

Antitumor Activity of 5-Aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines

William J. Houlihan,^{*,†} Paul G. Munder,^{*,‡} Dean A. Handley, Seung H. Cheon, and Vincent A. Parrino

Preclinical Research, Sandoz Research Institute, Sandoz Pharmaceuticals Corporation, East Hanover, New Jersey 07936, and Max-Planck-Institut für Immunbiologie, 78 Freiburg-Zaehringen, Germany

Received September 20, 1994[®]

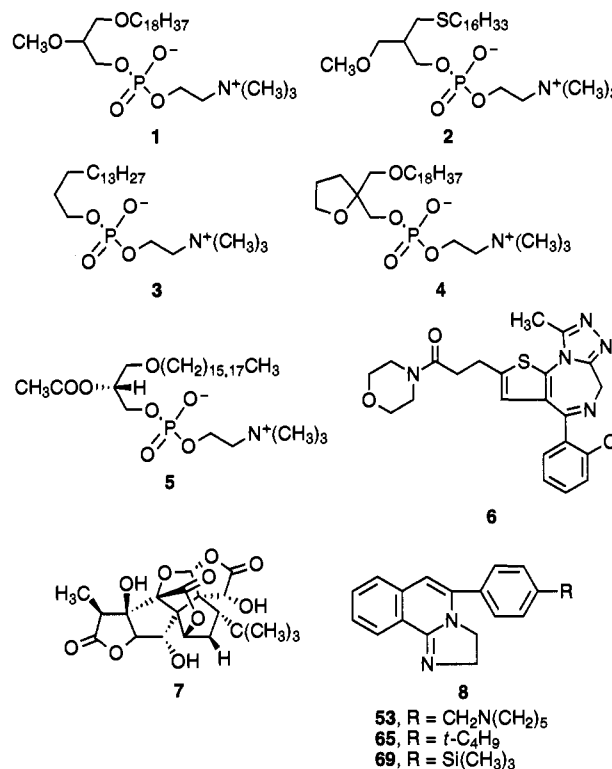
A series of 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines previously reported to be platelet activating factor (PAF) receptor antagonists were evaluated for potential antitumor activity. Several compounds, such as the 5-(4'-*tert*-butylphenyl) (**65**), 5-[4'-(trimethylsilyl)phenyl] (**69**), and 5-(4'-cyclohexylphenyl) (**71**) analogs showed very good cytotoxicity against several tumor cell lines. 5-[4'-(Piperidinomethyl)phenyl]-2,3-dihydroimidazo[2,1-*a*]isoquinoline (SDZ 62-434, **53**) was more effective on a milligram per kilogram basis than the clinical cytostatic agent edelfosine (**1**) in increasing survivors and decreasing tumor volume in the oral mouse Meth A fibrosarcoma assay. It was selected for further development and is currently in phase I clinical trials in cancer patients.

Introduction

A number of phospholipids, such as edelfosine¹ (ET-18-OCH₃, **1**), ilmofosine² (BM41-440, **2**), miltefosine³ (**3**), and (±)-2-[[hydroxy[[tetrahydro-2-[(octadecyloxy)methyl]furan-2-yl]methoxy]phosphinyl]oxy]-*N,N,N*-trimethylethanaminium hydroxide, inner salt⁴ SRI62-834, **4**), that are structurally related to platelet activating factor^{5,6} (PAF, **5**) have been shown to exhibit antitumor activity against a variety of murine and human tumor cell lines and in murine tumor models.⁷ These phospholipids exert a wide range of pharmacological activities such as direct cytotoxic effects against tumor cells,⁸ macrophage activation,⁹ changes in membrane structure and permeability,^{10,11} alteration of signal transduction,^{12,13} and the rate of uptake by endocytosis^{14,15} that may contribute to their antitumor activity. No direct correlation between direct or macrophage-induced cytotoxic effects and binding to the PAF receptor or other PAF-induced effects has been reported.¹⁶⁻¹⁸ Furthermore, the cytotoxic effects of edelfosine (**1**) and **4** on most human tumor cell lines could not be blocked by PAF receptor antagonists.¹⁹⁻²²

In our laboratories we have been evaluating the potential antitumor activity of noncharged PAF receptor antagonists, that is, those that do not contain a phospholipid moiety. We previously reported that there is no apparent relationship between PAF receptor antagonist activity and direct or macrophage-induced tumor cytotoxicity. Very potent noncharged PAF receptor antagonists such as the *s*-triazolo[3,4-*c*]thieno[2,3-*e*][1,4]-diazepine (**6**), WEB 2086 and ginkgolide B (**7**) failed to exhibit any activity when examined in several tumor cell lines.²³ However, one series of noncharged PAF receptor antagonists, the 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines^{24,25} (**8**), gave antitumor activity, with the 4'-(trimethylsilyl)^{23,26} (**69**), 4'-*tert*-butyl²⁶ (**65**), and 4'-(piperidinomethyl)²⁶⁻²⁹ (**53**) analogs displaying an *in vitro* cytotoxicity profile similar to edelfosine (**1**).

In this paper we report on the *in vitro* and *in vivo* antitumor activity of a series of 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines.



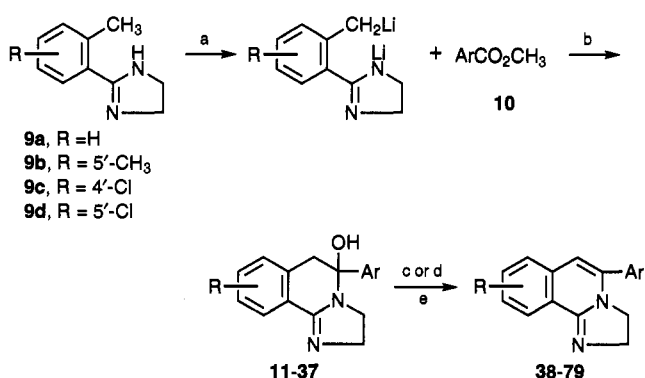
Chemistry

The synthesis of the 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines was carried out by the procedures given in Scheme 1. The 2-aryl-2(*1H*)-imidazolines **9a-d**, prepared from the appropriate benzonitrile and (2-aminoethyl)ammonium *p*-toluenesulfonate, were lithiated^{30,31} to the dilithio derivatives and then treated with a methyl aryl ester **10** to give the 5-aryl-2,3,5,6-tetrahydroimidazo[2,1-*a*]isoquinolin-5-ols **11-37** (Table 1). Dehydration to the 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines **38-79** (Table 2) was accomplished by refluxing in toluene in the presence of a catalytic amount of *p*-toluenesulfonic acid or refluxing acetic acid. When the 3',4'-dimethoxybenzyl ether **11** was dehy-

[†] Present address: The Charles A. Dana Research Institute, Drew University, Madison, NJ 07900-4000.

[‡] Max-Planck-Institut.

[®] Abstract published in *Advance ACS Abstracts*, January 1, 1995.

Scheme 1^a

- 9a, R = H
 9b, R = 5'-CH₃
 9c, R = 4'-Cl
 9d, R = 5'-Cl

^a Reagents/conditions: (a) *n*-BuLi, THF, 35 °C, 4 h; (b) -20±5 to 0 °C, *ca.* 4 h; (c) HOAc, reflux, 5 h; (d) *p*-toluenesulfonic acid, toluene, reflux; (e) HCl, EtOH.

drated, cleavage of the ether occurred to give the 4'-hydroxyphenyl derivative **46**.

Results and Discussion

In Vitro Studies. The direct cytotoxicity on Abelson 8.1 tumor cells for the 5-aryl-2,3-dihydroimidazo[2,1-a]-isoquinolines **38–79** at doses of 1, 3, and 5 μg/mL is given in Table 3.

Introduction of one or more F or Cl atoms (**39–45**), an OH (**46**), one to three OCH₃ groups (**47–49**), or a OCH₂O (**50**) failed to increase the cytotoxicity at the lowest dose (1 μg) relative to the unsubstituted 5-phenyl

compound **38**. At the highest dose (5 μg), the 4'-Cl and 2',4'- and 3',4'-Cl₂ derivatives **43–45** appeared to show increased cytotoxicity relative to **38** based on reduction of viable cells.

