

## Brief Articles

### A Selective Human H<sub>4</sub>-Receptor Agonist: (–)-2-Cyano-1-methyl-3-{(2*R*,5*R*)-5-[1*H*-imidazol-4(5)-yl]tetrahydrofuran-2-yl}methylguanidine

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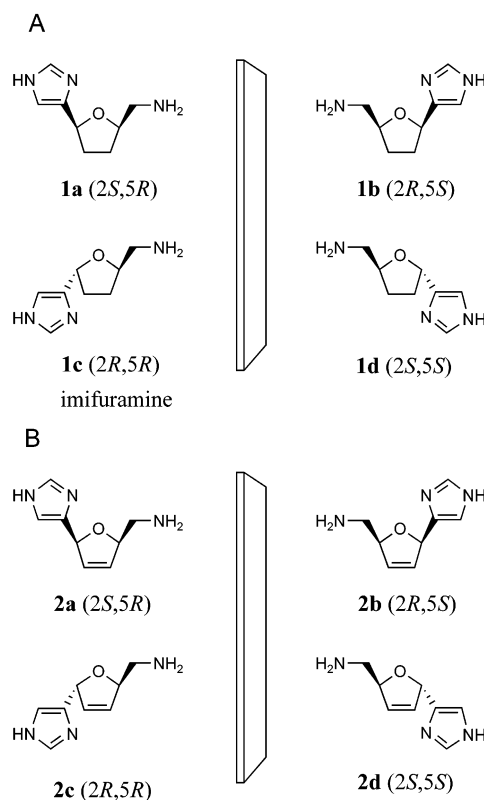
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A series of 16 compounds related to chiral 4(5)-(5-aminomethyltetrahydrofuran-2-yl)imidazoles (**1**) have been designed, synthesized, and examined *in vitro* by radioligand displacement studies and functional assays for both the human H<sub>3</sub>- and H<sub>4</sub>-receptors expressed in SK-N-MC cells. Among them, the (2*S*,5*S*)-isomer **1d** of amino compounds showed approximately 300-fold higher selectivity at the H<sub>3</sub>-receptor than the H<sub>4</sub>-receptor. On the other hand, (2*R*,5*S*)- and (2*R*,5*R*)-cyanoguanidines **3b** and **3c**, in which the amino group of the compounds **1b** and **1c** was substituted by the cyanoguanidino moiety, bound to the H<sub>4</sub>-receptor with a pEC<sub>50</sub> value of 6.65 and 7.11, respectively, and had >40-fold selectivities over the H<sub>3</sub>-receptor. As such, **3b** and **3c** are the first selective H<sub>4</sub> receptor agonists.

#### Introduction

A new histamine receptor, H<sub>4</sub>-receptor, was discovered by several groups in the quest for new G-protein-coupled receptors (GPCR).<sup>1–6</sup> The overall amino acid sequence showed approximately 37% homology between H<sub>3</sub>- and H<sub>4</sub>-receptors. However, the distribution of the human H<sub>4</sub>-receptor mRNA was entirely different from that of the H<sub>3</sub>-receptor.<sup>1,6–8</sup> Although little is known about its function *in vivo*, its expression in the bone marrow and eosinophils may elucidate a potential new role for histamine in the regulation of hemopoietic and immune functions.<sup>1,3</sup> To investigate the possible physiological function of the receptor, a specific ligand is required. Yet, most H<sub>3</sub>-receptor ligands are active at the H<sub>4</sub>-receptor as well. For example, the classical “selective” H<sub>3</sub>-receptor agonist, (*R*)- $\alpha$ -methylhistamine shows H<sub>4</sub>-agonistic activity, and thioperamide, the H<sub>3</sub>-antagonist prototype, has moderate affinities for the H<sub>4</sub>-receptor.<sup>1</sup> No ligands have so far been reported that can selectively target the human H<sub>4</sub>-receptor.

