

256. *Tautomeric Forms of Streptomycin.*

By DONALD P. YOUNG.

The salts, presumed to be of a tautomeric form of streptomycin, prepared by Heuser, Dolliver, and Stiller¹ are designated β -streptomycin salts. It is now shown that their difference from ordinary or α -streptomycin salts is a labile, chemical one. The β -streptomycin cation can exist in solution for a limited time, and the various β -salts are interconvertible by metathesis; but they revert completely to the α -form when their solutions are kept. This change can be followed polarimetrically. The only known way of forming β -streptomycin is by keeping ("ageing") the 4-ethyl-1-3'-ethylpentyloctyl sulphate (tergitate), or a closely related salt, in the solid state. The $\alpha \rightleftharpoons \beta$ changes in both directions are accelerated by bases, and retarded by acids. β -Streptomycin tergitate reacts with carbonyl reagents much more slowly than the α -form; hence it is thought that in β -streptomycin the aldehyde group is bound by cyclisation, possibly with the methylamino-group. β -Streptomycin salts are characteristically crystalline, but they are probably all solvated in that state.

HEUSER, DOLLIVER, and STILLER¹ prepared a series of streptomycin salts which they thought to be tautomeric with the normal salts. Their supposition now appears to be correct; it is therefore proposed to designate compounds obtained by their method as β -streptomycin salts, using α -streptomycin to denote the "ordinary" form of the antibiotic.

Heuser *et al.* precipitated streptomycin as the tergitate (4-ethyl-1-3'-ethylpentyloctyl sulphate; Tergitol-7 is the sodium salt). This when kept for a few days ("aged") became crystalline. It could then be converted into a sulphate, hydrochloride, etc., which unlike the corresponding α -streptomycin salts were crystalline. The most striking difference was shown by β -streptomycin sulphate, which was very sparingly soluble in water, whereas the α -sulphate is amorphous and extremely soluble. Formation of the β -sulphate in this way has been used commercially as a means of purifying the antibiotic. Heuser *et al.*

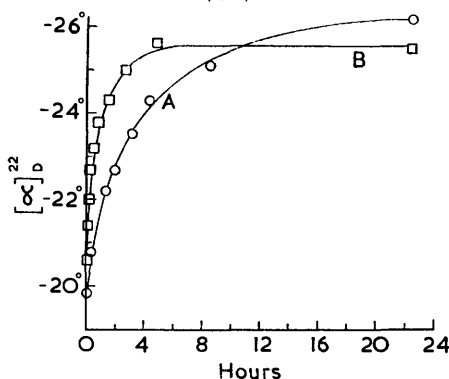
¹ Heuser, Dolliver, and Stiller, *J. Amer. Chem. Soc.*, 1953, **75**, 4013.

showed that their β -streptomycin salts further differed from the α -salts in distribution coefficient and the shape of the polarogram. By making use of these they demonstrated, in a roughly quantitative manner, that β -streptomycin salts reverted to the α -form in solution, and that this change was retarded by the presence of acids.

The clearest evidence² that the phenomenon was not merely one of allotropy or solvation is that solutions of β -streptomycin hydrochloride gave a precipitate of the β -sulphate on addition of sulphate ion, so that the β -streptomycin cation preserved its identity in solution.

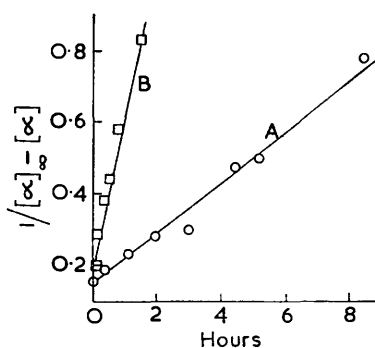
The observations of Heuser *et al.* have now been confirmed and extended. β -Streptomycin hydrochloride was converted, by precipitation of its freshly made aqueous solution with the appropriate sodium salt, into the biphenyl-4-sulphonate, 5-cyclohexyl-2-hydroxy-3-methylbenzoate, and 2-hydroxy-5-isobutyl-3-methylbenzoate. In the case of the first two of these, the α -streptomycin salts are also crystalline,^{3,4} but the pairs of salts had

FIG. 1. Mutarotation of β -streptomycin tergitate (c, 5).