Several members of the 4'-substituted aminomethyl-ene analogs **51–56** (type B, Table 3) showed a good level of cytotoxicity at both the 3 and 5 μg/mL doses with the diallylamino (**51**), pyrrolidino (**52**), piperidino (**53**), 2-methylpiperidino (**54**), and thiomorpholino (**56**) analogs giving a profile of activity similar to edelfosine (**1**).

A progressive increase in cytotoxicity was found as the size of the 4'-alkyl group increased from CH₃ to *t*-C₄H₉ (**58, 60–65**) with 3'- and 4'-*t*-C₄H₉ having nearly identical activity at all doses. Addition of a Cl at the 8- or 9-position or a CH₃ at the 9-position (**66–68**) of the 4'-*tert*-butyl derivative **65** gave compounds of similar activity. Replacing the CH₃ by the CF₃ group (**59**) resulted in loss of activity while substitution of the 4'-*t*-C₄H₉ by the lipophilic Si(CH₃)₃ (**69**), cyclopentyl (**70**), cyclohexyl (**71**), or phenyl (**72**) groups resulted in similar or slightly improved cytotoxicity.

Replacement of the phenyl group in **38** by 2'-furyl (**73**), 2'-thienyl (**74**), 1'- or 2'-naphthyl (**77, 78**) or tetramethyltetralin **79**, which can be regarded as a cyclic mimic of a 3',4'-di-*tert*-butylphenyl group, gave an increase in cytotoxicity, especially at the 3 and 5 μg/mL doses.

These findings suggest that introduction of a lipophilic group (cf. **60, 64, 69, 70**, etc.) in the phenyl ring of **38**

Table 1. 5-Aryl-2,3,5,6-tetrahydroimidazo[2,1-a]isoquinolin-5-ols (11–37)

no. ^a	type	R or NRR'	yield, %	mp, °C (recryst solv) ^b	formula ^d	anal. ^c
11	A	4'-OCH ₂ C ₆ H ₃ 3,4(OCH ₃) ₂	36	144–147 (A)	C ₂₆ H ₂₆ N ₂ O ₄	C,H,N
12	A	3',4'-(OCH ₃) ₂	46	194–196 (A)	C ₁₉ H ₂₀ N ₂ O ₃	C,H,N
13	A	3',4',5'-(OCH ₃) ₃	40	152 (B)	C ₂₀ H ₂₂ N ₂ O ₄	C,H,N
14	B	N(CH ₂ CH=CH ₂) ₂	36	150–151 (C)	C ₂₄ H ₂₇ N ₃ O	C,H,N
15	B	N(CH ₂) ₄	37	171–173 (D)	C ₂₂ H ₂₅ N ₃ O	C,H,N
16	B	N(CH ₂) ₆	43	168–170 (E)	C ₂₃ H ₂₇ N ₃ O	C,H,N
17	B	(±)-2-methylpiperidyl	42	oil	C ₂₄ H ₂₉ N ₃ O	C,H,N
18	B	morpholino	61	167–169 (C)	C ₁₉ H ₂₆ N ₃ O	C,H,N
19	B	thiomorpholino	49	oil	C ₁₉ H ₂₆ N ₃ OS	- ^d
20	B	N(C ₆ H ₁₁) ₂	45	180 (C)	C ₃₀ H ₃₉ N ₃ O	C,H,N
21	A	4'-C ₂ H ₅	48	177–179 (A)	C ₁₉ H ₂₀ N ₂ O	C,H,N
22	A	4'- <i>n</i> -C ₃ H ₇	26	183–185	C ₂₆ H ₂₂ N ₂ O	C,H,N
23	A	4'- <i>i</i> -C ₃ H ₇	56	179–181 (F)	C ₂₀ H ₂₂ N ₂ O	C,H,N
24	A	4'- <i>n</i> -C ₄ H ₉	43	179–180 (G)	C ₂₁ H ₂₄ N ₂ O	C,H,N
25	A	3'- <i>t</i> -C ₄ H ₉	42	172–174 (C)	C ₂₁ H ₂₄ N ₂ O	C,H,N
26	A	4'- <i>t</i> -C ₄ H ₉	48	187–188 (C)	C ₂₁ H ₂₄ N ₂ O	C,H,N
27	A	4'- <i>t</i> -C ₄ H ₉ ,8-Cl	46	198 (D)	C ₂₁ H ₂₃ ClN ₂ O	C,H,Cl,N
28	A	4'- <i>t</i> -C ₄ H ₉ ,9-Cl	38	189 (C)	C ₂₁ H ₂₃ ClN ₂ O	C,H,Cl,N
29	A	4'- <i>t</i> -C ₄ H ₉ ,9-CH ₃	35	168 (C)	C ₂₂ H ₂₆ N ₂ O	C,H,N
30	A	4'-Si(CH ₃) ₃	44	163 (C)	C ₂₀ H ₂₄ N ₂ O _{Si}	C,H,N
31	A	4'-cyclopentyl	37	198–200 (A)	C ₂₃ H ₂₆ N ₂ O	C,H,N
32	A	4'-cyclohexyl	35	214 (A)	C ₂₃ H ₂₆ N ₂ O	C,H,N
33	A	4'-C ₆ H ₅	41	167–169 (H)	C ₂₃ H ₂₀ N ₂ O	C,H,N
34	C	2'-furyl	46	160–162 (G)	C ₁₆ H ₁₄ N ₂ O ₂	C,H,N
35	C	1'-naphthyl	38	202 (H)	C ₂₁ H ₁₈ N ₂ O	C,H,N
36	C	2'-naphthyl	75	214 (H)	C ₂₁ H ₁₈ N ₂ O	C,H,N
37	C	1,1,4,4-tetramethyl-1,2,3,4-tetrahydronaphth-6'-yl	38	182–184 (G)	C ₂₆ H ₃₀ N ₂ O	C,H,N

^a The physical properties and ²H-NMR of type A where R = H, 2'-F, 4'-F, 2'-Cl, 3'-Cl, 4'-Cl, 2',4'-Cl₂, 3',4'-Cl₂, 4'-OCH₃, 3',4'-OCH₂O-, 4'-CH₃, 3'-CF₃ and type C where R = 2'-thiophenyl, 3'-pyridyl and 4'-pyridyl are given in ref 31. ^b Recrystallization solvents: A, EtOH; B, Et₂O-petroleum ether; C, CH₂Cl₂-pentane; D, CH₂Cl₂-petroleum ether; E, CH₂Cl₂; f, Et₂O-MeOH; G, EtOH-pentane; H, CH₂Cl₂-EtOH. ^c Unless otherwise stated, the analysis are within ±0.4% of the theoretical values. ^d All compounds had ¹H-NMR and MS consistent with assigned structures. Some representative ¹H-NMR are given in ref 41.

