Letters

Potent and Orally Bioavailable Non-Peptide Antagonists at the Human Bradykinin B₁ Receptor Based on a 2-Alkylamino-5-sulfamoylbenzamide Core

Timothy J. Ritchie,* Edward K. Dziadulewicz, Andrew J. Culshaw, Werner Müller,[†] Gillian M. Burgess,[‡] Graham C. Bloomfield,[§] Gillian S. Drake, Andrew R. Dunstan,[§] David Beattie,[§] Glyn A. Hughes, Pam Ganju, Peter McIntyre, Stuart J. Bevan, Clare Davis, and Mohammed Yaqoob

> Novartis Institute for Medical Sciences, 5 Gower Place, London WC1E 6BS, U.K.

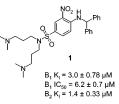
> > Received March 31, 2004

Abstract: The bradykinin B_1 receptor is rapidly induced after inflammation or tissue trauma and appears to play an important role in the maintenance of hyperalgesia in inflammatory conditions. Here, we describe the optimization process to identify novel, potent non-peptide human B_1 receptor antagonists based on a 2-alkylamino-5-sulfamoylbenzamide core. Optimized derivatives are selective, functional B_1 antagonists with low nanomolar affinity and exhibit oral bioavailability in animals.

Chronic pain affects a significant proportion of people worldwide and causes unnecessary suffering, disability, and a poor quality of life. There is a great medical need for new medicines to treat chronic nociceptive pain primarily because of the side effects that are frequently observed with existing treatments such as nonsteroidal antiinflammatory drugs (NSAIDs) and opioids.

The kinins are potent 9–10 amino acid peptide hormones involved in inflammatory, vascular, and pain processes and are released following inflammation, tissue damage, or other noxious stimulation.¹ In humans, the two endogenous kinins bradykinin (BK, Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg) and kallidin (KD, Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg) are formed by proteolytic cleavage of high- and low-molecularweight kininogen precursor proteins by plasma or tissue kallikrein (serine protease) enzymes, respectively.

The kinin peptides exert their biological effects by activating two distinct BK receptors (termed B_1 and B_2), which belong to the rhodopsin superfamily of G-proteincoupled receptors (GPCRs) but have low (36%) sequence homology to one another. Across species, there are also considerable differences in the amino acid sequences of the BK B_1 receptors. Chart 1. Bradykinin B₁ Antagonist Lead 1



While BK and KD have high affinity for constitutive BK B_2 receptors, these kinins have low affinity for BK B_1 receptors, which are generally present at low levels under normal conditions. However, B_1 receptors are rapidly induced and expressed in peripheral tissues and cells within 2–5 h after tissue injury or inflammation and are activated by the metabolic fragments des-Arg⁹-BK and des-Arg¹⁰-KD.²

Compelling evidence suggests that the BK B_1 receptor plays an important role in chronic nociceptive pain associated with inflammatory conditions.³ In animal models of persistent nociceptive pain, it has also been demonstrated that peptide B_1 receptor antagonists are effective in reversing inflammatory hyperalgesia.⁴ Thus, a potent, selective, orally bioavailable B_1 antagonist should have significant potential as a novel therapy for inflammatory pain.

Several classes of non-peptide antagonists for BK B_2 receptors have been disclosed in the scientific literature,^{5,6} and more recently the discovery of non-peptide antagonists for the B_1 receptor have been described.^{7,8} In this Letter, we describe our own efforts in the optimization of a micromolar lead compound to afford potent, selective, and orally bioavailable non-peptide BK B_1 receptor antagonists based on the 2-alkylamino-5-sulfamoylbenzamide core.

