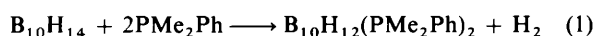


Identification of the *endo,exo* Isomer of 6,9-(PMe₂Ph)₂-*arachno*-B₁₀H₁₂ by Nuclear Magnetic Resonance Spectroscopy

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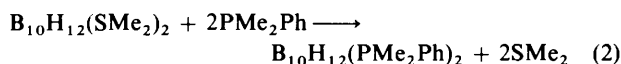
Reaction of PMe₂Ph with *nido*-B₁₀H₁₄ results in the formation of the previously unsuspected *endo,exo* isomer of *arachno*-B₁₀H₁₂(PMe₂Ph)₂ (ca. 85%) admixed with the expected *exo,exo* isomer (ca. 15%). Mild heating converts the *endo,exo* isomer into the *exo,exo* one. The mixture is readily analysed by multiple resonance and difference n.m.r. spectroscopy.

The *arachno* bis(ligand) decaborane compounds B₁₀H₁₂L₂, where L = NCR, SR₂, PR₃, etc., are generally believed to have the 6,9-*exo,exo*-bis(ligand) configuration (I),^{1,2} as established by single-crystal X-ray diffraction analysis for the bis(dimethyl sulphide) and bis(acetonitrile) compounds B₁₀H₁₂(SMe₂)₂ and B₁₀H₁₂(NCMe)₂.^{3,4} It has recently been shown that the related *arachno* anion [B₁₀H₁₂(PPh₂)₂]⁻ has a bridged structure [schematic configuration (II)]^{5,6} which effectively has the bidentate ligand PPh₂⁻ bound 6,9-*endo,endo* to the open decaboranyl cluster. This anion has long been recognized to be a species of particular stability,⁷⁻⁹ and although contributions to this stability will arise from the (Lewis-acid) chelating ability of the neutral B₁₀H₁₂ fragment of ten-vertex *arachno* geometry,⁶ it suggests that *endo,endo* and *endo,exo* isomers of B₁₀H₁₂L₂ [schematic configurations (III) and (IV) respectively] may also be stable providing that any steric interactions above the open face are not too severe. In this context we now report that the reaction of PMe₂Ph with B₁₀H₁₄, according to equation (1), in CD₂Cl₂ solution at <0 °C does in fact result in the initial formation of *endo,exo*-6,9-(PMe₂Ph)₂-*arachno*-B₁₀H₁₂ (ca. 85%) admixed with much smaller quantities (ca. 15%) of the symmetrical *exo,exo*-6,9 isomer.

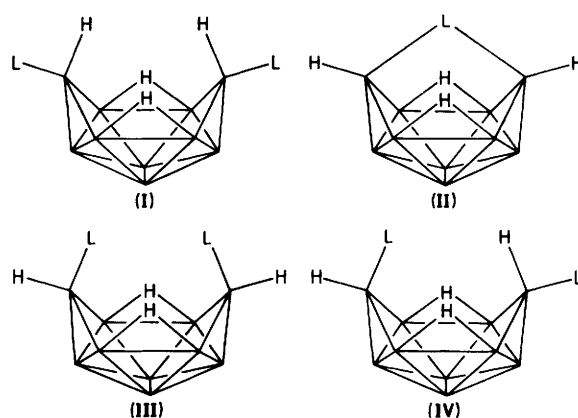


Results and Discussion

Addition of PMe₂Ph to a solution of *nido*-B₁₀H₁₄ in CD₂Cl₂ solution at ca. 200 K, followed by a slow warming to room temperature, resulted in the evolution of a colourless gas, presumably dihydrogen. The ¹¹B n.m.r. spectrum of the colourless solution [e.g. Figure 1(b)] showed that it contained two boron-containing compounds. Integrated n.m.r. spectroscopy [together with the subsequent identification (see below)] showed that these compounds were present in a molar ratio of 84:16, with only trace quantities of other boron-containing species being present. Interestingly, a similar product ratio was obtained when a sample of *exo,exo*-6,9-(SMe₂)₂B₁₀H₁₂ was treated with PMe₂Ph in CD₂Cl₂ solution [equation (2)]. In this case, however, the reaction occurred more slowly, being only ca. 50% complete after 24 h at ca. 293 K.



