, 3H in total, H-2" and 6"). Anal. $(C_{21}H_{17}ClO_6)$ C, H, Cl.

(±)-trans-Kehllactone (7). A solution of 6 (750 mg, 1.88 mmol) in dioxane (12 mL) and 0.5 N KOH (18 mL) was stirred at room temperature for 30 min. The mixture was acidified with 20% HCl and stirred further 1 h at room temperature. The mixture was concentrated under reduced pressure to give an aqueous solution, which was extracted by EtOAc. The EtOAc layer was dried over Na₂SO₄ and concentrated. The product was purified by preparative TLC [hexane-EtOAc (1: 2)] to afford 7^{13} (176 mg, 35.8% yield) as colorless needles (MeOH); mp 185-186 °C; IR (KBr) 3380 and 3425 (OH), 1700 (CO), 1600 (C=C) cm⁻¹; ¹H NMR δ 1.31, 1.54 (each s, 3H, 2'-(CH₃)₂), 2.78 (d, 1H, J = 4, 7 Hz, 3'-OH), 3.85 (dd, 1H, J = 4, 7 Hz, H-3'), 3.98 (d, 1H, J = 3.5 Hz, 4'-OH), 5.00 (dd, 1H, J =

3.5, 7 Hz, H-4'), 6.25 (d, 1H, J = 9.5 Hz, H-3), 6.79 (d, 1H, J = 8.5 Hz, H-6), 7.32 (d, 1H, J = 8.5 Hz, H-5), 7.66 (d, 1H, J = 9.5 Hz, H-4).

Preparation of (+)-cis-Kehllactone (5) and (-)-trans-**Kehllactone.** A solution of suksdorfin (1) (750 mg, 1.93 mmol) in dioxane (12 mL) and 0.5 N KOH (18 mL) was stirred at room temperature for 30 min. The reaction mixture was acidified with 20% HCl and stirred further 1 h at room temperature. The mixture was concentrated under reduced pressure to give an aqueous solution, which was extracted by EtOAc. The EtOAc layer was dried over Na₂SO₄ and concentrated to give a residue, which was subjected to silica gel chromatography. Elution with CHCl₃-MeOH-H₂O (10:1:0.1) afforded (+)-cis-kehllactone (30 mg) as colorless needles (benzene): mp 174-175 °C; $[\alpha]^{20}D + 87^{\circ} (c = 0.5, CHCl_3)$; IR and ¹H NMR were identical with 4. Further elution with same solvent system furnished (-)-trans-kehllactone (12 mg) as colorless needles (benzene); mp 182-185 °C; $[\alpha]^{20}$ _D -20.2° (c = 0.5, CHCl₃); IR and ¹H NMR were identical with 7. The structure comfirmation of these products was obtained by comparison of the physical and spectral data with those described in the literature.9,14

General Procedure for Synthesizing O-Acylkhellactones (8, 9, 11-17, 20-23, 31-34, 36-42) and (\pm) -trans-3'-(Acyloxy)-4'-azido- (45) and (\pm) -trans-3'-(Acyloxy)-4'-(alkylamido)dihydroseselins (49). To a solution of a corresponding alcohol (4-7, 19, 28-30, 35, 44, and 48) or amine (47) in anhydrous CH_2Cl_2 containing 20% dry pyridine was added an appropriate acyl chloride or anhydride (2.5-3 equiv) at 0 °C or at room temperature, respectively. The mixture was allowed to stand overnight, and the volatiles were evaporated in vacuo. Alternatively, the mixture was poured into water and extracted with CHCl₃. The organic layer was washed with water and brine, dried over anhydrous Na_2SO_4 , and concentrated in vacuo. The residue was chromatographed on preparative TLC plates or silica gel columns to afford the product.

3′,4′-Di-O-acetyl-cis-khellactone (8): yield 83% (starting with 20.0 mg of 4); crystallization from acetone gave colorless needles; mp 164–165 °C; HR-FAB MS calcd for $C_{18}H_{18}O_7$ 346.1053, found m/z 346.1054; positive FAB MS m/z 715 (2M + Na)⁺, 369 (M + Na)⁺; IR (KBr) 1750 (CO, lactone), 1720 (CO, ester), 1612 (C=C) cm⁻¹; ¹H NMR δ 1.41, 1.45 (each s, 3H, 2′-(CH₃)₂), 2.14, 2.10 (each s, 3H, OCOCH₃-3′,4′), 5.30 (d, J = 5.0 Hz, 1H, H-3′), 6.24 (d, J = 9.5 Hz, 1H, H-3), 6.54 (d, J = 5.0 Hz, 1H, H-4′), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.35 (d, J = 8.5 Hz, 1H, H-5), 7.60 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{18}H_{18}O_7^{-1}/_4H_2O$) C, H.

3',4'-Di-O-isovaleryl-cis-khellactone (9): yield 62% (starting with 25.5 mg of 4); amorphous; mp 126–128 °C; HR-FAB MS calcd for $C_{24}H_{30}O_7$ Na 453.1889, found m/z 453.1889 (M + Na)+; positive FAB MS m/z 883 (2M + Na)+, 453 (M + Na)+; IR (KBr) 1745 (CO, lactone), 1730 (CO, ester), 1608 (C=C) cm⁻¹; ¹H NMR δ 0.95–1.05 (m, 12H, 2 x isovaleryl (CH₃)₂), 1.41, 1.44 (each s, 3H, 2'-(CH₃)₂), 2.06–2.35 (m, 2H, 2 x CH), 2.23 (m, 4H, 2 x CH₂), 5.32 (d, J = 5.0 Hz, 1H, H-3'), 6.23 (d, J = 9.5 Hz, 1H, H-3), 6.55 (d, J = 8.5 Hz, 1H, H-4'), 6.79 (d, J = 8.5 Hz, 1H, H-6), 7.35 (d, J = 8.5 Hz, 1H, H-5), 7.58 (d, J = 9.5 Hz, 1H, H-4). Anal. $(C_{24}H_{30}O_7^{-1}/_4H_2O)$ C, H.

3',4'-Di-O-senecioyl-cis-khellactone (11): yield 30% (starting with 25.0 mg of 4); amorphous; mp 127–129 °C; IR (KBr) 1760 (CO, lactone), 1740 (CO, ester), 1650 (C=C, senecioyl), and 1615 (C=C) cm⁻¹; ¹H NMR δ 1.43, 1.47 (each s, 3H, 2'-(CH₃)₂), 1.88, 1.89, 2.15, 2.10 (each s, 3H, 2 x senecioyl (CH₃)₂), 5.36 (d, J = 5.0 Hz, 1H, H-3'), 5.65 (each s, 1H, 2 x senecioyl CH), 6.21 (d, J = 9.5 Hz, 1H, H-3), 6.63 (d, J = 5.0 Hz, 1H, H-4'), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.34 (d, J = 8.5 Hz, 1H, H-5), 7.58 (d, J = 9.5 Hz, 1H, H-4). Anal. (C₂₄H₂₆O₇) C, H.