An in-house collection of non-peptide bradykinin B_2 antagonists, generated in our previous research that afforded bradyzide and related analogues,^{9–11} was assayed for binding affinity at the cloned human bradykinin B_1 receptor expressed in HEK293 cells. This screening process identified the 4-alkylamino-3-nitrobenzene sulfonamide **1** (Chart 1) as an interesting lead, having single-digit micromolar binding affinity (expressed as the inhibition constant $K_I \pm$ SEM) at both human B_1 and B_2 receptors, expressed in HEK293 and Cos-7 cells, respectively. **1** also behaved as a functional antagonist at human B_1 receptors expressed in Cos-7 cells, inhibiting the Ca²⁺ efflux induced by des-Arg¹⁰-KD with IC₅₀ = 6.2 μ M.

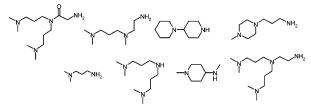
To rapidly investigate the importance of the bis-amine and the benzhydryl moieties in **1**, analogues were prepared. Thus, in separate reactions, eight different polar amines (see Chart 2) were coupled with commercially available 4-chloro-3-nitrobenzenesulfonyl chloride in THF in the presence of Et_3N to form the corresponding sulfonamide intermediates, which were each used crude for the subsequent nucleophilic aryl chloride displacement (DMSO at 120 °C) with nine

 $^{^{*}}$ To whom correspondence should be addressed. Phone: +44 20 7333 2168. Fax: +44 20 7387 4116. E-mail: tim.ritchie@pharma.novartis.com.

[†] Present address: Novartis Pharma AG, Basel, Switzerland. [‡] Present address: Pfizer Central Research, Sandwich, CT13 9NJ,

U.K. § Present address: Novartis Horsham Research Centre, Wimblehurst Road, Horsham RH12 5AB, U.K.

Chart 2. Amines Used for Initial Array of Analogues of 1 Step 1: amines for sulfonamide formation



Step 2: hydrophobic amines for aniline formation

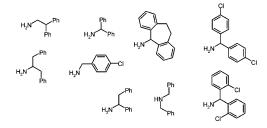
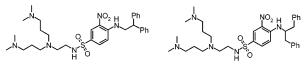
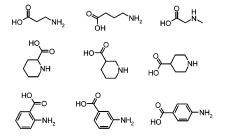


Chart 3. More Potent Analogues of 1 from First Array



2 $B_1 K_1 = 0.239 \pm 0.030 \mu M$ **3** $B_1 K_1 = 0.255 \pm 0.048 \mu M$

Chart 4. Amino Acid Linkers Used in Second Array



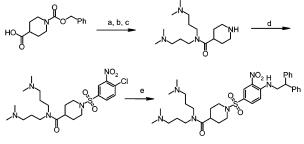
separate hydrophobic amines (Chart 2). The 71 new derivatives obtained by this process were screened for binding affinity at the human B_1 receptor and compared with that of 1.

The results indicated that while the majority of the members of the array were equiactive with **1** or less potent (particularly when the bis-amine motif was absent), a 10-fold increase in binding affinity was observed in two analogues where (i) an ethylene spacer had been introduced between the sulfonamide and the bis-amine and (ii) the benzhydryl group had been replaced by the 2,2-diphenylethyl or the dibenzylmethyl group (**2** and **3**, Chart 3). It is interesting that neither of these modifications, in the absence of the other, increased B₁ receptor binding affinity over that observed with **1**.

A subsequent array of analogues was prepared where the ethylene spacer in **2** and **3** was replaced with a variety of acyclic and cyclic amino acids (shown in Chart 4). For this array, 2,2-diphenylethylamine, dibenzylmethylamine, and 3,3-diphenylpropylamine were used as the hydrophobic amine partners. A representative synthesis for this array is shown in Scheme 1.

Results from the B_1 binding assay indicated that although the inclusion of methylaminoacetic acid, pip-

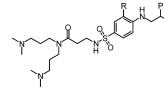
Scheme 1. Synthesis of Piperidine-4-carboxylic Acid Analogue $\mathbf{4}^a$



4 K₁ = 0.018 ± 0.003 μM

^{*a*} Conditions: (a) (COCl)₂, PhMe, DMF, room temp; (b) 3,3'iminobis(N,N-dimethylaminopropylamine), CH₂Cl₂; (c) H₂, 10% Pd/C, MeOH; (d) 4-chloro-3-nitrobenzenesulfonyl chloride, NEt₃, THF; (e) 2,2'-diphenylethylamine, DMSO, 150 °C.

