# Theoretical Study of 9- $\beta$ -D-Erythrofuranosyladenine and Corresponding Carbocyclic Analogues. Evidence for a Base-Activated Conformational Lock

Akin Akdağ, Cynthia M. Carver, Michael L. McKee,\* and Stewart W. Schneller\*

Department of Chemistry, Auburn University, Auburn, Alabama 36849 Received: July 8, 2002; In Final Form: September 3, 2002

The conformational surfaces of three nucleoside analogues have been investigated computationally, where an adenine is attached to a diol of tetrahydrofuran, a diol of cyclopentane, and a diol of cyclopentene. In each system, the lowest-energy conformer displays a conformational lock into the south position by an internal hydrogen bond between O2'H of the five-membered ring and the N3 nitrogen of adenine. When aqueous solvation is accounted for by the PCM method, the preference for the locked conformer is diminished. A pseudorotation angle of 9-(*trans*-2',*trans*-3'dihydroxycyclopentyl)adenine has been determined to be 176.8° by fitting the measured  ${}^{3}J_{\rm HH}$  values using PSEUROT which is in good agreement with the calculated value of 169.3°.

### Introduction

Nucleosides are the monomeric building blocks of nucleic acids and are composed of a heterocyclic base and ribofuranose unit. When the ring oxygen of the furanose group is replaced by a methylene, the class of compounds that results is referred to as carbocyclic nucleosides (CN).<sup>1</sup> Aristeromycin (1) is the carbocyclic nucleoside analog<sup>2</sup> of the naturally occurring adenosine (2) and has been found to display antiviral properties.<sup>3</sup> The clinical potential of 1 is, however, limited by its cytotoxicity, which has been attributed to metabolism to its 5'-phosphates (that is, nucleotides).<sup>4</sup>

Several years ago, efforts were undertaken to seek aristeromycin- based compounds with greater therapeutic possibilities via derivatives incapable of phosphorylation to the undesirable nucleotides.<sup>5</sup> With this in mind, several labs<sup>6,7</sup> prepared and assayed **3**, a compound lacking the C-4' hydroxymethylene of **1**. This proved very successful, as **3** was found to retain the significant antiviral properties of **1** but without cyctotoxicity.<sup>8,9</sup> This result led to **4**, a more natural-like furanose form of **3** (an erythrofuranose derivative). Surprisingly, **4** was found to be inactive.<sup>9</sup> If the C4'-C5' bond of aristeromycin is unsaturated, Neplanocin A is obtained (**5**), another well-known antiviral agent.<sup>10</sup> Very recently, the X-ray structure of the target enzyme *S*-adenosyl-L-homocysteine (AdoHcy) hydrolase<sup>11</sup> has been solved with **6** (replacing the CH<sub>2</sub>OH group of **5** with hydrogen) bound into the active site.<sup>12</sup> as two useful parameters for describing ring conformations.<sup>15</sup> The conformation of sugars as well as CNs can be described by the same parameters.<sup>16</sup> The variation of these parameters can be used to understand why certain compounds are more active than others. If a model compound has the same ring conformation as a parent natural compound, then it is more likely to bind to the natural receptor site in vivo. Of course, substituents play a very important role in determining the most stable conformer in bioactive compounds. For example, conformers can be locked into a certain position by the appropriate choice and location of substituent.<sup>17</sup> The tetrahydrofuran and cyclopentane rings are known to

attracted much attention.<sup>13,14</sup> Altona has proposed the pseu-

dorotation phase angle (P) and maximum torsion angle ( $\nu_{max}$ )

adopt two conformers: twisted and envelope. The passage from one envelope form to another can occur without going through the planar form by a process called pseudorotation.<sup>18</sup> This concept was first applied to five-membered sugar-containing nucleosides by Altona,<sup>15</sup> who also defined the pseudorotational phase angle (*P*) and maximum puckering ( $\nu_{max}$ ).

It was found<sup>16</sup> that nucleosides are mainly in the southern (C2'-endo) or in the northern (C3'-endo) hemispheres in the pseudorotation cycle (see below). In addition to the many

North

P=0



The analysis of conformations within biologically relevant five-membered rings, such as nucleosides and nucleotides, has



10.1021/jp021563v CCC: \$22.00 © 2002 American Chemical Society Published on Web 10/23/2002

<sup>\*</sup> To whom correspondence should be addressed.

TABLE 1: Relative Energies (kcal/mol) of A, B, and C Conformers at MMFF and DFT Levels of Theory

conformer	MMFF94	B3LYP/6-31G(d,p)// MMFF94	B3LYP/3-21G// B3LYP/3-21G	B3LYP/6-31G(d,p)// B3LYP/3-21G	B3LYP/6-31+G(d,p)// B3LYP/6-31+G(d,p)
A1	0.00	0.00	0.00	0.00	0.00
A2	3.68	6.25	10.06	6.37	→A1
A3	4.22	6.79	11.63	6.12	
A4	4.43	7.90	11.21	6.91	4.76
A5	4.55	9.45	12.09	8.81	
A6	5.19	6.87	12.90	6.79	
A7	5.21	9.46	15.20	8.07	
A8	5.33	6.05	8.04	5.64	4.68
A9	5.97	7.07	12.41	6.27	
A10	6.29	8.48	9.25	7.38	
A11	6.32	7.62	10.91	6.23	
A12	6.35	7.91	12.78	7.04	
A13	6.55	9.44	13.09	8.76	
A14	6.71	9.12	→A12	→A12	
B1	0.00	0.00	0.00	0.00	0.00
B2	0.08	-0.29	0.46	0.43	-0.40
B3	0.49	-4.99	-9.43	-5.40	-4.51
<b>B4</b>	0.66	-0.99	1.88	0.64	
B5	0.88	-4.33	→B3	→B3	
<b>B6</b>	1.19	-1.30	1.16	0.58	
B7	1.22	-1.42	-0.76	-0.77	
B8	2.03	0.13	→B1	→ <b>B</b> 1	
<b>B9</b>	2.38	2.61	-0.40	1.81	
B10	2.97	1.39	2.27	0.96	
B11	3.41	2.13	-0.20	0.87	
B12	4.07	2.07	4.49	2.96	
B13	4.46	3.24	3.28	4.99	
B14 D17	4.60	2.07	4.06	2.11	
B15	4.67	1.76	3.49	1.00	
B10 D17	4.84	2.65	2.82	3.25	
B17 D19	5.33	1.00	4.28	3.11 • <b>D</b> 16	
D10 D10	5.77	5.90	→ <b>D10</b>	B10 2.27	
D19 D20	5.80	1.95	1./3 • <b>D1</b> 5	2.37	
B20 B21	5.07	1.27	<b>D15</b> 2.05	2.02	
B21 B22	6.12	1.27	5.95 8.45	2.03	
D22 C1	0.18	4.31	0.45	0.00	
C1	0.00	-1.78	-1.21	-0.92	0.00
C3	0.04	-6.42	-7.38	-4.75	-4.03
C4	0.88	-1 51	→C1	→C1	4.05
C5	1.67	-1.35	$\rightarrow C3$	$\rightarrow$ C3	
C6	2.63	1.29	0.80	-0.80	0.08
Č7	2.70	0.03	1.32	0.28	0.00
C8	2.80	1.19	→C1	→C1	
C9	2.81	-0.21	0.69	-0.90	
C10	3.79	0.61	3.46	1.35	
-					