Inspection of the ¹¹B n.m.r. spectrum of the product suggested that the minor component [compound (I)] was the expected *exo,exo*-6,9 isomer of B₁₀H₁₂(PMe₂Ph)₂ [configur-



ation (I)]. This conclusion was also consistent with the results of selective ¹H-¹¹B} n.m.r. spectroscopy which related the observed ¹H resonances to their directly bound boron positions as summarized in the Table. Subtraction of the ¹¹B spectrum of a pure sample of compound (1) from that of the mixture (Figure 1) then more clearly revealed the ¹¹B spectrum of the major component [compound (2)]. These data for compound (2) are also summarised in the Table, together with the corresponding ¹H n.m.r. data which were again established by ¹H-¹¹B} spectroscopy. The Table also presents data for the *endo,endo* anionic species [B₁₀H₁₂(PPh₂)₂]⁻ (3), configuration (II), for comparison.

The similarity of the ¹¹B and ¹H nuclear shielding patterns (Figure 2) suggests that the borane clusters of compounds (1)–(3) have closely related structures. In particular, the intermediacy of the n.m.r. properties of the new compound (2) compared to those of compounds (1) and (3) indicates that the detailed character of its structure is also intermediate. The 1:1:2:2:1:1:2 ¹¹B intensity pattern shows that compound (2) has effective mirror-plane symmetry, rather than the C_{2v} symmetry of the other two species, and the retention of ³¹P coupling to ¹¹B(6) and ¹¹B(9) [these resonances being distinguished from those of ¹¹B(1,3) by the relative sharpness¹⁰ of the latter] shows that it still retains PMe₂Ph ligands directly bound to B(6) and B(9). These considerations suggest the *endo,exo*-6,9 configuration (IV) for the phosphine ligands in compound (2).

This structure is confirmed by the comparative ¹H(6) and ¹H(9) shielding behaviour among the three species. For the *exo,exo* compound (1) the *endo*-6,9 proton resonance is some 2 p.p.m. above the approximately linear (¹¹B, ¹H) shielding-correlation plot for boron atoms and their directly bound *exo*-

Table. Measured n.m.r. parameters for *exo,exo*-6,9-(PMe₂Ph)₂B₁₀H₁₂ (1), *endo,exo*-6,9-(PMe₂Ph)₂B₁₀H₁₂ (2), and [*endo,endo*-μ(6,9)-(PPh₂)₂B₁₀H₁₂]⁻ (3) at +21 °C

Assignment	Compound (2) ^a			Compound (1) ^a			Compound (3) ^b	
	δ(¹¹ B)/p.p.m.	¹ J(¹¹ B- ¹ H)/Hz	δ(¹ H) ^d /p.p.m.	δ(¹¹ B)/p.p.m.	¹ J(¹¹ B- ¹ H)/Hz	δ(¹ H) ^d /p.p.m.	δ(¹¹ B)/p.p.m.	δ(¹ H) ^d /p.p.m.
2,4	{ +7.4 (1 B) +1.2 (1 B)	143 134	{ +3.09 (1 H) ^e +2.37 (1 H) ^b	-3.0 (2 B)	131	+2.24 (2 H) ^f	+6.4 (2 B)	+2.50 (2 H) ^g
5,7,8,10	{ -14.4 (2 B) -18.4 (2 B)	138 140	{ +1.76 (2 H) +1.65 (2 H)	-19.0 (4 B)	133	{ +1.65 (4 H), -4.16 (2 H) ^j	-7.9 (4 B)	{ +1.95 (4 H), -4.19 (2 H) ^k
6	-29.3 (1 B) ⁱ	114	+1.01 (1 H)(<i>exo</i>) ^l	-35.2 (2 B) ^m	114	{ -0.94 (2 H) (<i>endo</i>) ⁱ	-26.8 (2 B) ⁿ	{ +0.77 (2 H) (<i>exo</i>)
9	ca. -34.5 (1 B) ^o	<i>o</i>	-1.41 (1 H)(<i>endo</i>) ^j					
1,3	ca. -34.5 (2 B)	145	+0.85 (2 H)	-38.8 (2 B)	141	+0.59 (2 H)	-26.9 (2 B)	+0.98 (2 H)

