

Design and Synthesis of Naphthalenic Dimers as Selective MT₁ Melatonergic Ligands

Carole Descamps-François,[§] Saïd Yous,^{*,§}
Philippe Chavatte,[§] Valérie Audinot,[†]
Anne Bonnaud,[†] Jean A. Boutin,[†]
Philippe Delagrangé,[†] Caroline Bennejean,[†]
Pierre Renard,[†] and Daniel Lesieur[§]

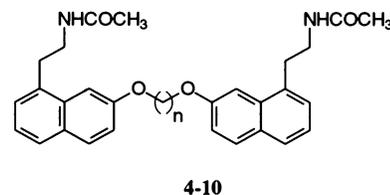
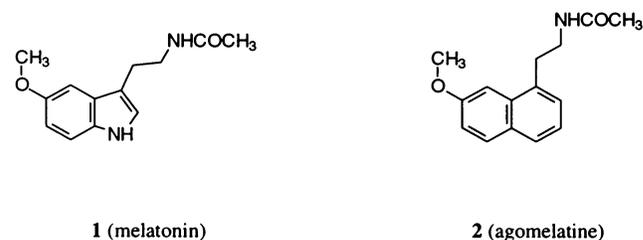
Faculté de Pharmacie, Laboratoire de Chimie
Thérapeutique, Université de Lille 2, 3 Rue du Professeur
Laguesse, BP 83, 59006 Lille Cedex, France, and Institut de
Recherches Internationales Servier,
Courbevoie Cedex, France

Received September 20, 2002

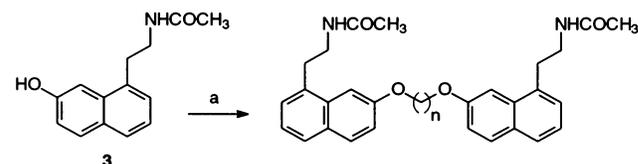
Abstract: We report the synthesis and binding properties at MT₁ and MT₂ receptors of the first example of agomelatine (*N*-[2-(7-methoxynaphth-1-yl)ethyl]acetamide) dimers in which two agomelatine moieties are linked together through their methoxy substituent by a polymethylene side chain according to the “bivalent ligand” approach. Some of these compounds behave as MT₁-selective ligands. The most selective one (**5**) behaves as an antagonist.

Introduction. Melatonin (*N*-acetyl-5-methoxytryptamine, **1**; Chart 1) is synthesized and released by the pineal gland during the dark period. Indeed, its synthesis is regulated by the day–night alternation and by the way melatonin transmits information about the photoperiod to the organism. This neurohormone plays an important role in the regulation of mammalian circadian rhythms and reproductive functions,¹ and it has been implicated in a number of pathological states suggesting its therapeutic application in several disorders.^{2,3} These effects are mediated through high-affinity G-protein-coupled receptors, two of which (MT₁ and MT₂) have been cloned in mammals, including humans.^{4–6} These two receptors are present in humans in different parts of the brain (suprachiasmatic nuclei, cortex, *pars tuberalis*, etc.) and at the periphery (kidney, adipocytes, retina, blood vessels, etc.).^{7–13} The functional physiological roles of these receptors are not well-known, and current research goals include the design of subtype-selective melatonin receptor agonists and antagonists, which will provide pharmacological tools to assess and better characterize the role of each receptor subtype. To date, only a few MT₂-selective ligands (agonists and antagonists) have been described^{14–20} and some of them have allowed the identification of several physiological responses mediated through activation of MT₂ receptors.¹⁹ But there is no selective agonist or antagonist for MT₁ receptors, and this is the reason that the physiological role of this subtype is still unknown. The aim of this study was to obtain such selective MT₁ ligands. Among the different methods currently available for medicinal chemists to design potent and selective receptor subtype ligands, the “bivalent ligands”

Chart 1. Chemical Structures of Melatonin Ligands



Scheme 1. Preparation of Dimeric Compounds **4–10**^a



compd	n	Yield (%)
4	2	62
5	3	56
6	4	74
7	5	55
8	6	60
9	8	74
10	10	76

^a Reagents: (a) Br(CH₂)_nBr, K₂CO₃, acetonitrile.

approach appears very promising. Since its first application by Portoghese^{21,22} in the field of opioid research, this concept has been successfully applied to numerous research areas and particularly to monoamine neurotransmitters such as norepinephrine,²³ dopamine,²⁴ and serotonin.²⁵ During our efforts to discover selective melatonin receptor subtype ligands, we decided to apply this bivalent ligand approach to agomelatine (**2**, Chart 1), the naphthalenic bioisostere of melatonin, which we have previously described²⁶ and which is, at the present time, in phase III clinical trials.