Table 2. Physical Properties of Compounds Listed in Table 3

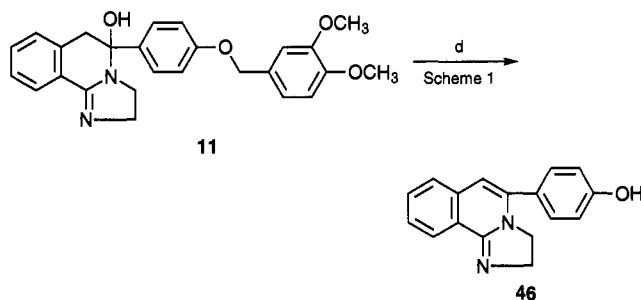
no.	method, yield, %	mp, °C (recryst solv) ^a	formula ^c	anal. ^b
38	D, 65	>250 (A)	C ₁₇ H ₁₄ N ₂ HCl	C, H, Cl, N
39	D, 70	>250 (A)	C ₁₇ H ₁₃ FN ₂ HCl	C, H, Cl, N
40	D, 62	150–151 (B)	C ₁₇ H ₁₃ FN ₂	C, H, N
41	D, 62	134–136 (B)	C ₁₇ H ₁₃ ClN ₂	C, H, Cl, N
42	D, 75	>250 (A)	C ₁₇ H ₁₃ ClN ₂ HCl	C, H, Cl, N
43	D, 88	>250 (A)	C ₁₇ H ₁₃ ClN ₂ HCl	C, H, Cl, N
44	D, 60	150–152 (B)	C ₁₇ H ₁₂ Cl ₂ N ₂	C, H, Cl, N
45	D, 53	158–160 (C)	C ₁₇ H ₁₂ Cl ₂ N ₂	C, H, Cl, N
46	D, 68	>260 (D)	C ₁₇ H ₁₄ N ₂ O·2H ₂ O	C, H, N
47	D, 40	126–128 (C)	C ₁₈ H ₁₆ N ₂ O	C, H, N
48	D, 45	158–160 (C)	C ₁₉ H ₁₈ N ₂ O ₂	C, H, N
49	D, 46	>250 (D)	C ₂₀ H ₂₀ N ₂ O ₃ HCl	C, H, Cl, N
50	D, 53	128–130 (B)	C ₁₈ H ₁₃ N ₂ O ₂	C, H, N
51	D, 72	228 (E)	C ₂₄ H ₂₅ N ₃ ·2HCl	C, H, Cl, N
52	D, 60	106–108 (F)	C ₂₂ H ₂₃ N ₃	C, H, N
53	D, 75	>250 (E)	C ₂₃ H ₂₅ N ₃ ·2HCl	C, H, Cl, N
54	D, 63	>250 (E)	C ₂₄ H ₂₇ N ₃ ·2HCl	C, H, Cl, N
55	D, 85	160–162 (C)	C ₂₂ H ₂₃ N ₃ O	C, H, N
56	D, 83	275–278 (D)	C ₂₂ H ₂₃ N ₃ S·2HCl	C, H, Cl, N, S
57	D, 80	273 (E)	C ₃₀ H ₃₇ N ₃ ·2HCl	C, H, Cl, N
58	D, 86	>250 (A)	C ₁₈ H ₁₆ N ₂ HCl	C, H, Cl, N
59	D, 70	>250 (A)	C ₁₈ H ₁₃ F ₃ N ₂ HCl	C, H, Cl, N
60	E, 75	>275 (E)	C ₁₉ H ₁₈ N ₂ HCl	C, H, Cl, N
61	E, 63	>260 (E)	C ₂₀ H ₂₀ N ₂ HCl	C, H, Cl, N
62	E, 52	>270 (E)	C ₂₀ H ₂₀ N ₂ HCl	C, H, Cl, N
63	E, 81	272–274 (E)	C ₂₁ H ₂₂ N ₂ HCl	C, H, Cl, N
64	E, 75	>275 (E)	C ₂₁ H ₂₂ N ₂ HCl	C, H, Cl, N
65	E, 82	>250 (E)	C ₂₁ H ₂₂ N ₂ HCl	C, H, Cl, N
66	E, 84	>250 (E)	C ₂₁ H ₂₁ ClN ₂ HCl	C, H, Cl, N
67	E, 65	>275 (E)	C ₂₁ H ₂₁ ClN ₂ HCl	C, H, Cl, N
68	E, 65	>275 (E)	C ₂₂ H ₂₄ N ₂ HCl	C, H, Cl, N
69	E, 74	>275 (E)	C ₂₀ H ₂₂ N ₂ Si·HCl	C, H, Cl, N
70	E, 39	>270 (E)	C ₂₂ H ₂₂ N ₂ HCl	C, H, Cl, N
71	E, 80	>270 (A)	C ₂₃ H ₂₄ N ₂ HCl	C, H, Cl, N
72	E, 58	>250 (E)	C ₂₃ H ₁₈ N ₂ HCl	C, H, N
73	D, 47	>315 (E)	C ₁₅ H ₁₂ N ₂ O·HCl	C, H, Cl, N
74	D, 46	98–100 (B)	C ₁₅ H ₁₂ N ₂ S	C, H, N
75	D, 58	>250 (E)	C ₁₆ H ₁₃ N ₃ HCl	C, H, Cl, N
76	D, 47	>250 (E)	C ₁₆ H ₁₃ N ₃ HCl	C, H, Cl, N
77	E, 70	>250 (E)	C ₂₁ H ₁₆ N ₂ HCl	C, H, Cl, N
78	E, 56	>250 (E)	C ₂₁ H ₁₆ N ₂ HCl	C, H, Cl, N
79	E, 81	>250 (A)	C ₂₅ H ₂₈ N ₂ HCl	C, H, Cl, N

^a Recrystallization solvents: A, CH₂Cl₂–EtOH; B, Et₂O–pentane; C, Et₂O; D, EtOH–Et₂O; E, EtOH; F, CH₂Cl₂–pentane.
^b Unless otherwise stated, the analyses are within ±0.4% of the theoretical values. ^c Satisfactory mass spectrum and ¹H-NMR were obtained for all compounds. Some representative ¹H-NMR are given in ref 41.

leads to more of an increase in cytotoxicity than adding one or more electron-rich substituents (cf. **40**, **44**, **48**, **49**). A straight or branched alkyl group or a cycloalkyl group with four to six carbon atoms (cf. **63**, **65**, **70**, **71**) or a phenyl group (**72**) confers good cytotoxicity while an increase in the lipophilic bulk to 10 or 14 carbon atoms (**78**, **79**) leads to a decline in cytotoxicity.

Several of the compounds that showed good cytotoxicity on the Abelson 8.1 tumor cells were evaluated on the YAC-1 tumor cell line (Table 4). The cytotoxicity on the YAC-1 tumor cells parallels that found on the Abelson 8.1 tumor cells. The 4'-*n*-C₄H₉ (**63**), 3'- and 4'-*t*-C₄H₉ (**64**, **65**), and 4'-cyclohexyl (**71**) analogs increased cytotoxicity relative to that of edelfosine (**1**) at 1 μg/mL.

The inhibition of [³H]PAF receptor binding on human platelets for a representative grouping of 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines is given in Table 3. Best inhibition (IC₅₀ 1–3 μM) occurred with the 4'-*t*-C₄H₉ (**65**) thiomorpholinomethyl (**56**), 4'-*t*-C₄H₉, 8-Cl (**66**), and morpholinomethyl (**55**) derivatives. No obvious correlations between these activities and direct cytotoxicity exist. A most striking lack of correlation



is the 4'-*t*-C₉H₉ (**65**), 4'-*t*-C₄H₉, 8-Cl (**66**), 4'-*t*-C₄H₉, 9-Cl (**67**), and 4'-*t*-C₄H₉, 9-CH₃ (**68**) analogs that have about the same cytotoxicity (2.0–4.2% viable cells at 1 μg/mL) but have IC₅₀ values of 0.9, 2.2, 40.4, and 31.8 μM respectively.

In Vivo Study. The 4'-piperidinomethyl (**53**) and 4'-*t*-C₄H₉ (**65**) analogs, representatives of the more cytotoxic polar and lipophilic compounds, were evaluated orally (po) in the mouse Meth A fibrosarcoma tumor model (Table 5). At a daily dose of 5 μg/mouse/day (ca. 0.25 mg/kg) both compounds showed a positive effect in increasing the number of survivors and decreasing tumor volume relative to controls. When the dose was increased to 50 μg/mouse/day (2.5 mg/kg), compound **53** gave a 100% (10/10 mice) survivor rate with all mice being tumor free, while **65** was less effective at the 5 μg/mouse dose in both the number of survivors and tumor volume on day 28 after tumor inoculation. In comparison with edelfosine (**1**), compound **53** appears to be more effective in both decreasing tumor size and increasing survivors.

Conclusion

The 5-aryl-2,3-dihydroimidazo[2,1-*a*]isoquinolines, originally designed as PAF receptor antagonists, also represent a novel class of potential antitumor agents.

