

Conformational Analysis of 3,4-Dimethyltetrahydro-1,3,4-oxadiazine

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Contrary to a recent report, the diequatorial conformation makes no appreciable contribution to the title compound. Evidence is presented that the preferred conformation is 3-axial-4-equatorial. The activation parameters or the conformational process in this compound are: ΔH^\ddagger 13.25 \pm 0.14 kcal mol⁻¹, ΔS^\ddagger +2.8 \pm 0.6 cal mol⁻¹ K⁻¹, possibly owing to slowing of a nitrogen inversion process.

KATRITZKY and his co-workers (hereafter referred to as KCO) have recently reported on the conformational analysis of some tetrahydro-1,3,4-oxadiazines.¹ For the NN'-dimethyl derivative (1) they give a free energy of activation of 12.6 kcal mol⁻¹ at the coalescence temperature (ca. 256 K) for a process which they describe as slow 'non-passing' ring inversion. They suggest, on the basis of comparison of N-methyl conformational free energy differences in related compounds, that the major conformation is (1ae), and they assign a minor doublet in the low-temperature, slow-inversion-limit spectrum, to the (1ee) conformation. Although this doublet would, on their interpretation, arise from ca. 20% of the minor conformation, no other lines attributable to this conformation were observed.

RESULTS

A full lineshape analysis of the ¹H n.m.r. spectrum of the coalescing AB quartet of the C-2 protons in compound (1) over a range of temperatures gives the activation parameters ΔH^\ddagger 13.25 \pm 0.14 kcal mol⁻¹, ΔS^\ddagger +2.8 \pm 0.6 cal mol⁻¹ K⁻¹ (Table). These agree well with the approximate free energy of activation measured by KCO (12.6 kcal

Rate data* for compound (1)

Temp. (°C)	k_{obs} from fitting	k_{calc} from activation parameters
+5	1 000	934
-5	340	368
-16	125	122
-24	54	51
-32	20	20.4
-40	8.0	7.6
-45	4.0	3.98

* These data lead to ΔH^\ddagger 13.25 \pm 0.14 kcal mol⁻¹ and ΔS^\ddagger +2.8 \pm 0.6 cal mol⁻¹ K⁻¹. For a ca. 10% solution in CDCl₃

mol⁻¹), attributed to slowing of 'non-passing' ring inversion.¹ However, as shown below, slow nitrogen inversion cannot be ruled out as the origin of these observations, and may indeed be responsible for the observed changes.

We agree with the assumption of KCO that the preferred conformation is (1ae). Experimental proof may be found in the C-2 proton geminal coupling constant at slow exchange, found to be 10.1 Hz at ca. -80 °C. This coupling is much closer to that found in the model compound (2) with an axial N-alkyl group (*J* 10.5 Hz) than in the model compound (3) with the N-methyl group constrained to be equatorial (*J* 7.5 Hz).² In addition, the chemical shift of the C-2 proton in the 2-*p*-nitrophenyl derivative (4) (δ 5.70) is much closer to that of the model (5) (δ 5.62) than to that of (6) (δ 4.21).

The minor doublet in the low-temperature spectra of compound (1) does not however arise from the diequatorial conformation as suggested by KCO. It comes from the equatorial C-5 proton. The triplet observed at ambient temperature for the C-5 protons at δ ca. 2.47 broadens as the temperature is lowered, and reappears at δ ca. 2.91 and 2.11 (Figure 1). Decoupling of the resonances due to the

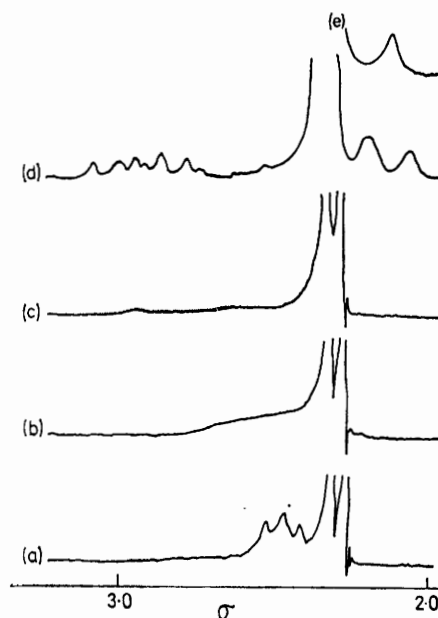


FIGURE 1 90 MHz ¹H N.M.R. spectra of 3,4-dimethyltetrahydro-1,3,4-oxadiazine: (a) 33.5°C; (b) 0°C; (c) -20°C; (d) -65°C; (e) -65°C with decoupling at δ ca. 2.91

