

## Synthesis and SAR of a New Series of COX-2-Selective Inhibitors: Pyrazolo[1,5-*a*]pyrimidines

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The synthesis and pharmacological activity of a series of bicyclic pyrazolo[1,5-*a*]pyrimidines as potent and selective cyclooxygenase-2 (COX-2) inhibitors are described. The new compounds were evaluated both in vitro (COX-1 and COX-2 inhibition in human whole blood) and in vivo (carrageenan-induced paw edema and air-pouch model). Modification of the pyrimidine substituents showed that 6,7-disubstitution provided the best activity and led to the identification of 3-(4-fluorophenyl)-6,7-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (**10f**) as one of the most potent and selective COX-2 inhibitor in this series.

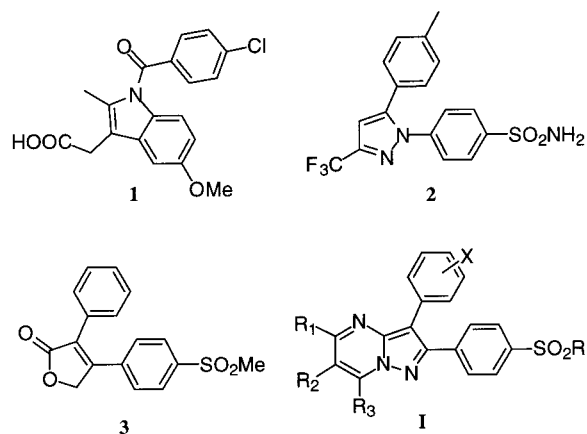
### Introduction

Nonsteroidal antiinflammatory drugs (NSAIDs) exert their antiinflammatory action mainly through inhibition of cyclooxygenase (COX), one of the key enzymes in the arachidonic acid (AA) cascade.<sup>1</sup> This enzyme bis-oxygenates AA to PGG<sub>2</sub>, which is subsequently degraded to vasoactive and inflammatory mediators such as prostaglandins (PGs), prostacyclin (PGI<sub>2</sub>), and thromboxane A<sub>2</sub>. The therapeutic use of NSAIDs has been associated with well-known side effects at the gastrointestinal level (mucosal damage, bleeding)<sup>2</sup> and, less frequently, at the renal level<sup>3</sup>, and thus their use in chronic diseases such as rheumatoid arthritis has caused complications.

At the beginning of the 1990s two COX isoforms were discovered:<sup>4</sup> one (COX-1) constitutively present in many tissues such as stomach, kidney, and platelets and the other (COX-2) cytokine inducible and expressed mainly in a wide range of inflammatory cells. This scenario led to the recognition<sup>5</sup> that selective COX-2 inhibitors could provide antiinflammatory agents devoid of the undesirable effects associated with classical, nonselective NSAIDs.

Several approaches based on modification of established nonselective agents have been studied for the design of COX-2-selective inhibitors,<sup>6</sup> in a line of research in which the crystal resolution of the structures of both enzymes has also been somewhat helpful.<sup>7</sup> Lengthening the carboxyl side chain of indomethacin<sup>8</sup> (**1**) or substituting it with other groups<sup>9</sup> and modifying the preferential inhibitor nimesulide to flosulide,<sup>10</sup> L-745337,<sup>11</sup> and NS-398<sup>12</sup> has led to selective compounds, although none of them is being pursued for a variety of reasons. Based on the structure of early known antiinflammatory agents such as flumizole, a wide range of diarylheterocycle COX-2-selective inhibitors have been described.<sup>13</sup> To obtain good activity and selectivity such compounds require the presence of a 4-methylsulfonylphenyl group attached to an unsaturated (generally five-membered) ring in which an additional vicinal phenyl ring is present. The methylsul-

fonyl group can only be replaced by an SO<sub>2</sub>NH<sub>2</sub>, whereas the second phenyl group accepts lower alkyl or halogen substituents, the most common being 4-fluoro and 4-methyl. Two compounds in this class, celecoxib<sup>14</sup> (**2**) and rofecoxib<sup>15</sup> (**3**), are already in the market for the treatment of acute pain, osteoarthritis, and rheumatoid arthritis and seem to be as effective as other established NSAIDs and to have much better gastric tolerance.<sup>16</sup> In addition they are undergoing clinical trials for the treatment of certain forms of cancer and Alzheimer's disease.

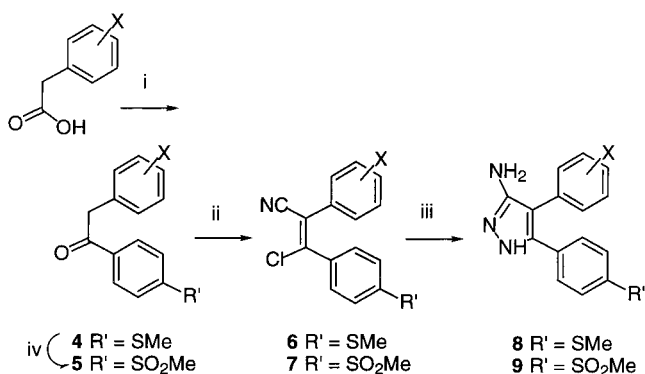


In a new approach to diarylheterocycle COX-2-selective inhibitors, we decided to investigate the attachment of an additional six-membered ring to the usual five-membered central ring. The pyrazolo[1,5-*a*]pyrimidine nucleus **I** was selected for the SAR study in view of the easier access to substituent variation in the extra ring in comparison to other possible frameworks, which in turn could be prepared once the key features of **I** were identified. Two similar approaches taken with imidazo[1,2-*a*]pyridines<sup>17</sup> and isoindoles<sup>18</sup> have appeared in the patent literature.

### Chemistry

The basic SAR around the pyrazolopyrimidines **I** was effected with the 3-(4-fluorophenyl)-2-(methylsulfo-

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Scheme 1<sup>a</sup>

a X = F; b X = Me; c X = 3,4-diF; d X = 2,4-diF

<sup>a</sup> (i) Thioanisole, AlCl<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -10 °C, 1 h; (ii) 1. POCl<sub>3</sub>/DMF, CHCl<sub>3</sub>, 80 °C, 18 h, 2. NH<sub>2</sub>OH·HCl, DMF, 20 °C, 4 h; (iii) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O, EtOH, 78 °C, 18 h; (iv) Oxone, MeOH/THF, 20 °C, 1 h.

nylphenyl) derivatives **10**, which could be prepared from amine **9a**. Amines **8** and **9** were obtained from ketones **4** and **5**<sup>19</sup> in a process involving reaction with POCl<sub>3</sub>/DMF followed by treatment with hydroxylamine in DMF to give a 3-chloropropenenitrile intermediate (**6**, **7**), which was cyclized with hydrazine in ethanol (Scheme 1).

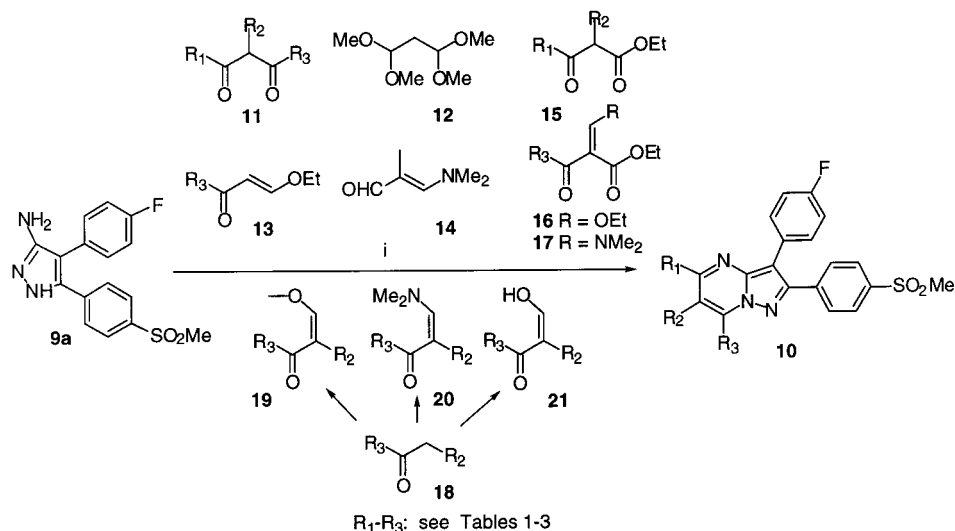
The cyclization of 3-aminopyrazoles with a 1,3-dicarbonyl compound is the usual method for preparing pyrazolopyrimidines.<sup>20</sup> When a nonsymmetrical reagent is used, usually isomeric mixtures are obtained. The proportion of isomers is quite erratic and depends on the substrate, the nature of the reagent, and the type of groups used to mask the carbonyl compounds.<sup>20a,b</sup>

As outlined in Scheme 2, compounds **10** were obtained by cyclization of **9a** with an appropriately substituted free or masked 1,3-dicarbonyl derivative (**11–17**, **19–21**), under conditions A–G indicated for each compound in Tables 1–3. When a free dicarbonyl derivative (**11** or **15**) was used, or in the case of the ethoxymethylene derivatives **13**, the reactions were run either in ethanol

or in toluene (methods B, D). In the remaining cases, the presence of masked carbonyl groups in the reactants required acidic conditions and one of methods A (HCl/ZnCl<sub>2</sub>/EtOH), C (HCl/EtOH), E (AcOH), or G (*p*-TsOH/toluene) was used. The 6-bromo derivative **10ab** (Table 2) was obtained from 3-bromo-4-diethylamino-3-buten-2-one (**20**, R<sub>2</sub> = Br, R<sub>3</sub> = Me) using HBr (method F), since the standard HCl gave its 6-chloro analogue, product of a halogen exchange.

The parent compound **10a** (Table 1) was obtained using the diprotected malonaldehyde **12** in the presence of ZnCl<sub>2</sub> (method A), while the 6-methyl **10d** was obtained from **14** under the conditions of method C. The isomeric 5-methyl **10b** and 7-methyl **10c** were obtained as a 1:4 mixture by reaction of **9a** with **13** (R<sub>3</sub> = Me) under the conditions of method B. The structural assignment was made on the basis of their <sup>1</sup>H and <sup>13</sup>C NMR spectra. The methyl groups of **10b** and **10c** showed proton shifts of 2.63 and 2.88 ppm, respectively, and carbon shifts of 26.5 and 18.5 ppm, respectively, in accordance with results previously reported for related compounds.<sup>20a,b</sup> The structural identity of **10b,c** was further confirmed by additional chemical transformations.<sup>20c,d</sup> Hence, **10s** was converted to **10b** upon hydrogenolysis with H<sub>2</sub> over Pd/C/NaOAc,<sup>20c</sup> whereas **10x** was converted to **10c** upon decarboxylation with H<sub>2</sub>SO<sub>4</sub>, using the same conditions described below for the preparation of **10j**.