3',4'-Bis-O-(*tert***-butylacetyl)-***cis***-khellactone** (12): yield 34% (starting with 53.0 mg of 4); crystallization from EtOH gave colorless needles; mp 133–134 °C; IR (KBr) 1745 (CO), 1605 (C=C) cm⁻¹; ¹H NMR δ 1.05, 1.60 (each s, 9H, 2 x butyl (CH₃)₃), 1.43, 1.45 (each s, 3H, 2'-(CH₃)₂), 2.20–2.37 (m, 4H, 2 x CH₂), 5.31 (d, J = 4.5 Hz, 1H, H-3'), 6.22 (d, J = 9.5 Hz, 1H, H-3), 6.55 (d, J = 4.5 Hz, 1H, H-4'), 6.78 (d, J = 8.5 Hz, 1H,

3',4'-Bis-O-(4-tert-butylbenzoyl)-cis-khellactone (13): yield 81% (starting with 48.0 mg of 4); amorphous; mp 255–256 °C; HR-FAB MS calcd for $C_{36}H_{38}O_7Na$ 805.2516, found m/z 805.2518 (M + Na)+; positive FAB MS m/z 805 (M + Na)+; IR (KBr) 1740 (CO), 1615 (C=C) cm⁻¹; ¹H NMR δ 1.30 (s, 18H, 2 x butyl (CH₃)₃), 1.50, 1.64 (each s, 3H, 2'-(CH₃)₂), 5.65 (d, J = 5.0 Hz, 1H, H-3'), 6.18 (d, J = 9.5 Hz, 1H, H-4), 6.88 (d, J = 8.5 Hz, 1H, H-6), 6.94 (d, J = 5.0 Hz, 1H, H-4'), 7.40 (d, J = 8.5 Hz, 1H, H-5), 7.58 (d, J = 9.5 Hz, 1H, H-4'), 7.33–7.35 and 7.79–7.83 (m, 8H, benzoyl H). Anal. ($C_{36}H_{38}O_7^{1/2}$ $_2H_2O$) C, H.

3',4'-Bis-O-((-)-3-menthyloxycarbonyl)-cis-khellactone (14): yield 50% (starting with 93.0 mg of 4); gum; HR-FAB MS calcd for $C_{36}H_{50}O_9Na$ 649.3352, found m/z 649.3350 (M + Na)+; positive FAB MS m/z 649 (M + Na)+; IR (KBr) 1750 (CO), 1610 (C=C) cm⁻¹; $[\alpha]^{20}_D$ -53.2° (c = 0.5, CHCl₃); ¹H NMR δ 0.79-0.97 (m, 22H, 6 x menthyl CH₃ and 2 x menthyl CH₂), 0.99-1.14 (m, 4H, 2 x menthyl CH₂), 1.38-1.42 (m, 2H, 2 x menthyl CH), 1.45, 1.47 (each s, 3H, 2'-(CH₃)₂), 1.65-1.69 (m, 4H, 2 x menthyl CH₂), 1.91-2.35 (m, 4H, 4 x menthyl CH), 4.58-4.63 (m, 2H, 2 x menthyl OCOCH), 5.10 (d, J = 5.0 Hz, 1H, H-3'), 6.22 (d, J = 9.5 Hz, 1H, H-3), 6.45 (d, J = 5.0 Hz, 1H, H-4'), 6.76 (d, J = 8.5 Hz, 1H, H-6), 7.32 (d, J = 8.5 Hz, 1H, H-5), 7.56 (d, J = 9.5 Hz, 1H, H-4).

3′,4′-Bis-O-((-)-3-menthyloxycarbonyl)-cis-khellactone (15): yield 37% (starting with 93.0 mg of 4); gum; HR-FAB MS calcd for $C_{36}H_{50}O_9Na$ 649.3352, found m/z 649.3353 (M + Na)⁺; positive FAB MS m/z 649 (M + Na)⁺; IR (KBr) 1760 (CO), 1613 (C=C) cm⁻¹; $[\alpha]^{20}D_-43.6^\circ$ (c=0.5, CHCl₃); 1H NMR δ 0.81-0.97 (m, 20H, 6 x menthyl CH₃, menthyl CH₂), 1.01-1.14 (m, 4H, 2 x menthyl CH₂), 1.33-1.49 (m, 4H, menthyl CH₂ and 2 x menthyl CH), 1.66-1.71 (m, 4H, 2 x menthyl CH₂), 1.98-2.17 (m, 4H, 4 x menthyl CH), 4.53-4.72 (m, 2H, 2 x menthyl OCOCH), 1.47, 1.48 (each s, 3H, 2′-(CH₃)₂), 5.10 (d, J=5.0 Hz, 1H, H-3′), 6.22 (d, J=9.5 Hz, 1H, H-3), 6.41 (d, J=5.0 Hz, 1H, H-4′), 6.76 (d, J=8.5 Hz, 1H, H-6), 7.34 (d, J=8.5 Hz, 1H, H-5), 7.57 (d, J=9.5 Hz, 1H, H-4). Anal. $(C_{36}H_{50}O_{9}^{-1/2}H_2O)$ C, H.

3′,4′-Di-O-(-)-camphanoyl-(+)-cis-khellactone (16): yield 27% (starting with 200.0 mg of 4); colorless needles (from EtOH); mp 200–202 ° C; $[\alpha]^{20}_{\rm D}$ +31.1° (c=0.5, CHCl₃); positive FAB MS m/z 623 (M + H)+, 425 (M – camphanic acid)+, 227 (M – 2 x camphanic acid)+; IR (KBr) 1790, 1745 (COO), 1605 (C=C) cm⁻¹; ¹H NMR δ 0.98, 1.01, 1.08, 1.10, 1.11, 1.12 (each s, 3H, camphanoyl CH₃), 1.45, 1.50 (each s, 3H, 2′-CH₃), 1.70, 1.94, 2.23, 2.50 (each m, 2H, camphanoyl CH₂), 5.39 (d, 1H, J=5 Hz, H-3′), 6.24 (d, 1H, J=9.5 Hz, H-3), 6.66 (d, 1H, J=5 Hz, H-4′), 6.82 (d, 1H, J=8.5 Hz, H-6), 7.41 (d, 1H, J=8.5 Hz, H-5), 7.62 (d, 1H, J=9.5 Hz, H-4). Anal. (C₃4H₃₈O₁₁) C, H.

3',4'-Di-O-(-)-camphanoyl-(-)-cis-khellactone (17): yield 80% (starting with 200.0 mg of 4); colorless needles (from EtOH); mp 242-244 °C; $[\alpha]^{20}_D$ -67.7° (c=0.5, CHCl₃); positive FAB MS m/z 623 (M + H)+, 425 (M - camphanic acid)+, 227 (M - 2 x camphanic acid)+; IR (KBr) 1780, 1750 (COO), 1605 (C=C) cm⁻¹; ¹H NMR δ 0.94, 1.04, 1.06, 1.12, 1.13, (each s, 18H in total, camphanoyl CH₃), 1.45, 1.56 (each s, 3H, 2'-CH₃), 1.70, 1.93, 2.10, 2.34, 2.55 (each m, 8H in total, camphanoyl CH₂), 5.47 (d, 1H, J=4.5 Hz, H-3'), 6.22 (d, 1H, J=9.5 Hz, H-3), 6.74 (d, 1H, J=4.5 Hz, H-4'), 6.82 (d, 1H, J=8.5 Hz, H-6), 7.40 (d, 1H, J=8.5 Hz, H-5), 7.61 (d, 1H, J=9.5 Hz, H-4). Anal. (C₃₄H₃₈O₁₁) C, H.