(A) In neutral methanol. (B) In 0.01N-methanolic sodium hydroxide.

FIG. 2. Second-order plot for mutarotation of β -streptomycin tergitate.



different X-ray diffraction patterns. These three new salts were then converted into β -streptomycin sulphate.

The reversion of β - to α -streptomycin salts in solution can be followed polarimetrically, most easily with the tergitate in methanol (50 mg./ml.), which showed a change of $[\alpha]_D^{20}$ from -19.8° to -26.4° . As ordinarily prepared, the β -tergitate reverted in this way to the α -form in a few days. In 0.01N-methanolic sodium hydroxide, the mutarotation was more rapid (Fig. 1); but in 0.01N-methanolic sulphuric acid there was no measurable alteration in $[\alpha]_D$ during 18 hr., and in 0.1N-methanolic acetic acid a change of only 0.2° in 24 hr., corresponding to *ca.* 3% conversion into the α -form. Further, if the β -tergitate was recrystallised from methanol-water containing a trace of sulphuric acid, it became stable in methanol solution, and its stability was not destroyed by subsequent recrystallisation from unacidified methanol. The "stabilised" tergitate did nevertheless revert rapidly to the α -form if a trace of sodium hydroxide was added to its solution. The change, surprisingly, was of the second order in all cases.

β -Streptomycin 5-cyclohexyl-2-hydroxy-3-methylbenzoate showed a similar, very rapid change in rotation in methanol solution as it reverted to the α -form. For the hydrochloride in aqueous solution the change was too small and too fast for accurate observation: $[\alpha]_D^{22}$ was -82.3° ten minutes after preparation (50 mg./ml.), falling to a final value of -79.1° (24 hr.) (at least three-quarters complete in 50 min.). Gravimetric estimation of the β -tautomer by precipitation as sulphate led to the same conclusion. The change

² Heuser, U.S.P. 2,663,464.

³ Ziegler, U.S.P. 2,857,376.

⁴ Ziegler, U.S.P. 2,857,375.

was appreciably retarded by 0.001*N*-concentration of hydrochloric acid, and in 0.01*N*-acid it occupied many hours; here again the reaction was approximately of the second-order (Fig. 3); the rate, however, varied with different batches of β -salt. In 5% aqueous toluene-*p*-sulphonic acid, the β -hydrochloride initially showed $[\alpha]_D^{18} -70.8^\circ$ changing, in the opposite sense, to -78.5° (final: 24 hr.).

Probably all crystalline β -streptomycin salts are solvated. The sulphate lost the equivalent of 7H₂O when dried; the dried salt dissolved in water, but the crystalline hydrate was soon precipitated. X-Ray examination of the dried material revealed a partly orientated structure. The hydrochloride crystallised from methanol with 2 mol. of methanol, which was readily driven off, leaving an amorphous substance. Recrystallised β -streptomycin tergitate is hydrated.¹

The "ageing" of streptomycin tergitate involves a spontaneous change from the α - to the β -form, accompanied by crystallisation. Precipitated α -streptomycin tergitate is soap-like and difficult to purify, but samples were conveniently obtained by keeping a methanolic solution of the β -tergitate for a few days and then removing the solvent. This left the α -tergitate as a glass, stable when dry. Only in presence of water did it change into the β -form. Like the $\beta \rightarrow \alpha$ change, this was strongly accelerated by a trace of alkali, and inhibited by aqueous acid.

So far, no other streptomycin salt except the related 4-ethyl-1-isobutyloctyl sulphate² (from Tergitol-4) has been found which changes from the α - into the β -form. In all streptomycin salts examined, there was no detectable amount of β -form in equilibrium with the α -tautomer in solution; solutions of α -streptomycin showed no measurable mutarotation, and it has proved impossible to obtain the β -sulphate from them by adding sulphate ion. It is therefore clear that a change towards the β -form could not occur in a solid phase in contact with a solvent containing appreciable amounts of dissolved streptomycin. Although the presence of water is necessary for the formation of β -streptomycin tergitate—presumably because the driving force is crystallisation of the hydrate—the tergitate is very insoluble in water; this combination of properties may be almost unique amongst streptomycin salts. Heuser *et al.*¹ thought that some α -tergitate was present at equilibrium, which may indeed arise from back-reaction in the aqueous phase.