In our previous study, we synthesized the respective four stereoisomers of the tetrahydrofurans (THFs) and dihydrofurans (DHF) containing imidazole and examined their H<sub>3</sub>-receptor pharmacology using the *in vivo* microdialysis method in rats. These compounds spatially



**Figure 1.**

arrange the imidazole and aminomethyl groups across the THF and DHF rings (Figure 1). Among them, only (+)-4(5)-[(2*R*,5*R*)-5-aminomethyltetrahydrofuran-2-yl]-

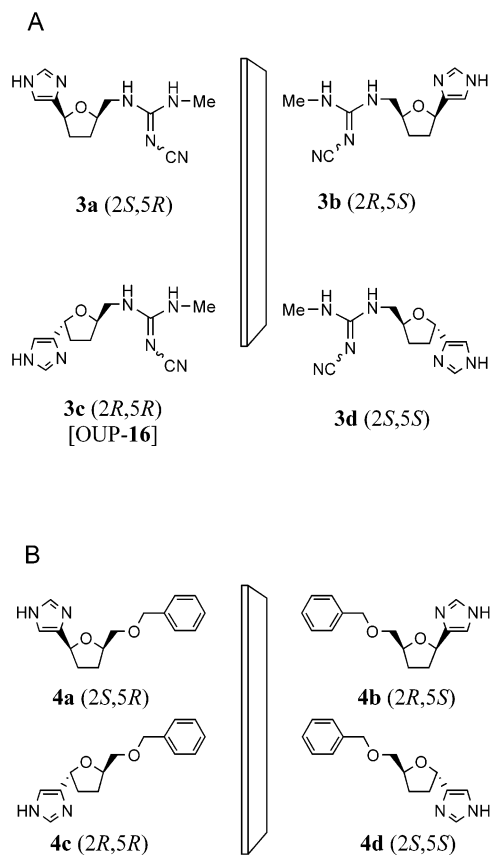
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**Figure 2.**

imidazole (imifuramine, **1c**) was the eutomer exhibiting H<sub>3</sub>-agonistic activity.<sup>9,10</sup>

In this study, we examined the binding affinity and functional activity for the human H<sub>3</sub>- (hH<sub>3</sub>-) and H<sub>4</sub>- (hH<sub>4</sub>-) receptors of a total of 16 tetrahydrofuranylimidazoles: THFs (Figure 1A), DHFs (Figure 1B), and newly synthesized cyanoguanidines (Figure 2A) and benzyl ethers (Figure 2B).

## Results

The competitive binding affinities ( $pK_i$  value) for the hH<sub>3</sub>-receptor of the DHFs (**2a–d**) and cyanoguanidines (**3a–d**) were significantly lower than that of the THFs (**1a–d**) and benzyl ethers (**4a–d**) (Table 1). The  $pK_i$  values of **1c**, **1d**, **4b**, and **4c** for the binding to the hH<sub>3</sub>-receptor were 10-fold higher than that of the other compounds, and were  $6.64 \pm 0.12$ ,  $6.66 \pm 0.14$ ,  $6.48 \pm 0.16$ , and  $6.61 \pm 0.14$ , respectively. The trans-isomers (**1c**, **1d**, **2c**, and **2d**) of amino compounds (THFs and DHFs) exhibited about 10-fold higher affinity than their cis-isomers (**1a**, **1b**, **2a**, and **2b**). In a functional H<sub>3</sub>-receptor assay, the compounds **1a**, **1b**, **1c**, **1d**, **2b**, **2c**, **4b**, and **4c** acted as full agonists ( $0.9 < \alpha < 1.0$ , Table 1). Among them, **1c** had the highest agonistic activity.

As shown in Table 2, **1c**, **3b**, **3c**, and **4b** competed for [<sup>3</sup>H] histamine binding to the hH<sub>4</sub>-receptor with  $pK_i$  values of  $6.05 \pm 0.04$ ,  $6.65 \pm 0.06$ ,  $6.90 \pm 0.17$ , and  $6.36 \pm 0.11$ , respectively. Moreover, the six compounds, **1a**, **3a**, **3b**, **3c**, **3d**, and **4b**, all showed agonist properties with high intrinsic activities ( $0.9 < \alpha < 1.0$ ). Among them, **3c** most potently inhibited the 1  $\mu$ M forskolin-stimulated responses with an apparent  $pEC_{50}$  value of  $7.11 \pm 0.05$  (Table 2).

**Table 1.**  $pEC_{50}$  Values and Affinity Values of Tetrahydrofuranylimidazoles for the Human H<sub>3</sub>-Receptor<sup>a</sup>