3'-O-Acetyl-4'-(α -methylbutyryl)-cis-khellactone (20): yield 54% (starting with 9.0 mg of 19); gum; HR-FAB MS calcd for C₂₁H₂₄O₇Na 411.1420, found m/z 411.1423 (M + Na)⁺; positive FAB MS m/z 411 (M + Na)⁺; IR (KBr) 1740 (CO), 1610 (C=C) cm⁻¹; ¹H NMR δ 0.94, 0.96 (each t, J=7.5 Hz, 3 /₂H, CH₂CH₃), 1.17, 1.22 (each d, J=7.0 Hz, 3 /₂H, CHCH₃), 1.45 (each s, 3H, 2'-(CH₃)₂), 1.45-1.80 (m, 2H, CH₂), 2.09 (s, 3H, COCH₃), 2.35-2.46 (m, 1H, CH), 5.30, 5.32 (each d, J=5.0 Hz, 1 /₂H, H-3'), 6.23 (d, J=9.5 Hz, 1H, H-3), 6.53, 6.54 (each d, J=5.0 Hz, 1 /₂H, H-4'), 6.80 (d, J=8.5 Hz, 1H, H-6), 7.36 (d, J=8.5 Hz, 1H, H-5), 7.59 (d, J=9.5 Hz, 1H, H-4).

3',4'-Bis-O-(tert-butylacetyl)-(+)-cis-khellactone (21): yield 58% (starting with 15.0 mg of 5); mp, IR, and 1 H NMR are the same as 12, $[\alpha]^{20}$ D -13° (c=0.5, CHCl₃).

3'-O-Acetyl-4'-O-(m-chlorobenzoyl)-trans-khellactone (22): yield 65% (starting with 46.0 mg of 6); crystallization from acetone gave colorless prisms; mp 205–206 °C; IR (KBr) 1740 (CO, lactone), 1720 (CO, ester), 1605 (C=C) cm⁻¹;

1H NMR δ 1.41, 1.52 (each s, 3H, 2'-(CH₃)₂), 2.11 (s, 3H, COCH₃), 5.41 (d, J = 3.5 Hz, 1H, H-3'), 6.20 (d, J = 9.5 Hz, 1H, H-3), 6.43 (d, J = 3.5 Hz, 1H, H-4'), 6.87 (d, J = 8.5 Hz, 1H, H-6), 7.33–7.38 (m, 1H, benzoyl H-5"), 7.42 (d, J = 8.5 Hz, 1H, H-5), 7.51 (d, J = 8.0 Hz, 1H, benzoyl H-4"), 7.60 (d, J = 9.5 Hz, 1H, H-4), 7.90–7.95 (m, 2H, benzoyl H-2",6"). Anal. (C₂₃H₁₉ClO₇) C, H, Cl.

3'-O-Isovaleryl-4'-O-(m-chlorobenzoyl)-trans-khellactone (23): yield 56% (starting with 50.0 mg of 6); amorphous; mp 144–146 °C; IR (KBr) 1730 (CO), 1610 (C=C) cm⁻¹; 1 H NMR δ 0.93 (d, J = 6.5 Hz, 6H, isovaleryl (CH₃)₂), 1.40, 1.51 (each s, 3H, 2'-(CH₃)₂), 2.04–2.11 (m, 1H, CH), 2.23–2.26 (m, 2H, CH₂), 5.42 (d, J = 4.0 Hz, 1H, H-3'), 6.20 (d, J = 9.5 Hz, 1H, H-3), 6.43 (d, J = 4.0 Hz, 1H, H-4'), 6.86 (d, J = 8.5 Hz, 1H, H-6), 7.32–7.37 (m, 1H, benzoyl H-5"), 7.40 (d, J = 8.5 Hz, 1H, H-5), 7.50 (d, J = 8.0 Hz, 1H, benzoyl H-4"), 7.60 (d, J = 9.5 Hz, 1H, H-4), 7.90–7.95 (m, 2H, benzoyl H-2",6"). Anal. (C₂₆H₂₅ClO₇) C, H, Cl.

3'-O-Benzyl-4'-O-acetyl-trans-khellactone (31): yield 80% (starting with 70.0 mg of 28); amorphous; mp 153–155 °C; HR-FAB MS calcd for $C_{23}H_{22}O_6$ 394.1416, found m/z 394.1413; positive FAB MS m/z 811 (2M + Na)+, 417 (M + Na)+; IR (KBr) 1730 (CO, lactone), 1710 (CO, ester), 1610 (C=C) cm^{-1}; H NMR δ 1.33, 1.38 (each s, 3H, 2'-(CH₃)₂), 2.13 (s, 3H, COCH₃), 3.58 (d, J = 2.5 Hz, 1H, H-3'), 4.75, 4.89 (each d, J = 12.0 Hz, 1H, ArCH₂), 6.24 (d, J = 9.5 Hz, 1H, H-3), 6.32 (d, J = 2.5 Hz, 1H, H-4'), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.28–7.41 (m, 6H, H-5 and benzyl ArH-2',3',4',5',6'), 7.61 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{23}H_{22}O_{6}$ 1/4H₂O) C, H.

3'-O-Methyl-4'-O-acetyl-trans-khellactone (32): yield 64% (starting with 12.2 mg of 29); amorphous; mp 148–150 °C; IR (KBr) 1735 (CO), 1608 (C=C) cm⁻¹; 1 H NMR δ 1.39, 1.49 (each s, 3H, 2'-(CH₃)₂), 2.14 (s, 3H, COCH₃), 3.40 (d, J=3.0 Hz, 1H, H-3'), 3.62 (s, 3H, OMe), 6.24 (d, J=9.5 Hz, 1H, H-3), 6.25 (d, J=3.0 Hz, 1H, H-4'), 6.80 (d, J=8.5 Hz, 1H, H-6), 7.35 (d, J=8.5 Hz, 1H, H-5), 7.62 (d, J=9.5 Hz, 1H, H-4). Anal. (C₁₇H₁₈O₆) C, H.

3'-O-(2''.Tetrahydropyranyl)-4'-O-isovaleryl-trans-khellactone (33): yield 67% (starting with 57.0 mg of 30); yellow oil; HR-FAB MS calcd for $C_{24}H_{30}O_7$ Na 453.1889, found m/z 453.1891 (M + Na)+; positive FAB MS m/z 453 (M + Na)+; IR (KBr) 1740 (CO), 1610 (C=C) cm⁻¹; ¹H NMR δ 0.96–1.00 (m, 6H, isovaleryl (CH₃)₂), 1.39, 1.47 (each s, 3H, 2'-(CH₃)₂), 142–1.52, 1.69–1.78 (m, 6H, tetrahydropyranyl CH₂-3",4",5"), 2.08–2.17 (m, 1H, CH), 2.21–2.25 (m, 2H, isovaleryl CH₂), 3.55–3.62, 3.87–3.99 (m, 2H, CH₂-6"), 3.84 (d, J = 2.5 Hz, 1H, H-3'), 4.84–4.86 (m, 1H, CH-1"), 6.21 (d, J = 9.5 Hz, 1H, H-3), 6.46 (d, J = 2.5 Hz, 1H, H-4'), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.33 (d, J = 8.5 Hz, 1H, H-5), 7.59 (d, J = 9.5 Hz, 1H, H-4').