It has been established that the β -streptomycin cation has an independent existence, and that its salts have the same composition as the corresponding α -streptomycin ones. The equilibrium between the two is sufficiently mobile to justify the term tautomerism, although, in the absence of molecular-weight determinations, reversible dimerisation cannot be excluded.

Mannosidostreptomycin shows the same phenomenon,² but dihydrostreptomycin and streptomycin oxime do not, which implicates the aldehyde group as the seat of the tautomerism. This was confirmed when it was found that β -streptomycin tergitate, in a metastable condition in methanolic 0.1*N*-acetic acid, reacted much more slowly than the α -salt with hydroxylamine and 4-phenylsemicarbazide (reactions followed polarimetrically) (cf. Fig. 4). Evidently, in β -streptomycin the aldehyde group is combined in such a way that it has become unreactive.

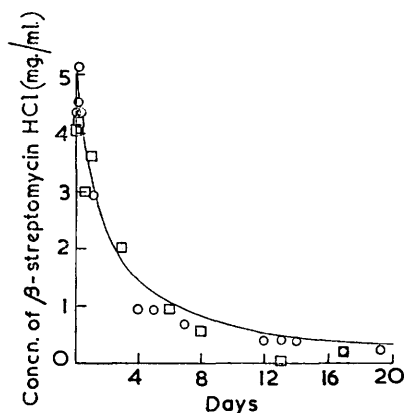
Heuser *et al.*¹ interpreted their polarographic results (which we have confirmed) in the opposite sense, namely, that it was the α -tautomer that had a bound (unreactive) aldehyde group. The extra polarographic wave on the more positive side with the α -form could, however, equally be interpreted as arising from an easily reducible group, *i.e.*, a free aldehyde group, in that tautomer.

The pronounced effect of bases on the $\alpha \rightleftharpoons \beta$ changes is manifested at the neutralisation point of the third basicity of the streptomycin cation. This suggests that the weakest basic group, *i.e.*, the NHMe-group, may also be concerned in the tautomerism. Possibly it combines with the aldehyde group to form a $\cdot\text{CH}(\text{OH})\cdot\text{NMe}\cdot$ structure; this would result in a 7-membered ring, which, from atomic models, is sterically possible (dimerisation of the type common amongst α -ketols is sterically unlikely). The fact that the $\beta \rightarrow \alpha$

reaction, resulting in the freeing of a bound aldehyde group, is of the second order is puzzling, but it need not indicate that the reaction is bimolecular. There is no appreciable difference in base-strength between α - and β -streptomycin.

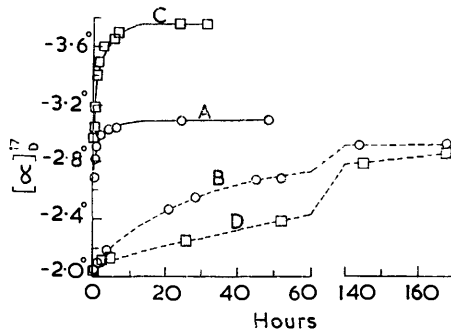
Other authors⁵ have obtained evidence, from polarographic and distribution experiments, of tautomerism of streptomycin in solution. Their results might at first sight be

FIG. 3. Change of β - into α -streptomycin hydrochloride in 0.01N-hydrochloric acid at 17°.



○ Polarimetric observations. □ Gravimetric observations. — Calculated second-order curve, $k = 0.0125 \text{ g. ml.}^{-1} \text{ day}^{-1}$.

FIG. 4. Reaction of streptomycin tergitates (0.6 mmole) with carbonyl reagents in 0.1N-acetic acid in methanol (20 ml.).



(A, C) α -, (B, D) β -Tergitate. (A, B) Hydroxylamine acetate (0.7 mmole); (C, D) 4-phenylsemicarbazide (0.6 mmole).

explained by an equilibrium between α - and β -streptomycin. However, it now appears that this equilibrium would be entirely on the side of the α -form; thus there may be more than two desmotropic forms of streptomycin.