experimental studies on conformers of nucleosides and modified nucleosides, several theoretical studies have also been reported.<sup>19</sup>

A theoretical investigation of 4, 3, and 6 (A, B, and C) was undertaken to determine the nature of the lowest-energy conformers. It is of considerable interest to know whether differences in the conformation of the cyclopentyl and furanose rings of 3 and 4, a feature that has been the source of variable activity in other nucleoside derivatives,<sup>1h</sup> might be responsible for the difference in activity between 3 and 4.

## Methods

**Experimental.** NMR spectra of **B** were recorded (Bruker DRX 500) in DMSO- $d_6$  with DCl added to simplify the spectra through deuterium exchange. The coupling constants were found by irradiating corresponding peaks and the assignments made by using COSY, HETCOR, and NOESY spectra.

**Computational.** A search of low-energy conformers was made for **A**, **B**, and **C** by using the MMFF molecular mechanics force field (MMFF94)<sup>20</sup> and the Monte Carlo method (with default options) as implemented in the Spartan 5.1 program.<sup>21</sup> Within an energy range of 7 kcal/mol, 14 conformers were

returned for **A**, 23 for **B**, and 10 for **C**. One conformer returned for **B** required breaking and reforming the five-membered ring and was, therefore, removed from consideration.

Each of the MMFF94-minimized conformers was subjected to a single-point calculation at the B3LYP/6-31G(d,p)//MMFF94 level using Gaussian 98.22 These structures were then fully optimized at the B3LYP/3-21G level, and single-point energies were obtained at the B3LYP/6-31G(d,p)//B3LYP/3-21G level.<sup>23</sup> Four conformers of A and three of B and C were selected for further optimization at the B3LYP/6-31+G(d,p) level. The energy ordering given by MMFF94 was used to designate the conformers of A (A1-14), B (B1-B22), and C (C1-C10). The conformational parameters for each conformer were found by using ConforMole.<sup>24</sup> Relative energies (kcal/mol) at different levels of theory are given for A, B, and C in Table 1, whereas structural data [pseudorotation phase angle (P), maximum torsion angle ( $\nu_{max}$ ), and base torsion angle ( $\chi$ )] for the same conformers are given in Table 2. Molecular plots of the conformers optimized at the B3LYP/6-31+G(d,p) level are given in Figures 1 (A conformers), 2 (B conformers), and 3 (C conformers)). Cartesian coordinates of conformers optimized

TABLE 2: Pseudorotation Phase Angle (*P*), Maximum Torsion Angle ( $v_{max}$ ) and Torsion Angle ( $\chi = C5'-C1'-N9-C4$ )<sup>*a*</sup> for Conformers of A, B, and C

	MMFF94		B3LYP/3-21G		B3LYP/ 6-31+G(d,p)				
confor.	P	$\nu_{\rm max}$	χ	Р	$\nu_{ m max}$	χ	Р	$\nu_{\rm max}$	χ
A1	176.8	37.2	174.1	154.8	41.8	168.2	169.3	36.2	169.9
A2	20.3	37.0	189.0	0.0	36.0	167.9		→A1	
A3	199.2	36.3	62.8	222.6	38.4	64.8			
A4	31.6	35.7	82.4	25.0	38.3	65.1	30.9	33.9	72.9
A5	15.3	38.3	187.1	2.3	39.0	164.7			
A6	181.2	37.2	66.8	161.1	41.7	73.2			
A7	181.4	38.3	212.1	159.0	40.7	217.4			
A8	207.8	38.3	65.3	233.2	48.0	73.3	223.5	40.7	68.6
A9	188.3	36.3	63.9	220.9	37.7	69.9			
A10	354.0	37.0	186.4	305.0	44.8	150.5			
A11	181.3	40.9	176.9	178.4	44.4	164.6			
A12	30.2	33.2	84.7	29.6	36.5	74.9			
A13	195.6	39.5	212.5	238.5	48.7	263.0			
A14	16.4	35.4	104.0		$\rightarrow$ A12	<b>7</b> 0 4			
BI	41.0	42.0	65.8	34.6	45.1	58.6	43.1	42.4	61.1
B2	205.6	41.2	56.0	225.9	44.1	63.2	189.4	40.8	57.5
B3	175.5	43.4	144.8	145.9	46.3	162.4	151.9	42.0	161.1
B4	195.4	42.5	60.4	213.5	46.1	64.6			
B5 D(	208.3	42.8	190.0	210.0	→B3	(7.2			
B0 D7	200.1	42.4	58.1	219.9	45.9	0/.3			
Б/ DQ	40.4	42.0	667	34.7	44.1 → <b>D1</b>	61.1			
	40.4	41.8	00./ 92.1	200 0		02 /			
D9 D10	215.5	12 5	60.1	200.0	41.4	61.2			
D10 R11	40.2	42.3	307.6	41.9	44.0	324.1			
B11 B12	200.1	43.1	245.2	231.0	40.2	24.1			
B12 B13	333.3	42.4	196.1	301.0	45.5	158.5			
B13 B14	34.2	42.5	258.9	32.0	44.4	244.8			
B15	37.7	42.5	222.6	42.7	44.3	226.1			
B16	339.2	39.7	200.3	301.4	43.9	166.9			
B17	204.5	42.6	233.9	231.1	44.0	229.4			
B18	328.9	40.1	196.3		→B16				
B19	175.0	43.9	307.2	142.8	48.0	332.6			
B20	38.5	42.1	247.5		→B15				
B21	31.5	42.3	241.8	32.1	43.2	239.7			
B22	192.2	45.2	59.3	165.7	49.6	56.1			
C1	348.2	18.0	71.2	334.6	15.0	69.4			
C2	347.6	18.4	74.2	345.6	20.6	69.9	349.3	17.7	66.1
C3	161.1	20.5	181.8	158.7	32.4	172.3	160.8	26.7	175.3
C4	346.6	16.2	72.4		→C1				
C5	346.3	19.8	193.3		→C3				
C6	345.5	24.6	206.1	165.6	21.6	234.9	164.6	26.4	237.7
C7	344.4	22.8	208.4	342.3	28.3	214.0			
C8	347.8	16.8	77.9		→C1				
C9	344.2	22.6	204.3	344.8	21.0	195.8			
C10	160.0	18.6	62.2	159.7	32.0	74.1			