Table 1. Binding Affinity of Nitro Group Replacements



	l	
compd	R	$B_1 K_I (\mu M)$
5	NO ₂	0.144 ± 0.022
6	Cl	>1
7	C(O)NMe ₂	0.136 ± 0.022
8	C(O)morpholine	0.012 ± 0.003

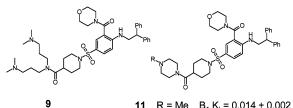
eridine-2-carboxylic acid, and the aromatic amino acids as spacers was not useful, incorporation of the 3aminopropionic acid, piperidine-3-carboxylic acid, and piperidine-4-carboxylic acid spacers increased receptor binding affinity relative to the simple ethylene spacer. In particular, the piperidine-4-carboxylic acid derivative of **2** (**4**, Scheme 1) exhibited $K_{\rm I} = 0.018 \ \mu M$.

In parallel to the optimization of the sulfonamidebis-amine spacer group, analogues were prepared in which the undesirable aromatic nitro group was replaced with alternative substituents. In this investigation, the 3-aminopropionic acid spacer was used with the 2,2-diphenylethyl group (Table 1). While the chloro derivative **6** had low affinity for B_1 receptors, the tertiary amide **7** exhibited a binding affinity similar to that of the parent nitro analogue **5**. Although primary and secondary amide derivatives were found to be inactive (as was the parent carboxylic acid), the utility of a tertiary amide as a nitro replacement was confirmed when morpholine was used in place of dimethylamine. This change resulted in **8**, which had an affinity similar to that of **4**.

Subsequently a hybrid molecule was prepared, which combined the optimal amino acid spacer (piperidine-4-carboxylic acid used in **4**) and nitro replacement (morpholine amide used in **8**). This analogue (**9**, Chart 5) was found to be a highly potent B₁ ligand, with a binding $K_{\rm I}$ of 0.001 μ M. **9** also behaved as a functional antagonist in Cos-7 cells, with an IC₅₀ of 0.012 μ M.

As discussed above, analogues prepared during the initial optimization of **1** that lacked the bis-amine motif tended to lose binding affinity at the B_1 receptor. With the discovery of **9**, we decided to reexplore the bis-amine



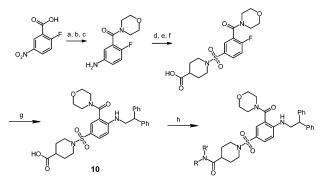


12

 $B_1 K_1 = 0.001 \pm 0.0001 \ \mu M$

 $\begin{array}{ll} {\sf R} = {\sf Me} & {\sf B}_1 \, {\sf K}_1 = 0.014 \pm 0.002 \ \mu {\sf M} \\ {\sf R} = {\sf Pr}^i & {\sf B}_1 \, {\sf K}_1 = 0.011 \pm 0.002 \ \mu {\sf M} \end{array}$

Scheme 2^a



 a Conditions: (a) (COCl)₂, PhMe, DMF, room temp; (b) morpholine, NEt₃, THF; (c) Zn, AcOH; (d) NaNO₂, HCl, AcOH; (e) SO₂, CuCl₂, AcOH, H₂O; (f) isonipecotic acid, Na₂CO₃, dioxane–water; (g) 2,2-diphenylethylamine, DMSO, 120 °C; (h) isopropyl chloroformate, NMM, THF, -40 °C, 40 min, followed by amine.

region to find a smaller monobasic amine to replace the iminobis (N, N)-dimethylpropylamine).

To expedite this, the advanced intermediate **10** was prepared according to the synthetic route shown in Scheme 2. The carboxyl group of **10** was then activated via the corresponding mixed anhydride and coupled with a range of cyclic and acyclic amines.