Chemistry. Dimers **4–10** (Chart 1) were obtained in one step by condensation of 2 equiv of *N*-[2-(7-hydroxynaphth-1-yl)ethyl]acetamide (**3**)²⁷ with the appropriate dibromoalkane in the presence of potassium carbonate in acetonitrile (Scheme 1).

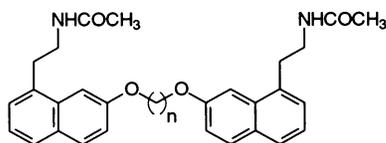
Pharmacology. The compounds were evaluated for their binding affinity for human MT₁ and MT₂ receptors stably transfected in human embryonic kidney (HEK 293) cells, using 2-[¹²⁵I]iodomelatonin as radioligand.

The [³⁵S]GTPγS binding assay used to determine the functional activity of the compounds was difficult to handle using the transfected HEK 293 cell lines, while reliable results were obtained using Chinese hamster

* To whom correspondence should be addressed: Phone and fax: +33(0)320964375. E-mail: syous@phare.univ-lille2.fr.

[§] Université de Lille 2.

[†] Institut de Recherches Internationales Servier.

Table 1. MT₁ and MT₂ Receptor Binding Affinities of Compounds **1**, **2**, and **4–10**^a**4-10**

compd	<i>n</i>	<i>K</i> _i ± SEM (nM) MT ₁	<i>K</i> _i ± SEM (nM) MT ₂	ratio MT ₂ /MT ₁
1		0.14 ± 0.03	0.41 ± 0.04	3
2		0.06 ± 0.01	0.27 ± 0.04	4
4	2	2.05 ± 0.76	182 ± 48.20	88
5	3	0.50 ± 0.03	112 ± 16.70	224
6	4	0.60 ± 0.12	72.70 ± 22.20	120
7	5	0.05 ± 0.01	1.74 ± 0.26	35
8	6	0.08 ± 0.01	2.33 ± 0.02	30
9	8	0.05 ± 0.01	0.18 ± 0.01	36
10	10	0.03 ± 0.00	0.04 ± 0.01	1.3

^a Concentration–response curves were analyzed by nonlinear regression comparing a one-site and a two-site analysis. All the curves were found to be monophasic with a Hill number close to unity (not shown). Binding affinities (nM) are expressed as the mean *K*_i ± SEM of at least three independent experiments. The selectivity ratio of MT₁ to MT₂ receptors is calculated for the compounds.

ovarian (CHO) cell lines stably expressing the human MT₁ or MT₂ receptors. At each receptor, binding affinities were verified for more than 50 selective and nonselective molecules, using either the transfected HEK 293 or the CHO cell lines. Indeed, the correlations between affinities in HEK 293 and CHO cells are highly significant (*r* = 0.98) (unpublished data) for both MT₁ and MT₂ receptors.

Results and Discussion. The chemical structures, binding affinities, and MT₁/MT₂ selectivity ratios of the new compounds **4–10** are reported in Table 1. The agonist (EC₅₀) or antagonist (*K*_B) potencies and efficacies (expressed relative to that of melatonin taken at 100%) in the [³⁵S]GTPγS binding assay are shown in Table 2.

Comparison of the relative MT₁ and MT₂ binding affinities of both melatonin (**1**) and agomelatine (**2**) shows that the MT₁ affinity of **2** is about 2-fold lower than that of melatonin, leading to respective MT₂/MT₁ selectivity ratios of 4 versus 3. These results justify our choice of agomelatine as the starting monomer for the assembly of bivalent ligands. On the other hand, we decided to link the two agomelatine moieties through their 7-methoxy rather than their 3-acetamido substituent according to previously described²⁸ structure–affinity relationships, which showed that homologation of the former was more favorable than that of the latter.