<sup>*a*</sup> This torsion angle describes the orientation of the base  $(90-270^{\circ} \text{ anti}, 0-90^{\circ} \text{ and } 270-360^{\circ} \text{ syn})$ .

at the B3LYP/3-21G and B3LYP/6-31+G(d,p) levels are available as Supporting Information.

The PSEUROT program<sup>25</sup> calculates  ${}^{3}J_{\text{HH}}$  values from the generalized Karplus equation and compares them with experimental values to determine the endocyclic dihedral angles from exocyclic H–C–C–H dihedral angles. The program assumes the coexistence of two conformations, south and north.<sup>26</sup> The relationship between exocyclic dihedral angles and endocyclic dihedral angles is given as  $\varphi_{\text{exo}} = A\varphi_{\text{endo}} + B$ , where the *A* and *B* values<sup>15c,15d</sup> are found by fixing  $\nu_{\text{max}} = 38$  and varying *P* values in increments of 30° for the entire pseudorotational cycle and optimizing at the B3LYP/3-21G level (Figure 4). The other  $\nu$  values were found from the formula,  $\nu_i = \nu_{\text{max}} \cos(P + 4\pi i/5)$ .<sup>15b</sup> For values of P along the pseudorotation circle, the optimization was started with the base in the anti and syn orientation. The plot in Figure 4 was made from the energies of structures in the lower-energy orientation for each value of



Figure 1. B3LYP/6-31+G(d,p) optimized conformers A1, A4, and A8.

*P*. For angles of *P* between  $60^{\circ}$  and  $240^{\circ}$ , the energy of the ring was lower with the base in the anti orientation (see Supporting Information for table of energies). From these structures, the *A* and *B* values were found (Table 3).

## **Results and Discussion**

**Conformational Analysis of A.** At the B3LYP/6-31G(d,p)// MMFF94 level, the most stable conformer A1 (south/anti) was calculated to be 6.05 kcal/mol more stable (Table 1) than the next lowest-energy conformer, A8 (south/syn). At the B3LYP/ 3-21G//B3LYP/3-21G level, A1 was 8.04 kcal/mol more stable than A8. The DFT optimization moved the ring conformation of A1 toward the east ( $P = 176.8^{\circ} \rightarrow 154.8^{\circ}$ ), whereas A8 moved toward the west ( $P = 207.8^{\circ} \rightarrow 223.2^{\circ}$ ). Conformer A14 collapsed to A12 when optimized at the B3LYP/3-21G level, indicating that the barrier between them disappears at the DFT level. Comparing relative energies at the B3LYP/6-31G(d,p) level when B3LYP/3-21G geometries are used rather than MMFF94 geometries, the largest change in relative energies is about 1.4 kcal/mol (Table 1).

The next step in refinement is to optimize several conformers at the B3LYP/6-31+G(d,p) level. Because of the significant amount of computer time required, only four conformers of **A** were chosen, two south (anti and syn; **A1** and **A8**) and two north (anti and syn; **A2** and **A4**). During the course of geometry optimization, conformer **A2** collapsed to **A1**, indicating that the activation barrier between them disappeared when a larger basis set was applied. The energy difference between **A1** and **A8** decreased to 4.68 kcal/mol. The lower energy of **A1** is due to an "internal conformational lock"<sup>27</sup> created by a hydrogen bond



Figure 2. B3LYP/6-31+G(d,p) optimized conformers B1, B2, and B3.

between N3 in the adenine base and the hydrogen of O2' on the 5 ring (see standard atomic labeling below). The base is



syn in both A4 (north,  $P = 30.9^{\circ}$ ) and A8 (south,  $P = 223.5^{\circ}$ ) and the relative energies are very similar (4.76 and 4.68 kcal/ mol, respectively), which indicates that, without the "internal lock", north and south conformers have similar stabilities.

At each level of theory, conformer A1 remained lowest in energy because of the two consecutive hydrogen bonds (O3'H···O2' and O2'H···N3; see Figure 1 and Table 1), which also locks A1 into a south conformer. Although the level of theory used for geometry optimization did not affect the orientation of the base (see dihedral angle  $\chi$  in Table 2), the pseudorotation angle changed by up to 49.0° in going from MMFF94 to B3LYP/3-21G. In the case of A2, optimization at the B3LYP/6-31+G(d,p) level resulted in the collapsed to A1, a 169° change in pseudorotation angle (0.0°  $\rightarrow$  169.3°).

The second lowest energy conformer, **A8**, is in the southern part of pseudorotational cycle with a syn orientation of the base. There are two relatively short nonbonded CH···N interactions



Figure 3. B3LYP/6-31+G(d,p) optimized conformers C2, C3, and C6.



**Figure 4.** Pseudorotation potential energy surface of **B** optimized at the B3LYP/3-21G level. Empty circles indicate an anti orientation of the base, whereas filled circles indicate a syn orientation of the base.