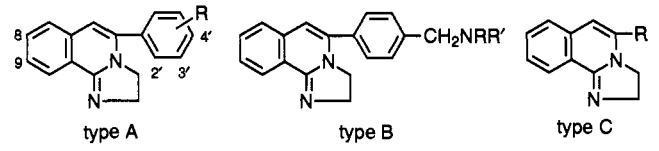
Several members, such as the 3'- and 4'-*t*-C₄H₉, 4'-Si(CH₃)₃, 4'-cyclopentyl, and 4'-cyclohexylphenyl (**64**, **65**, **69**–**71**), gave an *in vitro* cytotoxicity against two tumor cell lines that was considerably more effective than edelfosine (**1**).

The 4'-piperidinomethyl analog **53** was more effective, on a milligram per kilogram basis, than edelfosine (**1**) in increasing survivors and decreasing tumor volume in the oral mouse Meth A fibrosarcoma assay.

Compound **53** (SDZ 62–434) has now entered phase I clinical trials under the auspice of the Cancer Research Campaign (CRC) of Great Britain. Further investigations to profile the potential antitumor activity of (**53**) have been completed,³² and additional studies on the 4'-*t*-C₄H₉ analog **65** are in progress. No obvious correlations between inhibition of [³H] PAF binding to human platelets and direct tumor cell cytotoxicity were found.

Experimental Section

General. Melting points were determined on a Thomas-Hoover melting point apparatus and are uncorrected. Nuclear magnetic resonance (NMR) data for ¹H-NMR were taken on JEOL-FX-90 (90 MHz) or Bruker (200 MHz) spectrometers and are reported in δ (ppm) downfield from tetramethylsilane (TMS). Mass spectra (MS) were obtained on a Finnigan MAT 4600 GC/MS instrument applying a desorption chemical ionization method using ammonia (or isobutane) as the reagent gas. Elemental analyses for carbon, hydrogen, and nitrogen were determined with a Carlo Erba Instruments Model

Table 3. Direct Cytotoxicity on Abelson 8.1 Tumor Cells and Inhibition of [³H]PAF Binding to Human Platelet Receptor


no. ^a	type	R or NRR'	% viable cells at 72 h, ^{b,c} μg/mL			[³ H]PAF binding IC ₅₀ μM ^d
			1	3	5	
38	A	H	90	70	48	11.2
39	A	2'-F	85	62	31	5.7
40	A	4'-F	82	58	30	14.2
41	A	2'-Cl	72	17	4	8.1
42	A	3'-Cl	85	60	30	5.4
43	A	4'-Cl	80	19	1	37.2
44	A	2',4'-Cl ₂	68	5.0	1.2	82.0
45	A	3',4'-Cl ₂	68	5.3	1.4	17.0
46	A	4'-OH	80	68	53	7.2
47	A	4'-OCH ₃	72	28	25	55.0
48	A	3',4'-(OCH ₃) ₂	78	59	37	22.0
49	A	3',4',5'-(OCH ₃) ₃	110	124	120	>100
50	A	3',4'-OCH ₂ O-	68	32	10	19.0
51	B	N(CH ₂ CH=CH ₂) ₂	39	2.0	1.2	9.2
52	B	N(CH ₂) ₄	62	10	1.3	59.4
53	B	N(CH ₂) ₅	72	12	3	7.5
54	B	(±)-2-methylpiperidyl	51	3.1	2.4	29.4
55	B	morpholino	73	46	28	2.8
56	B	thiomorpholino	51	3.1	2.4	1.1
57	B	N(C ₆ H ₁₁) ₂	100	80	51	13.1
58	A	4'-CH ₃	81	38	7	5.4
59	A	3'-CF ₃	92	50	23	19.2
60	A	4'-C ₂ H ₅	36	3.0	2.0	6.3
61	A	4'- <i>n</i> -C ₃ H ₇	2.8	1.5	1.3	nt
62	A	4'- <i>i</i> -C ₃ H ₇	4.6	1.4	1.4	nt
63	A	4'- <i>n</i> -C ₄ H ₉	2.0	1.4	1.3	nt
64	A	3'- <i>t</i> -C ₄ H ₉	2.0	0.5	0.5	4.0
65	A	4'- <i>t</i> -C ₄ H ₉	3.3	1.4	1.3	0.9
66	A	4'- <i>t</i> -C ₄ H ₉ ,8-Cl	3.7	1.7	1.5	2.2
67	A	4'- <i>t</i> -C ₄ H ₉ ,9-Cl	4.2	2.0	1.9	40.4
68	A	4'- <i>t</i> -C ₄ H ₉ ,9-CH ₃	2.0	2.0	2.0	31.8
69	A	4'-Si(CH ₃) ₃	2.0	0.5	0.5	6.5
70	A	4'-cyclopentyl	1.5	1.3	1.2	nt
71	A	4'-cyclohexyl	1.4	1.3	1.3	nt
72	A	4'-C ₆ H ₅	2.1	1.6	1.4	3.4
73	C	2'-furyl	24	3.1	2.3	100
74	C	2'-thienyl	83	29	2.3	15.2
75	C	3'-pyridyl	100	95	93	100
76	C	4'-pyridyl	100	96	98	47.9
77	C	1'-naphthyl	60.7	1.6	1.4	nt
78	C	2'-naphthyl	56.1	3.8	1.7	nt
79	C	1,1,4,4-tetramethyl-1,2,3,4-tetrahydronaphth-6'-yl	100	2.8	1.8	nt
1	edelfosine		70.0	9.5	2.8	

^a Compounds 40, 44, 45, 47, 48, 50, 52, 55, and 74 are free bases; all other compounds are HCl salts. ^b Values are averages of quadruplicate assays. Average errors: ±0.6% at 0.5–10% viable cells, ±1.5% at 11–25% viable cells and ±4.2% at >25% viable cells. ^c Viable cells were measured by alkaline phosphatase activity. ^d nt denotes not tested.

EA1108 elemental analyzer and are within ±0.4% of theory unless noted otherwise. If not otherwise specified, chemicals and reagents were obtained from the Aldrich Chemical Co. Solvents were of reagent grade and dried prior to use. Reaction progress and purity of final products were determined on E. Merck silica gel 60 chromatography plates. Column chromatography was carried out using E. Merck silica gel CH83 (0.06–0.20 mesh) with the indicated eluents.

Cell Lines. The tumor cell line Abelson 8.1 was obtained from A. W. Harris (Walter and Eliza Hall Institute of Medical Research, Melbourne, Australia) and the YAC-1 from G. Klein (Department of Tumor Biology, Karolinska Institute, Stockholm, Sweden). The cells were grown in stationary suspension culture in Dulbecco Modified Eagle's Medium (DMEM) and 10% heat-inactivated fetal calf serum (FCS) supplemented with 50 μM 2-mercaptoethanol, 100 units of penicillin, and 1 μg of streptomycin. The Meth A fibrosarcoma cells were originally induced in Balb/c mice by administering methylcholanthrene according to the procedure of Old *et al.*³³

Tumor Cell Cytotoxicity. Abelson 8.1 tumor cells (1 × 10³ cells/well) in DMEM and 10% FCS were placed in flat-bottom microtiter plastic plates (Nunc Roskilde Denmark) and incubated with 1–3 μg/mL of test substance dissolved in water for 24–72 h. The number of tumor cells present was determined by measuring alkaline phosphatase activity by a modified procedure of Culvenor,³⁴ the tumor cell plates were centrifuged at 500g for 10 min, and the supernatant was flicked off. Without further washing, 100 μL of buffer containing 20 μL of diethanolamine, 2 μM magnesium chloride hexahydrate, 2.5 μM *p*-nitrophenyl phosphate, and 10 mg of Triton X-100 was added. The samples were incubated for 60 min at room temperature, and the enzymatic activity was terminated by the addition of 100 μL of 0.5 N sodium hydroxide. The absorbance was then measured at 405 nm using a Titertak Multiskan apparatus and compared to non-drug-treated cells. The same assay procedure was used to determine YAC-1 tumor cells.