compound	config	functional activity		binding $pK_i$
		$pEC_{50}$	efficacy ( $\alpha$ )	
histamine		$8.39 \pm 0.06$	1.00	$7.47 \pm 0.11$
( <i>R</i> )- $\alpha$ -methyl-histamine		$9.91 \pm 0.05$	$0.85 \pm 0.05$	
<b>1a</b>	2 <i>S</i> ,5 <i>R</i>	$6.09 \pm 0.05$	$0.95 \pm 0.04$	$5.77 \pm 0.03$
<b>1b</b>	2 <i>R</i> ,5 <i>S</i>	$6.11 \pm 0.08$	$1.06 \pm 0.04$	$5.69 \pm 0.05$
<b>1c</b> [imifuramine]	2 <i>R</i> ,5 <i>R</i>	$7.35 \pm 0.07$	$1.04 \pm 0.05$	$6.64 \pm 0.12$
<b>1d</b>	2 <i>S</i> ,5 <i>S</i>	$6.98 \pm 0.05$	$0.91 \pm 0.04$	$6.66 \pm 0.14$
<b>2a</b>	2 <i>S</i> ,5 <i>R</i>	<4		$4.70 \pm 0.20$
<b>2b</b>	2 <i>R</i> ,5 <i>S</i>	$4.57 \pm 0.15$	$0.93 \pm 0.06$	$4.94 \pm 0.08$
<b>2c</b>	2 <i>R</i> ,5 <i>R</i>	$6.57 \pm 0.10$	$1.00 \pm 0.05$	$5.99 \pm 0.10$
<b>2d</b>	2 <i>S</i> ,5 <i>S</i>	$5.55 \pm 0.13$	$0.75 \pm 0.06$	$5.60 \pm 0.09$
<b>3a</b>	2 <i>S</i> ,5 <i>R</i>	NE	<0.1	$5.09 \pm 0.08$
<b>3b</b>	2 <i>R</i> ,5 <i>S</i>	$4.99 \pm 0.08$	$0.43 \pm 0.03$	$5.15 \pm 0.17$
<b>3c</b> [OUP-16]	2 <i>R</i> ,5 <i>R</i>	$5.50 \pm 0.08$	$0.79 \pm 0.06$	$5.66 \pm 0.09$
<b>3d</b>	2 <i>S</i> ,5 <i>S</i>	<4		$4.73 \pm 0.13$
<b>4a</b>	2 <i>S</i> ,5 <i>R</i>	$5.01 \pm 0.07$	$0.86 \pm 0.04$	$5.16 \pm 0.19$
<b>4b</b>	2 <i>R</i> ,5 <i>S</i>	$6.72 \pm 0.15$	$1.06 \pm 0.05$	$6.48 \pm 0.16$
<b>4c</b>	2 <i>R</i> ,5 <i>R</i>	$7.04 \pm 0.10$	$0.97 \pm 0.04$	$6.61 \pm 0.14$
<b>4d</b>	2 <i>S</i> ,5 <i>S</i>	$5.02 \pm 0.11$	$0.66 \pm 0.05$	$4.98 \pm 0.12$

<sup>a</sup> The  $pEC_{50}$  values were determined by the inhibition of the forskolin-stimulated (1  $\mu$ M) cAMP production, expressing the human H<sub>3</sub>-receptor. All values shown are means  $\pm$  SEM of at least four experiments. H<sub>3</sub>-receptor competition binding was performed using [<sup>3</sup>H] *N*<sup>2</sup>-methylhistamine (1 nM).

**Table 2.**  $pEC_{50}$  Values and Affinity Values of Tetrahydrofuranylimidazoles for the Human H<sub>4</sub>-Receptor<sup>a</sup>