 TABLE 3: A and B Values (u and d Represents Up and Down Relative to the Adenine)

exocylic dihedral	Λ	B	<b>P</b> <sup>2</sup>	exocylic dihedral	Δ	R	<b>P</b> <sup>2</sup>
angle $(\psi)$	л	Б	Λ	angle $(\psi)$	л	Б	Λ
H1'-H2'	1.16	123.4	0.994	H1′-H5'u	1.17	-125.7	0.983
H2'-H3'	1.07	-4.6	0.992	H3'-H4'd	1.14	-120.4	0.998
H1'-H5'd	1.17	-5.4	0.998	H3'-H4'u	1.05	0.1	0.998

(2.588 and 2.584 Å, Figure 1 and Table 4) in **A8** which are reminiscent of the CH···N type interactions found in the crystal structures of nucleic acids.<sup>28</sup> Two of the short CH···N contacts C3'H···N3 (**A4**) and C2'H···N3 (**A8**) involve a sp<sup>3</sup> hybridized carbon with a OH substituents. It is known that electronegative substituents attached to carbon increase the strength of CH···N

TABLE 4: Length (Å) and the Angle (degrees) of Hydrogen Interactions for Conformers of A, B, and C

conformer	bond type	length	angle
A1	O2'H····N3	1.907	113.7
	O2'H•••O3'	2.147	152.1
A4	O2'H•••O3'	2.078	118.4
	C3'H•••N3	2.541	123.9
A8	O2'H•••O3'	2.085	119.1
	C2'H···N3	2.588	119.5
	C4'H•••N3	2.584	126.9
B1	O2'H•••O3'	2.114	113.3
B2	O2'H•••O3'	2.173	113.3
B3	O3'H•••O2'	2.124	113.6
	O2'H…N3	1.810	155.6
C2	O2'H•••O3'	1.969	123.3
C3	O3'H•••O2'	2.065	117.1
	O2'H…N3	1.942	152.7
C6	O2'H•••O3'	2.103	118.7
	C1'H•••N3	2.547	107.4

hydrogen bonding by increasing the hydrogen acidity.<sup>29,30</sup> In a recent analysis of neutron diffraction structures of CH···N hydrogen bonded systems,<sup>31</sup> the mean H···N distance was found to be  $\bar{r} = 2.43$  Å (range 2.32–2.50 Å) and the mean CH···N angle was found to be  $\bar{\alpha} = 150^{\circ}$  (range 139–166°). Because the CH···N distances and angles in A (Table 4) are out of this observed range, the interactions are likely to be weaker than normal CH···N hydrogen bonds but perhaps sufficient to stabilize conformers A4 and A8 with a syn orientation of the base. In this respect, it is interesting to note that an AIM analysis of CH···O hydrogen bonding has recently been carried out on several nucleosides.<sup>32</sup> Such an analysis of the CH···N interaction in A might reveal whether it should be considered as a hydrogen bond.

The conformational analysis of **A** has been done experimentally by Kline et al.<sup>14e</sup> From an analysis of the cyclopentyl <sup>1</sup>H– <sup>1</sup>H coupling constants determined in water using PSEUROT, they reported a dominant South conformer (95%) with a pseudorotation angle of  $180.1^{\circ}$ .<sup>33</sup> The pseudorotation angle of **A1** by the B3LYP/6-31+G(d,p) level is in good agreement (169.3°). In the gas phase, only one conformer is expected. However, solution effects will likely diminish the preference for the locked conformer (see discussion of solvation effects below).

Conformational Analysis of B. In contrast to A, single-point energies at the B3LYP/6-31G(d,p)//MMFF94 level of theory show significant differences in relative energies. Specifically, B3 and B5 were stabilized by 5.48 and 5.21 kcal/mol relative to B1, respectively. When the conformers B1-B22 were optimized at the B3LYP/3-21G level, B3 became the lowestenergy conformer (9.43 kcal/mol lower than B1) and B5 collapsed to B3. The adenine base is syn in B1 and anti in B3. B1 is in a north conformation, whereas B2 and B3 are in south conformations. At the B3LYP/6-31G(d,p)//B3LYP/3-21G level, the ordering is B3 < B7 < B1 < B2 at relative energies of -5.40, -0.77, 0.00, and 0.43 kcal/mol, respectively. Because **B7** and **B1** appear to have very similar values of *P*,  $v_{\text{max}}$ , and  $\chi$ , only conformers **B1**, **B2**, and **B3** were optimized at the B3LYP/6-31+G(d,p) level. At this level, B3 is 4.51 kcal/mol lower in energy than **B1**. The *P*,  $v_{max}$ , and  $\chi$  values of **B3** are 151.9°, 42.0°, and 161.1° which are very similar to the lowest conformer A1 (169.3°, 36.2°, and 169.9°) which indicates that A and B both have nearly the same conformation which is about 4.5 kcal/mol more stable than the next lowest energy conformer.

The parameters for the hydrogen bonding in **B** are given in Table 4. It can be seen that the O2'H···N3 hydrogen bond is shorter in **B3** than in A1 (1.810 versus 1.907 Å) which may be

TABLE 5: Chemical Shifts ( $\delta$ , ppm) and <sup>1</sup>H-<sup>1</sup>H Coupling Constants (<sup>3</sup>J<sub>HH</sub>, Hz) for Compound B<sup>*a*</sup>

	chemical shift	coupling constants ( ${}^{3}J_{\rm HH}$ , Hz)				
nuclei	(δ, ppm)	H1′	H2′	H3′		
H1′	4.7(6.02)					
H2′	4.3(4.94)	9.31(6.7)				
H3′	4.0(4.54)		3.97(4.6)			
H4'u	2.1(4.56)			4.91(3.8)		
H4′d	1.6(4.09)			1.82(1.7)		
H5'u	2.0	9.2				
H5′d	2.2	9.2				

<sup>*a*</sup> The experimental values for **A** are in parentheses. Reference 14e.

TABLE 6: Comparison of Geometric Parameters for X-ray Structure of A and Calculated Values (B3LYP/6-31+G(d,p)) of A1

	X-ray		
parameters <sup>b</sup>	unit A	unit B	theory (A1)
Р	168.9	117.0	169.3
$\nu_{ m max}$	44.4	40.6	36.2
χ	$246.1^{c}$	69.4	169.9
C1'-C2'	1.508	1.519	1.547
C2'-C3'	1.532	1.541	1.536
C3'-C4'	1.514	1.536	1.526
C4'-O4'	1.458	1.438	1.452
O4'-C1'	1.422	1.409	1.409
C1'-N9	1.458	1.447	1.468
C2'-O2'	1.404	1.415	1.405
C3'-O3'	1.415	1.420	1.420

<sup>*a*</sup> Serianni, A. S. unpublished results. Unit cell  $(P2_12_12_1)$  contains two molecules of **A**. <sup>*b*</sup> Pseudorotation angle (*P*), pseudorotation amplitude ( $\nu_{max}$ ), torsion angle ( $\chi = C5'-C1'-N9-C4'$ ), and bond distances (Å). <sup>*c*</sup> The corresponding value from X-ray in aristeromycin (**1**) is 246.5 (-113.5). Kishi, T.; Muroi, M.; Kusaka, T.; Nishikawa, M.; Kamiya, K.; Mizuno, K. *Chem. Pharm. Bull.* **1972**, 20, 940. For the X-ray structure of adenosine (**2**), see: Lai, T. F.; Marsh, R. E. *Acta Crystallogr.* **1972**, *B28*, 1982.

due to the greater puckering in cyclopentane ring ( $\nu_{max} = 42.4^{\circ}$ ) compared to the tetrahydrofuran ring ( $\nu_{max}=36.2^{\circ}$ ; see Figure 2 and Table 2).