It soon became apparent that it was possible to replace the bis-amine with a number of cyclic and acyclic monoamines to afford analogues that retained a respectable level of B_1 binding affinity. As anticipated, the oral absorption profiles of some of the monoamines were superior to those of 9. For example, while 9 could not be detected in plasma after oral administration to rats, the two piperazine derivatives 11 and 12 (Chart 5) were found to exhibit reasonable oral absorption. This improvement was thought to be due to the reduction in overall molecular weight and the lower basicity of the piperazine nitrogens (p K_a of 6.5 and 7.6 for **11** and **12**, respectively). Thus, after an oral dose of 7 mg/kg, administered as a suspension in 0.5% methyl cellulose, the hydrochloride salts of **11** and **12** exhibited C_{max} of 913 and 196 nM and oral bioavailabilities of 42% and 15%, respectively. After intravenous administration to rats (0.7 mg/kg), the half-lives for 11 and 12 were estimated at 161 and 37 min, respectively. In dogs, a 20 mg/kg dose of 11 exhibited an oral bioavailability of 35% ($C_{\text{max}} = 10.3 \,\mu\text{M}$) when administered as an aqueous hydroxypropyl β -cyclodextrine inclusion complex.

11 and **12** also exhibited at least 500-fold selectivity for human bradykinin B_1 receptors over a panel of 30 G-protein-coupled receptors (including human BK B_2 receptors), 7 ion channels (including sodium, potassium, and calcium channels), and cyclooxygenase enzymes.

In conclusion, we have described the optimization of a micromolar lead ${\bf 1}$ to afford a novel series of non-

peptide antagonists for the human bradykinin B_1 receptor. The optimized derivatives are potent and selective antagonists with reasonable oral bioavailability in animals and as such have potential as a novel therapy for pain associated with inflammatory conditions.

Acknowledgment. The authors thank Devnandan Chatterjee, Wai Tsang, Lee Edwards, Frédéric Ratel, and Lucile Fisher for invaluable technical support, Christian Guenat for obtaining the HRMS data, and Olivier Kretz and Guy Taccard for conducting the dog pharmacokinetics study. The authors also thank Christopher Snell, Michael Brown, Peter Kipfer, Julian Arbuckle, Reg Docherty, Steven Phagoo, Ximena Nuñez, Wai Lee, Andrew Davis, Michael Webb, and Elsa Phillips.

Supporting Information Available: ¹H NMR spectra (400 MHz), HPLC and HRMS characterization data for all compounds, elemental analyses for **1**, **11**, and **12**, and details of human B₁ receptor binding and functional bioassays. This material is available free of charge via the Internet at http:// pubs.acs.org.