Comparison of the relative MT₁ and MT₂ binding affinities of the bivalent ligands (**4–10**) shows that as

the number of methylene groups in the linking chain increases from 2 to 10, the MT₁ binding affinities increase from 2.05 to 0.03 nM whereas the MT₂ affinities increase from 182 to 0.04 nM. Compound **10**, with 10 methylene groups, is the most potent with MT₁ and MT₂ affinities respectively 2-fold and 15-fold better than agomelatine, but this compound is not selective (MT₂/MT₁ = 1.3). These results suggest that for both MT₁ and MT₂ receptors compound **10** binds at two binding sites located on two neighboring receptors and that the distance between these sites should coincide with the optimal distance (10 methylene units) between the dimer headgroups. Recently homodimerization of melatonin receptors has been reported.²⁹ Consequently, another hypothesis could be that this molecule binds at each binding site of the homodimerized receptors.

Compounds **7–9**, which respectively possess five, six, and eight methylene groups, have the same MT₁ affinity and the same (**9**) or a slightly higher MT₂ affinity as agomelatine. We can therefore assume that in these cases, the headgroups can still bind to both sites, but the fit is reduced as a result of a less favorable connecting chain conformation that is more sensitive in the case of the MT₂ than the MT₁ subtype. This difference in sensitivity could explain the appearance of weak MT₁ selectivity of these compounds (MT₂/MT₁ = 30).

Compounds **4–6** with less than five methylene groups in the linker show a slightly lower (10- to 30-fold) MT₁ affinity but a strongly decreased (300- to 500-fold) MT₂ affinity compared to agomelatine. In these cases, the dimer is too short to bridge both MT₂ sites located on two separate or homodimerized receptors. This hypothesis is probably the same for MT₁ binding sites. Nevertheless, these compounds retain good (nM) MT₁ affinity and behave as potent MT₁-selective ligands. The optimal distance for MT₁ selectivity between the dimer headgroups is obtained with three methylene units (MT₂/MT₁ = 224), but the two homologous derivatives with two and four methylene units also show good selectivity ratios (88 and 120, respectively). These results suggest that in these cases steric factors could be predominant: the voluminous seven-substituent of the monomer is able to take place in or near the MT₁ but not in or near the MT₂ receptor pocket. The MT₁ selectivity could be due to the choice of the position of the spacer attachment to agomelatine and to the bulkiness of the substituent.

The functional activity of the most selective MT₁ ligand (**5**) has been evaluated on both receptors in comparison with agomelatine. The full agonist activity is confirmed for agomelatine, whereas compound **5** behaves as an antagonist on both receptors.

Table 2. Activity Values of Compounds **1**, **2**, and **5**^a

compd	MT ₁			MT ₂		
	EC ₅₀ ± SEM (nM)	<i>E</i> _{max} ± SEM (%)	<i>K</i> _B ± SEM (nM)	EC ₅₀ ± SEM (nM)	<i>E</i> _{max} ± SEM (%)	<i>K</i> _B ± SEM (nM)
1 (melatonin)	2.24 ± 0.35	110 ± 2	nd	0.49 ± 0.04	104 ± 6	nd
2	1.56 ± 0.44	101.3 ± 5.7	nd	0.1 ± 0.04	91 ± 7	nd
5	inactive	<10	5.32 ± 0.95	inactive	<10	143 ± 25

^a Concentration–response curves were analyzed by nonlinear regression. Agonist potency was expressed as EC₅₀ ± SEM (nM), while the maximal efficacy *E*_{max} ± SEM was expressed as a percentage of that observed with 1 μM (=100%) melatonin. Antagonist potency to inhibit the effect of melatonin (30 and 3 nM, respectively, for MT₁ and MT₂ receptors) was expressed as *K*_B ± SEM. Data are the mean of at least three independent experiments. Inactive means no dose–response effect, and nd means not determined.

The respective binding (affinities) and functional (potencies) profiles of agonists are conserved for each receptor. However, for hMT₁, a shift in the order of potency (5- to 10-fold) is seen for reference agonists including melatonin (**1**) and agomelatine (**2**), probably due to less efficient coupling at this receptor. This shift is also observed for the antagonist, compound **5**. This explains why the ratio of selectivity is lower in the [³⁵S]-GTPγS assay compared to the binding assay.

Acknowledgment. The authors thank Dr. C. Bochu (Laboratoire d'Application de Résonance Magnétique Nucléaire de Lille 2) for his aid with the interpretation of the ¹H NMR spectra.