The 5,7-dimethyl derivative **10e** was prepared from **9a** and acetylacetone (**11**, R<sub>2</sub> = H, R<sub>1</sub> = R<sub>3</sub> = Me), whereas the isomeric **10f,g** were obtained in different proportions depending on the reactant employed. Thus, the α-hydroxymethylene derivative **21** (R<sub>2</sub> = R<sub>3</sub> = Me) led to **10f,g** in a 1:9 ratio, while the α-methoxymethylene derivative **19** (R<sub>2</sub> = R<sub>3</sub> = Me)<sup>21</sup> led to **10f,g** in a 9:1 ratio. In both cases the most abundant isomer was isolated by silica gel chromatography in 62% (**10g**) and 70% (**10f**) yield. The assignment of the structure to **10g** was based on its unambiguous synthesis by hydrogenolysis of the 7-chloropyrazolopyrimidine **10t** over Pd/

Scheme 2<sup>a</sup>

<sup>a</sup> (i) Method A: HCl, ZnCl<sub>2</sub>, EtOH, 78 °C, 1 h; Method B: piperidine, EtOH, 78 °C, 18 h; Method C: HCl, EtOH, 78 °C, 1–18 h; Method D: toluene, 111 °C, 8 h; Method E: AcOH, 117 °C, 1 h; Method F: HBr, EtOH, 78 °C, 1 h; Method G: *p*-TsOH, toluene, 111 °C, 8 h. Specific methods and reagents used are indicated in Tables 1–3.

**Table 1.** Methylpyrazolopyrimidines of Formula 10

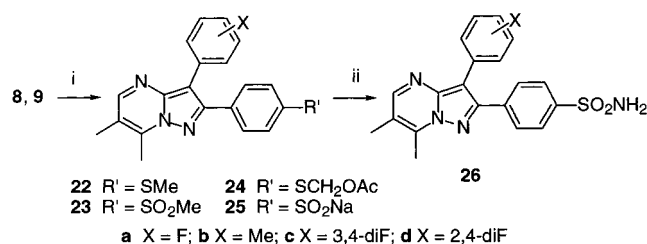
							% Inh HWB <sup>c</sup>			IC <sub>50</sub> Whole Cell <sup>d</sup>	
reactant, yield, isomeric							COX-1	COX-2		COX-1	COX-2
compd	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	method <sup>a</sup>	%	ratio <sup>b</sup>	10 μM	10 μM	1 μM	(μM)	(μM)
<b>10a</b>	H	H	H	<b>12</b> , A	53	-	35.0	36.2	-	> 10	4.201
<b>10b</b>	Me	H	H	<b>13</b> , B	11	8:2	19.0	28.7	-	> 10	0.504
<b>10c</b>	H	H	Me	<b>13</b> , B	44	8:2	50.5	97.2	55.2	> 10	0.012
<b>10d</b>	H	Me	H	<b>14</b> , C	63	-	8.2	78.3	25.6	> 10	0.267
<b>10e</b>	Me	H	Me	<b>11</b> , B	54	-	8.3	93.1	17.1	> 10	0.154
<b>10f</b>	H	Me	Me	<b>19</b> , C	70	9:1	47.5	96.4	72.8	> 10	0.012
<b>10g</b>	Me	Me	H	<b>21</b> , D	62	1:9	32.6	57.2	-	> 10	2.133
<b>10h</b>	Me	Me	Me	<b>11</b> , D	53	-	61.3	100	56.1	> 10	0.192
<b>2</b>							34.5	94.6	67.0	5.1	0.080
<b>1</b>							100 <sup>e</sup>	100	67.9	0.003	0.009

<sup>a</sup> Numbers indicate the type of reagent used as indicated in Scheme 2 and letters the cyclization method used for the preparation of compounds, as described in the Experimental Section. <sup>b</sup> Isomeric ratio of 7- vs 5-monosubstituted, 6,7- vs 5,6-disubstituted, or 5,7- vs 7,5-disubstituted compounds. <sup>c</sup> Percentage of inhibition of COX-1 (10 μM) or COX-2 (10 and 1 μM) activity in human whole blood. Assays were performed in duplicate or repeated, so that standard errors were inferior to 10%. <sup>d</sup> IC<sub>50</sub> for whole cell inhibition in U-937 cells (COX-1) and 143982 cells (COX-2). Assays were performed in duplicate. <sup>e</sup> 82.3% inhibition at 1 μM.

C/NaOAc.<sup>20c</sup> It is noteworthy that the <sup>1</sup>H NMR spectra of the two isomers were characterized by the chemical shift of the 7-methyl (**10f**) at 2.85 ppm and the 5-methyl (**10g**) at 2.60 ppm. The downfield shift generally observed for the 7- over the 5-substituted derivatives allowed the identification of the remaining compounds of Tables 2 and 3.

As can be seen from the results outlined in Tables 2 and 3, where the observed isomeric ratios are indicated for every compound, the 7-substituted and 6,7-disubstituted isomers are favored over their 5-substituted and 5,6-disubstituted counterparts. This result is consistent with the proposed mechanism for this reaction, in which the most reactive carbonyl group (in this case the masked aldehyde function) first interacts with the amino group of the aminopyrazole, after which cyclization takes place.

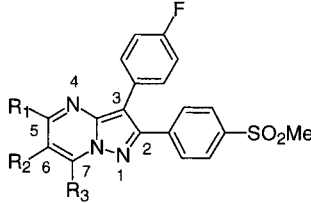
Some compounds were obtained by further transformations of the pyrimidine substituents. Hence, the 7-hydroxy derivative **10q** was converted to the methoxy **10r** with methyl sulfate and to the chloro **10s** on treatment with POCl<sub>3</sub>. Compound **10t** was prepared from its hydroxyl counterpart in an analogous way. Standard ester hydrolysis provided acids **10n,x,af**. Decarboxylation at position 6 by refluxing in H<sub>2</sub>SO<sub>4</sub> for 18 h provided access to 7-monosubstituted derivatives such as **10j**. The preparation of the monohydroxymethyl derivatives **10y,ad** was more difficult. Attempts to reduce **10w** with LiAlH<sub>4</sub> or NaBH<sub>4</sub> were unsuccessful,

**Scheme 3<sup>a</sup>**

<sup>a</sup> (i) **19** (R<sub>2</sub> = R<sub>3</sub> = Me), HCl, EtOH, 78 °C, 4 h; (ii) 1. MCPBA, CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 2 h, 2. Ac<sub>2</sub>O, AcONa, 140 °C, 8 h, 3. MMPP, CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 20 °C, 18 h, 4. 1 N NaOH, THF/MeOH, 20 °C, 1 h, 5. NH<sub>2</sub>OSO<sub>3</sub>H, AcONa, H<sub>2</sub>O, 20 °C, 18 h.

since partial saturation of the pyrimidine ring was observed while the ethoxycarbonyl group was unaffected. However, using diisobutylaluminum hydride (DIBAL) in THF, **10y** could be prepared in good yield. Various approaches were studied for the preparation of **10ad**, but only the oxidation of **10f** with KMnO<sub>4</sub>/NEt<sub>3</sub>/H<sub>2</sub>SO<sub>4</sub><sup>22</sup> allowed its isolation in moderate yields. The fluoromethyl derivative **10aa** was obtained upon treatment of **10y** with diethylaminosulfur trifluoride (DAST).

Other 6,7-dimethyl sulfones (**23**) were prepared from the corresponding amines **9** as described for **10f**. The sulfonamides **26** (Table 4, Scheme 3) were prepared from the methylthio derivatives **22** using a procedure<sup>23</sup> which involves monooxidation to the sulfoxide, Pummerer rearrangement to the acetoxymethylthio compound **24**, and oxidation with magnesium monoperoxy-

**Table 2.** Pyrazolopyrimidines of Formula 10


The chemical structure of Pyrazolopyrimidine 10 is shown above the table. It features a pyrazolo[1,5-a]pyrimidine core. The pyrimidine ring is numbered 1 to 7, and the pyrazole ring is numbered 1 to 2. Substituents include R<sub>1</sub> at position 5, R<sub>2</sub> at position 6, and R<sub>3</sub> at position 7. The pyrazole ring has a 4-fluorophenyl group at position 3 and a 4-methylsulfonylphenyl group at position 2.

compd	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	reactant, yield,		isomeric ratio <sup>b</sup>	mp <sup>c</sup> , °C	formula <sup>d</sup>	% Inh HWB <sup>e</sup>		
				method <sup>a</sup>	%				COX-1 10 μM	COX-2 10 μM	COX-2 1 μM
<b>10i</b>	H	H	CF <sub>3</sub>	<b>13</b> , B	46	10:0	213	C <sub>20</sub> H <sub>13</sub> F <sub>4</sub> N <sub>3</sub> O <sub>2</sub> S	0.0	90.6	3.5
<b>10j</b>	H	H	Pr	- <sup>g</sup>	27	-	194	C <sub>22</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	37.3	56.1	32.2
<b>10k</b>	H	H	Ph	<b>20</b> , C	35	10:0	231	C <sub>25</sub> H <sub>18</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	5.7	90.4	0
<b>10l</b>	CF <sub>3</sub>	H	Me	<b>11</b> , B	70	10:0	223	C <sub>21</sub> H <sub>15</sub> F <sub>4</sub> N <sub>3</sub> O <sub>2</sub> S	2.5	57.2	-
<b>10m<sup>f</sup></b>	COOEt	H	Me	<b>11</b> , B	68	5:5	98-99	C <sub>23</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>4</sub> S.0.5H <sub>2</sub> O	8.0	41.0	-
<b>10n<sup>f</sup></b>	COOH	H	Me	- <sup>g</sup>	78	-	-	C <sub>21</sub> H <sub>16</sub> FN <sub>3</sub> O <sub>4</sub> S.2H <sub>2</sub> O	33.5	24.6	-
<b>10o</b>	Bu	H	Me	<b>11</b> , B	31	5:5	57-61	C <sub>24</sub> H <sub>24</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	86.2 <sup>h</sup>	100	51.5
<b>10p</b>	Me	H	Bu	<b>11</b> , B	33	5:5	183	C <sub>24</sub> H <sub>24</sub> FN <sub>3</sub> O <sub>2</sub> S	0.0	58.3	11
<b>10q</b>	Me	H	OH	<b>15</b> , D	54	10:0	198-200	C <sub>20</sub> H <sub>16</sub> FN <sub>3</sub> O <sub>3</sub> S.H <sub>2</sub> O	8.1	21.9	-
<b>10r</b>	Me	H	OMe	- <sup>g</sup>	25	-	213	C <sub>21</sub> H <sub>18</sub> FN <sub>3</sub> O <sub>3</sub> S.0.75H <sub>2</sub> O	0.0	0.0	-
<b>10s</b>	Me	H	Cl	- <sup>g</sup>	11	-	230-232	C <sub>20</sub> H <sub>15</sub> ClFN <sub>3</sub> O <sub>2</sub> S.0.25H <sub>2</sub> O	32.5	14.3	-
<b>10t</b>	Me	Me	Cl	- <sup>g</sup>	88	-	253	C <sub>21</sub> H <sub>17</sub> ClFN <sub>3</sub> O <sub>2</sub> S.H <sub>2</sub> O	1.2	45.8	-
<b>10u</b>	H	Et	Me	<b>19</b> , C	45	9:1	166-170	C <sub>22</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	91.8 <sup>i</sup>	100	50.3
<b>10v</b>	H	COMe	Me	<b>20</b> , E	80	10:0	238	C <sub>22</sub> H <sub>18</sub> FN <sub>3</sub> O <sub>3</sub> S	71.1	100	60.8
<b>10w</b>	H	CO <sub>2</sub> Et	Me	<b>17</b> , C	50	9.5:0.5	201	C <sub>23</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>4</sub> S	0	100	40.5
<b>10x</b>	H	COOH	Me	- <sup>g</sup>	78	-	287-292	C <sub>21</sub> H <sub>16</sub> FN <sub>3</sub> O <sub>4</sub> S.1.25CHCl <sub>3</sub>	8.3	56.3	-
<b>10y</b>	H	CH <sub>2</sub> OH	Me	- <sup>g</sup>	28	-	268-270	C <sub>21</sub> H <sub>18</sub> FN <sub>3</sub> O <sub>3</sub> S.0.75H <sub>2</sub> O	54.4	77.7	26.1
<b>10z</b>	H	CF <sub>3</sub>	Me	<b>20</b> , E	36	8:2	208-215	C <sub>21</sub> H <sub>15</sub> F <sub>4</sub> N <sub>3</sub> O <sub>2</sub> S	100 <sup>j</sup>	100	69.0
<b>10aa</b>	H	CH <sub>2</sub> F	Me	- <sup>g</sup>	75	-	218-219	C <sub>21</sub> H <sub>17</sub> F <sub>2</sub> N <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	7.3	84.6	4.4
<b>10ab</b>	H	Br	Me	<b>20</b> , F	31	7:3	255-256	C <sub>25</sub> H <sub>15</sub> BrFN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	83.8 <sup>k</sup>	100	84.3
<b>10ac</b>	H	Me	Et	<b>19</b> , C	36	9:1	237	C <sub>22</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	82.8 <sup>l</sup>	100	62.3
<b>10ad</b>	H	Me	CH <sub>2</sub> OF	- <sup>g</sup>	33	-	226-227	C <sub>21</sub> H <sub>18</sub> FN <sub>3</sub> O <sub>3</sub> S.0.5CH <sub>2</sub> Cl <sub>2</sub>	42.7	100	56.5
<b>10ae</b>	H	Me	Ph	<b>20</b> , C	51	10:0	219	C <sub>26</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>2</sub> S.0.5H <sub>2</sub> O	0	12.3	-
<b>10af</b>	H	CO <sub>2</sub> H	Pr	- <sup>g</sup>	70	-	257	C <sub>23</sub> H <sub>20</sub> FN <sub>3</sub> O <sub>4</sub> S.0.5H <sub>2</sub> O	7.4	0.0	31.9