Relative energies at the B3LYP/6-31G(d,p)//B3LYP/3-21G level (Table 1) suggest that **B** has only one dominant conformation (the "locked" conformation). Under the one-state assumption and using the experimental proton coupling constants obtained in DMSO (Table 5), PSEUROT6.3 gives  $P = 176.8^{\circ}$  and  $\nu_{\text{max}} = 41.9^{\circ}$  with an RMS error of 0.6 Hz.

Thibaudeau et al.<sup>16</sup> have compared the X-ray and solution phase structures of aristeromycin (1). In aqueuous solution, 1 has  $P = 136.1^{\circ}$  and  $\nu_{max} = 37.4^{\circ}$  compared to the X-ray value of 89.0° and 40.8°, respectively. The ratio of syn:anti in solution is about 46:56, whereas the base is anti in the X-ray structure (torsion angle,  $\chi = 246.1^{\circ}$ ). The authors<sup>16</sup> conclude that the "solution- and the solid-state structure of aristeromycin are indeed different".

In the course of their study, Thibaudeau et al.<sup>16</sup> reparametrized the Haasnoot–Altona Karplus equation to obtain a better fit between calculated and experiment coupling constants for aristeromycin (1). When these parameters are used in the PSEUROT program for **B** (and assuming one state), the RMS is reduced to 0.5 Hz with very similar values of *P* and  $v_{\text{max}}$ (176.3° and 41.2°, respectively).

In the X-ray structure of **A**, there are two molecules per unit cell (Table 6) with pseudorotation angles (*P*) of 168.9° and 117.0° which can be compare to  $P = 169.3^{\circ}$  for **A1**. The base torsion angles ( $\chi$ ) are 246.1° (anti) and 69.4° (syn) which can be compared to  $\chi = 169.9^{\circ}$  (syn) in **A1**. Although there is some



**Figure 5.** (a) *P* versus relative energy (kcal/mol) at B3LYP/3-21G for A1–A14. (b) *P* versus relative energy (kcal/mol) at B3LYP/3-21G for B1–B22. (c) *P* versus relative energy (kcal/mol) at B3LYP/3-21G for C1–C10. The lowest energy conformer has a very similar *P* value for A1 ( $P = 154.8^{\circ}$ ), B3 ( $P = 145.9^{\circ}$ ), and C3 ( $P = 158.7^{\circ}$ )

similarity between the calculated structure (A1) and each of the two molecules in the unit cell (unit A and unit B), neither unit in the X-ray structure displays the hydrogen lock between O2'H of the sugar and N3 of the adenine base. Thus, similar to aristeromycin (1), it appears that the conformer of A is different in X-ray and gas/solution phases.

**Conformational Analysis of C.** A total of 10 conformers were returned from the MMFF94 search which were reduced to seven by optimizing at the B3LYP/3-21G level of theory. From these structures, conformers **C2**, **C3**, and **C6** were further optimized at the B3LYP/6-31+G(d,p) level (Figure 3). The low

 TABLE 7: Relative Energies (kcal/mol) of

 Modified Nucleotides in Water by

 PCM/B3LYP/6-31+G(d,p)//B3LYP/6-31+G(d,p)<sup>a</sup>

	gas phase ( $\epsilon = 1.0$ )	water ( $\epsilon = 78.39$ )
A1	0.00	0.00
A4	4.76	2.49
A8	4.68	1.38
B1	4.51	1.80
B2	4.11	0.93
B3	0.00	0.00
C2	4.03	3.75
C5	0.00	0.00
C6	3.95	0.16

<sup>a</sup> Dielectric constant is given in parentheses.

number of conformers can be accounted for by the presence of the double bond in the five-member ring.

The unsaturated five-membered ring in C was analyzed in the same manner as the saturated five-membered rings of A and B which allowed direct comparison of results. The lowestenergy conformer of C (C3) displayed the same O2'H···N3 lock as found in A1 and B3 (Figures 1 and 2). The C3 conformer, which is in the southern hemisphere ( $P = 160.8^{\circ}$ ), is 4.03 kcal/ mol more stable than the next lowest conformer. The maximum puckering ( $v_{max} = 26.7^{\circ}$ ) in C3 is less than in the corresponding saturated rings of A1 (36.2°) and B3 (42.0°), which is due to the stiffening of the ring by the double bond. The second lowest energy conformer (C2) is in the northern hemisphere (P =349.3°,  $v_{max} = 17.7^{\circ}$ ) with a syn orientation of the base. The third lowest conformer (C6) is only 0.12 kcal/mol higher than C2 and is in the southern hemisphere ( $P = 164.6^{\circ}$ ,  $v_{max} = 26.4^{\circ}$ ) with an anti orientation of the base.

It is interesting to point out that the conformer with the base in an pseudoaxial position (C2) has a  $v_{\text{max}}$  value about 10° smaller than the conformers with the base in an pseudoequatorial position (C3 and C6). It is possible that the  $\pi_{C4'-C5'}$  electrons can conjugate with the  $\sigma^*_{C1'-N9}$  orbital in C2 which may flatten the ring.<sup>34</sup>

**General Considerations.** As discussed above, **A**, **B**, and **C** all have similar structures for the lowest energy conformation. The absence of other conformations close in energy to the locked conformer is emphasized by plotting conformer energies as a function of pseudrotation angle (Figure 5). Houseknecht et al.<sup>24c</sup> report the conformational analysis of 3-O-methyl- $\alpha$ -D-arabino-furanoside and found a significantly different distribution which suggests that the nature of substituents in the five-membered ring has a large effect on the conformational preferences.