3'.O-(2"-Tetrahydropyranyl)-4'-O-acetyl-trans-khellactone (34): yield 16% (starting with 190.0 mg of 30); crystallization from acetone gave colorless needles; mp 169–170 °C; IR (KBr) 1740 (CO) and 1605 (C=C) cm⁻¹; ¹H NMR δ 1.41, 1.60 (each s, 3H, 2'-(CH₃)₂), 2.13 (s, 3H, COCH₃), 144–1.58, 1.67–1.77 (m, 6H, tetrahydropyranyl CH₂-3",4",5"), 3.55–3.63, 3.93–3.96 (m, 2H, CH₂-6"), 3.98 (d, J=2.5 Hz, 1H, H-3'), 5.06–5.08 (m, 1H, CH-1"), 6.15 (d, J=2.5 Hz, 1H, H-4'), 6.25 (d, J=9.5 Hz, 1H, H-3), 6.83 (d, J=8.5 Hz, 1H, H-6), 7.36 (d, J=8.5 Hz, 1H, H-5), 7.62 (d, J=9.5 Hz, 1H, H-4). Anal. (C₂₁H₂₄O₇) C, H.

3'-O-Acetyl-4'-O-isovaleryl-trans-khellactone (**36**): yield 50% (starting with **3**0.0 mg of **35**); crystallization from acetone gave colorless needles; mp 175–176 ° C; IR (KBr) 1748 (CO, lactone), 1730 (CO, ester), 1610 (C=C) cm⁻¹; ¹H NMR δ 0.98, 0.99 (each d, J=6.5 Hz, 3H, isovaleryl (CH₃)₂), 1.38, 1.45 (each s, 3H, 2'-(CH₃)₂), 2.10 (s, 3H, COCH₃), 2.11–2.21 (m, 1H, CH), 2.24–2.35 (m, 2H, CH₂), 5.29 (d, J=4.0 Hz, 1H, H-3'), 6.22 (d, J=4.0 Hz, 1H, H-4'), 6.24 (d, J=9.5 Hz, 1H, H-3), 6.83

(d, J = 8.5 Hz, 1H, H-6), 7.38 (d, J = 8.5 Hz, 1H, H-5), 7.62 (d, J = 9.5 Hz, 1H, H-4). Anal. $(C_{21}H_{24}O_7)$ C, H.

3',4'-Di-O-acetyl-trans-khellactone (37): yield 68% (starting with 20.0 mg of 7); crystallization from acetone gave colorless needles; mp > 260 °C; HR-FAB MS calcd for $C_{18}H_{18}O_7$ -Na 369.0950, found m/z 369.0951 (M + Na)+; positive FAB MS m/z 369 (M + Na)+; IR (KBr) 1758 (CO, lactone), 1735 (CO, ester), 1612 (C=C) cm⁻¹; ¹H NMR δ 1.37, 1.45 (each s, 3H, 2'-(CH₃)₂), 2.11, 2.14 (each s, 3H, OCOCH₃-3',4'), 5.29 (d, J = 4.5 Hz, 1H, H-3'), 6.22 (d, J = 4.5 Hz, 1H, H-4'), 6.25 (d, J = 9.5 Hz, 1H, H-6), 7.37 (d, J = 8.5 Hz, 1H, H-5), 7.61 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{18}H_{18}O_7$ ³/₄H₂O) C, H.

3',4'-Di-O-(-)-camphanoyl-(-)-trans-khellactone (38): yield 51% (starting with 100.0 mg of 7); colorless needles (from EtOH); mp 249-251 °C; $[\alpha]^{20}_{\rm D}+18.4^{\circ}$ (c=0.5, CHCl₃); positive FAB MS m/z 623 (M + H)+, 425 (M - camphanic acid)+, 227 (M - 2 x camphanic acid)+; IR (KBr) 1790, 1770, 1750 (COO), 1610 (C=C) cm⁻¹; 1 H NMR δ 0.97, 0.98, 1.00, 1.08, 1.09, 1.12 (each s, 3H, camphanoyl CH₃), 1.41, 1.50 (each s, 3H, 2'-CH₃), 1.66, 1.93, 2.07, 2.46, 2.50 (each m, 8H in total, camphanoyl CH₂), 5.39 (d, 1H, J=3.5 Hz, H-3'), 6.24 (d, 1H, J=9.5 Hz, H-3), 6.30 (d, 1H, J=3.5 Hz, H-4'), 6.86 (d, 1H, J=8.5 Hz, H-6), 7.42 (d, 1H, J=8.5 Hz, H-5), 7.63 (d, 1H, J=9.5 Hz, H-4). Anal. (C₃₄H₃₈O₁₁) C, H.

3',4'-Di-O-(-)-camphanoyl-(+)-trans-khellactone (39): yield 19% (starting with 100.0 mg of 7); colorless needles (from EtOH); mp 253-254 °C; $[\alpha]^{20}_{\rm D}$ -42.0° (c=0.5, CHCl₃); positive FAB MS m/z 623 (M + H)+, 425 (M - camphanic acid)+, 227 (M - 2 x camphanic acid)+; IR (KBr) 1800, 1750, 1735 (COO), 1605 (C=C) cm⁻¹; ¹H NMR δ 0.99, 1.06, 1.07, 1.09, 1.10 (each s, 18H in total, camphanoyl CH₃), 1.41, 1.50 (each s, 3H, 2'-CH₃), 1.68, 1.92, 2.12, 2.49 (each m, 2H, camphanoyl CH₂), 5.40 (d, 1H, J=3.5 Hz, H-3'), 6.26 (d, 1H, J=9.5 Hz, H-3), 6.29 (1H, d, J=3.5 Hz, H-4'), 6.84 (1H, d, J=8.5 Hz, H-6), 7.41 (1H, d, J=8.5 Hz, H-6), 7.64 (1H, d, J=9.5 Hz, H-4). Anal. (C₃₄H₃₈O₁₁) C, H.

3',4'-Bis-O-(phenyloxycarbonyl)-trans-khellactone (40): yield 38% (starting with 54.0 mg of 7); amorphous; mp 134–135 °C; HR-FAB MS calcd for $C_{28}H_{22}O_9Na$ 525.1162, found m/z 525.1163 (M + Na)+; positive FAB MS m/z 1027 (2M + Na)+, 525 (M + Na)+; IR (KBr) 1775 (CO, lactone), 1740 (CO, ester), 1610 (C=C) cm⁻¹; ¹H NMR δ 1.51, 1.59 (each s, 3H, 2'-(CH₃)₂), 5.33 (d, J = 4.5 Hz, 1H, H-3'), 6.30 (d, J = 9.5 Hz, 1H, H-3), 6.38 (d, J = 4.5 Hz, 1H, H-4'), 6.86 (d, J = 8.5 Hz, 1H, H-6), 7.15–7.44 (m, 11H, H-5 and 2 x phenyl Hs), 7.64 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{28}H_{22}O_9^{-1}/_2H_2O$) C, H.