<sup>a,b,e</sup> See footnotes a–c of Table 1. <sup>c</sup> Melting points correspond generally to chromatographed products. <sup>d</sup> Elemental analyses for C, H, N, and S were within 0.4% of the theoretical values indicated. <sup>f</sup> Mixture of regioisomers. <sup>g</sup> See text and Experimental Section for method of synthesis. <sup>h</sup> 6.8% inhibition at 1 μM. <sup>i</sup> 0% inhibition at 1 μM. <sup>j</sup> 53.9% inhibition at 1 μM. <sup>k</sup> 18.9% inhibition at 1 μM. <sup>l</sup> 44.6% inhibition at 1 μM.

phthalate (MMPP), followed by treatment with NaOH to give the sodium sulfinate **25**. The reported process involved subsequent treatment with sulfonyl chloride and ammonia, but we found that on treatment of **25** with hydroxylamine *O*-sulfonic acid and NaOAc in H<sub>2</sub>O, the sulfonamides **26** were produced in one step and in somewhat better yields.

Triazines **29** and **33** (Table 5) were prepared as described in Scheme 4. Formation of the diazonium salt of amine **9a** with NaNO<sub>2</sub> followed by treatment<sup>24</sup> with the in situ generated β-keto acids from **28** afforded the [1,2,4]triazines **29**. The [1,3,5]triazines **33** were obtained from **9a** upon treatment<sup>25</sup> with a suitable ethyl imide **30**, followed by boiling in a triethyl orthoacylate **32**.

**Table 3.** Cyclopyrazolopyrimidines of Formula 10

% Inh HWB<sup>e</sup>

reactant, yield, isomeric

mp<sup>c</sup>,

COX-1

COX-2

compd

R<sub>2</sub>

R<sub>3</sub>

method<sup>a</sup>

%

ratio<sup>b</sup>

°C

formula<sup>d</sup>

10 μM

10 μM

1 μM

**10ag**

**20**, E

30

10:0

145-149

C<sub>23</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>3</sub>S.0.9CHCl<sub>3</sub>

0

95.3

30.4

**10ah**

**21**, G

42

5:5

193-194

C<sub>23</sub>H<sub>20</sub>FN<sub>3</sub>O<sub>2</sub>S.0.75H<sub>2</sub>O

40.4

100

57.7

**10ai**

**20**, E

30

10:0

232-233

C<sub>22</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>3</sub>S.0.3CHCl<sub>3</sub>

16.6

75.0

0

**10aj**

**21**, G

67

9.5:0.5

223-224

C<sub>22</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>2</sub>S.0.75H<sub>2</sub>O

0

90.4

48.3

**10ak**

**20**, E

18

10:0

217-219

C<sub>21</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>2</sub>S

7.0

90.7

14.8

**10al**

**20**, E

44

10:0

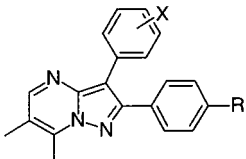
229-232

C<sub>21</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>2</sub>S

0

93.6

25.0

<sup>a-e</sup> See footnotes a–e of Table 2.**Table 4.** 6,7-Dimethylpyrazolopyrimidines of Formulas 23 and 26


compd <sup>a</sup>	R	R'	%Inh. HWB <sup>b</sup>		
			COX-1 10 μM	COX-2 10 μM	COX-2 1 μM
<b>26a</b>	4-F	SO <sub>2</sub> NH <sub>2</sub>	22.0	99.1	22.5
<b>23b</b>	4-Me	SO <sub>2</sub> Me	54.2	100	87.6
<b>26b</b>	4-Me	SO <sub>2</sub> NH <sub>2</sub>	33.5	100	65.7
<b>23c</b>	3,4-diF	SO <sub>2</sub> Me	36.3	100	100
<b>26c</b>	3,4-diF	SO <sub>2</sub> NH <sub>2</sub>	68.2	100	97.2
<b>23d</b>	2,4-diF	SO <sub>2</sub> Me	80.8	100	61.1

<sup>a</sup> Physicochemical properties described in the Experimental Section. <sup>b</sup> See footnote c of Table 1.

Other bicycles (Table 6) were prepared as described in Schemes 5 and 6. Imidazo[4,5-*c*]pyridines **38** were obtained from the corresponding 4-hydroxy-3-nitropyridines **34** upon treatment with POCl<sub>3</sub>, followed by nucleophilic displacement with 4-fluoroaniline, reduction to the amine **37** with SnCl<sub>2</sub> in EtOH, and final cyclization with 4-methylsulfonylbenzoyl chloride in

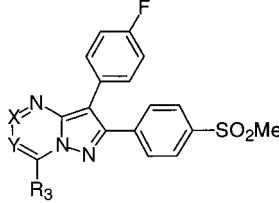
pyridine. On the other hand, the α-bromo ketone **39**, obtained from ketone **5a** upon treatment with bromine in CH<sub>2</sub>Cl<sub>2</sub>/AcOH, was reacted with the corresponding arylamines **40**, **42**, and **44** in DMF at 60 °C<sup>26</sup> to give imidazo[1,2-*b*]pyridazines **41**, imidazo[1,2-*a*]pyrazines **43**, and imidazo[1,2-*a*]pyrimidines **45**. Amines **40**, **42**, and **44** were commercially available or were prepared according to reported procedures (see Experimental Section). The Glaxo-reported bicycle **46** was also prepared from **39** as described.<sup>17</sup>

## Results and Discussion

The new compounds were tested in vitro for their ability to inhibit COX-1 and COX-2 activity in a human whole blood (HWB) assay.<sup>27</sup> Percent inhibition at a 10 μM concentration was initially determined, and active compounds were also tested at 1 μM (Tables 1–5). All compounds showing more than 60% inhibition at 10 μM were tested in vivo in the rat carrageenan-induced paw edema assay (CPE)<sup>28</sup> at a 30 mg/kg dose; some of them were also tested in the carrageenan-induced air-pouch model<sup>29</sup> to estimate PG production at a 1 mg/kg dose.

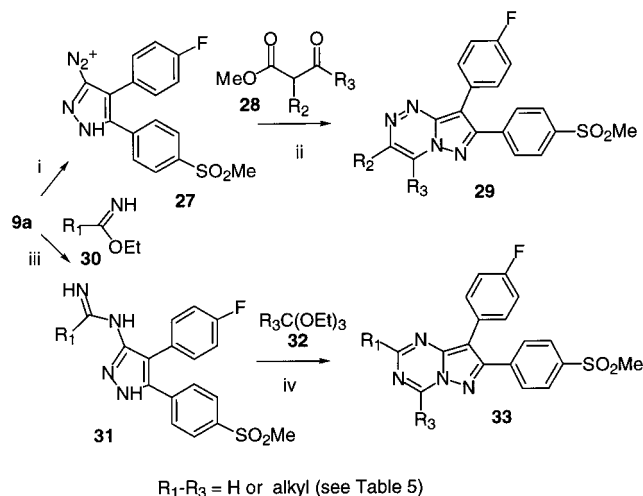
The main SAR was carried out with the 3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl) derivatives **10**, which are characterized by a 4-methylsulfonylphenyl group vicinal to a phenyl ring similarly to other COX-2-selective inhibitors. The gradual introduction of methyl groups into the poorly active parent unsubstituted compound (**10a**) was initially explored. In this initial series the activity in human cell lines expressing COX-1



**Table 5.** PyrAZolotriazines of Formulas **29** and **33**


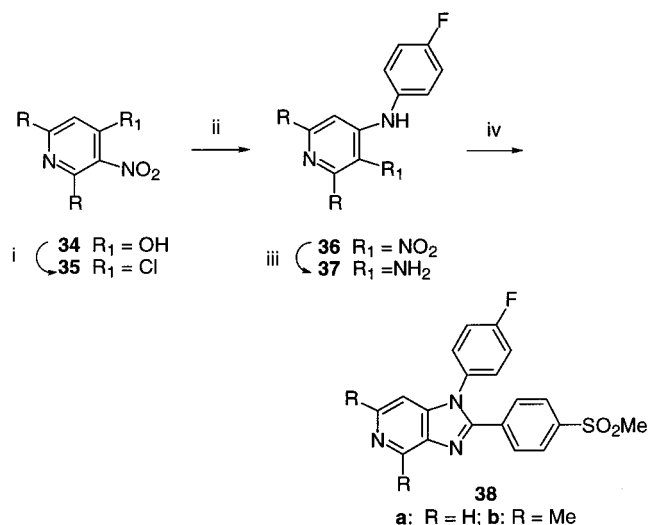
**29** X = N; Y = CR<sub>2</sub>;  
**33** X = CR<sub>1</sub>; Y = N

Compd <sup>a</sup>	R <sub>1,2</sub>	R <sub>3</sub>	%Inh. HWB <sup>b</sup>		
			COX-1 10 μM	COX-2 10 μM	COX-2 1 μM
<b>29a</b>	Me	Me	1.2	100	40.5
<b>29b</b>	H	Pr	29.3	100	43.9
<b>29c</b>	Et	Me	61.9	100	74.9
<b>29d</b>	Me	Pr	18.7	95.0	17.9
<b>29e</b>	H	Bu	69.8	89.7	0.9
<b>29f</b>	Et	Et	88.7	100	50.9
<b>33a</b>	Me	Me	4.75	90.3	18.1
<b>33b</b>	Me	Bu	7.7	54.6	-