EXPERIMENTAL

Samples for analysis and bioassay were dried at 60°/0.1 mm. for 8 hr.; this did not always completely remove water. Nitrogen analyses, whether by the Dumas or the Kjeldahl method, often gave low results for streptomycin derivatives. Bioassays were by *Bacillus subtilis* NTCC 8236. Rotations were measured with 2-dm. tubes.

β -Streptomycin tergitate, m. p. 155–158°, was prepared by precipitation, "ageing," and recrystallisation, as described by Heuser *et al.*¹ Under the polarising microscope it appeared as small flat birefringent needles.

Change of β - into α Streptomycin Tergitate.—A solution of β -tergitate in methanol was kept until the rotation ceased to rise. The apparent pH of the solution remained unchanged (within 0.1 unit). Evaporation under a vacuum left the α -tergitate as a glass, soluble in carbon tetrachloride and acetone in which the β -tergitate is insoluble; such solutions did not crystallise, even with added water. With aqueous guanidine sulphate and butanol it gave no crystalline streptomycin sulphate.

Evaporation of a solution of β -tergitate in 0.01N-methanolic sulphuric acid after 18 hr. left unchanged β -tergitate, which crystallised. This was convertible into β -streptomycin sulphate.

Conversion of α - into β -Streptomycin Tergitate.—(a) *Under neutral conditions.* α -Streptomycin tergitate, obtained as above, remained unchanged for months in a closed vessel. When covered with distilled water, the glass softened to a jelly, then after 3 days became friable and opaque. After 7 days it was dried, and then consisted of birefringent crystals, m. p. 149°. In methanol $[\alpha]_D$ changed from -19.9° to -24.1° . By evaporation of this solution and addition of water, the cycle could be repeated.

(b) *In presence of alkali.* α -Streptomycin tergitate was covered with 0.01N-aqueous sodium

⁵ Bricker and Vail, *J. Amer. Chem. Soc.*, 1951, **73**, 585; Titus and Fried, *J. Biol. Chem.*, 1948, **174**, 57.

hydroxide. In $\frac{1}{2}$ hr. it had peptised to a thick colloidal suspension, but after 1 hr. this coagulated to waxy masses, leaving a mobile supernatant liquid. The solid was crystalline, with m. p. $\sim 155^\circ$.

(c) *Under acid conditions.* The α -tergitate, after 10 days under 0.01N-hydrochloric acid, was unchanged.

"Stabilised" β -Streptomycin Tergitate.—Crude β -tergitate (15 g.) was dissolved in methanol (100 ml.) containing sulphuric acid (0.02 ml.) at $30-40^\circ$. The solution was cooled and water (10 ml.) added. After some hours at 0° the crystals (11.2 g.) were collected and recrystallised from methanol-water. They then had m. p. 145° , $[\alpha]_D^{28} -21.6^\circ$ in methanol, rising to -22.6° after 12 days.

Reaction of Streptomycin Tergitates with Hydroxylamine.—To streptomycin tergitate (1 g.) in 0.1N-methanolic acetic acid (20 ml.) was added a solution (1 ml.) of hydroxylamine acetate [from hydroxylamine hydrochloride (1.02 g.) and sodium acetate (1.25 g.) in methanol, filtered, and made up to 25 ml.].

When the reaction with the α -tergitate was complete (48 hr.; Fig. 4), 0.9N-aqueous methylamine sulphate (2.2 ml.) was added, and the precipitated *oxime sulphate* (0.38 g.) was collected (Found: C, 32.9; H, 6.4; N, 14.2; S, 6.8. $C_{21}H_{40}N_8O_{12} \cdot 1\frac{1}{2}H_2SO_4 \cdot 3H_2O$ requires C, 32.9; H, 6.2; N, 14.1; S, 6.1%). This was biologically almost inactive (potency found, 28 u./mg.).

The reaction with β -tergitate was not complete after 7 days. The product at that point was converted into sulphate, which from its assay (80 u./mg.) was oxime containing an appreciable amount of unchanged streptomycin.