3',4'-Bis-O-(tert-butylacetyl)-trans-khellactone (41): yield 59% (starting with 170.0 mg of 7); crystallization from MeOH gave colorless needles; mp 118–120 °C; IR (KBr) 1745 (CO, lactone), 1730 (CO, ester), 1605 (C=C) cm⁻¹; 1 H NMR 0 1.02, 1.05 (each s, 9H, 2 x butyl (CH₃)₃), 1.38, 1.45 (each s, 3H, 2'-(CH₃)₂), 2.22–2.33 (m, 4H, 2 x CH₂), 5.28 (d, 1 J = 3.5 Hz, 1H, H-3'), 6.18 (d, 1 J = 3.5 Hz, 1H, H-4'), 6.23 (d, 1 J = 9.5 Hz, 1H, H-3), 6.82 (d, 1 J = 8.5 Hz, 1H, H-6), 7.36 (d, 1 J = 8.5 Hz, 1H, H-5), 7.60 (d, 1 J = 9.5 Hz, 1H, H-4'). Anal. (C₂₆H₃₄O₇) C, H.

3',4'-Di-O-isovaleryl-trans-khellactone (42): yield 37% (starting with 98.0 mg of 7); crystallization from acetone gave colorless needles; mp 110–111 °C; HR-FAB MS calcd for $C_{24}H_{30}O_7$ Na 453.1889, found m/z 453.1889 (M + Na)⁺; positive FAB MS m/z 453 (M + Na)⁺; IR (KBr) 1750 (CO, lactone), 1730 (CO, ester), 1608 (C=C) cm⁻¹; ¹H NMR δ 0.89–0.99 (m, 12H, 2 x isovaleryl (CH₃)₂), 1.37, 1.44 (each s, 3H, 2'-(CH₃)₂), 2.10–2.30 (m, 6H, 2 x isovaleryl CH₂ and CH), 5.28 (d, J = 4.0 Hz, 1H, H-3'), 6.19 (d, J = 4.0 Hz, 1H, H-4'), 6.24 (d, J = 9.5 Hz, 1H, H-3), 6.82 (d, J = 8.5 Hz, 1H, H-6), 7.37 (d, J = 8.5 Hz, 1H, H-5), 7.61 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{24}H_{30}O_7$:H₂O) C, H.

Mixtures of (±)-trans- and (±)-cis-3'-Acetoxy-4'-azidodihydroseselins (45): yield 88% (starting with 61.0 mg of 44); amorphous; mp 126-127 °C; IR (KBr) 2120 (N₃), 1740 (CO), 1610 (C=C) cm⁻¹; ¹H NMR δ 1.39, 1.43, 1.45, 1.46 (each s, $^{3}/_{2}$ H, 2'-(CH₃)₂), 2.10, 2.23 (each s, $^{3}/_{2}$ H, COCH₃), 4.91 (d, J = 3.0 Hz, $^{1}/_{2}$ H, trans-H-3'), 5.16 (d, J = 5.0 Hz, $^{1}/_{2}$ H, cis-H-3'), 5.18 (d, J = 3.0 Hz, $^{1}/_{2}$ H, trans-H-4'), 5.28 (d, J = 5.0 Hz, $^{1}/_{2}$ H, cis-H-4'), 6.28 (d, J = 9.5 Hz, $^{1}/_{2}$ H, cis-H-3), 6.30 (d, J = 9.5

(±)-trans-3'-Acetoxy-4'-isovaleramidodihydroseselin (49): yield 58% (starting with 20.0 mg of 48); crystallization from methanol gave prisms; mp 244–245 °C; HR-FAB MS calcd for $C_{21}H_{25}NO_6Na$ 410.1579, found m/z 410.1577 (M + Na)+; positive FAB MS m/z 410 (M + Na)+; IR (KBr) 1750 (CO, lactone), 1735 (CO, ester), 1638 (CO, amide), 1608 (C=C) cm⁻¹; ¹H NMR δ 0.90, 0.95 (each d, J = 6.5 Hz, 3H, isovaleryl (CH₃)₂), 1.32, 1.42 (each s, 3H, 2'-(CH₃)₂), 1.75–2.09 (m, 1H, CH₃), 5.29 (t, J = 7.5 Hz, 1H, H-4'), 5.40 (d, J = 7.5 Hz, 1H, H-3'), 5.60 (d, J = 7.5 Hz, 1H, NH), 6.23 (d, J = 9.5 Hz, 1H, H-3), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.32 (d, J = 8.5 Hz, 1H, H-5), 7.59 (d, J = 9.5 Hz, 1H, H-4).

Procedure for Synthesizing 10, 18, 19. To a solution of 4 (150.0 mg, 0.57 mmol) and dry pyridine (1 mL) in anhydrous CH_2Cl_2 (5 mL) was added (S)-(+)- α -methylbutyric anhydride (1 mL, 5.02 mmol) at room temperature. The reaction mixture was stirred overnight and was then concentrated in vacuo. The residue was column chromatographed on silica gel to give 18 (12.0 mg), 19 (27.0 mg), and 10 (135.0 mg).

3',4'-Bis-O-((+)- α -methylbutyryl)-cis-khellactone (10): yield 55% (starting with 150.0 mg of 4); crystallization from acetone gave colorless prisms; mp 121–123 °C; HR-FAB MS calcd for $C_{24}H_{30}O_7Na$ 453.1889, found m/z 453.1889 (M + Na)+; positive FAB MS m/z 453 (M + Na)+; IR (KBr) 1755 (CO, lactone), 1730 (CO, ester), 1605 (C=C) cm⁻¹; ¹H NMR δ 0.90–0.96 (m, 6H, 2 x CH₂CH₃), 1.16–1.23 (m, 6H, 2 x CHCH₃), 1.41, 1.41, 1.45, 1.46 (each s, 3 /₂H, 2'-(CH₃)₂), 1.51–1.78 (m, 4H, 2 x CH₂), 2.34–2.43 (m, 2H, 2 x CH), 5.33 (t, J = 5.0 Hz, 1H, H-3'), 6.22 (d, J = 9.5 Hz, 1H, H-3), 6.57 (t, J = 5.0 Hz, 1H, H-4'), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.36 (d, J = 8.5 Hz, 1H, H-5), 7.58 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{24}H_{30}O_7H_2O$) C, H.

4'-O-((+)-α-Methylbutyryl)-cis-khellactone (18): yield 6% (starting with 150.0 mg of 4); amorphous; mp 168–169 °C; IR (KBr) 3520 (OH) 1710 (CO), 1610 (C=C) cm⁻¹; ¹H NMR δ 0.92–0.99 (m, 3H, CH₂CH₃), 1.20, 1.25 (each d, J=7.0 Hz, 3 /₂H, CHCH₃), 1.41–1.49 (m, 6H, 2'-(CH₃)₂), 1.50–1.81 (m, 4H, 2 x CH₂), 2.46, 2.51 (each d, J=7.0 Hz, 1 /₂H, CH), 2.80 (t, J=5.0 Hz, OH, D₂O exchangeable), 4.05 (m, 1H, H-3'), 6.23 (d, J=9.5 Hz, 1H, H-3), 6.38, 6.39 (each d, J=4.5 Hz, 1 /₂H, H-4'), 6.79 (d, J=8.5 Hz, 1H, H-6), 7.35 (d, J=8.5 Hz, 1H, H-5), 7.60 (d, J=9.5 Hz, 1H, H-4). Anal. (C₁₉H₂₂O₆) C, H.