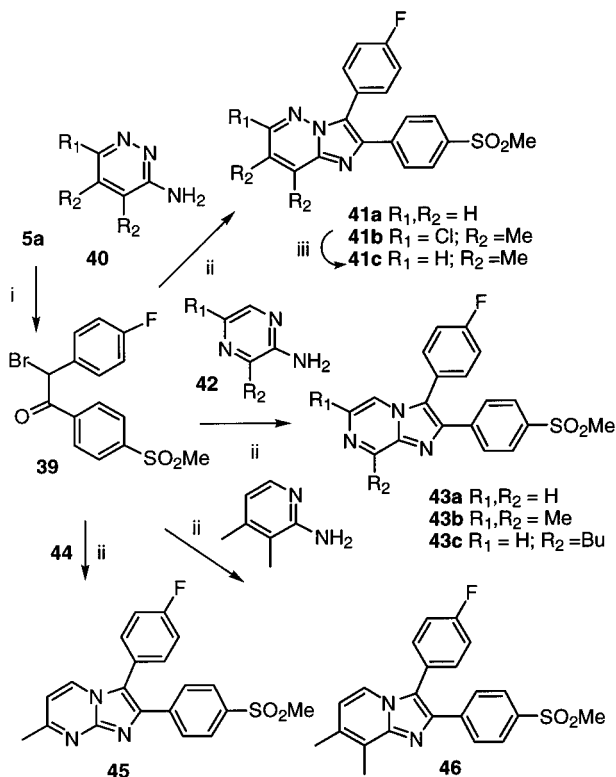
<sup>a,b</sup> See footnotes a,b of Table 4.**Scheme 4<sup>a</sup>**

<sup>a</sup> (i) NaNO<sub>2</sub>, H<sub>2</sub>O, 0 °C, 1 h; (ii) 1. KOH, H<sub>2</sub>O, 0 °C, 24 h, 2. NaOAc, HCl, H<sub>2</sub>O, 0–20 °C, 1 h; (iii) AcOH, 20 °C, 18 h; (iv) reflux, 18 h.

and COX-2<sup>30</sup> was also evaluated (Table 1). While substitution at the 5- or 6-position provided some increase in whole cell activity, a substantial gain was observed for the 7-methyl derivative **10c**. Its activity was maintained with a second methyl group in position 6 (**10f**) and impaired by introduction of additional methyl groups at the 5 (**10e**) or 5,6 (**10h**) positions. These results, together with the poor activity of the 5,6-disubstituted compound **10g**, indicated the need for substitution at position 7, which then tolerated a second substituent at position 6. The whole cell results cor-

**Scheme 5<sup>a</sup>**

<sup>a</sup> (i) POCl<sub>3</sub>, DMF, toluene, 110 °C, 7 h; (ii) 4-fluoroaniline, NEt<sub>3</sub>, CHCl<sub>3</sub>, 67 °C, 48 h; (iii) SnCl<sub>2</sub>, EtOH, 78 °C, 45 min; (iv) 4-methylsulfonylbenzoyl chloride, pyridine, 115 °C, 18 h.

**Scheme 6<sup>a</sup>**

<sup>a</sup> (i) Br<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, AcOH, 20 °C, 18 h; (ii) DMF, 60 °C, 18–36 h; (iii) H<sub>2</sub>, Pd/C, TEA, 200 psi, 18 h.

related quite well with those obtained in the whole blood assay. The most potent compounds in the whole cell assay, **10c,f**, gave also the highest inhibitions in HWB. When their IC<sub>50</sub> in HWB were calculated (**10c**: COX-1 = 10 μM, COX-2 = 0.6 μM; **10f**: COX-1 > 10 μM, COX-2 = 0.08 μM) the superior activity shown at 1 μM by **10f** was confirmed. This compound had excellent in vitro activity, which compared well with celecoxib (**2**), whether in whole cells (Table 1) or in HWB (**2** IC<sub>50</sub>: COX-1 = 13 μM, COX-2 = 0.6 μM).

Among the methylated derivatives only compound **10f** showed some in vivo activity. In the air-pouch model it

showed 74% inhibition of PGE<sub>2</sub> production at 1 mg/kg, approximately one-third of that shown by **2** (93.4% and 71.3% inhibition at 1 and 0.3 mg/kg, respectively). In the CPE model **10f** provided around 20% of edema inhibition at 30 as well as at 10 mg/kg, in comparison to 43% shown by **2** at 10 mg/kg. In initial pharmacokinetic evaluation in the rat, compound **10f** had a short half-life (1 h) and poor bioavailability.

With the aim of improving these results and keeping the 7-position substituted, we introduced further variation. Changing the methyl of **10c** for CF<sub>3</sub> (**10i**) or lengthening to propyl (**10j**) or phenyl (**10k**) clearly diminished activity. Replacing the 5-methyl of **10e** by CF<sub>3</sub>, COOEt, or COOH (**10l–n**) was clearly detrimental, while lengthening it to butyl provided the more active albeit less selective compound **10o**. Substitution of the 7-methyl of **10e** by butyl, hydroxy, methoxy, or chloro (**10p–t**) produced a complete loss in activity. Next, some 6,7-disubstituted analogues of **10f** were prepared and evaluated. Replacing the 6-methyl group by ethyl (**10u**), acetyl (**10v**), CF<sub>3</sub> (**10z**), or Br (**10ab**) maintained activity but provided some decrease in selectivity, whereas replacement by COOH, COOEt, CH<sub>2</sub>OH, or CH<sub>2</sub>F (**10w–y,aa**) was more detrimental. Elongation of the 7-methyl group to ethyl provided a loss in selectivity (**10ac**), while the 7-phenyl (**10ae**) completely lost potency, as in the case of **10k**. The 7-hydroxymethyl derivative, which was prepared together with **10y** as a possible metabolite of **10f**, retained its potency. Among these derivatives only the ethyl ester **10w** and the monohydroxylated **10y** showed some in vivo activity in the CPE test (20.2 and 17.3% inhibition at 30 mg/kg, respectively), somewhat inferior to that of **10f**.

Since 6,7-disubstitution seemed to provide the best activity, several 6,7-cyclo derivatives (Table 3) were prepared. The naked cyclohexyl (**10ah**) and cyclopentyl (**10aj**) derivatives had a profile similar to that of **10f**, while substitution by heteroatoms was detrimental. Despite this low in vitro potency, the cyclohexanone **10ag** and cyclopentanone **10ai** were the only ones displaying activity in the CPE test (22.4 and 26.3% inhibition at 30 mg/kg, respectively).

These results showed a restricted SAR around the pyrimidine substituents, which precluded their further optimization. Maintaining the 6,7-dimethyl groups, we decided to introduce some changes in the nature of the aromatic ring and explore changing the SO<sub>2</sub>Me for an SO<sub>2</sub>NH<sub>2</sub> group, a switch that in some instances has been shown to improve in vivo activity through an improvement of bioavailability.<sup>14</sup> In this case, the sulfonamide **26a** was equipotent to its sulfone counterpart **10f** in the whole cell assay (IC<sub>50</sub>: COX-1 > 10  $\mu$ M, COX-2 = 0.019  $\mu$ M), although in the HWB assay **26a** showed less potency than expected. This result contrasted with that generally observed in the diaryl C5-heterocycle class, where this group usually provides more potency albeit less selectivity.<sup>14</sup> Although the 4-methyl and difluoro derivatives **23b–d** and **26b,c** were highly potent in the HWB assay, none of them showed improved in vivo results and only compound **26b** showed some activity in the CPE test (22.8% inhibition at 10 mg/kg).

To improve in vivo results, the introduction of a second nitrogen atom in the pyrimidine ring was envisaged and triazines **29** and **33** were prepared (Table 5).

**Table 6.** Heterocycle Variations: Compounds of Schemes 5 and 6

Compd <sup>a</sup>	% Inh. HWB <sup>b</sup>		
	COX-1	COX-2	
	10 $\mu$ M	10 $\mu$ M	1 $\mu$ M
<b>38a</b>	34.3	21.3	-
<b>38b</b>	0	4.3	-
<b>41a</b>	47.0	61.2	-
<b>41b</b>	0	100	35.4
<b>41c</b>	0	100	37.3
<b>43a</b>	6.0	42.5	-
<b>43b</b>	0	82.4	0
<b>43c</b>	9.8	0	-
<b>45</b>	13.5	50.7	-
<b>46</b>	0	66.7	59.3

<sup>a,b</sup> See footnotes a,b of Table 4.

The HWB activity of the 6,7-dimethyl-1,3,5-triazine **29a** was inferior to that of **10f**, although in this case it was improved upon lengthening to the 6-ethyl derivative **29c**. However, a second ethyl group (**29f**) diminished selectivity. The 7-propyl **29b** was more active than its pyrimidine counterpart **10j**, but further lengthening (**29e**) or introduction of a methyl group (**29d**) failed to improve this result. On the other hand, the 1,3,4-triazines **33** lost activity in relation to their pyrimidine counterparts. Among these compounds only **29a,b** showed in vivo activity, but again neither of them improved the potency of **10f**.

Finally, other bicyclic frameworks, which eventually could provide the desired in vivo properties, were evaluated (Table 6). As in the pyrazolopyrimidine series, the unsubstituted imidazopyridine compound **38a** was inactive, but in this case the introduction of two methyl groups at the 6- and 7-positions (**38b**) impaired activity even further. As far as the imidazopyrazines were concerned, the poor activity of the parent **43a** could not be improved upon introduction of alkyl groups, and compounds **43b,c** were much less potent than their pyrazolopyrimidine counterparts **10c,e**. Moreover, around 10 different substituted imidazopyrimidines were prepared (results not shown), and as in the case of **45**, none of them showed significant COX-2 inhibitory activity.

Among these modifications only the imidazopyridazine framework **41** showed any activity. The presence of the two methyl groups was also important in this case, since **41c** was much more active than the parent **41a**. However, none of these derivatives showed any in vivo activity. For purposes of comparison, the imidazopyridine **46** described in the patent literature<sup>17</sup> was prepared but exhibited less potency both in vitro and in vivo than its counterpart **10f**.

Altogether these results indicate that a nitrogen atom at position 4 (**10f** vs **46**) increases activity, whereas it is merely tolerated at position 5 (**29**). On the contrary,

its presence in position 6 (compounds **33**, **38**, and **43**) or 7 (**45**) is clearly detrimental.

## Conclusion

In summary, the tight SAR around the pyrazolo[1,5-*a*]pyrimidine nucleus indicated that 6,7-disubstitution provided the best activity and led to identification of the 6,7-dimethylated derivative **10f** as one of the most potent and selective COX-2 inhibitor in this series. While **10f** exhibited a remarkable in vitro potency and selectivity profile in relation to reference compounds, it was less active in vivo due to low oral bioavailability. In vivo activity could not be improved by introducing more polar groups in the side chains (Tables 1, 2), changing the methylsulfonyl to sulfonamide groups (Table 4), or introducing more nitrogen atoms to the nucleus (Table 6). The pyrazolo[1,5-*a*]pyrimidine and imidazo[1,2-*b*]pyridazine were the best bicyclic frameworks among those studied, indicating that the presence of a nitrogen atom at position 4 of the central core was positive, at position 5 was tolerated, and at position 6 or 7 was clearly detrimental. Further efforts to discover novel orally active COX-2-selective inhibitors related to these structures are underway and will be reported in due course.