Solvation Effects. A major limitation of the present calculations is that they refer to the gas phase (dielectric constant,  $\epsilon =$ 1.0), whereas NMR measurements are usually performed in water ( $\epsilon = 78.39$ ) or DMSO ( $\epsilon = 46.70$ ). In the presence of solvent molecules, the specific hydrogen-bond lock O2'H···N3 will weaken or may completely disappear. To evaluate the effect of solvation, single-point calculations were made with the polarized continuum model (PCM) solvation method<sup>36</sup> at the PCM/B3LYP/6-31+G(d,p)//B3LYP/6-31+G(d,p) for conformers of A, B, and C (Table 7). For A and B, the locked conformers (A1 and B3) are still the lowest energy by about 1 kcal/mol which suggests that the lowest energy conformers are unchanged in the presence of solvation. For C, the locked conformer C5 is only 0.2 kcal/mol more stable than C6. Both conformers are in the south, but there may be nearly free rotation of the base around the C1'-N9 bond. It is interesting to point out that in aristeromycin (1) the ratio of syn to anti in water has been determined to be 44:56 from NOE enhancements.<sup>16</sup>

#### Conclusions

The lowest energy conformer of three nucleosides containing an adenine base (**A**, **B**, and **C**) each displays an internal hydrogen bond between O2'H of the five-membered ring and the N3 nitrogen of the adenine base. The very short hydrogen bond O2'H···N3 distances in **A**, **B**, and **C** of 1.907, 1.810, and 1.942 Å, respectively, indicate a conformational lock that could have biological significance. In the gas phase, the locked conformer is over 4 kcal/mol more stable than the next lowest conformer. When aqueous solvation is modeled using PCM, the preference is reduced to about 1 kcal/mol for **A** and **B** and to about 0.2 kcal/mol for **C**. Given the known difference in the antiviral properties of **B** and **C** versus **A** (**B** and **C** are active, **A** is inactive), we conclude that the conformational lock proposed with structures **A1**, **B3**, and **C3** does not account for the observed biological property correlation.

Acknowledgment. Computer time was provided by the Alabama Supercomputer Network and Maui High Performance Computer Center. M.L.M. would like to thank Sun Microsystems Computer Corporation for the award of an Academic Equipment Grant. S.W.S. would like to acknowledge the support of the Department of Health and Human Services (Grant AI48495). We thank Professor Cornelis Altona for helpful discussions about PSEUROT6.3, Professor Katherine L. Seley for taking NMR, and Professor Peter Livant for helping to interpret NMR and NOESY spectra. We would also like to acknowledge Professor Anthony Serianni for providing the X-ray structure of 9- $\beta$ -D-erythrofuranosyladenine (A) and P. R. McCarren for providing a copy of the ConforMole program.

**Supporting Information Available:** Relative energies (kcal/ mol) at the MMFF94 level and total energies (hartrees) at DFT levels (Table S1, 1 page). Relative energies (kcal/mol) and total energies (hartrees) at the B3LYP/3-21G//B3LYP/3-21G level of structures on the pseudorotation potential energy surface of **B** (Table S2, 1 page). Cartesian coordinates for relevant structures optimized for **A**, **B**, and **C** conformers at the B3LYP/ 3-21G and B3LYP/6-31+G(d,p) levels of theory (Tables S3 and S4, 29 pages). This material is available free of charge via the Internet at http://pubs.acs.org.

#### **References and Notes**

(a) De Clercq, E. Nucleosides Nucleotides 1998, 17, 625. (b) De Clercq, E. Nucleosides Nucleotides 1994, 13, 1271. (c) Perigaud, C.; Gosselin, G.; Imbach, J. L. Nucleosides Nucleotides 1992, 11, 903. (d) Huryn, D. M.; Okabe, M. Chem. Rev. 1992, 92, 1745. (e) Marquez, V. E.; Lim, M. I. Med. Res. Rev. 1986, 6, 1. (f) Marquez, V. E. Carbocyclic Nucleosides. In Advances in Antivirial Drug Design; De Clercq, E., Ed.; JAI Press: Greenwich, CT, 1997; Vol. 2, pp 89–145. (g) Crimmins, M. T. Tetrahedron 1998, 54, 9229. (h) Herdewijn, P. Drug Discovery Today, 1997, 2, 235.

(2) (±)-Aristeromycin was synthesized<sup>2a,b</sup> prior to isolation and structure proof of the biologically active<sup>2c</sup> (-) enantiomer.<sup>2d,e</sup> (a) Shealy, Y. F.; Clayton, J. D. J. Am. Chem. Soc. **1966**, 88, 3885. (b) Shealy, Y. F.; Clayton, J. D. J. Am. Chem. Soc. **1969**, 91, 3075. (c) Herdewijn, P.; Balzarini, J.; De Clercq, E.; Vanderhaeghe, H. J. Med. Chem. **1985**, 28, 1385. (d) Kusaka, T.; Yamamoto, H.; Shibata, M.; Muroi, M.; Kishi, T.; Mizuno, K. J. Antibiot. (Tokyo) **1968**, 21, 255. (e) Kishi, T.; Muroi, M.; Kusaka, T.; Nishikawa, M.; Kamiya, K.; Mizuno, K. Chem. Pharm. Bull. **1972**, 20, 940.

(3) (a) De Clercq, E. *Biochem. Pharmacol.* **1987**, *36*, 2567. (b) Cools, M.; De Clercq, E. *Biochem. Pharmacol.* **1989**, *38*, 1061.

(4) (a) Bennett, L. L., Jr.; Allan, P. W.; Hill, D. L. *Mol. Pharmacol.* **1968**, *4*, 208. (b) Hill, D. L.; Straight, S.; Allan, P. W.; Bennett, L. L., Jr. *Mol. Pharmacol.* **1971**, *7*, 375. (c) Bennett, L. L., Jr.; Brockman, R. W.; Rose, L. M.; Allan, P. W.; Shaddix, S. C.; Shealy, Y. F.; Clayton, J. D. *Mol. Pharmacol.* **1985**, *27*, 666.

(5) (a) For a leading reference, see: Rajappan, V. P.; Hegde, V. R.; Schneller, S. W. Synth. Commun. 2001, 31, 2849. (b) Siddiqi, S. M.; Chen, X.; Rao, J.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1995, 38, 1035. (c) Siddiqi, S. M.; Chen, X.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1994, 37, 551. (d) Siddiqi, S. M.; Schneller, S. W. Nucleosides Nucleotides 1993, 12, 185.