3'-O-((+)-α-Methylbutyryl)-cis-khellactone (19): yield 14% (starting with 150.0 mg of 4); gum; HR-FAB MS calcd for $C_{19}H_{22}O_6Na$ 369.1314, found m/z 369.1314 (M + Na)⁺; positive FAB MS m/z 369 (M + Na)⁺; IR (KBr) 3480 (OH), 1730 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 0.93-0.98 (m, 3H, CH₂CH₃), 1.21 (d, J=7.0 Hz, 3H, CHCH₃), 1.40, 1.49 (each s, 3H, 2'-(CH₃)₂), 1.51-1.81 (m, 2H, CH₂), 2.46-2.55 (m, 1H, CH), 2.98-2.31 (m, 1H, OH, D₂O exchangeable), 5.14 (d, J=5.0 Hz, 1H, H-3'), 5.42 (s, br, 1H, H-4'), 6.26 (d, J=9.5 Hz, 1H, H-3), 6.78 (d, J=8.5 Hz, 1H, H-6), 7.34 (d, J=8.5 Hz, 1H, H-5), 7.64 (d, J=9.5 Hz, 1H, H-4').

General Procedure for Synthesizing 3'-O-Alkyl-4'-O-acyl-trans-khellactones (24, 25). To a mixture of 6 and Ag₂O (catalytic amount) in DMF was added a corresponding alkyl halide at 10–12 °C. After stirring for 2.5–4 h, the reaction mixture was kept in a refrigerator overnight. The catalyst was filtered, and the filtrate was washed with water and brine and then concentrated in vacuo. The residue was purified by column chromatography on silica gel to give the product.

3'-O-Methyl-4'-O-(m-chlorobenzoyl)-trans-khellactone (24): yield 94% (starting with 1.36 g of 6); amorphous; mp 173–175 ° C; IR (KBr) 1730 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.45, 1.53 (each s, 3H, 2'-(CH₃)₂), 3.50 (d, J = 2.5 Hz, 1H, H-3'), 3.67 (s, 3H, OMe), 6.20 (d, J = 9.5 Hz, 1H, H-3, 6.50 (d, J = 2.5 Hz, 1H, H-4'), 6.85 (d, J = 8.5 Hz, 1H, H-6), 7.33–7.40 (m, 2H, H-5 and benzoyl H-5"), 7.51–7.54 (m, 1H, H-4"), 7.61 (d, J = 9.5 Hz, 1H, H-4), 7.90–7.95 (m, 2H, H-2",6"). Anal. (C₂₂H₁₉ClO₆) C, H, Cl.

3'-O-Benzyl-4'-O-(m-chlorobenzoyl)-trans-khellactone (25): yield 56% (starting with 136.0 mg of 6); crystallization from acetone gave colorless needles; mp 168–170 ° C; IR (KBr) 1730 (CO, lactone), 1710 (CO, ester), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.42 (s, 6H, 2'-(CH₃)₂), 3.70 (d, J = 2.5 Hz, 1H, H-3'), 4.83, 4.97 (each d, J = 12.0 Hz, 1H, ArCH₂), 6.23 (d, J = 9.5 Hz, 1H, H-3), 6.60 (d, J = 2.5 Hz, 1H, H-4'), 6.87 (d, J = 8.5 Hz, 1H, H-6), 7.30–7.44 (m, 2H, H-5 and benzoyl H-5"), 7.52–7.55 (m, 1H, H-4"), 7.62 (d, J = 9.5 Hz, 1H, H-4), 7.91–7.97 (m, 2H, H-2",6"). Anal. (C₂₈H₂₃ClO₆) C, H, Cl.

3'-O-(2"'-Tetrahydropyranyl)-4'-O-(m-chlorobenzoyl)-trans-khellactones (26, 27). A solution of 6 (100.0 mg, 0.25 mmol) and tosic acid monohydrate (20.0 mg, 0.018 mmol) in 3,4-dihydro-2H-pyran (1 mL, 10.96 mmol) was stirred at room temperature for 30 min. The reaction mixture was neutralized by methanolic NaOMe and concentrated in vacuo. The residue was separated by preparative TLC to give 26 (56.0 mg) and 27 (66.0 mg) [R_f 0.17 and 0.22, respectively; hexane—ethyl acetate (3:1)].

26: yield 46% (starting from 100.0 mg of **6**); crystallization from acetone gave colorless needles; mp 216–217 ° C; IR (KBr) 1730 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.47, 1.48 (each s, 3H, 2'-(CH₃)₂), 1.56–1.76 (m, 6H, tetrahydropyranyl CH₂-3",4",5"), 3.52–3.58, 3.83–3.90 (m, 2H, CH₂-6"), 3.98 (d, J = 3.0 Hz, 1H, H-3'), 4.86–4.88 (m, 1H, CH-1"), 6.18 (d, J = 9.5 Hz, 1H, H-3), 6.71 (d, J = 3.0 Hz, 1H, H-4'), 6.83 (d, J = 8.5 Hz, 1H, H-6), 7.31–7.50 (m, 3H, H-5 and benzoyl H-4"',5"'), 7.58 (d, J = 9.5 Hz, 1H, H-4), 7.90–7.95 (m, 2H, H-2"',6"'). Anal. (C₂₆H₂₅ClO₇) C, H, Cl.

27: yield 54.5% (starting from 100.0 mg of **6**); crystallization from acetone gave colorless needles; mp 191–194 °C; IR (KBr) 1715 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.47, 1.56 (each s, 3H, 2'-(CH₃)₂), 1.54–1.78 (m, 6H, tetrahydropyranyl CH₂-2",3",4"), 3.58–3.62, 3.92–3.98 (m, 2H, CH₂-6"), 4.08 (d, J=2.0 Hz, 1H, H-3'), 5.17 (d, J=5.5 Hz, 1H, CH-1"), 6.21 (d, J=9.5 Hz, 1H, H-3), 6.39 (d, J=2.0 Hz, 1H, H-4'), 6.86 (d, J=8.5 Hz, 1H, H-6), 7.33–7.53 (m, 3H, H-5, benzoyl H-4",5"'), 7.61 (d, J=9.5 Hz, 1H, H-4), 7.88–7.94 (m, 2H, benzoyl H-2"',6"'). Anal. (C₂₆H₂₅ClO₇) C, H, Cl.

General Procedure for Synthesizing 3'-O-Alkyl-trans-khellactones (28-30). To a solution of a corresponding ester (24-27) in dioxane (6-15 mL) was added KOH (0.5 N, 7-20 mL) dropwise. After stirring for 15 min to 1 h, the reaction mixture was acidified with dilute HCl and extracted with CHCl₃ or ethyl acetate. The organic fraction was worked up as described above (in the synthesis of 8, 9) to afford the corresponding product.