## Experimental Section

Melting points were determined with a Mettler FP 80 central processor melting point apparatus and are uncorrected. Infrared spectra were recorded on a Perkin-Elmer 983 spectrophotometer. <sup>1</sup>H (80 MHz) and <sup>13</sup>C (20.1 MHz) NMR spectra were recorded on a Brücker AC 80 spectrometer and <sup>1</sup>H (300 MHz) NMR spectra were recorded on a Brücker Avance DPX-300 spectrometer. They are reported in ppm on the  $\delta$  scale, from the indicated reference. Combustion analyses were performed with a Carlo Erba 1106 analyzer. Liquid chromatography was performed with a forced flow (flash chromatography) of the indicated solvent system on SDS silica gel Chromagel 60 ACC (230–400 mesh). Analytical thin-layer chromatography (TLC) was performed with Macherey-Nagel 0.25-mm silica gel SIL G-25 plates.

**3-Chloro-2-(4-fluorophenyl)-3-(4-methylsulfonylphenyl)propenenitrile (7a).** A mixture of DMF (33.5 mL, 40 mmol) and POCl<sub>3</sub> (14.25 mL, 155 mmol) was stirred at 0 °C under argon for 15 min. To this solution, **5a**<sup>19</sup> (15.2 g, 52 mmol) in CHCl<sub>3</sub> (310 mL) was added dropwise and the mixture was heated at 80 °C overnight. The mixture was cooled at 0 °C and NH<sub>2</sub>OH·HCl (10.5 g, 154 mmol) in DMF (29.25 mL) was added and stirred for 4 h at room temperature. The suspension was poured over saturated NaHCO<sub>3</sub> solution and extracted with CHCl<sub>3</sub>. The organic phase was dried and concentrated to a crude product which was used directly in the next step (20 g): <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 3.10 (s, 3 H), 7.06 (m, 2 H), 7.23 (m, 2 H), 7.50 (d, *J* = 7 Hz, 2 H), 7.90 (d, *J* = 7 Hz, 2 H).

**3-Amino-4-(4-fluorophenyl)-5-(4-methylsulfonylphenyl)pyrazole (9a).** To a solution of crude **7a** (20 g, 52 mmol) in EtOH (546 mL), NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O (5.4 mL, 104 mmol) was added and the mixture was stirred at reflux overnight. The solvent was concentrated and the residue poured over a mixture of H<sub>2</sub>O and AcOEt. The organic phase was extracted with AcOEt and the combined organic extracts were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **9a** as a white solid (9.0 g, 52%): mp 192–193 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 3.06 (s, 3 H), 3.92 (s, 3 H), 7.05 (m, 2 H), 7.20 (m, 2 H), 7.52 (d, *J* = 7 Hz, 2 H), 7.80 (d, *J* = 7 Hz, 2 H).

**3-(4-Fluorophenyl)-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10a).** Method A. A mixture of **9a** (0.41 g, 1.33 mmol), EtOH (1 mL), concentrated HCl (2.6 mL) and ZnCl<sub>2</sub> (0.08 g, 0.57 mmol) was heated to reflux and 1,1,3,3-

tetramethoxypropane (**12**; 0.164 mL, 1 mmol) in EtOH (0.4 mL) was added. After 1 h at reflux the solution thus obtained was poured over ice and basified with NH<sub>4</sub>OH. The mixture was extracted with AcOEt and the organic phase dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10a** as a white solid (0.34 g, 76%): mp 184–185 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 3.06 (s, 3 H), 6.93 (dd, *J* = 4 Hz, *J* = 7 Hz, 1 H), 7.13 (m, 2 H), 7.48 (m, 2 H), 7.86 (d, *J* = 7 Hz, 2 H), 7.95 (d, *J* = 7 Hz, 2 H), 8.49 (dd, *J* = 1.8 Hz, *J* = 4 Hz, 1 H), 8.69 (dd, *J* = 1.8 Hz, *J* = 7 Hz, 1 H). Anal. (C<sub>19</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S.

**3-(4-Fluorophenyl)-5,7-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10e).** Method B. A mixture of **9a** (0.51 g, 1.7 mmol), acetylacetone (**11**, R<sub>1</sub>, R<sub>3</sub> = Me, R<sub>2</sub> = H; 0.176 g, 1.7 mmol), 1 drop of piperidine and EtOH (8 mL) was refluxed for 18 h. The solvent was concentrated and the residue was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10e** as a white solid (0.48 g, 80%): mp 220 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 2.59 (s, 3 H), 2.81 (s, 3 H), 3.07 (s, 3 H), 6.67 (s, 1 H), 7.07 (m, 2 H), 7.47 (m, 2 H), 7.87 (d, *J* = 7 Hz, 2 H), 7.93 (d, *J* = 7 Hz, 2 H). Anal. (C<sub>21</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S.

Using the same procedure but starting from 4-ethoxy-3-buten-2-one (**13**, R<sub>3</sub> = Me), a 1:4 mixture of **10b,c** was obtained.

**10b:** 11% yield; mp 209–216 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 2.63 (s, 3 H, 5-Me), 3.09 (s, 3 H, SO<sub>2</sub>Me), 6.78 (d, *J* = 7.2 Hz, 1 H, H<sub>6</sub>), 7.09 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.48 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.82 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 7.94 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 8.56 (d, *J* = 7.2 Hz, 1 H, H<sub>7</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>  $\delta$  TMS) 26.51 (CH<sub>3</sub>, 5-Me), 45.90 (CH<sub>3</sub>, SO<sub>2</sub>Me), 109.44 (C), 110.64 (CH), 117.11 (CH, d, *J*<sub>F-C</sub> = 20 Hz), 128.54 (C), 129.73 (CH), 131.03 (CH), 133.19 (CH, d, *J*<sub>F-C</sub> = 8 Hz), 135.52 (CH, C<sub>7</sub>), 140.14 (C), 141.59 (C), 147.88 (C), 152.97 (C), 161.43 (C), 163.45 (C, d, *J*<sub>F-C</sub> = 245 Hz). Anal. (C<sub>20</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S.

**10c:** 44% yield; mp 192 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 2.88 (s, 3 H, 7-Me), 3.09 (s, 3 H, SO<sub>2</sub>Me), 6.80 (d, *J* = 4 Hz, 1 H, H<sub>6</sub>), 7.10 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.48 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.89 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 7.96 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 8.46 (d, *J* = 4 Hz, 1 H, H<sub>5</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>  $\delta$  TMS) 18.53 (CH<sub>3</sub>, 7-Me), 45.90 (CH<sub>3</sub>, SO<sub>2</sub>Me), 110.16 (CH), 110.49 (C), 117.22 (CH, d, *J*<sub>F-C</sub> = 20 Hz), 128.75 (C), 128.96 (CH), 131.14 (CH), 133.05 (CH, d, *J*<sub>F-C</sub> = 8 Hz), 140.24 (C), 141.61 (C), 147.58 (C), 148.59 (C), 150.85 (CH, C<sub>5</sub>), 152.35 (C), 163.65 (C, d, *J*<sub>F-C</sub> = 245 Hz). Anal. (C<sub>20</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>2</sub>S·H<sub>2</sub>O) C, H, N, S.

Compound **10b** was also obtained by the following procedure: A mixture of **10s** (0.13 g, 0.35 mmol), EtOH (10 mL), Pd/C (5%) (0.075 g) and NaOAc (0.070 g, 0.85 mmol) was hydrogenated at atmospheric pressure for 1 h. The suspension thus obtained was filtered over Celite, concentrated, and recrystallized twice from AcOEt to give **10b** as a white solid (0.098 g, 71%).

Compound **10c** was also obtained on decarboxylation of **10j** using the conditions described below for **10j**.

**3-(4-Fluorophenyl)-6,7-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10f).** Method C. A mixture of **9a** (0.4 g, 1.2 mmol), 3-methyl-4-methoxy-3-buten-2-one<sup>21</sup> (**19**, R<sub>2</sub> = R<sub>3</sub> = Me; 0.176 g, 1.2 mmol), HCl (0.1 mL) and EtOH (8 mL) was refluxed for 4 h. The solvent was concentrated and the residue was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10f** as a white solid (0.330 g, 70%): mp 202 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>  $\delta$  TMS) 2.42 (s, 3 H, 6-Me), 2.85 (s, 3 H, 7-Me), 3.08 (s, 3 H, SO<sub>2</sub>Me), 7.12 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.48 (m, 2 H, C<sub>6</sub>H<sub>4</sub>F), 7.88 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 7.94 (d, *J* = 7 Hz, 2 H, C<sub>6</sub>H<sub>4</sub>SO<sub>2</sub>Me), 8.36 (s, 1 H, H<sub>5</sub>). Anal. (C<sub>21</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S. Additionally, 0.039 g of **10f** was obtained (8%).

The same procedure was used for the preparation of **10d** and **23b–d** starting from **9b–d** respectively. **10d:** mp 234 °C; Anal. (C<sub>20</sub>H<sub>16</sub>FN<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S. **23b:** mp 244 °C; Anal. (C<sub>22</sub>H<sub>21</sub>N<sub>3</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S. **23c:** mp 230–232 °C; Anal. (C<sub>21</sub>H<sub>17</sub>F<sub>2</sub>N<sub>3</sub>O<sub>2</sub>S·0.5H<sub>2</sub>O) C, H, N, S. **23d:** mp 189–194 °C; Anal. (C<sub>21</sub>H<sub>17</sub>F<sub>2</sub>N<sub>3</sub>O<sub>2</sub>S) C, H, N, S.



**3-(4-Fluorophenyl)-5,6-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10g).** Method D. A mixture of **9a** (0.30 g, 0.9 mmol), 4-hydroxy-3-methyl-3-buten-2-one (**21**,  $R_2 = R_3 = \text{Me}$ ; 0.11 g, 0.9 mmol) and toluene (2 mL) was refluxed for 2 h. The solvent was concentrated and the residue was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10g** as a white solid (0.22 g, 62%): mp 242–243 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.37 (s, 3 H, 6-Me), 2.60 (s, 3 H, 5-Me), 3.10 (s, 3 H,  $\text{SO}_2\text{Me}$ ), 7.12 (m, 2 H,  $\text{C}_6\text{H}_4\text{F}$ ), 7.48 (m, 2 H,  $\text{C}_6\text{H}_4\text{F}$ ), 7.84 (d,  $J = 7$  Hz, 2 H,  $\text{C}_6\text{H}_4\text{SO}_2\text{Me}$ ), 7.95 (d,  $J = 7$  Hz, 2 H,  $\text{C}_6\text{H}_4\text{SO}_2\text{Me}$ ), 8.42 (s, 1 H,  $\text{H}_7$ ). Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_2\text{S} \cdot \text{CHCl}_3$ ) C, H, N, S. Additionally, 0.024 g of **10f** was obtained (6%).

The same procedure was used for the preparation of **10h**: mp 202 °C; Anal. ( $\text{C}_{22}\text{H}_{20}\text{FN}_3\text{O}_2\text{S} \cdot 0.5\text{H}_2\text{O}$ ) C, H, N, S.

Compound **10g** was also obtained on hydrogenation of **10t** over Pd/C/NaOAc as described above for the preparation of **10b**.