Supporting Information Available: Experimental section, including chemistry and pharmacology information. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Reiter, R. J. Pineal melatonin: cell biology of its synthesis and of its physiological interactions. *Endocr. Rev.* **1991**, *12*, 151–180.
- Mahle, C. D.; Watson, A. J. Melatonin receptors: potential targets for central nervous system disorders. *Expert Opin. Invest. Drugs* **1997**, *6*, 399–406.
- Li, P. K.; Witt-Enderby, P. A. Melatonin receptors as potential targets for drug discovery. *Drugs Future* **2000**, *25*, 945–957.
- Reppert, S. M.; Weaver, D. R.; Ebisawa, T. Cloning and characterisation of a mammalian melatonin receptor that mediates reproductive and circadian responses. *Neuron* **1994**, *13*, 1177–1185.
- Reppert, S. M.; Godson, C.; Mahle, C. D.; Weaver, D. R.; Slaugenhaupt, S. A.; Gusella, J. F. Molecular characterisation of a second melatonin receptor expressed in human retina and brain: The Mel_{1b} melatonin receptor. *Proc. Natl. Acad. Sci. U.S.A.* **1995**, *92*, 8734–8738.
- Dubocovich, M. L.; Cardinali, D. P.; Delagrangé, P.; Krause, D. N.; Strosberg, A. D.; Sugden, D.; Yocca, F. D. *The IUPHAR Compendium of Receptor Characterisation and Classification*; IUPHAR Media: London, 2000; pp 271–277.
- Mazzucchelli, C.; Pannacci, M.; Nonno, R.; Lucini, V.; Fraschini, F.; Stankov, B. M. The melatonin receptor in the human brain: cloning experiments and distribution studies. *Brain Res. Mol. Brain Res.* **1996**, *39*, 117–126.
- Weaver, D. R.; Reppert, S. M. The Mel_{1a} melatonin receptor gene is expressed in human suprachiasmatic nuclei. *NeuroReport* **1996**, *8*, 109–112.
- Al-Ghoul, W. M.; Herman, M. D.; Dubocovich, M. L. Melatonin receptor subtype expression in human cerebellum. *NeuroReport* **1998**, *9*, 4063–4068.
- Song, Y.; Chan, C. W.; Brown, G. M.; Pang, S. F.; Silverman, M. Studies of the renal action of melatonin: evidence that the effects are mediated by 37 kDa receptors of the Mel_{1a} subtype localised primarily to the basolateral membrane of the proximal tubule. *FASEB J.* **1997**, *11*, 93–100.
- Brydon, L.; Petit, L.; Delagrangé, P.; Strosberg, A. D.; Jockers, R. Functional expression of mt2 (mel_{1b}) melatonin receptors in human p3z6 adipocytes. *Endocrinology* **2001**, *142*, 4264–4271.
- Morgan, P. J.; Barrett, P.; Howell, H. E.; Helliwell, R. Melatonin receptors: localization, molecular pharmacology and physiological significance. *Neurochem. Int.* **1994**, *24*, 101–146.
- Doolen, S.; Krause, D. N.; Dubocovich, M. L.; Duckles, S. P. Melatonin mediates two distinct responses in vascular smooth muscle. *Eur. J. Pharmacol.* **1998**, *345*, 67–69.
- Dubocovich, M. L.; Masana, M. I.; Jacob, S.; Sauri, D. M. Melatonin receptor antagonists that differentiate between the human Mel_{1a} and Mel_{1b} recombinant subtypes are used to assess the pharmacological profile of the rabbit retina ML1 presynaptic heteroreceptor. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1997**, *355*, 365–375.
- Teh, M.; Sugden, D. Comparison of the structure–activity relationships of melatonin receptor agonists and antagonists: lengthening the N-acyl side-chain has differing effects on potency on *Xenopus* melanophores. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1998**, *358*, 522–528.
- Sugden, D.; Yeh, L. K.; Teh, M. T. Design of subtype selective melatonin receptor agonists and antagonists. *Reprod., Nutr., Dev.* **1999**, *39*, 335–44.