Table 5. Mouse Meth A Fibrosarcoma Assay:^a Oral Administration

no.	dose, $\mu\text{g}/\text{mouse}$, po ($n = 10$)	tumor volume, % control				survivors ^b day 28
		day 7	day 14	day 21	day 28	
control		100	100	100	100	0/10
53	5.0	102	32	19	27	5/10
	50.0	70	14	0.3	0	10/10
65	5.0	80	43	26	44	3/10
	50.0	90	53	41	61	1/10
1	5.0	86	27	12	18	6/10
	50.0	64	18	9	9	7/10

^a At an average weight of 20 g/mouse: 5.0 $\mu\text{g}/\text{mouse}$ is ca. 0.25 mg/kg and 50 $\mu\text{g}/\text{mouse}$ is ca. 2.5 mg/kg. ^b All survivors are tumor free as measured by calipers and were observed for 120 days after completion of the assay.

Table 4. Direct Cytotoxicity on YAC-1 Tumor Cells^a

no.	R or NRR' ^b	% viable cells at 72 h, $\mu\text{g}/\text{mL}$		
		1	3	5
61	4'- <i>n</i> -C ₃ H ₇	18.0	1.0	0.9
62	4'- <i>i</i> -C ₃ H ₇	46.0	3.9	2.1
63	4'- <i>n</i> -C ₄ H ₉	4.0	1.6	1.1
64	3'- <i>t</i> -C ₄ H ₉	4.2	1.7	1.0
65	4'- <i>t</i> -C ₄ H ₉	3.4	2.7	2.2
69	4'-Si(CH ₃) ₃	11.6	2.6	2.3
70	4'-cyclopentyl	10.5	1.7	1.6
71	4'-cyclohexyl	3.6	1.6	1.4
72	4'-C ₆ H ₅	10.8	3.5	2.8
53	CH ₂ N(CH ₂) ₅	92.0	53.0	21.0
1	edelfosine	68.0	8.4	2.5

^a See footnotes *b* and *c* in Table 3. ^b See formulas type A and type B in Table 3.

Mouse Meth A Fibrosarcoma Assay. Ten CBF₁ mice, 10–12 weeks of age, were implanted with 10⁵ Meth A sarcoma cells to serve as control. Ten other CBF₁ mice were implanted with 10⁵ Meth A sarcoma cells and on day 1 after implant were each treated per os (po) with 5.0 or 50 $\mu\text{g}/\text{mouse}$ of 1, 53, or 65, and daily drug treatment continued for 27 days. On days 7, 15, 21, and 28 after tumor implant, the entire tumor volume was calculated by the equation $V = \frac{2}{3}\pi AB(AB + B/2)$, where *A* and *B* are measured tumor diameters.

PAF Receptor Assay. Methodology for evaluation in PAF receptor binding studies has previously been described.³⁵ Human platelets prepared for the receptor binding were

incubated with [9,10-³H₂]PAF (49 Ci mM⁻¹) at a final concentration of 1.5 nM for 1 h at 24 °C. Free ligand was separated by centrifugation, and the platelets were counted for 1 min in a liquid scintillation counter. Specific binding was calculated as the difference in counts per minute between bound and nonspecifically (calculated using 0.37 mM cold PAF) bound [³H]PAF.

2-Aryl-2-(1*H*)-imidazolines³⁶ 9a-d: 2-(2',5'-Dimethylphenyl)imidazoline(9b). A mixture of 2,5-dimethylbenzotrile (7.87 g, 0.06 mol) and (2-aminoethyl)ammonium *p*-toluenesulfonate (13.96 g, 0.06 mol) was refluxed for ca. 6 h, allowed to cool to ca. 40 °C, poured with stirring into H₂O (100 mL), treated with 6 N NaOH (80 mL, 0.48 mol), and then allowed to stir overnight at room temperature. The resultant solid was filtered off, dissolved in CH₂Cl₂, washed with H₂O (twice with 100 mL), filtered through Celite, and then concentrated *in vacuo* to give 7.84 g (75%) of 9b: mp 108–110 °C (CH₂Cl₂/petroleum ether); MS *m/z* 175 (MH⁺). Anal. (C₁₁H₁₄N₂) C, H, N.

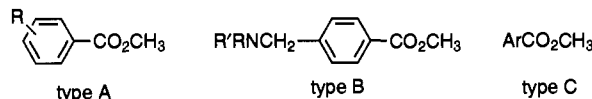
2-(4'-Chloro-2'-methylphenyl)imidazoline (9c): obtained as a white solid (45%); mp 97–98 °C (CH₂Cl₂/pentane); MS *m/z* 195 (MH⁺). Anal. (C₁₀H₁₁ClN₂) C, H, Cl, N.

2-(5'-Chloro-2'-methylphenyl)imidazoline (9d): obtained as a white solid (60%); mp 96–97 °C (C₂H₅OH/pentane); MS *m/z* 195 (MH⁺). Anal. (C₁₀H₁₁ClN₂) C, H, Cl, N.

Methyl Aryl Esters 10a-s. The 3,4-dimethoxy- and 3,4,5-trimethoxybenzoic acid methyl esters, methyl furoate, 4-isopropyl-, 4-*tert*-butyl, and 4-phenylbenzoic acid, 4-ethyl-, 4-propyl-, and 4-butylbenzoyl chloride, and 1- and 2-naphthoyl chloride were obtained from Aldrich. The 4-(trimethylsilyl)-, ³⁷4-cyclopentyl-, ³⁸ and 4-cyclohexylbenzoic³⁹ acids and 5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-2-naphthoic acid⁴⁰ were prepared by cited procedures.

Method A (10a-g). A stirred mixture of methyl *p*-toluate (67.5 g, 0.45 mol), anhydrous NaHCO₃ (45.3 g, 0.54 mol), and CCl₄ (750 mL) was irradiated with a high-intensity light source (150 W reflector Westinghouse lamp) and treated dropwise with a solution of Br₂ (23.05 mL, 0.45 mol) in CCl₄ (100 mL) at such a rate that color faded immediately. The mixture was then refluxed for ca. 2 h, allowed to stand overnight at room temperature, filtered, and then concentrated *in vacuo* to give 104.3 g of crude methyl 4-(bromomethyl)toluate (80% CH₂Br by ¹H-NMR).

A stirred solution of crude bromo ester (22.9 g, ca. 0.08 mol as CH₂Br) in THF (200 mL) was treated dropwise at room temperature with a solution of piperidine (13.6 g, 0.16 mol) in

Table 6. Methyl Aryl Esters 10

no.	type	R or NRR'	method, ^a yield, %	mp or bp (mmHg), °C	formula	anal.
10a	B	N(CH ₂ CH=CH ₂) ₂	A, 52	145–147 (0.2)	C ₁₅ H ₁₉ NO ₂	C,H,N
10b	B	N(CH ₂) ₄	A, 65	130–135 (0.3)	C ₁₃ H ₁₇ NO ₂	C,H,N
10c	B	N(CH ₂) ₅	A, 70	128–133 (0.3)	C ₁₄ H ₁₉ NO ₂	C,H,N
10d	B	(±)-2-methylpiperidyl	A, 42	135–140 (0.2)	C ₁₅ H ₂₁ NO ₂	C,H,N
10e	B	morpholino	A, 61	130–132 (0.3)	C ₁₃ H ₁₇ NO ₃	C,H,N
10f	B	thiomorpholino	A, 46	134–138 (0.3)	C ₁₃ H ₁₇ NO ₂ S	C,H,N
10g	B	N(C ₆ H ₁₁) ₂	A, 56	99–101	C ₂₁ H ₃₁ NO ₂	C,H,N
10h	A	4'-C ₂ H ₅	B, 75	90–93 (0.13) ^b	C ₁₀ H ₁₂ O ₂	
10i	A	4'- <i>n</i> -C ₃ H ₇	B, 95	138–140 (16)	C ₁₁ H ₁₄ O ₂	
10j	A	4'- <i>i</i> -C ₃ H ₇	B, 80	135–138 (18)	C ₁₁ H ₁₄ O ₂	
10k	A	4'- <i>n</i> -C ₄ H ₉	B, 82	146–148 (18) ^c	C ₁₂ H ₁₆ O ₂	
10l	A	3'- <i>t</i> -C ₄ H ₉	B, 63	69–72 (0.2)	C ₁₂ H ₁₆ O ₂	
10m	A	4'-Si(CH ₃) ₃	B, 79	80–82 (0.2)	C ₁₁ H ₁₆ O ₂ Si	
10n	A	4'-cyclopentyl	B, 82	oil	C ₁₃ H ₁₆ O ₂	
10o	A	4'-cyclohexyl	B, 50	45–46 ^d	C ₁₄ H ₁₈ O ₂	C,H
10p	A	4'-C ₆ H ₅	B, 61	111–112 ^e	C ₁₄ H ₁₂ O ₂	
10q	C	1-naphthyl	B, 75	128–130 (0.2)	C ₁₂ H ₁₀ O ₂	
10r	C	2-naphthyl	B, 72	76–78	C ₁₂ H ₁₀ O ₂	
10s	C	1,1,4,4-tetramethyl-1,2,3,4-tetrahydronaphth-6'-yl	B, 93	75–77 (0.1) ^f	C ₁₆ H ₂₂ O ₂	