**6-Acetyl-3-(4-fluorophenyl)-7-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10v).** Method E. A mixture of **9a** (0.30 g, 0.9 mmol), 3-dimethylaminomethylene-2,4-pentanedione (0.13 g, 0.9 mmol; **20**,  $R_3 = \text{Me}$ ,  $R_2 = \text{COMe}$ ), obtained by reaction of 2,4-pentanedione and 1.6 equiv of dimethylformamide dimethylacetal in THF at room temperature overnight) and AcOH (1.5 mL) was refluxed for 1 h. The solvent was eliminated and the residue was dissolved in  $\text{CHCl}_3$  and washed with saturated  $\text{NaHCO}_3$  and NaCl solution. The organic phase was dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10v** as a white solid (0.14 g, 37%): mp 238 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.73 (s, 3 H), 3.09 (s, 3 H), 3.24 (s, 3 H), 7.14 (m, 2 H), 7.48 (m, 2 H), 7.90 (d,  $J = 7$  Hz, 2 H), 7.97 (d,  $J = 7$  Hz, 2 H), 8.90 (s, 1 H). Anal. ( $\text{C}_{22}\text{H}_{18}\text{FN}_3\text{O}_3\text{S}$ ) C, H, N, S.

**6-Bromo-3-(4-fluorophenyl)-7-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10ab).** Method F. A mixture of **9a** (0.4 g, 1.2 mmol), 3-bromo-4-diethylamino-3-buten-2-one (**20**,  $R_2 = \text{Br}$ ,  $R_3 = \text{Me}$ ), obtained by bromination with  $\text{Br}_2$  in  $\text{CHCl}_3$  of 4-diethylamino-3-buten-2-one; 0.340 g, 1.6 mmol), HBr (0.1 mL) and EtOH (4 mL) was refluxed for 1 h. The solvent was concentrated and the residue was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10ab** as a white solid (0.170 g, 31%): mp 255–256 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 3.01 (s, 3 H), 3.08 (s, 3 H), 7.13 (m, 2 H), 7.47 (m, 2 H), 7.89 (d,  $J = 7$  Hz, 2 H), 7.94 (d,  $J = 7$  Hz, 2 H), 8.52 (s, 1 H). Anal. ( $\text{C}_{25}\text{H}_{15}\text{BrFN}_3\text{O}_2\text{S} \cdot 0.5\text{H}_2\text{O}$ ) C, H, N, S.

**3-(4-Fluorophenyl)-6,7,8,9-tetrahydro-2-(4-methylsulfonylphenyl)-6-oxopyrazolo[1,5-*a*]quinazoline (10ag).** Method E. Following the same procedure described for the preparation of **10v** but starting from 1,3-cyclohexanedione, compound **10ag** was obtained as a sole isomer in 30% yield: mp 145–149 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.41 (m, 2 H), 2.78 (m, 2 H), 3.10 (s, 3 H), 3.58 (m, 2 H), 7.14 (m, 2 H), 7.47 (m, 2 H), 7.89 (d,  $J = 7$  Hz, 2 H), 7.97 (d,  $J = 7$  Hz, 2 H), 9.05 (s, 1 H).

**3-(4-Fluorophenyl)-6,7,8,9-tetrahydro-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]quinazoline (10ah).** Method G. A mixture of **9a** (0.5 g, 1.5 mmol), 2-hydroxymethylcyclohexanone<sup>31</sup> (**21**; 0.189 g, 1.5 mmol), *p*-toluenesulfonic acid (0.011 g) and toluene (19 mL) was refluxed for 1 h. The solvent was concentrated and the residue chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10ah** as a white solid (0.27 g, 43%), together with its regioisomer 3-(4-fluorophenyl)-6,7,8,9-tetrahydro-2-(4-methylsulfonylphenyl)pyrazolo[5,1-*b*]quinazoline (0.26 g, 42%): mp 193–194 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.0 (m, 4 H), 2.85 (m, 2 H), 3.07 (s, 3 H), 3.21 (m, 2 H), 7.10 (m, 2 H), 7.47 (m, 2 H), 7.87 (d,  $J = 7$  Hz, 2 H), 7.93 (d,  $J = 7$  Hz, 2 H), 8.32 (s, 1 H). Anal. ( $\text{C}_{23}\text{H}_{20}\text{FN}_3\text{O}_2\text{S} \cdot 0.75\text{H}_2\text{O}$ ) C, H, N, S.

**3-(4-Fluorophenyl)-2-(4-methylsulfonylphenyl)-7-propylpyrazolo[1,5-*a*]pyrimidine (10j).** A mixture of **10af** (0.124 g, 0.27 mmol) and 40%  $\text{H}_2\text{SO}_4$  (1.2 mL) was refluxed for 18 h. After cooling, the mixture was poured over  $\text{H}_2\text{O}$  and basified with 2 N NaOH. The aqueous phase was extracted

with  $\text{CHCl}_3$  and the organic phase was dried and concentrated to a crude product, which was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10j** as a white solid (0.03 g, 27%): mp 194 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 1.15 (t,  $J = 7$  Hz, 3 H), 2.02 (m, 2 H), 3.10 (s, 3 H), 3.26 (t,  $J = 7$  Hz, 2 H), 6.80 (d,  $J = 4.5$  Hz, 1 H), 7.14 (m, 2 H), 7.49 (m, 2 H), 7.90 (d,  $J = 7$  Hz, 2 H), 7.96 (d,  $J = 7$  Hz, 2 H), 8.49 (d,  $J = 4.5$  Hz, 1 H). Anal. ( $\text{C}_{22}\text{H}_{20}\text{FN}_3\text{O}_2\text{S} \cdot 0.5\text{H}_2\text{O}$ ) C, H, N, S.

**3-(4-Fluorophenyl)-5-methyl-2-(4-methylsulfonylphenyl)-7-methoxypyrazolo[1,5-*a*]pyrimidine (10r).** A mixture of **10q** (0.51 g, 1.36 mmol), acetone (4.2 mL),  $\text{Me}_2\text{SO}_4$  (0.13 mL, 1.36 mmol),  $\text{K}_2\text{CO}_3$  (0.186 g, 1.36 mmol) and KI (0.042 g, 0.2 mmol) was refluxed for 2 h. The solvent was concentrated and the residue dissolved in a mixture  $\text{CHCl}_3$ – $\text{H}_2\text{O}$ . The organic phase was extracted with  $\text{CHCl}_3$  and the combined organic extracts were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10r** as a white solid (0.18 g, 25%): mp 213 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.40 (s, 3 H), 3.14 (s, 3 H), 3.88 (s, 3 H), 6.10 (s, 1 H), 7.03 (m, 2 H), 7.37 (m, 2 H), 7.61 (d,  $J = 7$  Hz, 2 H), 8.04 (d,  $J = 7$  Hz, 2 H). Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_3\text{S} \cdot 0.75\text{H}_2\text{O}$ ) C, H, N, S.

**7-Chloro-3-(4-fluorophenyl)-5-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10s).** A mixture of **10q** (0.33 g, 0.9 mmol) and  $\text{POCl}_3$  (1 mL) was refluxed for 1 h. After concentration the residue was dissolved in ice– $\text{H}_2\text{O}$  and basified with  $\text{NH}_4\text{OH}$ . The mixture was extracted with AcOEt and the organic phase was dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10s** as a white solid (0.04 g, 11%): mp 230–232 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.65 (s, 3 H), 3.09 (s, 3 H), 6.96 (s, 1 H), 7.13 (m, 2 H), 7.48 (m, 2 H), 7.87 (d,  $J = 7$  Hz, 2 H), 7.96 (d,  $J = 7$  Hz, 2 H). Anal. ( $\text{C}_{20}\text{H}_{15}\text{ClFN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S.

Using the same procedure from the corresponding hydroxy derivative [3-(4-fluorophenyl)-5,6-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidin-7-ol, obtained from **9a** and **15** ( $R_1, R_2 = \text{Me}$ ) under conditions of method D: mp 293–298 °C; Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_3\text{S} \cdot \text{H}_2\text{O}$ ) C, H, N, S] **10t** was obtained in 88% yield.

**3-(4-Fluorophenyl)-7-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine-6-carboxylic Acid (10x).** A mixture of **10w** (0.23 g, 0.54 mmol), KOH (0.13 g, 1.4 mmol),  $\text{H}_2\text{O}$  (1 mL) and EtOH (10 mL) is heated at reflux overnight. The solvent was concentrated and the residue poured over a mixture of  $\text{H}_2\text{O}$  and AcOEt. The aqueous phase was washed with AcOEt and acidified with 6 N HCl. This was extracted with  $\text{CHCl}_3$ , which was dried and concentrated to afford **10x** as a white solid (78%): mp 287–292 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3 + \text{CD}_3\text{OD}$   $\delta$  TMS) 3.09 (s, 3 H), 3.25 (s, 3 H), 4.28 (s, 1 H +  $\text{H}_2\text{O}$ ), 7.07 (m, 2 H), 7.42 (m, 2 H), 7.89 (d,  $J = 7$  Hz, 2 H), 7.91 (d,  $J = 7$  Hz, 2 H), 8.97 (s, 1 H). Anal. ( $\text{C}_{21}\text{H}_{16}\text{FN}_3\text{O}_4\text{S} \cdot 1.25\text{CHCl}_3$ ) C, H, N, S.

The same procedure was used for the preparation of **10n** from **10m**.

**3-(4-Fluorophenyl)-6-hydroxymethyl-7-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10y).** To a solution of **10w** (0.2 g, 0.44 mmol) in THF (4 mL) at  $-78$  °C, a solution of DIBAL (1.24 mL, 1.24 mmol) 1 M in THF was added and the mixture stirred at  $-78$  °C for 2 h and at room temperature overnight.  $\text{H}_2\text{O}$  was added and the solution extracted with AcOEt. The organic phase was dried and concentrated to a crude product, which was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10y** as a yellow solid (0.05 g, 28%): mp 268–270 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3 + \text{CD}_3\text{OD}$   $\delta$  TMS) 2.95 (s, 3 H), 3.11 (s, 3 H), 3.49 (s, 1 H +  $\text{H}_2\text{O}$ ), 4.79 (s, 2 H), 7.13 (m, 2 H), 7.46 (m, 2 H), 7.89 (d,  $J = 7$  Hz, 2 H), 7.94 (d,  $J = 7$  Hz, 2 H), 8.51 (s, 1 H). Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_3\text{S} \cdot 0.75\text{H}_2\text{O}$ ) C, H, N, S.

**3-(4-Fluorophenyl)-6-fluoromethyl-7-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10aa).** A mixture **10y** (0.2 g, 0.48 mmol), DAST (0.067 g, 0.52 mmol) and  $\text{CH}_2\text{Cl}_2$  (8 mL) was stirred at room temperature for 2 h. The solution thus obtained was washed with  $\text{H}_2\text{O}$  and the

organic phase was dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10aa** as a white solid (0.15 g, 75%): mp 218–219 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 2.99 (d, *J* = 3 Hz, 3 H), 3.09 (s, 3 H), 3.57 (d, *J* = 48 Hz, 2 H), 7.13 (m, 2 H), 7.47 (m, 2 H), 7.89 (d, *J* = 7 Hz, 2 H), 7.94 (d, *J* = 7 Hz, 2 H), 8.50 (s, 1 H). Anal. (C<sub>21</sub>H<sub>17</sub>F<sub>2</sub>N<sub>3</sub>O<sub>2</sub>S·0.5H<sub>2</sub>O) C, H, N, S.