^a In CDCl₃ solution at +21 °C, this work. ^b In (CD₃)₂CO solution, data from ref. 5. ^c ± 8 Hz; measured from ¹¹B spectrum with line narrowing to achieve baseline separation of doublet components. ^d δ(¹H) related to directly bound B atoms by ¹H-¹¹B(selective) spectroscopy; fine structure in ¹H spectra (footnotes e—i and k) is observed under conditions of complete ¹¹B(broad-band noise) decoupling. ^e Doublet, ³J(³¹P-¹H) ca. 25 Hz. ^f Doublet, ³J(³¹P-¹H) ca. 24 Hz. ^g Doublet, ³J(³¹P-¹H) ca. 18 Hz. ^h Doublet, ³J(³¹P-¹H) ca. 28 Hz. ⁱ Possible doublet, ³J(³¹P-¹H) ca. 7 Hz. ^j No doublet structure observed, i.e. ³J(³¹P-¹H) < ca. 5 Hz. ^k Doublet, ³J(³¹P-¹H) ca. 52 Hz. ^l ¹J(³¹P-¹¹B)(*endo*) ca. 120 Hz. ^m ¹J(³¹P-¹¹B)(*exo*) ca. 105 Hz. ⁿ ¹J(³¹P-¹¹B)(*endo*) 97 Hz. ^o ¹J(³¹P-¹¹B)(*exo*) and ¹J(¹¹B-¹H)(*endo*) both very approximately 100 Hz; not measurable more accurately due to overlap with ¹¹B(1,3) in the same compound and ¹¹B(6,9) of the *exo,exo* isomer.



Figure 1. 128-MHz ¹¹B N.m.r. spectra of isomers of B₁₀H₁₂(PMe₂Ph)₂ in CD₃C₆D₅ solution at 294 K. Trace (a) shows the ¹¹B-¹H(broad-band noise) spectrum of the *exo,exo* isomer (1) [obtained by thermal isomerization of the mixture which gave trace (b)], and trace (b) shows the ¹¹B-¹H(broad-band noise) spectrum of a mixture of the *exo,exo* and *endo,exo* isomers (1) and (2) respectively. Trace (c) is a weighted subtraction of spectrum (a) from (b), and trace (d) is a similar subtraction of the equivalent ¹¹B spectra recorded without ¹H decoupling. Traces (c) and (d) respectively therefore represent the ¹¹B-¹H(broad-band noise) and ¹¹B spectra of the *endo,exo* isomer (2)

hydrogen atoms (Figure 2, top). This is a diagnostic of *endo*-terminal hydrogen character. ¹¹F For the *endo,endo* species (3) the *exo*-6,9 proton resonances lie close to the correlation plot, as expected from their *exo* nature. The *endo,exo*-6,9-bis(phosphine) compound (2), however, with a corresponding *exo,endo*-6,9-proton configuration, has one proton of each type.

A definitive precise spectroscopic assignment at the 2,4 and 5,7,8,10 positions in compound (2) is not possible on present evidence, although consideration of the shielding trends among compounds (1)—(3) suggests that ¹¹B(2) adjacent to the *endo*-phosphine-substituted vertex designated B(6) resonates at δ +7.4 p.p.m. and that the ¹¹B(5,7) resonance similarly is that at δ -14.4 p.p.m. In principle, precise assignments for the polyhedral borane species may be obtained by two-dimensional [¹¹B-¹¹B]-COSY¹¹⁻¹⁴ n.m.r., but in practice the resonance lines for the mixture of compounds (1) and (2) were so broadened by rapid ¹¹B relaxation that no correlations could be observed at ambient temperatures. Higher temperatures are often used to increase ¹¹B relaxation times and thus render ¹¹B-¹¹B cross-correlations more observable,^{11,15-17} but in this case the *endo,exo* isomer (2) was found to be insufficiently thermally robust, and heating in CD₃C₆D₅ solution for a few minutes resulted in conversion into the symmetrical *exo,exo* isomer (1). The lack of robustness has so far precluded the mutual separation of these two isomers. They have very similar solubility and chromatographic properties, and there is a slow conversion of the asymmetric species (2) into the symmetric species (1) under the separatory conditions that we have previously found satisfactory for systems of this nature.