^a See the Experimental Section for a detailed description of the general methods A and B. ^b Lit.⁴² oil. ^c Lit.⁴³ bp 69–70 °C (0.13 mm). ^d Lit.⁴⁴ mp 47–48 °C. ^e Lit.⁴⁵ mp 112 °C. ^f Lit.⁴⁰ bp 161–164 °C (7.8 mm).

THF (20 mL) and then stirred ca. 48 h. The solids were filtered off, the filtrate was concentrated *in vacuo*, and the residue was dissolved in CH₂Cl₂ (300 mL) and washed with 2 N HCl (300 mL). The acid layer was treated with 2 N NaOH until basic to litmus paper, extracted with CH₂Cl₂ (400 mL), dried (MgSO₄), filtered, and concentrated *in vacuo*, and the residue was distilled to give 13.1 g of methyl 4-(1-piperidinylmethyl)benzoate (**10c**) (Table 6).

Method B. A solution of 4-(trimethylsilyl)benzoic acid (8.8 g, 0.045 mol), thionyl chloride (8.0 g, 0.073 mol), and CH₂Cl₂ was stirred and refluxed for ca. 8 h and then concentrated *in vacuo*. The residue was added to sodium methoxide (0.054 mol) in CH₃OH (80 mL), stirred at room temperature for ca. 48 h, and concentrated *in vacuo*, and the residue was treated with H₂O (20 mL) and CH₂Cl₂ (50 mL). The organic layer was separated, dried (MgSO₄), filtered, and concentrated *in vacuo*, and the residue was distilled to give 7.4 g of methyl 4-(trimethylsilyl)benzoate (**10m**, Table 6).

Method C. Methyl 4-[(3',4'-Dimethoxyphenyl)methyl]oxy]benzoate (10t). A stirred mixture of methyl 4-hydroxybenzoate (19.8 g, 0.13 mol), 50% NaH (7.5 g, 0.16 mol), and 3,4-dimethoxybenzyl chloride (21.2 g, 0.16 mol) in DMF (500 mL) under N₂ was heated at 60 °C for 4 h and concentrated *in vacuo*, and the residue was treated with H₂O (50 mL) and CH₂Cl₂ (200 mL). The organic layer was washed with H₂O, dried (MgSO₄), filtered, and concentrated *in vacuo* to give 24.0 g (62%) of **10t**: mp 94–5 °C (C₂H₅OH/pentane); MS *m/z* 303 (MH⁺). Anal. (C₁₇H₁₈O₅) C, H.

5-Aryl-2,3,5,6-tetrahydroimidazo[2,1-a]isoquinolin-5-ols (11–37). A stirred solution of 2-(*o*-methylphenyl)imidazoline (8.0 g, 0.05 mol) in THF (200 mL) under N₂ was treated dropwise with 1.6 M *n*-BuLi in hexane (105 mL, 0.15 mol) and then heated at 35 °C for ca. 4 h. The mixture was then immersed in a dry ice–acetone bath, cooled to an internal temperature of –25 °C, and treated dropwise with a solution of methyl aryl ester (0.10 mol) in THF (50 mL) at such a rate that the internal temperature did not exceed –20 °C. After an additional 3 h at –25 °C, the reaction mixture was allowed to warm to 0 °C, treated with saturated NH₄Cl solution (30 mL), and allowed to stand overnight at room temperature. The mixture was then concentrated *in vacuo* and treated with CH₂Cl₂ (200 mL) and H₂O (100 mL), the CH₂Cl₂ layer was separated, washed with H₂O, dried (MgSO₄), and filtered, and the filtrate was concentrated *in vacuo*. The resultant residue was then crystallized from the appropriate solvent given in Table 1.

5-Aryl-2,3-dihydroimidazo[2,1-a]isoquinolines (38–79). **Method D.** A solution of 5-aryl-2,3,5,6-tetrahydroimidazo[2,1-a]isoquinolin-5-ol (0.01 mol) in acetic acid (40 mL) was stirred and refluxed under N₂ for ca. 5 h. The acetic acid was removed *in vacuo*, and the residue was treated with H₂O (50 mL) and then made alkaline with 2 N Na₂CO₃ (35 mL). The mixture was extracted with CH₂Cl₂ (2 × 100 mL), and the organic layer was washed with H₂O (50 mL), dried (MgSO₄), filtered, and concentrated *in vacuo* to give the free base. The HCl salt was prepared by dissolving the base in saturated HCl/EtOH, evaporating *in vacuo*, and crystallizing from the appropriate solvent (cf. Table 2).

Method E. A mixture of 5-aryl-2,3,5,6-tetrahydroimidazo[2,1-a]isoquinolin-5-ol (2 g), *p*-toluenesulfonic acid (0.2 g), and toluene (50 mL) were stirred and refluxed for ca. 12 h with continuous removal of H₂O (Dean–Stark trap). The toluene was removed *in vacuo*, and the residue was dissolved in CH₂Cl₂, washed with H₂O, saturated NaHCO₃ solution, and saturated NaCl solution, dried (MgSO₄), filtered, and evaporated *in vacuo* to give the free base. The HCl salt was prepared as given in method D.

Acknowledgment. The authors wish to thank Dr. Michael Shapiro and Ms. Ann Archinal for NMR spectra, Mr. Eric Roos for elemental analysis, and Mrs. Bertha Owens and Mr. Robert Suzuki for mass spectra. We also like to thank Miss Danielle Padula for typing the manuscript.