**3-(4-Fluorophenyl)-7-hydroxymethyl-6-methyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine (10ad).** A mixture of **10f** (0.2 g, 0.5 mmol), 6 N H<sub>2</sub>SO<sub>4</sub> (0.6 mL) and CHCl<sub>3</sub> (0.5 mL) was added to a suspension of KMnO<sub>4</sub> (0.079 g, 0.5 mmol), NEt<sub>3</sub> (0.07 g, 0.5 mmol), H<sub>2</sub>O (0.1 mL) and CHCl<sub>3</sub> (2 mL) and it was stirred at room temperature for 3 days. The mixture was filtered over a silica gel/CaCl<sub>2</sub> (2.5 g/0.5 g) mixture and washed thoroughly with CHCl<sub>3</sub>. The organic phase was washed with a saturated NaHCO<sub>3</sub> solution, dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **10ad** as a white solid, which was recrystallized from CH<sub>2</sub>Cl<sub>2</sub> (0.07 g, 33%): mp 226–227 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 1.55 (s, 1 H), 2.25 (s, 3 H), 3.09 (s, 3 H), 5.20 (s, 2 H), 7.13 (m, 2 H), 7.47 (m, 2 H), 7.89 (m, 4 H), 8.42 (s, 1 H). Anal. (C<sub>21</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>3</sub>S·0.5CH<sub>2</sub>Cl<sub>2</sub>) C, H, N, S.

**3-(4-Fluorophenyl)-2-(4-methylsulfonylphenyl)-7-propylpyrazolo[1,5-*a*]pyrimidine-6-carboxylic Acid (10af).** A mixture of ethyl 3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)-7-propylpyrazolo[1,5-*a*]pyrimidine-6-carboxylate [obtained from **9a** and **16** (R<sub>3</sub> = Pr) under conditions of method C; 0.26 g, 0.54 mmol], KOH (0.13 g, 1.4 mmol), H<sub>2</sub>O (1 mL) and EtOH (10 mL) was heated at reflux overnight. The solvent was concentrated and the residue poured over a mixture of H<sub>2</sub>O and AcOEt. The aqueous phase was washed with AcOEt and acidified with 6 N HCl. This was extracted with CHCl<sub>3</sub>, which was dried and concentrated to afford **10af** as a white solid (70%): mp: 257 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 1.20 (t, *J* = 7 Hz, 3 H), 1.98 (m, 2 H), 3.12 (s, 3 H), 3.85 (t, *J* = 7 Hz, 2 H), 7.17 (m, 2 H), 7.48 (m, 2 H), 7.91 (d, *J* = 7 Hz, 2 H), 8.00 (d, *J* = 7 Hz, 2 H), 9.09 (s, 1 H), 10.63 (s, 1 H). Anal. (C<sub>23</sub>H<sub>20</sub>FN<sub>3</sub>O<sub>4</sub>S·0.5H<sub>2</sub>O) C, H, N, S.

**4-[3-(4-Fluorophenyl)-6,7-dimethylpyrazolo[1,5-*a*]pyrimidin-2-yl]benzenesulfonamide (26a).** To a solution of **22a** (0.37 g, 1.0 mmol, obtained from **8a** as described for **10f**) in CH<sub>2</sub>Cl<sub>2</sub> (2.5 mL) was added *m*-chloroperbenzoic acid (0.32 g, 1.0 mmol) at 0 °C and the mixture was stirred for 2 h at room temperature. The suspension was washed with 1 N NaOH and the aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic phases were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford 3-(4-fluorophenyl)-6,7-dimethyl-2-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*]pyrimidine as a white solid (0.32 g, 82%): <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 2.41 (s, 3 H), 2.75 (s, 3 H), 2.84 (s, 3 H), 7.10 (m, 2 H), 7.46 (m, 2 H), 7.67 (d, *J* = 7 Hz, 2 H), 7.83 (d, *J* = 7 Hz, 2 H), 8.35 (s, 1 H).

The previous compound was treated with Ac<sub>2</sub>O (2.6 mL) and AcONa (0.26 g, 3.2 mmol) at reflux for 8 h. The solvent was eliminated and the crude 2-[4-(acetoxymethylthio)phenyl]-3-(4-fluorophenyl)-5,7-dimethylpyrazolo[1,5-*a*]pyrimidine (**24a**) thus obtained was used directly in the next step.

A solution of the previous compound in CH<sub>2</sub>Cl<sub>2</sub> (2.6 mL) and MeOH (1.7 mL) at 0 °C was treated with hexahydrated magnesium monoperoxophthalate (0.58 g, 0.97 mmol) and the mixture stirred at room temperature overnight. Saturated NaHCO<sub>3</sub> solution was added and the mixture extracted with CH<sub>2</sub>Cl<sub>2</sub>. The solvent was eliminated and the residue dissolved in THF (2.6 mL) and MeOH (1.3 mL) and cooled to 0 °C. 1 N NaOH (0.84 mL) was added and the mixture stirred at room temperature for 1 h. The solvent was eliminated and the residue vacuum-dried to afford crude sodium 4-[3-(4-fluorophenyl)-5,7-dimethylpyrazolo[1,5-*a*]pyrimidin-2-yl]benzenesulfonate (**25a**; 0.28 g).

A mixture of the previous crude, H<sub>2</sub>O (3.6 mL), AcONa (0.063 g, 0.76 mmol) and hydroxylamine *O*-sulfonic acid (0.086 g, 0.76 mmol) was stirred at room temperature overnight. The suspension thus obtained was filtered and washed with AcOEt

and H<sub>2</sub>O. The phases were separated and the aqueous phase was extracted with AcOEt. The combined organic phases were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **26a** as a white solid (0.083 g, 30%): mp 228–229 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> + CD<sub>3</sub>OD δ TMS) 2.39 (s, 3 H), 2.81 (s, 3 H), 4.09 (s, 2 H + H<sub>2</sub>O), 7.07 (m, 2 H), 7.40 (m, 2 H), 7.74 (d, *J* = 7 Hz, 2 H), 7.85 (d, *J* = 7 Hz, 2 H), 8.30 (s, 1 H). Anal. (C<sub>20</sub>H<sub>17</sub>FN<sub>4</sub>O<sub>2</sub>S·0.75H<sub>2</sub>O) C, H, N, S.

The same procedure was used to obtain **26b**: mp 281 °C; Anal. (C<sub>21</sub>H<sub>20</sub>N<sub>4</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S. **26c**: mp 250–252 °C; Anal. (C<sub>20</sub>H<sub>16</sub>F<sub>2</sub>N<sub>4</sub>O<sub>2</sub>S·0.5H<sub>2</sub>O) C, H, N, S.

**8-(4-Fluorophenyl)-3,4-dimethyl-7-(4-methylsulfonylphenyl)pyrazolo[5,1-*c*][1,2,4]triazine (29a).** A mixture of ethyl 2-methylacetoacetate (0.21 mL, 1.5 mmol) and KOH (0.13 g, 1.9 mmol) in H<sub>2</sub>O (1 mL) was stirred at 0 °C for 24 h. After adjusting the pH to 6 with concentrated HCl, a suspension of **27** [prepared from **9a** (0.5 g, 1.5 mmol) by reaction at 0 °C with NaNO<sub>2</sub> (0.104 g, 1.5 mmol) in H<sub>2</sub>O (0.27 mL)] in HCl (0.37 mL) and H<sub>2</sub>O (0.27 mL) was added under the surface followed by AcONa (0.38 g, 4.6 mmol). The mixture was stirred at 0 °C for 1 h and allowed to reach room temperature. CHCl<sub>3</sub> was added and the phases were separated. The aqueous phase was extracted with CHCl<sub>3</sub> and the combined organic phases were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **29a** as a white solid (0.100 g, 17%): mp 219 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 2.91 (s, 3 H), 2.93 (s, 3 H), 3.11 (s, 3 H), 7.17 (m, 2 H), 7.63 (m, 2 H), 7.94 (d, *J* = 7 Hz, 2 H), 7.99 (d, *J* = 7 Hz, 2 H). Anal. (C<sub>20</sub>H<sub>17</sub>FN<sub>4</sub>O<sub>2</sub>S·2H<sub>2</sub>O) C, H, N, S.

The same procedure was used to obtain **29b**: mp 164 °C; Anal. (C<sub>21</sub>H<sub>19</sub>FN<sub>4</sub>O<sub>2</sub>S) C, H, N, S. **29c**: mp 213–214 °C; Anal. (C<sub>21</sub>H<sub>19</sub>FN<sub>4</sub>O<sub>2</sub>S) C, H, N, S. **29d**: mp 221–224 °C; Anal. (C<sub>22</sub>H<sub>21</sub>FN<sub>4</sub>O<sub>2</sub>S) C, H, N, S. **29e**: mp 170–176 °C; Anal. (C<sub>22</sub>H<sub>21</sub>FN<sub>4</sub>O<sub>2</sub>S·0.5H<sub>2</sub>O) C, H, N, S. **29f**: mp 186–190 °C; Anal. (C<sub>22</sub>H<sub>21</sub>FN<sub>4</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S.

**8-(4-Fluorophenyl)-2,4-dimethyl-7-(4-methylsulfonylphenyl)pyrazolo[1,5-*a*][1,3,5]triazine (33a).** A mixture of ethyl acetimidate hydrochloride (**30**, R<sub>1</sub> = Me; 0.25 g, 2 mmol), K<sub>2</sub>CO<sub>3</sub> (0.28 g, 2 mmol) and CH<sub>3</sub>CN (5 mL) was stirred for 5 min and the suspension thus obtained was filtered. To the resulting solution a mixture of **9a** (0.3 g, 0.9 mmol) in AcOH (0.05 mL) was added and stirring continued overnight. The suspension thus obtained was filtered and the solvent was concentrated to give **31a** (R<sub>1</sub> = Me; 0.3 g). This compound was dissolved in ethyl orthoacetate (**32**, R<sub>2</sub> = Me; 1 mL) and heated at reflux overnight. The solvent was concentrated to a crude product, which was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **33a** as a white solid (0.075 g, 21%): mp 190 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 2.71 (s, 3 H), 3.02 (s, 3 H), 3.11 (s, 3 H), 7.13 (m, 2 H), 7.45 (m, 2 H), 7.86 (d, *J* = 7 Hz, 2 H), 7.99 (d, *J* = 7 Hz, 2 H). Anal. (C<sub>20</sub>H<sub>17</sub>FN<sub>4</sub>O<sub>2</sub>S) C, H, N, S.

The same procedure was used to obtain **33b**: mp 177–183 °C; Anal. (C<sub>23</sub>H<sub>23</sub>FN<sub>4</sub>O<sub>2</sub>S·1.25H<sub>2</sub>O) C, H, N, S. **33c**: mp 170–173 °C; Anal. (C<sub>22</sub>H<sub>21</sub>FN<sub>4</sub>O<sub>2</sub>S·0.25H<sub>2</sub>O) C, H, N, S.

**4-Chloro-2,6-dimethyl-3-nitropyridine (35b).** A mixture of 2,6-dimethyl-4-hydroxy-3-nitropyridine (**34b**; 4.6 g, 27.3 mmol), POCl<sub>3</sub> (2.5 mL), DMF (2 mL) and toluene (20 mL) was heated at 110 °C for 7 h. After cooling to 0 °C, the mixture was basified with Na<sub>2</sub>CO<sub>3</sub> and the solid thus obtained filtered and dried to afford **35b** as an oil (3.54 g, 69%): <sup>1</sup>H NMR (CDCl<sub>3</sub> δ TMS) 2.56 (s, 3 H), 2.57 (s, 3 H), 7.18 (s, 1 H).