(6) (a) Seley, K. L.; Schneller, S. W.; Korba, B. J. Med. Chem. 1998, 41, 2168. (b) Seley, K. L.; Zhang, L.; Hagos, A.; Quirk, S. J. Org. Chem. 2002, 67, 3365.

(7) Wolfe, M. S.; Lee, Y.; Bartlett, W. J.; Borcherding, D. R.; Borchardt, R. T. J. Med. Chem. **1992**, 35, 1782.

(8) Yuan, C.-S.; Liu, S.; Wnuk, S. F.; Robbins, M. J.; Borchardt, R. T. Adv. Antiviral Drug Design 1996, 2, 41.

(9) Schneller, S. W.; Kern, E. R.; De Clercq, E. Unpublished results.
(10) (a) Obara, T.; Shuto, S.; Saito, Y.; Snoeck, R.; Andrei, G.; Balzarini,
J.; De Clercq, E.; Matsuda, A. J. Med. Chem. 1996, 39, 3847. (b)
Borcherding, D. R.; Scholtz, S. A.; Borchardt, R. T. J. Org. Chem. 1987, 52, 5457. (c) Kitade, Y.; Kozaki, A.; Yatome, C. Tetrahedron Lett. 2001, 42, 433.

(11) (a) Guillerm, G.; Guillerm, D.; Vandenplas-Witkowki, C.; Rogniaux, H.; Carte, N.; Leize, E.; Dorsselaer, A. V.; De Clercq, E.; Lambert, C. J. Med. Chem. 2001, 44, 2743. (b) Komoto, J.; Huang, Y.; Gomi, T.; Ogawa, H.; Takata, Y.; Fujioka, M.; Takusagawa, F. J. Biol. Chem. 2000, 275, 32147. (c) Hu, Y.; Komoto, J.; Huang, Y.; Gomi, T.; Ogawa, H.; Takata, Y.; Fujioka, M.; Takusagawa, F. Biochemistry 1999, 38, 8323. (d) Turner, M. A.; Yuan, C.-S.; Borchardt, R. T.; Hershfield, M. S.; Smith, G. D.; Howell, P. L. Nature Struct. Biol. 1998, 5, 369. (e) Hasobe, M.; Liang, H.; Ault-Riche, D. B.; Borcherding, D. R.; Wolfe, M. S.; Borchardt, R. T. Antiviral Chem. Chemother. 1993, 4, 245. (f) Wolfe, M. S.; Borchardt, R. T. J. Med. Chem. 1991, 34, 1521. (g) Yeh, J. C.; Borchardt, R. T.; Vedani, A. J. Comput.-Aid. Mol. Des. 1991, 5, 213.

(12) (a) Turner, M. A.; Yang, X.; Yin, D.; Kuczera, K.; Borchardt, R. T.; Howell, P. L. *Cell. Biochem. Biophys.* **2000**, *33*, 101. (b) Hu, Y.; Yang, X.; Yin, D. H.; Mahadevan, J.; Kuczera, K.; Schowen, R. L.; Borchardt, R. T. *Biochemistry* **2001**, *40*, 15143.

(13) (a) González-Moa, M. J.; Terán, C.; Mosquera, R. A. Int. J. Quantum Chem. 2002, 86, 67. (b) Marquez, V. E.; Siddiqui, M. A.; Ezzitouni, A.; Russ, P.; Wang, J.; Wagner, R. W.; Matteucci, M. D. J. Med. Chem. 1996, 39, 3739. (c) Thibaudeau, C.; Plavec, J.; Chattopadhyaya, J. J. Am. Chem. Soc. 1994, 116, 8033. (d) Koole, L. H.; Buck, H. M.; Nyilas, A.; Chattopadhyaya, J. Cham. 1987, 65, 2089. (e) Pearlman, D. A.; Kim, S.-H. J. Biomol. Struct. Dyn. 1985, 3, 99. (f) Cyr, N.; Perlin, A. S. Can. J. Chem. 1979, 57, 2504.

(14) (a) Cloran, F.; Carmichael, I.; Serianni, A. S. J. Am. Chem. Soc.
2001, 123, 4781. (b) Cloran, F.; Zhu, Y.; Osborn, J.; Carmichael, I.; Serianni, A. S. J. Am. Chem. Soc. 2000, 122, 6435. (c) Kennedy, J.; Wu, J.; Drew, K.; Carmichael, I.; Serianni, A. S. J. Am. Chem. Soc. 1997, 119, 8933. (d) Podlasek, C. A.; Stripe, W. A.; Carmichael, I.; Shang, M.; Basu, B.; Serianni, A. S. J. Am. Chem. Soc. 1996, 118, 1413. (e) Kline, P. C.; Serianni, A. S. J. Org. Chem. 1992, 57, 1772. (f) Serianni, A. S.; Barker, R. J. Org. Chem. 1984, 49, 3292.

(15) (a) Hoffmann, R. A.; van Wijk, J.; Leeflang, B. R.; Kamerling, J. P.; Altona, C.; Vliegenthart, J. F. G. J. Am. Chem. Soc. 1992, 114, 3710.
(b) Altona, C.; Sundaralingam, M. J. Am. Chem. Soc. 1972, 94, 8205. (c) Haasnoot, C, A. G.; de Leeuw, F. A. A. M.; de Leeuw, H. P. M.; Altona, C. Org. Magn. Reson. 1981, 15, 43. (d) Houseknecht, J. B.; Altona, C.; Hadad, C. M.; Lowary, T. L. J. Org. Chem. 2002, 67, 4647. (e) Cremer, D.; Pople, J. A. J. Am. Chem. Soc. 1975, 97, 1354. (f) Wu, A.; Cremer, D.; Auer, A. A.; Gauss, J. J. Phys. Chem. A 2002, 106, 657.

(16) Thibaudeau, C.; Kumar, A.; Bekiroglu, S.; Matsuda, A.; Marquez, V. E.; Chattopadhyaya, J. J. Org. Chem. **1998**, 63, 5447.