Streptomycin Oxime Tergitate.—To a solution of α -streptomycin sulphate (7.3 g.) and hydroxylamine hydrochloride (1.04 g.) in water (50 ml.), sodium hydroxide was added as necessary to keep the pH at 6. After 3 hr., a slight excess of Tergitol-7 was added. The pasty precipitate was centrifuged off and washed with water. It did not crystallise and was dried to a glass (Found: C, 53.8; H, 9.1; N, 6.4; S, 6.0. $C_{21}H_{40}N_8O_{12} \cdot 3C_{17}H_{26}O_4S \cdot 2H_2O$ requires C, 53.1; H, 8.8; N, 6.9; S, 5.9%). With methylamine sulphate in aqueous methanol, it gave the oxime sulphate (potency 48 u./mg.).

Reaction of Streptomycin Tergitates with 4-Phenylsemicarbazide.—Streptomycin tergitate (1.0 g.) and 4-phenylsemicarbazide (0.10 g.) were dissolved in 0.1N-methanolic acetic acid (total volume, 20 ml.).

The reaction with α -tergitate was complete in 24 hr. (Fig. 4). 0.9N-Aqueous methylamine sulphate (2.2 ml.) was added to this solution to precipitate *streptomycin 4-phenylsemicarbazone sulphate*, which was washed with methanol and dried. It was biologically inactive (Found: potency, 20 u./mg.; C, 36.7; H, 6.2; N, 14.0; S, 5.3. $C_{28}H_{46}N_{10}O_{12} \cdot 1\frac{1}{2}H_2SO_4 \cdot 3H_2O$ requires C, 36.7; H, 6.1; N, 15.3; S, 5.3%). The reaction with β -tergitate was still incomplete (cf. Fig. 4) after 4 weeks, when the $[\alpha]_D^{17}$ had reached -35.4° .

β -Streptomycin Hydrochloride.—The hydrochloride, prepared in methanol¹ and air-dried, formed birefringent rhombic prisms. On drying at $60^\circ/0.1$ mm. for 8 hr., it lost 9% in weight (corresponding to 2MeOH) and became amorphous, although still apparently retaining water (Found: C, 34.8, 35.2; H, 6.7, 6.4; Cl, 15.0, 15.2; N, 13.4, 13.8%; potency, 861 u./mg. Calc. for $C_{21}H_{39}N_7O_{12} \cdot 3HCl \cdot H_2O$: C, 34.6; H, 6.2; Cl, 15.0; N, 13.8%; potency, 820 u./mg.). The methanol-free material was soluble in methanol; a 5% solution soon deposited crystals of the methanol solvate, which was almost insoluble. The crystalline compound gave a diffuse X-ray pattern, but it became amorphous during the exposure.

Conversion of β - into α -Streptomycin Hydrochloride (Fig. 3).—Solutions of the β -hydrochloride (50 mg./ml.) were observed polarimetrically, and the amount of β -streptomycin was estimated at intervals gravimetrically: an aliquot part was treated with an equal volume of 0.5N-guanidine sulphate (acidified to pH 3) and kept at 0° for 30 min. The precipitated β -sulphate was filtered off (filter-stick), washed with 0.1N-sulphuric acid, ethanol, and acetone, briefly dried *in vacuo*, and weighed.

β -Streptomycin Tergitate from β -Hydrochloride.—A fresh solution of the β -hydrochloride in water with a trace of hydrochloric acid was treated with Tergitol-7. The tergitate was precipitated immediately in crystalline condition (m. p. 148–149°).

β -Streptomycin Biphenyl-4-sulphonate.—A hot solution of sodium biphenyl-4-sulphonate (1.90 g.) in water (50 ml.) was added to a freshly prepared one of the β -hydrochloride (1.60 g.) in water (50 ml.). The β -streptomycin salt was precipitated immediately (yield 68%); it had m. p. 187–189° (decomp.) (Found: C, 51.4; H, 5.7; N, 7.5; S, 7.5. $C_{21}H_{39}N_7O_{12} \cdot 3C_{12}H_{10}O_3S \cdot 3H_2O$ requires

C, 51.1; H, 5.6; N, 7.3; S, 7.2%). With the equivalent amount of guanidine sulphate in aqueous methanol, it gave β -streptomycin sulphate (92%), which was crystallised from water; its identity was checked by its infrared spectrum.