References

- (1) (a) Westphal, O. *ET 18 OCH₃: Wissenschaftliche Zusammenfassung publizierter und nicht-publizierter Erkenntnisse zum Stand der synthetischen Etherlipid ET 18 OCH₃, Ein Beitrag zum Stand der Wissenschaft*; Max-Planck-Institut fuer Immunobiologie: 78 Freiburg-Zaehringen, Germany, 1987; 104 pp. (b) Khanavkar, B.; Ulbrich, F.; Gatzemeier, U.; Meyer-Schwickerath, E.; Lorenz, J.; Schreml, W.; Brugger, R.; Schick, H. D.; von Pawel, J.; Nordstrom, R.; Drings, P. Treatment of Non-Small Cell Lung Cancer With the Alkyliso phospholipid Edelfosine. *Contrib. Oncol.* **1989**, *37*, 224–235.
- (2) Herrmann, D. B. J.; Neumann, H. A.; Heim, M. E.; Berdel, W. E.; Fromm, M.; Andreesen, R.; Queisser, W.; Boerner, D.; Sterz, R.; Besenfelder, E.; Bicker, U. Short and Long-Term Tolerability Study of the Thioether Phospholipid Derivative Ilmofofosine in Cancer Patients. *Contrib. Oncol.* **1989**, *37*, 236–247.
- (3) (a) Eibl, H., Hilgard, P., Unger, C., Eds. *Alkylphosphocholines: New Drugs in Cancer Therapy*; Krager: Basel, Switzerland, 1992. (b) Unger, C.; Eibl, H.; Nagel, G. A.; Von Heyden, H. W.; Breiser, A.; Engel, J.; Stekar, J.; Peukert, M.; Hilgard, P.; Berger, M. *Contrib. Oncol.* **1989**, *37*, 219–223.
- (4) Houlihan, W. J.; Lee, M. L.; Munder, P. G.; Nemecek, G. M.; Handley, D. A.; Winslow, C. M.; Happy, J.; Jaeggi, C. Antitumor Activity of SRI 62-834, a Cyclic Ether Analog of ET-18-OCH₃. *Lipids* **1987**, *22*, 884–890.
- (5) Baumann, W. J. *Platelet-Activating Factor and Structurally Related Alkyl Ether Lipids*; American Oil Chemists' Society: Champaign, IL, 1992.
- (6) Braquet, P., Ed. *Handbook of PAF and PAF Antagonists*; CRC Press: Boca Raton, FL, 1991.
- (7) For reviews on phospholipid antitumor agents, see: (a) Berdel, W. E.; Munder, P. G. Antitumor Analogs of PAF. In *Platelet-Activating Factor and Related Lipid Mediators*; Snyder, F., Ed.; Plenum Press: New York, 1987; pp 449–467. (b) Modest, E. J.; Berens, M. E.; Piantadosi, C.; Nosedá, A. Pharmacological Effects and Anticancer Activity of New Ether Phospholipid Analogs. In *The Pharmacological Effect of lipids III, Role of Lipids in Cancer Research*; Kabara, J. J., Ed.; The American Oil Chemists' Society; Champaign, IL, 1989; pp 330–337. (c) Munder, P. G.; Westphal, O. Antitumor and Other Biomedical Activities of Synthetic Ether Lyso-phospholipids. In *1939–1989: Fifty Years Progress in Allergy*; Waksman, B. H., Ed.; Karger: New York, 1990; pp 206–235. (d) Houlihan, W. J.; Workman, P.; Lohmeyer, M.; Cheon, S. H. Phospholipid Antitumor Agents. *Med. Res. Rev.* **1995**, unpublished results.
- (8) Berdel, W. E. Membrane-interactive Lipids as Experimental Anticancer Drugs. *Br. J. Cancer* **1991**, *64*, 208–211.
- (9) Berdel, W. E.; Bausert, W. R.; Weltzien, H. U.; Modolell, M. L.; Widmann, K. H.; Munder, P. G. The Influence of Alkyl Lyso-phospholipids and Alkyl Lyso-phospholipid-Activated Macrophages on the Development of Metastasis of 3-Lewis Lung Carcinoma. *Eur. J. Cancer* **1980**, *16*, 1199–1204.
- (10) Nosedá, A.; Godwin, P. L.; Modest, E. J. Effects of Antineoplastic Ether Lipids on Model and Biological Membranes. *Biochim. Biophys. Acta* **1988**, *945*, 92–100.
- (11) Dive, C.; Watson, J. V.; Workman, P. Multiparametric Flow Cytometry of the Modulation of Tumor Cell Membrane Permeability by Developmental Antitumor Ether Lipid SRI 62–834 in EMT6 Mouse Mammary Tumor and HL60 Human Promyelocytic Leukemia cells. *Cancer Res.* **1990**, *51*, 799–806.
- (12) Seewald, M. J.; Olsen, R. A.; Sehgal, I.; Melder, D. C.; Modest, E. J.; Powis, G. Inhibition of Growth Factor-Dependent Inositol Phosphate Ca²⁺ Signaling by Antitumor Ether Lipid Analogues. *Cancer Res.* **1990**, *50*, 4458–4463.
- (13) Ueberall, F.; Oberhuber, H.; Maly, K.; Zaknun, J.; Demuth, L.; Grunicke, H. H. Hexadecylphosphocholine Inhibits Inositol Phosphate Formation and Protein Kinase C Activity. *Cancer Res.* **1991**, *51*, 807–812.
- (14) Bazill, G. W.; Dexter, T. M. Role of Endocytosis in the Action of Ether Lipids on WEHI-3B, HL60 and FDCP-Mix A4 Cells. *Cancer Res.* **1990**, *50*, 7505–7512.
- (15) Workman, P. Antitumor Ether Lipids: Endocytosis as a Determination of Cellular Sensitivity. *Cancer Cells* **1991**, *3*, 315–317.
- (16) Berdel, W. E.; Korth, R.; Reichert, A.; Houlihan, W. J.; Bicker, U.; Nomura, H.; Vogler, W. R.; Benveniste, J.; Rastetter, J. Lack of Correlation Between Cytotoxicity of Agonists and Antagonists of Platelet Activating Factor (PAF-acether) in Neoplastic Cells and Modulation of [³H]-PAF-acether binding to Platelets from Humans *in vitro*. *Anticancer Res.* **1987**, *7*, 1181–1188.
- (17) Houlihan, W. J.; Lee, M. L.; Munder, P. G.; Winslow, C. M.; Cheon, S. H.; D'Aries, F. J.; DeLillo, A. K.; Jaeggi, C. S.; Mason, R. B.; Parrino, V. A. Cyclic Oxygen Analogues of Alkyl-lyso-phospholipids. *Synthesis and Neoplastic Cell Growth Inhibitory Properties*. *J. Lipid Mediators* **1990**, *2*, 295–307.
- (18) Kudo, I.; Nojima, S.; Chang, H. W.; Yanoshita, R.; Hidetoshi, H.; Hayashi, H.; Kondo, E.; Nomura, H.; Inoue, K. Antitumor Activity of Synthetic Alkylphospholipids with or without PAF Activity. *Lipids* **1987**, *22*, 862–867.