C-6 protons or the axial C-5 proton produces the expected changes in the equatorial C-5 proton signal [Figure 1(e)]. There is no broadening and resharping of the methyl peaks as the temperature is lowered, as would occur were the explanation of KCO correct. Moreover, the value of the supposed equilibrium constant can be calculated to be 0.17, in reasonable agreement with that reported (0.22). Finally, the non-appearance of any other lines due to a diequatorial conformation in such a simple spectrum must argue against its presence. An upper limit of 5% can be placed on the proportion of the diequatorial conformation on the basis of our estimate of detectability, giving a free energy difference at 250 K of >1.5 kcal mol⁻¹.

DISCUSSION

Our purpose in preparing and examining this compound was two-fold: to determine its preferred conform-

¹ I. J. Ferguson, A. R. Katritzky, and D. M. Read, *J.C.S. Perkin II*, 1976, 1861.

² F. G. Riddell and J. M. Lehn, *J. Chem. Soc. (B)*, 1968, 1224.

oxadiazines, we must add $2.3 - 0.4 = 1.9$ kcal mol⁻¹ to each barrier.* Thus, probable barriers in the oxadiazine series become 1, 13.4; 2, 12.1; 3, 12.7; and 4, 14.6 kcal mol⁻¹. The order in which they would be expected to 'freeze out' is therefore 4, 1, 3, 2. Thus, possibility 1, 3, 4 for the observed process mentioned above becomes a likely contender, and the activation energy of the most rapid process and the last to be frozen out, 3, is 12.7 kcal mol⁻¹, in good agreement with the experimental results.

EXPERIMENTAL

Preparation of 3,4-Dimethyltetrahydro-1,3,4-oxadiazine (1).—1,2-Dimethylhydrazine (1.42 g) was added to a solution of ethylene oxide (1.1 g) in ethanol (10 ml) at 0 °C. The solution was allowed to warm to room temperature overnight. Removal of volatile material yielded crude 1-(2-hydroxyethyl)-1,2-dimethylhydrazine (1.98 g, 81%) as an oil, which was used without further purification. This product (0.67 g) and paraformaldehyde (0.19 g) were heated under reflux in benzene (10 ml) for 2.5 h. Distillation yielded 3,4-dimethyltetrahydro-1,3,4-oxadiazine as a liquid, b.p. 119° (0.4 g, 55%) (Found: *m/e* 116.0953. Calc. for C₅H₁₂N₂O: *M*, 116.0950).

Preparation of 3,4-Dimethyl-2-p-nitrophenyl-1,3,4-oxadiazine (4).—*p*-Nitrobenzaldehyde (0.36 g) and the crude hydroxyethyl dimethylhydrazine (0.25 g) were heated under

reflux in benzene (10 ml) for 2 h. Evaporation yielded a solid which was recrystallised twice from light petroleum (b.p. 60–80°); yield 0.18 g (32%), m.p. 94–97° (Found: *m/e* 237.1091. Calc. for C₁₁H₁₅N₃O₃: *M*, 237.1114).

N.m.r. spectra were recorded with a Perkin-Elmer R32 spectrometer operating at 90 MHz fitted with the standard variable-temperature equipment and frequency counter. Calibration of the temperature control unit over its entire range revealed all temperatures to be within ± 1.5 °C of the control settings and reproducible to within ± 0.5 °C. The temperature in the sample was uniform to within ± 0.5 °C over the bottom centimetre of the tube. The effective chemical shifts and coupling constants were obtained by extrapolation of values obtained down to -90 °C (supercooled solution) and were calibrated with the frequency counter. The effective transverse relaxation times were obtained from measurements of linewidth at half height ($W_{\frac{1}{2}}$) in the regions of slow (-50 to -90 °C) and rapid (+20 to +40 °C) exchange, by assuming a linear variation of $W_{\frac{1}{2}}$ with temperature.

Computations were performed on the University Elliott 4130 computer using the program DNMRS. Errors on the enthalpy and entropy of activation are 95% confidence limits from a least-squares fit of the Eyring rate equation.

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* This only alters the magnitudes of the barriers, not their relative values.