In a similar manner were prepared β -streptomycin 5-cyclohexyl-2-hydroxy-3-methylbenzoate (85%), m. p. 170—175° (decomp.; rapid heating), $[\alpha]_D^{19}$ -22.0° (5 min.), -26.7° (190 min.), -27.0° (27½ hr.) in MeOH (*c* 5) (Found: C, 57.9, 57.6; H, 7.7, 7.6; N, 7.2, 7.2. $C_{21}H_{39}N_7O_{12}, 3C_{14}H_{18}O_3$ requires C, 57.7; H, 7.3; N, 7.6%), and the 2-hydroxy-5-isobutyl-3-methylbenzoate (83%), m. p. 90—95° (Found: C, 55.2; H, 7.4; N, 7.5%; potency, 437 u./mg. $C_{21}H_{39}N_7O_{12}, 3C_{12}H_{18}O_3, 2H_2O$ requires C, 55.1; H, 7.4; N, 7.9%; potency, 469 u./mg.). Both these were converted in good yield into β -streptomycin sulphate.

From α -streptomycin sulphate were obtained α -streptomycin biphenyl-4-sulphonate³ (85%), m. p. 187.5—189° (charred) (Found: C, 50.2; H, 5.5; N, 6.6; S, 7.4%; potency, 466, 453 u./mg. Calc. for $C_{21}H_{39}N_7O_{12}, 3C_{12}H_{10}O_3, 4H_2O$: C, 50.4; H, 5.8; N, 7.2; S, 7.1%; potency, 432 u./mg.), and 5-cyclohexyl-2-hydroxy-3-methylbenzoate⁴ (99%), m. p. 160—165° (charred), 93° (decomp.; in preheated bath), $[\alpha]_D^{21}$ -27.4° in MeOH (*c* 5) (Found: C, 55.6; H, 7.7; N, 6.3%; potency, 448, 510 u./mg. Calc. for $C_{21}H_{39}N_7O_{12}, 3C_{14}H_{18}O_3, 2H_2O$: C, 56.1; H, 7.4; N, 7.4%; potency, 441 u./mg.). The X-ray patterns of these two were similar to, but distinguishable from, those of the corresponding β -salts. α -Streptomycin hydroxyisobutylmethylbenzoate formed a gum.

β -Streptomycin Sulphate from Hydrochloride.— β -Streptomycin hydrochloride (5.0 g.) was dissolved in water (100 ml.), and 0.9N-methylamine sulphate (25 ml.) immediately added. The β -sulphate that crystallised was filtered off, washed with a little water and then with ethanol, and air-dried (2.9 g.). When dried at 60°/0.1 mm. for 8 hr., this product lost 15% in weight (corresponding to 7H₂O), and was then anhydrous (Found: C, 34.8; H, 6.1; N, 12.7; S, 6.2%; potency, 817 u./mg. Calc. for $C_{21}H_{39}N_7O_{12}, 1\frac{1}{2}H_2SO_4$: C, 34.6; H, 5.8; N, 13.5; S, 6.6%; potency, 798 u./mg.). It then dissolved in water, but the solution rapidly afforded crystals of the hydrate. This recrystallised from 0.01N-sulphuric acid (dissolution at 40°) at 0°. The hydrate gave a sharp X-ray pattern; the dried β -sulphate gave diffuse rings, suggesting that part of the crystalline orientation remained. The infrared spectrum (KBr disc) was indistinguishable from that of α -streptomycin sulphate, in which no C=O band is detectable.

Thanks are expressed to Dr. C. C. F. Blake for X-ray powder diagrams, to Mr. A. R. Philpotts for infrared spectra, to Mr. R. J. Thompson for polarography, and to Dr. R. H. Hall for his interest.

THE DISTILLERS COMPANY LIMITED, RESEARCH AND DEVELOPMENT DEPARTMENT,
GREAT BURGH, EPSOM, SURREY.

[Received, September 1st, 1960.]