3'-O-Benzyl-trans-khellactone (28): yield 78% (starting with 117.0 mg of 25); gum; HR-FAB MS calcd for $C_{21}H_{21}O_5$ 353.1389, found m/z 353.1389 (M + H)+; positive FAB MS m/z 727 (2M + Na)+, 375 (M + Na)+; IR (KBr) 3440 (OH), 1740 (CO), 1605 (C=C) cm⁻¹; ¹H NMR δ 1.33, 1.44 (each s, 3H, 2'-(CH₃)₂), 3.64 (d, J = 5.0 Hz, 1H, H-3'), 4.71, 4.95 (each d, J = 11.5 Hz, 1H, ArCH₂), 5.21 (d, J = 5.0 Hz, 1H, H-4'), 6.25 (d, J = 9.5 Hz, 1H, H-3), 6.78 (d, J = 8.5 Hz, 1H, H-6), 7.28-7.39 (m, 6H, H-5 and benzyl ArH-2",3",4",5",6"), 7.65 (d, J = 9.5 Hz, 1H, H-4).

3'-O-Methyl-trans-khellactone (29): yield 36% (starting with 100.0 mg of 24); amorphous; mp 184 °C sublimed; HR-FAB MS calcd for $C_{15}H_{17}O_5$ 277.1076, found m/z 277.1078; (M + H)+ Positive FAB MS m/z 575 (2M + Na)+, 299 (M + Na)+; IR (KBr) 3390 (OH), 1700 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.37, 1.46 (each s, 3H, 2'-(CH₃)₂), 3.39 (d, J = 4.5 Hz, 1H, H-3'), 3.60 (s, 3H, OMe), 5.12 (d, J = 4.5 Hz, 1H, H-4'), 6.25 (d, J = 9.5 Hz, 1H, H-3), 6.78 (d, J = 8.5 Hz, 1H, H-6), 7.31 (d, J = 8.5, 1H, H-5), 7.65 (d, J = 9.5 Hz, 1H, H-4). Anal. ($C_{15}H_{16}O_5$ -1/4H₂O) C, H.

3'-O-(2"-Tetrahydropyranyl)-trans-khellactone (30): yield 41% (starting with 850.0 mg of mixture of 26); amorphous; mp 126~130 °C; HR-FAB MS calcd for $C_{19}H_{22}O_6Na$ 369.1314, found m/z 369.1315 (M + Na)+; positive FAB MS m/z 369 (M + Na)+; IR (KBr) 3420 (OH), 1728 (CO), 1610 (C=C) cm⁻¹; ¹H NMR δ 1.29, 1.47 (each s, 3H, 2'-(CH₃)₂), 1.50~1.59, 1.81~1.85 (m, 6H, tetrahydropyranyl CH₂-2",3",4"), 3.59~3.62, 4.02~4.06 (m, 2H, CH₂-6"), 3.73 (d, J = 5.5 Hz, 1H, H-3'), 4.67 (d, J = 4.5 Hz, 1H, CH-1"), 5.10 (d, J = 5.5 Hz,

1H, H-4'), 6.55 (d, J=9.5 Hz, 1H, H-3), 6.77 (d, J=8.5 Hz, 1H, H-6), 7.31 (d, J=8.5 Hz, 1H, H-5), 7.63 (d, J=9.5 Hz, 1H, H-4). Anal. ($C_{19}H_{22}O_{6'}^{-1}/_4H_2O$) C, H.

4'-O-Isovaleryl-*trans***-khellactone** (35). A solution of 33 (48.0 mg, 0.11 mmol) in AcOH (20%, aqueous) was refluxed for 30 min. The reaction mixture was cooled and extracted with CHCl₃. The organic layer was worked up as described above (in the synthesis of **8**, **9**) to afford **35**: yield 46%; crystallization from methanol gave colorless prisms; mp 185–187 °C; IR (KBr) 3420 (OH), 1740 (CO, lactone), 1700 (CO, ester), 1605 (C=C) cm⁻¹; ¹H NMR δ 0.98, 1.00 (each d, J = 6.5 Hz, 3H, isovaleryl (CH₃)₂), 1.39, 1.46 (each s, 3H, 2'-(CH₃)₂), 2.11–2.25 (m, 1H, CH), 2.27–2.40 (m, 2H, CH₂), 3.13 (d, J = 4.5 Hz, 1H, OH), 3.91 (t, J = 4.5 Hz, 1H, H-3'), 6.09 (d, J = 4.5 Hz, 1H, H-4'), 6.23 (d, J = 9.5 Hz, 1H, H-3), 6.81 (d, J = 8.5 Hz, 1H, H-6), 7.35 (d, J = 8.5 Hz, 1H, H-5), 7.61 (d, J = 9.5 Hz, 1H, H-4). Anal. (C₁₉H₂₂O₆) C, H.

3',4'-Di-O-methyl-trans-khellactone (43). To a mixture of 6 (32.0 mg, 0.08 mmol) and NaH (12 mg, 0.5 mmol) in DMF (1 mL) was added MeI (0.5 mL, 8 mmol) at 0 ° C. After the mixture was stirred for 5 h, the temperature was raised to room temperature. The reaction mixture was then neutralized with dilute HCl and extracted with ethyl acetate. The organic fraction was worked up as described above (in the synthesis of 8, 9) to afford 43: yield 43%; crystallization from acetone gave colorless prisms; mp 164-166 ° C; IR (KBr) 1720 (CO), 1605 (C=C) cm⁻¹; ¹H NMR δ 1.41, 1.49 (each s, 3H, 2'-(CH₃)₂), 3.35 (d, J=2.5 Hz, 1H, H-3'), 3.53 (s, 3H, OMe-3'), 3.73 (s, 3H, OMe-4'), 4.57 (d, J=2.5 Hz, 1H, H-4'), 6.24 (d, J=9.5 Hz, 1H, H-3), 6.76 (d, J=8.5 Hz, 1H, H-6), 7.29 (d J=8.5 Hz, 1H, H-6), 7.61 (d, J=9.5 Hz, 1H, H-4). Anal. (C₁₆H₁₈O₅) C, H

Mixture of (\pm) -trans- and (\pm) -cis-3'-Hydroxy-4'-azi-dodihydroseselins (44). To a solution of 4 (100.0 mg, 0.38mmol) and NaN₃ (50.0 mg, 0.76 mmol) in CHCl₃ (6 mL) was added trifluoroacetic acid (3 mL) dropwise at room temperature. After the mixture was stirred overnight, additional NaN₃ (25 mg, 0.38 mmol) was added, and the reaction was continued for another night. The reaction mixture was washed with water and brine, dried over Na₂SO₄, and concentrated on a rotary evaporator. The residue was purified by preparative TLC to give 44: yield 92%; amorphous; 162 °C dec; HR-FAB MS calcd for $C_{14}H_{14}N_3O_4$ 288.0984, found m/z 288.0984 (M + Na)+; positive FAB MS m/z 597 (2M + Na)+, 310 (M + Na)+; IR (KBr) 3540 (OH), 2100 (N₃), 1720 (CO), 1608 (C=C) cm⁻¹; ¹H NMR δ 1.37, 1.48 (each s, 3H, 2'-(CH₃)₂), 2.31 (d, J = 5.5Hz, $^{3}/_{4}$ H, trans-OH, D₂O exchangeable), 2.57 (d, J = 8.9 Hz, $^{1}/_{4}$ H, cis-OH, D₂O exchangeable, 3.82 (t, J = 5.5 Hz, $^{3}/_{4}$ H, trans-H-3'), 3.94 (dd, J = 8.9 Hz, J = 5.5 Hz, $\frac{1}{4}$ H, H-3'), 4.82 $(d, J = 5.5 \text{ Hz}, 1H, trans-H-4'), 5.17 (d, J = 5.5 \text{ Hz}, \frac{1}{4}H, cis-$ H-4'), 6.28 (d, J = 9.5 Hz, $^{1}/_{4}$ H, cis-H-3), 6.30 (d, J = 9.5 Hz, $^{3}/_{4}H$, trans-H-3), 6.78 (d, J = 8.5 Hz, $^{1}/_{4}H$, cis-H-6), 6.79 (d, J $= 8.5 \text{ Hz}, \frac{3}{4}\text{H}, trans-\text{H-6}), 7.35 (d, J = 8.5 \text{ Hz}, \frac{3}{4}\text{H}, trans-\text{H-6})$ 5), 7.35 (d, J = 8.5 Hz, $\frac{1}{4}$ H, cis-H-5), 7.63 (d, J = 9.5 Hz, 1H, H-4). Anal. $(C_{14}H_{13}N_3O_4\cdot 1/8H_2O)$ C, H, N.