(17) Examples and references therein: (a) Moon, H. R.; Kim, H. O.;
Chun, M. W.; Jeong, L. S. J. Org. Chem. 1999, 64, 4733. (b) Shin, K. J.;
Moon, H. R.; George, C.; Marquez, V. E. J. Org. Chem. 2000, 65, 2172.
(c) Ezzitouni, A.; Russ, P.; Marquez, V. E. J. Org. Chem. 1997, 62, 4870.
(d) Meldgaard, M.; Wengel, J. J. Chem. Soc. Perkin Trans. 1 2000, 3539.
(e) Moon, H. R.; Ford, H., Jr.; Marquez, V. E. Org. Lett. 2000, 2, 3793.
(18) Kilpatrick, J. E.; Pitzer, K. S.; Spitzer, R. J. Am. Chem. Soc. 1947, 69, 2483.

(19) Examples and references therein: (a) Pelmenschikov, A.; Hovorun, D. M.; Shishkin, O. V.; Leszczynski, J. J. Chem. Phys. 2000, 113, 5986.
(b) Hocquet, A.; Leulliot, N.; Ghomi, M. J. Phys. Chem. B. 2000, 104, 4560. (c) Brameld, K. A.; Goddard, W. A. J. Am. Chem. Soc. 1999, 121, 985. (d) Foloppe, N.; MacKerell, A. D., Jr. J. Phys. Chem. B 1998, 102, 6669. (e) Polak, M.; Mohar, B.; Kobe, J.; Plavec, J. J. Am. Chem. Soc. 1998, 120, 2508. (f) Franchetti, P.; Marchetti, S.; Cappellacci, L.; Jayaram, H. N.; Yalowitz, J. A.; Goldstein, B. M.; Barascut, J.-L.; Dukhan, D.; Imbach, J.-L.; Grifantini, M. J. Med. Chem. 2000, 43, 1264.

(20) (a) Halgren, T. A J. Comput. Chem. 1996, 17, 490. (b) Halgren, T. A. J. Comput. Chem. 1996, 17, 520. (c) Halgren, T. A. J. Comput. Chem. 1996, 17, 553. (d) Halgren, T. A.; Nachbar, R. B. J. Comput. Chem. 1996, 17, 587. (e) Halgren, T. A. J. Comput. Chem. 1996, 17, 616. (f) Halgren, T. A. J. Comput. Chem. 1999, 20, 720. (g) Halgren, T. A. J. Comput. Chem. 1999, 20, 730.

(21) SPARTAN, version 5.1; Wavefunction, Inc.: Irvine, CA.

(22) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, J. A., Jr.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Gonzalez, C.; Challacombe, M.; Gill, P. M. W.; Johnson, B. G.; Chen, W.; Wong, M. W.; Andres, J. L.; Head-Gordon, M.; Replogle, E. S.; Pople, J. A. *Gaussian 98*; Gaussian, Inc.: Pittsburgh, PA, 1998.

(23) It is interesting to note that in a recent study of the pseudorotational flexibility of tetrahydrofuran the B3LYP method was found to be "the most promising quantum mechanical method for modeling biomolecules containing the furanose ring". Strajbl, M.; Florián, J. *Theor. Chem. Acc.* **1998**, *99*, 166.

(24) (a) McCarren, P. R., The Ohio State University. (b) McCarren, P. R.; Gordon, M. T.; Lowary, T. L.; Hadad, C. M. *J. Phys. Chem. A* **2001** *105*, 5911. (c) Houseknecht, J. B.; McCarren, P. R.; Lowary, T. L.; Hadad, C. M. *J. Am. Chem. Soc.* **2001**, *123*, 8811.

(25) van Wijk, J.; Haasnoot, C. A. G.; de Leeuw, F. A. A. M.; Huckriede,
B. D.; Hoekzema, A. W.; Altona, C. *PSEUROT 6.3.*; Leiden Institute of Chemistry, Leiden University: Leiden, The Netherlands, 1999.

(26) de Leeuw, F. A. A. M.; Altona, C. J. Comput. Chem. 1983, 4, 428.

(27) (a) The term "conformational lock" is used to indicate that just one conformation is preferred which may be due to one or several types of interactions. In the present case, the lock, which exists in the gas phase, may be substantially weakened in solution. The term "conformational lock" should not be confused with locks created by structural modification<sup>27b</sup> such as ring locks which should persist in solution. (b) Ravi, R. G.; Kim, H. S.; Servos, J.; Zimmermann, H.; Lee, K.; Maddileti, Boyer, J.; Harden, T. K.; Jacobson, K. A. *J. Med. Chem.* **2002**, *45*, 2090 and references therein.

(28) Brandl, M.; Lindauer, K.; Meyer, M.; Sühnel, J. Theor. Chem. Acc. 1999, 101, 103.

(29) Gu, Y.; Kar, T.; Scheiner, S. J. Mol. Struct. 2000, 552, 17.

(30) Wetmore, S. D.; Schofield, R.; Smith, D. M.; Radom, L. J. Phys. Chem. A 2001, 105, 8718.

(31) Mascal, M. Chem. Commun. 1998, 303.

(32) (a) Hocquet, A. Phys. Chem. Chem. Phys. 2001, 3192. (b) Louit, G.; Hocquet, A.; Ghomi, M. Phys. Chem. Chem. Phys. 2002, 3843.

(33) In a recent theoretical/experimental study<sup>19e</sup> of a derivative of **1** where a pyrazolopyridine moiety replaces the adenine base, the theoretical  $P/v_{\text{max}}$  values were 156.8°/42.2° (south) compared to the experimental fit from  ${}^{3}J_{\text{HH}}$  coupling constants of 137°/37° (south).

(34) A NBO<sup>35</sup> second order perturbation theory analysis of the Fock matrix of **C2** reveals that the interaction energy of the  $\pi$ (C4'-C5') donor orbital with the  $\varrho$ \*(C1'-N9) acceptor orbital is 5.9 kcal/mol at the B3LYP/ 6-31+G(d,p) level.

(35) (a) Reed, A. E.; Weinstock, R. B.; Weinhold, F. J. Chem. Phys. **1985**, 83, 735. (b) Reed, A.; Curtiss, L. A.; Weinhold, F. Chem. Rev. **1988**, 88, 899. (c) Weinhold, F. A. In *Encyclopedia of Computational Chemistry*; Schleyer, P. v. R., Ed.; Wiley Publishers: New York, 1998; Vol. 3, pp 1792–1810.

(36) (a) Miertus, S.; Scrocco, E.; Tomasi, J. Chem. Phys. 1981, 55, 117.
(b) Benzi, C.; Improta, R.; Scalmani, G.; Barone, V. J. Comput. Chem. 2002, 23, 341. (c) Cramer, C.; Truhlar, D. Chem. Rev. 1999, 99, 2161.