**2,6-Dimethyl-4-(4-fluorophenylamino)-3-nitropyridine (36b).** To a solution of **35b** (3.54 g, 19 mmol) in CHCl<sub>3</sub> (3 mL) was added, at 0 °C under argon, a solution of 4-fluoroaniline (2.2 mL, 23 mmol) and NEt<sub>3</sub> (3.7 mL) in CHCl<sub>3</sub> (6.5 mL) and the mixture was refluxed for 48 h. The solvent was eliminated and the residue was dissolved with AcOEt and H<sub>2</sub>O. The phases were separated and the aqueous phase was extracted with AcOEt. The combined organic phases were dried and concentrated to a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **36b** as a yellow solid (4.0 g, 81%): mp 128 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>



$\delta$  TMS) 2.35 (s, 3 H), 2.75 (s, 3 H), 6.52 (s, 1 H), 7.20 (m, 4 H), 8.60 (s, 1 H).

**3-Amino-2,6-dimethyl-4-(4-fluorophenylamino)pyridine (37b).** A mixture of **36b** (2.0 g, 7.6 mmol),  $\text{SnCl}_2$  (7.2 g, 38 mmol) and EtOH (54 mL) was refluxed for 45 min. The solvent was eliminated and the residue was dissolved with  $\text{H}_2\text{O}$  and basified with 25% NaOH. The aqueous phase was extracted with  $\text{CHCl}_3$  and the organic phase was dried and concentrated to afford **37b** as an oil (1.0 g, 56%):  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.34 (s, 3 H), 2.44 (s, 3 H), 3.19 (s, 2 H), 5.91 (s, 1 H), 6.63 (s, 1 H), 7.04 (m, 4 H).

**4,6-Dimethyl-1-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)imidazo[4,5-c]pyridine (38b).** A mixture of **37b** (1.0 g, 4.0 mmol), 4-methylsulfonylbenzoyl chloride (0.98 g, 4.5 mmol) and pyridine (16 mL) was refluxed overnight. The solvent was eliminated and the residue was dissolved with  $\text{H}_2\text{O}$  and extracted with  $\text{CHCl}_3$ . The organic phase was washed with 0.1 N NaOH, dried and concentrated to afford a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **38b** as a white solid (0.46 g, 29%): mp 259 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.62 (s, 3 H), 2.94 (s, 3 H), 3.06 (s, 3 H), 6.85 (s, 1 H), 7.28 (m, 4 H), 8.76 (d,  $J = 7$  Hz, 2 H), 7.91 (d,  $J = 7$  Hz, 2 H). Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S.

The same procedure was used to obtain **38a**: mp 218–221 °C; Anal. ( $\text{C}_{19}\text{H}_{14}\text{FN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S.

**2-Bromo-2-(4-fluorophenyl)-1-(4-methylsulfonylphenyl)ethanone (39).** To a solution of **5a** (5 g, 17.1 mmol) in  $\text{CH}_2\text{Cl}_2$  (44 mL) was added a solution of  $\text{Br}_2$  (0.98 mL, 19.2 mmol) in AcOH (107 mL) and the mixture was stirred at room temperature overnight. The solvent was eliminated and the residue was dried to afford **39** as a white solid (6.35 g, 100%): mp 113–116 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 3.13 (s, 3 H), 6.34 (s, 1 H), 7.22 (m, 2 H), 7.58 (m, 2 H), 8.09 (d,  $J = 7$  Hz, 2 H), 8.21 (d,  $J = 7$  Hz, 2 H).

**3-Amino-6-chloro-4,5-dimethylpyridazine (40b).** A mixture of 3,6-dichloro-4,5-dimethylpyridazine<sup>33</sup> (0.5 g, 2.8 mmol), 30% aqueous  $\text{NH}_3$  (20 mL) and THF (5 mL) was stirred in a pressure vessel at 150 °C for 36 h. The solvent was eliminated and the residue was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **40b** as an oil (0.22 g, 50%):  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.14 (s, 3 H), 2.33 (s, 3 H), 4.81 (s, 2 H).

**6-Chloro-7,8-dimethyl-3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)imidazo[1,2-b]pyridazine (41b).** A mixture of **40b** (0.22 g, 1.4 mmol), **39** (0.52 g, 1.4 mmol) and DMF (3 mL) was kept at 60 °C overnight. The solvent was eliminated and the residue was dissolved with  $\text{H}_2\text{O}$  and extracted with  $\text{CHCl}_3$ . The organic phase was washed with saturated  $\text{NaHCO}_3$  solution, dried and concentrated to afford a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **41b** as a white solid (0.46 g, 29%): mp 192–194 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.44 (s, 3 H), 2.74 (s, 3 H), 3.05 (s, 3 H), 7.22 (m, 2 H), 7.54 (m, 2 H), 7.94 (s, 4 H). Anal. ( $\text{C}_{21}\text{H}_{17}\text{ClFN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S.

The same procedure was used to obtain **41a**: mp 216 °C; Anal. ( $\text{C}_{19}\text{H}_{14}\text{FN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S.

**7,8-Dimethyl-3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)imidazo[1,2-b]pyridazine (41c).** A mixture of **41b** (0.15 g, 0.3 mmol), 10% Pd/C (0.03 g),  $\text{NET}_3$  (0.14 mL, 1 mmol) and THF (12 mL) was hydrogenated at 200 psi overnight. The mixture was filtered over Celite and the solvent was eliminated to afford a crude product. This was chromatographed on silica gel (hexane–AcOEt mixtures) to afford **41c** as a white solid (0.095 g, 69%): mp 216–217 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 2.38 (s, 3 H), 2.68 (s, 3 H), 3.05 (s, 3 H), 7.22 (m, 2 H), 7.54 (m, 2 H), 7.90 (m, 4 H), 8.10 (s, 1 H). Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_2\text{S} \cdot \text{H}_2\text{O}$ ) C, H, N, S.

**3-(4-Fluorophenyl)-2-(4-methylsulfonylphenyl)imidazo[1,2-a]pyrazine (43a).** The same procedure described for **41b** starting from 2-aminopyrazine (**42a**) was used to obtain **43a**: mp 229–231 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$   $\delta$  TMS) 3.04 (s, 3 H), 7.33 (m, 2 H), 7.47 (m, 2 H), 8.87 (m, 6 H), 9.19 (s, 1 H). Anal. ( $\text{C}_{19}\text{H}_{14}\text{FN}_3\text{O}_2\text{S} \cdot 0.5\text{H}_2\text{O}$ ) C, H, N, S.

Analogously, **43b,c** and **45** were obtained using, respectively, 2-amino-3,5-dimethylpyrazine (**42b**, obtained from 2-chlo-

ro-3,5-dimethylpyrazine in a manner similar to that described in ref 34), 2-amino-3-butylpyrazine (**42c**)<sup>35</sup> and 2-amino-4-methylpyrimidine (**44**). **43b**: mp 239–242 °C. Anal. ( $\text{C}_{21}\text{H}_{18}\text{FN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S. **43c**: Anal. ( $\text{C}_{23}\text{H}_{22}\text{FN}_3\text{O}_2\text{S} \cdot 0.25\text{H}_2\text{O}$ ) C, H, N, S. **45**: mp 272–273 °C; Anal. ( $\text{C}_{20}\text{H}_{16}\text{FN}_3\text{O}_2\text{S}$ ) C, H, N, S.

#### Inhibition of COX-1 and COX-2 Activities in HWB.<sup>27</sup>

The tests were performed with heparinized human whole blood from healthy donors who had not taken NSAIDs for 1 week nor alcohol or xanthines within 24 h of blood collection. COX-1 activity was measured in spontaneously clotting human blood for 5 h at 37 °C and stimulated with A23187 (5  $\mu\text{M}$ ) for 30 min longer. The reaction was stopped at 0 °C with EGTA and MeOH. After centrifugation the supernatant was removed and stored at –70 °C. TXB<sub>2</sub> levels were determined by enzyme immunoassay kit (RPN 220, Amersham). COX-2 activity was stimulated by adding lipopolysaccharide 026:B6 (25  $\mu\text{L}/\text{mL}$ ) at 37 °C for 24 h. Aspirin (20  $\mu\text{g}/\text{mL}$ ) was used to inhibit COX-1 activity. After centrifugation, the samples were stored at –70 °C. PGE<sub>2</sub> levels were determined by enzyme immunoassay kit (RPN 222, Amersham).

**Inhibition of COX-1 and COX-2 Activities in Human Cell Lines.** The production of PGE<sub>2</sub> after AA stimulation by lines expressing h-COX-1 (U-937 from human histiocytic lymphoma) and h-COX-2 (143.98.2 from human osteosarcoma) was used as a cell-based assay to evaluate COX-1 and COX-2 inhibition, respectively. Briefly, osteosarcoma cells were cultured in 1 mL of media in 24-well multidishes until confluent ( $2 \times 10^5$  cells/well during 24 h). U-937 cells were grown in flasks and resuspended to a final density of  $3 \times 10^6$  cells/mL/well. The cells were resuspended in HBSS without  $\text{Ca}^{2+}/\text{Mg}^{2+}$ . 10  $\mu\text{L}$  of a DMSO solution of test compound or DMSO vehicle was added. The samples were then incubated for 15 min at 37 °C (5%  $\text{CO}_2$  and 95% humidity) before the addition of AA (final concentration: 10  $\mu\text{M}$ ) for 10 min. The reactions were stopped by the addition of indomethacin (8 mM, 30  $\mu\text{L}$ ). PGE<sub>2</sub> levels in supernatant were determined by specific EIA (Amersham). All assays were performed in triplicate.

#### Inhibition of Carrageenan-Induced Rat Paw Edema.

Male Sprague–Dawley rats (175–200 g) were used. Edemas were produced by injecting 0.1 mL of a solution of 1%  $\lambda$ -carrageenan in the hindpaw. Paw volumes were measured by water displacement with a plethysmometer (UGO BASILE) before and 1, 2, 3, 4 and 5 h after treatment. The compounds were administered orally as a suspension in carboxymethyl cellulose and Tween-80 at 1% (10 mL/kg), 0.5 h before carrageenan injection and after being hydrated with  $\text{H}_2\text{O}$  (5 mL). The percentages of inhibition were calculated by comparing the areas under the curve of treated and control animals.

**Air-Pouch Model of Inflammation: Determination of PGE<sub>2</sub> in Exudate and Stomach.** Male Lewis rats (175–200 g) were used. Air cavities were produced by a subcutaneous injection of sterile air (20 mL) into the intracapsular area. Every 2 days air (10 mL) was injected into the cavity to keep the space open. 7 days after the first injection,  $\lambda$ -carrageenan (Sigma) in saline (2 mL of a 1% solution) was injected into the air pouch to produce an inflammatory reaction. The compounds were administered by oral route as a suspension in carboxymethyl cellulose and Tween-80 at 1% (10 mL/kg), 0.5 h before carrageenan injection. The animals were killed 6 h later and the exudate volume was measured. Cells were pelleted by centrifugation at 1200g for 5 min at 4 °C, and PGE<sub>2</sub> was determined in the supernatant by specific ELISA. Immediately after the exudates were collected, the stomachs were excised and frozen at –70 °C. On the day of analysis, stomachs were homogenized in 70% ethanol. The homogenates were centrifuged (1000g, 10 min, 4 °C) and the supernatants were dried under  $\text{N}_2$  stream and resuspended in ELISA buffer for PGE<sub>2</sub> determination.

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**Supporting Information Available:** Combustion analyses and  $^1\text{H}$  NMR spectra for all compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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