 (\pm) -cis-3'-Hydroxy-4'-acetamidodihydroseselin (46) and (\pm) -trans-3'-Hydroxy-4'-isovaleramidodihydroseselin (48). A mixture of 45 (140.0 mg, 0.48 mmol) and Pd/CaCO₃ (90.0 mg, poisoned with Pb) in EtOH (5 mL) was stirred under an H₂ atmosphere for 2 days. The catalyst was filtered, and the filtrate was concentrated and dried in vacuo. The residue was dissolved in pyridine (0.1 mL) and CH₂Cl₂ (5 mL), and then isovaleryl chloride (0.1 mL, 0.8 mmol) was added. After 16 h, the reaction mixture was extracted with CHCl₃, and the organic fraction was worked up as described above (in the synthesis of 8, 9) to afford 46 (46.0 mg) and 48 (34.0 mg).

46: yield 31%; crystallization from EtOH gave prisms; mp 260-262 °C; HR-FAB MS calcd for $C_{16}H_{17}NO_5Na$ 326.1004, found m/z 326.1011 (M + Na)+; positive FAB MS m/z 326 (M + Na)+; IR (KBr) 3360 (OH), 3300 (NH), 1730 (CO), 1650 (CO, amide), 1615 (C=C) cm⁻¹; ¹H NMR δ 1.39, 1.47 (each s, 3H, 2'-(CH₃)₂), 2.15 (s, 3H, COCH₃), 3.88 (d, J = 4.5 Hz, 1H, H-3'), 5.57 (d, J = 4.5 Hz, 1H, H-4'), 5.90 (d, J = 7.5 Hz, 1H, NH), 6.24 (d, J = 9.5 Hz, 1H, H-3), 6.80 (d, J = 8.5 Hz, 1H, H-6), 7.32 (d, J = 8.5 Hz, 1H, H-5), 7.60 (d, J = 9.5 Hz, 1H, H-4).

48: yield 34%; crystallization from EtOH gave prisms; mp

Acknowledgment. This investigation was supported by grant AI-33066 from the National Institute of Allergies and Infectious Diseases awarded to K. H. Lee.

232–234 °C; IR (KBr) 3520 (OH), 3270 (NH), 1720 (CO), 1655 (CO, amide), 1608 (C=C) cm⁻¹; ¹H NMR δ 0.95–1.03 (m, 6H, isovaleryl (CH₃)₂), 1.26, 1.52 (each s, 3H, 2'-(CH₃)₂), 2.06–2.30 (m, 3H, isovaleryl CH₂ and CH), 3.99 (d, J = 7.0 Hz, 1H, H-3'), 5.08 (dd, J = 7.0 Hz, J = 5.0 Hz, 1H, H-4'), 5.31 (s, 1H, OH, D₂O exchangeable), 5.91 (d, J = 5.0 Hz, 1H, NH, D₂O exchangeable), 6.25 (d, J = 9.5 Hz, 1H, H-3), 6.81 (d, J = 8.5 Hz, 1H, H-6), 7.34 (d, J = 8.5 Hz, 1H, H-5), 7.62 (d, J = 9.5 Hz, 1H, H-4). Anal. (C₁₉H₂₃NO₅) C, H, N.

Anti-HIV Assay. The T cell line, H9, and the promonocytic cell line, U937, were maintained separately in continuous culture with complete medium (RPMI 1640 with 10% fetal calf serum [FCS]) at 5% CO2 and 37 ° C. Cell lines were used in experiments only when in log phase of growth, whereas uninfected peripheral blood mononuclear cells (PBMCs) were first stimulated with PHA (1 μ g/mL) for 3 days. All cell targets were incubated with HIV-1 (IIIB isolated, TCID₅₀ 10⁴ IU/mL, at a multiplicity of infection of 0.1-0.01 IU/cell) for 1 h at 37 ° C and 5% CO2. The cell lines and PBMCs were washed thoroughly to remove unadsorbed virions and resuspended at 4×10^5 cells/mL in complete medium or complete medium with 10% v/v interleukin 2 (IL-2), respectively. Aliquots (1 mL) were placed in wells of 24-well culture plates containing an equal volume of test compound (diluted in the appropriate culture medium). Each test compound had its toxicity assessed by determining the number of compound-exposed uninfected cells that remained after 4 days at 37 °C and 5% CO2. A p24 antigen ELISA assay was used to determine the level of virus released in the medium of the HIV-infected cultures. The p24 antigen assay uses a HIV-1 anti-p24 specific monoclonal antibody as the capture antibody coated on 96well plates. Following a sample incubation period, rabbit serum containing antibodies for HIV-1 p24 is used to tag any p24 "captured" onto the microtiter well surface. Peroxidase conjugated goat anti-rabbit serum is then used to tag HIV-1 p24 specific rabbit antibodies that have complexed with captured p24. The presence of p24 in test samples is then revealed by addition of substrate. The cut-off for the p24 ELISA assay is 12.5 pg/mL. P24 in the culture medium was quantitated against a standard curve containing known amounts of p24. The effective (EC₅₀) and inhibitory (IC₅₀) concentrations for anti-HIV activity and cytotoxicity, respectively, were determined.

HIV-1 Reverse Transcriptase Assay. HIV-1 reverse transcriptase microassay was adapted from refs 15 and 16. Briefly, 10 mL of virion associated HIV-1 IIIB reverse transcriptase in 1% Triton X-100 was mixed with 50 mL of a reaction cocktail containing 50 mM Tri-HCl (pH 7.8), 75 mM KCl, 2 mM dithiothreitol, 5 mM MgCl2, poly(A) (5 mg/mL; Pharmacia), oligo(dT) (0.25 unit/mL; Pharmacia), 0.05% Nonidet P40, and [32P]dTTP (10 mCi/mL) in the presence of various concentrations of test compounds. After incubating 1 h at 37 °C, 40 mL of the reaction mixture was applied to a Schleicher & Schuell NA 45 membrane saturated with 2 x SSC (0.3 M NaCl, 30 mM sodium citrate, pH 7.0) in a Schleicher & Schuell Minifold over one sheet of GB003 filter paper. Each well of the minifold was washed four times with 2 x SSC. Autoradiography was performed, and radioactivity was quantified with a Packard Matrix (Meriden, CT) 9600 direct β counter.

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