References

- Regoli, D.; Barabé, J. Pharmacology of Bradykinin and Related Kinins. *Pharmacol. Rev.* 1980, *32*, 1–46.
- (2) Marceau, F.; Hess, F.; Bacharov, D. R. The B₁ Receptor for Kinins. *Pharmacol. Rev.* **1998**, *50*, 357–85.
- (3) Couture, R.; Harrison, M.; Vianna, R. M.; Cloutier, F. Kinin Receptors in Pain and Inflammation. *Eur. J. Pharmacol.* 2001, 429, 161–176.
- (4) Belichard, P.; Landry, M.; Faye, P.; Bachvarov, D. R.; Bouthillier, J.; Pruneau, D.; Marceau, F. Inflammatory hyperalgesia induced by zymosan in the plantar tissue of the rat: effect of kinin receptor antagonists *Immunopharmacology* **2000**, *46*, 139–147.
- (5) Lynnsan in the plantal discue of the fat. enerce of klimin receptor antagonists *Immunopharmacology* 2000, 46, 139–147.
 (5) Heitsch, H. Non-Peptide Antagonists and Agonists of the Bradykinin B₂ Receptor. *Curr. Med. Chem.* 2002, *9*, 913–928.
 (6) Gurrath, M. Peptide-Binding G Protein-Coupled Receptors: New York and States and St
- (6) Gurrath, M. Peptide-Binding G Protein-Coupled Receptors: New Opportunities for Drug Design. *Curr. Med. Chem.* 2001, *8*, 1605– 1648.
- Bock, M. G.; Hess, J. F.; Pettibone, D. J. Bradykinin-1 receptor antagonists. *Annu. Rep. Med. Chem.* **2003**, *38*, 111–120.
 Gougat, J.; Ferrari, B.; Sarran, L.; Planchenault, C.; Poncelet, New York, New York,
- (8) Gougat, J.; Ferrari, B.; Sarran, L.; Planchenault, C.; Poncelet, M.; Maruani, J.; Alonso, R.; Cudennec, A.; Croci, T.; Guagnini, F.; Urban-Szabo, K.; Martinolle, J.-P.; Soubrie, P.; Finance, O.; Le Fur, G. SSR240612 [(2R)-2-[((3R)-3-(1,3-benzodioxol-5-yl)-3-{[(6-methoxy-2-naphthyl)sulfonyl]amino}-propanoyl)amino]-3-(4-{[2R,6.S)-2,6-dimethyl-piperidinyl]methyl}phenyl)-N-isopropyl-N-methylpropanamide hydrochloride], a new nonpeptide antagonist of the bradykinin B1 receptor: Biochemical and pharmacological characterization. J. Pharmacol. Exp. Ther. 2004, 309 (2), 661-669.
- (9) Dziadulewicz, E. K.; Ritchie, T. J.; Hallett, A.; Snell, C. R.; Ko, S. Y.; Wrigglesworth, R.; Hughes, G. A.; Dunstan, A. R.; Bloomfield, G. C.; Drake, G. S.; Brown, M. C.; Lee, W.; Burgess, G. M.; Davis, C.; Yaqoob, M.; Perkins, M. N.; Campbell, E. A.; Davis, A. J.; Rang, H. P. 1-(2-Nitrophenyl)thiosemicarbazides: A Novel Class of Potent, Orally Active Non-Peptide Antagonist for the Bradykinin B₂ Receptor. *J. Med. Chem.* **2000**, *43*, 769–771.
- (10) Burgess, G. M.; Perkins, M. N.; Rang, H. P.; Campbell, E. A.; Brown, M. C.; McIntyre, P.; Urban, L.; Dziadulewicz, E. K.; Ritchie, T. J.; Hallett, A.; Snell, C. R.; Wrigglesworth, R.; Lee, W.; Davis, C.; Phagoo, S. B.; Davis, A. J.; Phillips, E.; Drake, G. S.; Hughes, G. A.; Dunstan, A.; Bloomfield, G. C. Bradyzide, a Potent Non-Peptide B₂ Bradykinin Receptor Antagonist with Long-Lasting Oral Activity in Animal Models of Inflammatory Hyperalgesia. Br. J. Pharmacol. **2000**, *129*, 77–86.
- Hyperalgesia. Br. J. Pharmacol. 2000, 129, 77–86.
 (11) Dziadulewicz, E. K.; Ritchie, T. J.; Hallett, A.; Snell, C. R.; Davies, J. W.; Wrigglesworth, R.; Dunstan, A. R.; Bloomfield, G. B.; Drake, G. S.; McIntyre, P.; Brown, M. C.; Burgess, G. M.; Lee, W.; Davis, C.; Yaqoob, M.; Phagoo, S. B.; Phillips, E.; Perkins, M. N.; Campbell, E. A.; Davis, A. J.; Rang, H. P. Nonpeptide Bradykinin B₂ Receptor Antagonists: Conversion of Rodent-Selective Bradyzide Analogues into Potent, Orally-Active Human Bradykinin B₂ Receptor Antagonists. J. Med. Chem. 2002, 45, 2160–2172.

JM049747G