- Faust, R.; Garratt, P. J.; Jones, R.; Yeh, L. K.; Tsotinis, A.; Panoussoupoulou, M.; Calogeropoulou, T.; Teh, M. T.; Sugden, D. Mapping the melatonin receptor. 6. Melatonin agonists and antagonists derived from 6*H*-isoindolo[2,1-*α*]indoles, 5,6-dihydroindolo[2,1-*α*]isoquinolines, and 6,7-dihydro-5*H*-benzo[*c*]azepino[2,1-*α*]indoles. *J. Med. Chem.* **2000**, *43*, 1050–1061.
- Spadoni, G.; Balsamini, C.; Diamantini, G.; Tontini, A.; Tarzia, G.; Mor, M.; Rivara, S.; Plazzi, P. V.; Nonno, R.; Lucini, V.; Pannacci, M.; Rfaschini, F.; Stankov, B. M. 2-*N*-Acylaminoalkylindoles: design and quantitative structure–activity relationship studies leading to MT₂-selective melatonin antagonists. *J. Med. Chem.* **2001**, *44*, 2900–2912.
- Masana, M. I.; Dubocovich, M. L. Melatonin receptor signaling: finding the path through the dark. *Sci. STKE* **2001**, *6*, 107.
- Wallez, V.; Durieux-Poissonnier, S.; Chavatte, P.; Boutin, J. A.; Audinot, V.; Nicolas, J. P.; Bennejean, C.; Delagrangé, P.; Renard, P.; Lesieur, D. Synthesis and structure–activity relationships of novel benzofuran derivatives as MT₂ melatonin receptor selective ligands. *J. Med. Chem.* **2002**, *45*, 2788–2800.
- Portoghese, P. S.; Ronsisvalle, G.; Larson, D. L.; Yim, C. B.; Sayre, L. M.; Takemori, A. E. Opioid agonist and antagonist bivalent ligands as receptor probes. *Life Sci.* **1982**, *31*, 1283–1286.
- Erez, M.; Takemori, A. E.; Porthoghese, P. S. Narcotic antagonist potency of bivalent ligands which contain β-naltrexamine. Evidence for bridging between proximal recognition sites. *J. Med. Chem.* **1982**, *25*, 847–849.
- Turnheim, K.; Kraupp, O. Pulmonary and systemic circulatory effect and β-adrenergic selectivity of hexaprenaline, salbutamol, oxyfedrine and isoproterenol. *Eur. J. Pharmacol.* **1971**, *15*, 231–239.
- Fisher, L. E.; Rosenkranz, R. P.; Clark, R. D.; Muchowski, J. M.; McClelland, D. L.; Michel, A.; Caroon, J. M.; Galeazi, E.; Eyles, R.; Whiting, R. L. *N,N*-6-Bis-[2-(3,4-dihydroxybenzyl)pyrrolidinyl]hexane, a potent, selective, orally active dopamine analog with hypotensive and diuretic activity. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 2371–2376.
- Le Boulluec, K.; Matson, R. J.; Mable, C. D.; McGovern, R. T.; Nowak, H. P.; Gentile, A. J. Bivalent indoles exhibiting serotonergic binding affinity. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 123–126.
- Yous, S.; Andrieux, J.; Howell, H. E.; Morgan, P. J.; Renard, P.; Pfeiffer, B.; Lesieur, D.; Guardiola-Lemaître, B. Novel naphthalenic ligands with high affinity for melatonin receptor. *J. Med. Chem.* **1992**, *35*, 1484–1486.
- Depreux, P.; Lesieur, D.; Mansour, H. A.; Morgan, P.; Howell, H. E.; Renard, P.; Caignard, D.-H.; Pfeiffer, B.; Delagrangé, P.; Guardiola, B.; Yous, S.; Demarque, A.; Adam, G.; Andrieux, J. Synthesis and structure–activity relationships of novel naphthalenic and bioisosteric related amidic derivatives as melatonin receptor ligands. *J. Med. Chem.* **1994**, *37*, 3231–3239.
- Lesieur, D.; Leclerc, V.; Chavatte, P.; Marot, C.; Renard, P.; Guardiola-Lemaître, B. La mélatonine, prototype pertinent pour l'innovation thérapeutique (Melatonin, a pertinent prototype for therapeutic innovation). *Thérapie* **1998**, *53*, 429–437.
- Ayoub, M. A.; Couturier, C.; Lucas-Meunier, E.; Angers, S.; Fossier, P.; Bouvier, M.; Jockers, R. Monitoring of ligand-independent dimerization and ligand-induced conformational changes of melatonin receptors in living cells by bioluminescence resonance energy transfer. *J. Biol. Chem.* **2002**, *277*, 21522–21528.

JM0255872