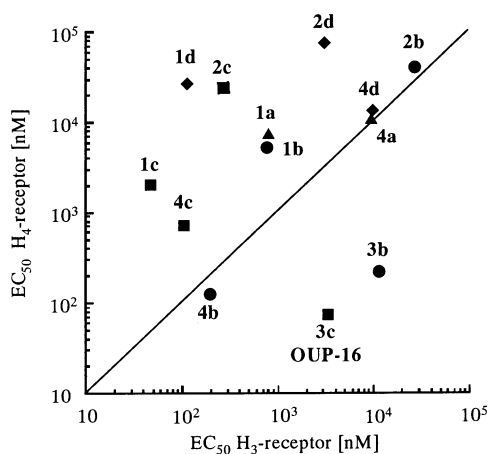
compound	config	functional activity		$pK_i$
		$pEC_{50}$	efficacy ( $\alpha$ )	
histamine		$7.68 \pm 0.05$	1.00	
( <i>R</i> )- $\alpha$ -methylhistamine		$6.26 \pm 0.07$	$1.01 \pm 0.01$	
<b>1a</b>	2 <i>S</i> ,5 <i>R</i>	$5.12 \pm 0.05$	$1.02 \pm 0.07$	$5.19 \pm 0.07$
<b>1b</b>	2 <i>R</i> ,5 <i>S</i>	$5.26 \pm 0.07$	$0.88 \pm 0.06$	$5.60 \pm 0.10$
<b>1c</b> [imifuramine]	2 <i>R</i> ,5 <i>R</i>	$5.70 \pm 0.05$	$0.70 \pm 0.01$	$6.05 \pm 0.04$
<b>1d</b>	2 <i>S</i> ,5 <i>S</i>	$4.51 \pm 0.01$	$0.60 \pm 0.04$	$4.89 \pm 0.06$
<b>2a</b>	2 <i>S</i> ,5 <i>R</i>	$4.12 \pm 0.08$	$0.71 \pm 0.05$	$4.92 \pm 0.16$
<b>2b</b>	2 <i>R</i> ,5 <i>S</i>	$4.39 \pm 0.04$	$0.74 \pm 0.04$	$4.66 \pm 0.12$
<b>2c</b>	2 <i>R</i> ,5 <i>R</i>	$4.62 \pm 0.08$	$0.76 \pm 0.04$	$4.93 \pm 0.03$
<b>2d</b>	2 <i>S</i> ,5 <i>S</i>	$4.07 \pm 0.20$	$0.85 \pm 0.11$	<4
<b>3a</b>	2 <i>S</i> ,5 <i>R</i>	$5.12 \pm 0.06$	$1.07 \pm 0.01$	$5.09 \pm 0.07$
<b>3b</b>	2 <i>R</i> ,5 <i>S</i>	$6.65 \pm 0.03$	$1.01 \pm 0.01$	$6.65 \pm 0.06$
<b>3c</b> [OUP-16]	2 <i>R</i> ,5 <i>R</i>	$7.11 \pm 0.05$	$0.99 \pm 0.01$	$6.90 \pm 0.17$
<b>3d</b>	2 <i>S</i> ,5 <i>S</i>	$4.67 \pm 0.03$	$1.06 \pm 0.02$	$4.69 \pm 0.12$
<b>4a</b>	2 <i>S</i> ,5 <i>R</i>	$4.94 \pm 0.03$	$0.82 \pm 0.02$	$4.87 \pm 0.07$
<b>4b</b>	2 <i>R</i> ,5 <i>S</i>	$6.87 \pm 0.05$	$0.91 \pm 0.02$	$6.36 \pm 0.11$
<b>4c</b>	2 <i>R</i> ,5 <i>R</i>	$6.13 \pm 0.10$	$0.46 \pm 0.02$	$5.98 \pm 0.12$
<b>4d</b>	2 <i>S</i> ,5 <i>S</i>	$4.89 \pm 0.08$	$0.49 \pm 0.05$	$4.67 \pm 0.05$

<sup>a</sup> The  $pEC_{50}$  values were determined by the inhibition of the forskolin-stimulated (1  $\mu$ M) cAMP production, expressing the human H<sub>4</sub>-receptor. All values shown are means  $\pm$  SEM of at least four experiments. H<sub>4</sub>-receptor competition binding was performed using [<sup>3</sup>H] histamine (10 nM).

As shown in Figure 3, **1c**, **1d**, **2c**, **2d**, **3b**, and **3c** exhibited receptor selectivity for either the hH<sub>3</sub>- or hH<sub>4</sub>-receptor. Amino compounds **1c**, **1d**, **2c**, and **2d** showed selective H<sub>3</sub>-agonistic activity, which was approximately 45-, 300-, 89-, and 30-fold higher than for the H<sub>4</sub>-receptor, respectively. In contrast, the cyanoguanidine analogues **3b** and **3c** exhibited full agonistic activities at the H<sub>4</sub>-receptor with 45- and 41-fold higher potency than at the H<sub>3</sub>-receptor, respectively.

## Conclusions

The substitution of an amino group of tetrahydrofuranylimidazoles with a cyanoguanidine moiety led to a decrease in the agonistic activity at the H<sub>3</sub>-receptor and an increase in the H<sub>4</sub>-receptor selectivity. The **3b**



**Figure 3.** Relation of the functional activity between the H<sub>3</sub>- and H<sub>4</sub>-receptors. Data for tetrahydrofuranylimidazoles in Tables 1 and 2 are plotted as H<sub>3</sub> EC<sub>50</sub> values (ordinate, Table 1) versus H<sub>4</sub> EC<sub>50</sub> values (abscissa, Table 2). Compound numbers correspond with those in Tables 1 and 2. ●, (2*R*,5*S*); ▲, (2*S*,5*R*); ■, (2*R*,5*R*); ◆, (2*S*,5*S*).

and **3c** [OUP-16], having the 2*R*-configuration,<sup>11</sup> were highly selective compounds at the H<sub>4</sub>-receptor. On the other hand, **1d**, having the 2*S*,5*S*-configuration, behaved as the most selective H<sub>3</sub>-receptor agonist in our series. The present results suggest that the stereochemistry of the tetrahydrofuranylimidazoles is useful for the investigation of selective ligands for hH<sub>3</sub>- and hH<sub>4</sub>-receptors and that the 2*R*-configured isomers of cyanoguanidines exhibit a high agonistic activity for the H<sub>4</sub>-receptor.