Experimental

Sample preparations were carried out under dry nitrogen by the addition of stoichiometric quantities of PMe₂Ph [equation (1)], scale ca. 10 mmol in ca. 0.5 cm³ solvent] to solutions of B₁₀H₁₄ or B₁₀H₁₂(SMe₂)₂ in CD₂Cl₂ or CD₃C₆D₅ at ca. 203 K, followed by slow warming to room temperature. N.m.r. spectroscopy was carried out in the Fourier-transform mode at 2.35 and 9.40 T using commercially available instrumentation. ¹H-¹¹B N.m.r. spectroscopy at 2.35 T used a JEOL FX-100 instrument equipped with a ¹H-¹H probe in which the formal ¹H-irradiation system had been tuned to accept the ¹¹B resonance frequency (32 MHz). That at 9.40 T used a Bruker AM-400 instrument equipped with a ¹¹B-¹H probehead that

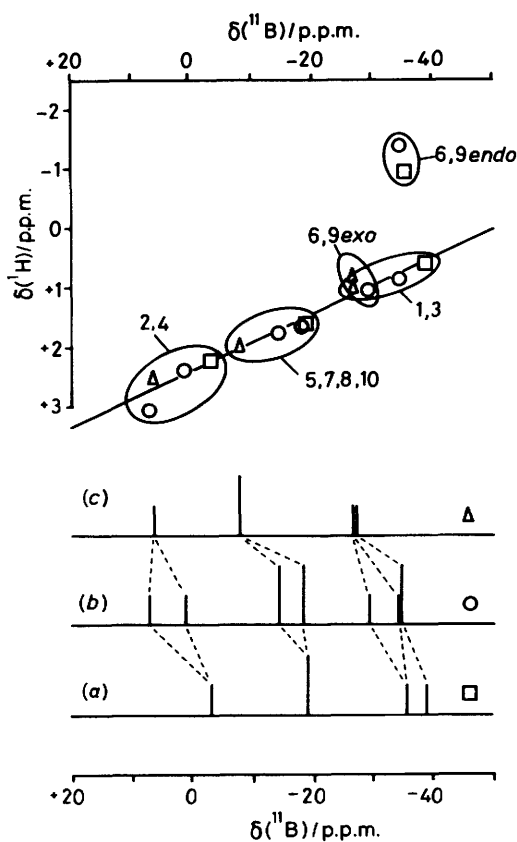


Figure 2. (a)–(c) Stick representations of the chemical shifts and relative intensities in the ^{11}B n.m.r. spectra of compounds (1) (*exo,exo*), (2) (*endo,exo*), and (3) (*endo,endo*), respectively, which illustrate the intermediate behaviour of (2). The upper diagram is a plot of $\delta(^1\text{H})$ versus $\delta(^{11}\text{B})$ for directly bound atoms in compounds (1) (\square), (2) (\circ), and (3) (\triangle); the line drawn has gradient $\delta(^1\text{H})/\delta(^{11}\text{B}) = 1/21$

was used inversely: ^1H observation (400 MHz) was effected via the 'irradiation' coils and $\{^{11}\text{B}\}$ decoupling (128 MHz) via the 'observation' coils. In both cases $\gamma B_2(^{11}\text{B})/2\pi$ (γ = magnetogyric ratio, B_2 = irradiating field strength) for the selective decoupling experiments was *ca.* 300 Hz [estimated using residual splittings in ^1H - $\{^{11}\text{B}(\text{off-resonance})\}$ experiments on NaBH_4 solutions].¹⁸ The indicated irradiation power in the AM-400 configuration under these conditions was about 0.15 W (Bird Thurline 43 Wattmeter), although precise adjustment

of the power level was not critical in the experiments reported here. The n.m.r. spectroscopy was otherwise routine; ^{11}B chemical shifts are given to lower field (higher frequency) of Ξ 32.083 971 MHz [nominally $\text{BF}_3(\text{OEt}_2)$ in CDCl_3],¹¹ measured using residual solvent-proton resonances as internal secondary standards relative to those of SiMe_4 (Ξ 100 MHz).

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