- (19) Bazill, G. W.; Dexter, T. M. An Antagonist to Platelet Activating Factor Counteracts the Tumouricidal Action of Alkyl Lysophospholipids. *Biochem. Pharmacol.* **1989**, *38*, 374–377.
- (20) Lazenby, C. M.; Thompson, M. G.; Hickman, J. A. Elevation of Leukemic Cell Intracellular Calcium by Ether Lipids. *Cancer Res.* **1990**, *50*, 3327–3330.
- (21) Workman, P.; Donaldson, J.; Lohmeyer, M. Platelet-Activating Factor (PAF) Antagonist WEB 2086 does not Modulate the Cytotoxicity of PAF or Antitumor Alkyl Lysophospholipids ET-18-O-Methyl and SRI 62-834 in HL-60 Promyelocytic Leukemia Cells. *Biochem. Pharmacol.* **1991**, *41*, 319–322.
- (22) Lohmeyer, M.; Workman, P. Lack of Stereospecificity in the Membrane-Damaging and Cytotoxic Potency of the Antitumor Ether Lipid SRI 62-834. *7th NCI-EORTC Symposium on New Drugs in Cancer Therapy*, Amsterdam, March 17–20, 1992, Abstr. 36, p 67.
- (23) Houlihan, W. J.; Munder, P. G.; Larson, D. Cytotoxic Activity of Non-Charged Platelet-Activating Factor (PAF) Antagonists Against Murine Tumor Cell Lines. *Prostaglandins* **1988**, *35*, 817.
- (24) Houlihan, W. J. *WO Pat.* 88/06157, 1988.
- (25) Houlihan, W. J. 5-Aryl-2,3-dihydroimidazo[2,1-a]isoquinolines: Platelet Activating Factor (PAF) Receptor antagonists. *Drugs Future* **1990**, *15*, 355–359.
- (26) Danhauser-Riedl, S.; Felix, S. B.; Houlihan, W. J.; Zafferani, M.; Steinhäuser, G.; Oberberg, D.; Kalvelage, H.; Busch, R.; Rastetter, J.; Berdel, W. E. Some Antagonists of Platelet Activating Factor are Cytotoxic for Human Malignant Cell Lines. *Cancer Res.* **1991**, *51*, 43–48.
- (27) Houlihan, W. J.; Munder, P. G.; Berdel, W. E.; Nemecek, G. M.; Schmitt, G.; Winslow, C. M. SDZ 62-434 A Novel Imidazo [2,1-a] Isoquinoline PAF Receptor Antagonist with in vitro and in vivo Antitumor Activity. *Proc. Am. Assoc. Cancer Res.* **1991**, *32*, 407.
- (28) Brunton, V. G.; Workman, P. In Vitro Antitumor Activity of the Novel Imidazoisoquinoline SDZ 62-434. *Br. J. Cancer* **1993**, *67*, 489–495.
- (29) Koenigsmann, M.; Zafferani, M.; Danhauser-Riedl, S.; Reufi, B.; Houlihan, W. J.; Thiel, E.; Berdel, W. E. Lack of Therapeutic Effects of Platelet Activating Factor Antagonists in WEHI-3B Leukemia, Human Xenotransplanted Colorectal and Lung Cancer and Lewis-Lung Tumor in vivo. *Cancer Lett.* **1992**, *67*, 145–156.
- (30) Houlihan, W. J.; Parrino, V. A. Directed Lithiation of 2-Phenyl- and 2-(o-methylphenyl)imidazoline. *J. Org. Chem.* **1982**, *47*, 5177–5180.
- (31) Houlihan, W. J.; Gogerty, J. H.; Parrino, V. A.; Ryan, E. Antidepressant Activity of 5-Aryl-2,3,5,6-tetrahydroimidazo [2,1-a]isoquinolin-5-ols. *J. Med. Chem.* **1983**, *26*, 765–768.
- (32) Houlihan, W. J.; Munder, P. G.; Handley, D. A.; Nemecek, G. A. Preclinical Pharmacology and Possible Mechanism of Action of the Novel Antitumor Agent (5-(4'-Piperidinomethylphenyl)-2,3-dihydroimidazo [2,1-a]isoquinoline (SDZ 62-434). *Arzneimittel-Forsch.*, in press.
- (33) Old, L. J.; Boyse, E. A.; Clarke, D. A.; Carswell, E. Antigenic Properties of Chemically-Induced Tumors. *Ann. N.Y. Acad. Sci.* **1962**, *101*, 80–92.
- (34) Culvenor, J. G.; Harris, A. W.; Mandel, T. E.; Whitelaw, A.; Ferber, E. Alkaline Phosphatase in Hematopoietic Tumor Cell Lines of the Mouse: High Activity in Cells of the B-lymphoid Lineage. *J. Immunol.* **1981**, *126*, 1974–1977.
- (35) Handley, D. A.; Van Valen, R. G.; Melden, M. K.; Houlihan, W. J.; Saunders, R. N. Biological Effects of the Orally Active Platelet Activating Factor Receptor Antagonist SDZ 64-412. *J. Pharmacol. Exp. Ther.* **1988**, *247*, 617–623.
- (36) Oxley, P.; Short, W. F. Amidines. Part VI. Preparation of 2-Substituted 4:5-dihydroxyoxalines and Ring Homologues from Cyanides and Alkylenediamines. *J. Chem. Soc.* **1947**, 497–505.
- (37) Deans, F. B.; Eaborn, C. Aromatic Reactivity, III. Cleavage of Substituted Phenyltrimethylsilanes by Sulfuric Acid in Acetic Acid-Water. *J. Chem. Soc.* **1959**, 2299–2303.
- (38) Monteils, Y. Préparation de Dérivés Halogénés par Condensation de Composés Aromatiques et Éthyléniques en Présence d'Acide Sulfurique Concentré. *Bull. Soc. Chim. Fr.* **1951**, 637–641.
- (39) Dauben, W. G.; Tanabe, M. The Hydrogenation of 4-Hydroxybiphenylcarboxylic Acids. *J. Am. Chem. Soc.* **1953**, *75*, 4969–4973.
- (40) Myhre, P. C.; Schubert, W. M. Isolation and Proof of Structure of 1,1,4,4-Tetramethyl-6-tert-butyl-1,2,3,4-tetrahydronaphthalene. *J. Org. Chem.* **1960**, *25*, 708–711.
- (41) ¹H-NMR data of selected compounds from Tables 1 and 2. The solvent is DMSO-*d*₆, unless indicated otherwise. **23**: δ 1.21 (d, 6H), 2.80–3.78 (m, 6H), 6.15 (s, 1H), 7.11–7.55 (m, 7H), 7.98 (d, 1H). **31**: δ 1.44–2.15 (m, 8H), 3.00 (m, 1H), 3.01–3.96 (m, 6H), 7.38–7.96 (m, 7H), 8.30 (d, 1H). **32**: δ 1.05–1.76 (m, 10H), 3.80–4.36 (m, 6H), 7.30–7.86 (m, 8H). **36**: δ 3.06–3.87 (m, 6H), 6.32 (s, 1H), 7.12–8.11 (m-11H). **37**: δ 1.31 (s, 12H) 1.72 (s, 4H), 3.03–3.93 (m, 6H), 7.05–7.55 (m, 6H), 8.08 (s, 1H). **39**: (D₂O): δ 3.90 (t, 2H), 4.30 (t, 2H), 7.02 (s, 1H), 7.15–8.18 (m, 8H). **42**: (CF₃CO₂D) δ 3.98 (m, 2H), 4.35 (m, 2H), 6.86 (s, 1H), 7.03–8.13 (m, 8H). **58**: (CF₃CO₂D) δ 2.52 (s, 3H), 4.35 (μ, 2H), 5.65 (μ, 2H), 7.17 (s, 1H), 7.42 (s, 4H), 7.65–8.28 (μ, 4H). **61**: δ 0.95 (t, 3H), 1.67 (m, 2H), 2.68 (t, 2H), 7.18 (s, 1H), 7.47 (A₂B₂, 4H), 7.63–8.02 (m, 3H), 8.48 (d, 1H). **62**: (D₂O) δ 1.18 (d, 6H), 2.92 (quintet, 1H), 3.95 (t, 2H), 4.37 (t, 2H), 6.83 (s, 1H), 7.33 (s, 4H), 7.52–7.95 (m, 4H). **63**: 0.98 (t, 3H), 1.17–1.72 (m, 4H), 2.65 (t, 2H), 4.02 (t, 2H), 4.48 (t, 2H), 7.13 (s, 1H), 7.43 (A₂B₂, 4H), 7.50–7.82 (m, 3H), 8.48 (d, 1H). **70**: 1.43–2.11 (m, 8H), 3.05 (m, 1H), 4.02 (t, 2H), 4.53 (t, 2H), 7.12 (s, 1H), 7.46 (A₂B₂, 4H), 7.52–7.96 (m, 3H) 8.51 (d, 1H). **71**: (D₂O) 1.08–1.82 (m, 10H), 3.08 (m, 1H), 3.98 (t, 2H), 4.38 (t, 3H), 6.92 (s, 1H), 7.38 (s, 4H), 7.58–8.00 (m, 4H). **77**: (D₂O) δ 3.92 (m, 4H), 6.87 (s, 1H), 7.32–8.05 (m, 11H). **78**: δ 4.11 (t, 2H), 4.63 (t, 2H), 7.35 (s, 1H), 7.56–8.32 (m, 10H), 8.68 (d, 1H). **79**: δ 1.28 (s, 12H), 1.75 (s, 4H), 4.09 (t, 2H), 4.60 (t, 2H), 7.17 (s, 1H), 7.20–7.82 (m, 6H), 8.43 (d, 1H).
- (42) Lau, C. K.; Tardif, S.; Dufresne, C.; Scheigetz, J. Reductive Deoxygenation of Aryl Ketones by tert-Butylamine-Borane and Aluminum Chloride. *J. Org. Chem.* **1989**, *54*, 491–494.
- (43) Ebert, G. W.; Rieke, R. D. Preparation of Aryl, Alkynyl and Vinyl Organocopper Compounds by the Oxidative Addition of Zero Valent Copper to Carbon-Halogen Bonds. *J. Org. Chem.* **1988**, *53*, 4482–4488.
- (44) Schubert, H.; Jaenecke, G.; Taubert, H. The Catalytic Hydrogenation of Aromatic-Substituted Imidazoles. V. (p-Biphenyl) imidazoles. *J. Prakt. Chem.* **1961**, *15*, 86–104.
- (45) Salijoughian, M.; Morimoto, H.; Dorsky, A. M.; Rapoport, H.; Andres, H. A new and efficient synthesis of monotriomethyl iodide. *J. Labelled Compd.* **1989**, *27*, 767–776.

JM9406211