## Experimental Section

SK-N-MC cells expressing the hH<sub>3</sub>-<sup>12</sup> or hH<sub>4</sub>-receptor<sup>1</sup> were maintained in Eagle's minimal essential medium (BioWhittaker, Verviers, Belgium) supplemented with 10% fetal calf serum (Integro, Zaandam, The Netherlands), 50 IU/mL penicillin, nonessential amino acids solution, 2 mM L-glutamine, 50 μg/mL streptomycin, and 50 μg/mL sodium pyruvate (Invitrogen, Breda, The Netherlands) under the selection of 600 μg/mL G<sub>418</sub> disulfate (Calbiochem, Amsterdam, The Netherlands) at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub> in air by the method described by Wieland et al.<sup>13</sup> Cells were detached from the dishes with 0.05% trypsin-EDTA (Invitrogen).

SK-N-MC cells stably expressing the hH<sub>3</sub>-<sup>12</sup> or hH<sub>4</sub>-receptors<sup>1</sup> were grown overnight in 96-well plates before the assay. To start the assay, the cells were incubated for 6 h with 1 μM forskolin (Sigma-Aldrich, St. Louis, MO) and respective tetrahydrofuranylimidazoles at 37 °C. Thereafter, the medium was aspirated, and cells were incubated overnight in a refrigerator with 100 μL of assay buffer (100 mM NaH<sub>2</sub>PO<sub>4</sub>, 100 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 8, 2 mM MgSO<sub>4</sub>, 0.1 mM MnCl<sub>2</sub>, 0.5% Triton, and 40 mM β-mercaptoethanol).<sup>13</sup> Transcription of β-galactosidase was determined by 4 mM *o*-nitrophenyl-β-D-galactopyranoside. Absorbance was quantified on a microplate reader at 420 nm.

The binding affinity of the hH<sub>3</sub>- and hH<sub>4</sub>-receptors was determined with 1.0 nM [<sup>3</sup>H] *N*<sup>ε</sup>-methylhistamine (82 Ci/mmol, PerkinElmer Life Sciences, Zaventem, Belgium) and 10 nM [<sup>3</sup>H] histamine (23.3 Ci/mmol, PerkinElmer Life Sciences), respectively. The cell pellets were harvested, washed, and homogenized in incubation buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 7.4). The cell homogenates (hH<sub>3</sub>-: 131 ± 11 fmol/mg of protein, hH<sub>4</sub>-: 166 ± 26 fmol/mg of protein) were incubated for 1 h at 37 °C with each radioligand in incubation buffer, pH 7.4, with or without competing ligands. Then, the membranes were filtered through the GF/C filters pretreated with 0.3% poly-

ethylenimine. The filter was washed three times with ice-cold washing buffer (hH<sub>3</sub>-: 25 mM Tris HCl, 145 mM NaCl, pH 7.4, hH<sub>4</sub>-: 50 mM Tris HCl, pH 7.4) and the radioactivity was determined by liquid scintillation counting. Nonspecific binding was defined with 1 μM thioperamide as the competing ligand.

Protein concentrations were determined spectrophotometrically by a Packard Argus 400 Microplate Reader using the Bradford reagent,<sup>14</sup> with bovine serum albumin as a standard.

The value of p*K*<sub>i</sub> and pEC<sub>50</sub> was obtained by fitting these data to a sigmoidal relation using GraphPad Prism (GraphPad Software, San Diego, CA). The intrinsic activities were calculated in comparison with the effects of histamine (100 μM).

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**Supporting Information Available:** Synthetic procedures, spectral data, and Scheme 1 for compounds **3a–d** and **4a–d**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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- In case of cyanoguanidines (A) in Figure 2, the carbon bonding to imidazole is numbered as the 2-position of the THF ring for convenience in order to clarify the mutual configurational relationships among the compounds in Figures 1 and 2. The numbering system of THF in the title compounds (OUP-16) is properly employed.

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