

## Notes

### Proton Nuclear Magnetic Resonance and Optical Spectra of Six-co-ordinated High-spin ( $S = 2$ ) Bis(tetrahydrofuran)(3,7,12,17-tetramethyl-8,13-divinylporphyrin-2,18-dipropionate)iron(II) encapsulated in Aqueous Detergent Micelles

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Proton n.m.r. (500 MHz) and optical spectral studies have been made on the six-co-ordinated high-spin iron(II) haem [ $\text{FeL}(\text{thf})_2$ ] ( $\text{L} = 3,7,12,17$ -tetramethyl-8,13-divinylporphyrin-2,18-dipropionate, thf = tetrahydrofuran) encapsulated in aqueous hexadecyltrimethylammonium bromide micelles. The results are compared with those of the five-co-ordinated high-spin iron(II) haem reported previously. The optical spectrum of the six-co-ordinated species show the Soret band at 415 nm and two other bands at 540 and 564 nm, while for the five-co-ordinated species the Soret band shifts to longer wavelength and a single broad band at 555 nm was obtained. The n.m.r. spectra show a larger downfield shift for the methyl- and *meso*-proton resonances and a smaller spread of the four methyl resonances for the six-co-ordinated complex as compared to those of five-co-ordinated high-spin haems.

In deoxyhaemoglobin and deoxymyoglobin the high-spin ( $S = 2$ ) iron(II) ion is found in a five-co-ordinated square-pyramidal geometry above the plane of the porphyrin and displaced towards the fifth ligand.<sup>1,2</sup> On binding a sixth ligand the iron atom is pulled into the plane of the porphyrin, where it is found as a low-spin species in octahedral geometry.<sup>1,2</sup> The out-of-plane displacement of the iron atom in the deoxyproteins was believed to be due to the large size of high-spin ( $S = 2$ ) iron(II) as compared to that of the  $\text{N}_4$  hole in the porphyrin plane.<sup>2</sup> This led to the general belief that the high-spin state is always associated with the out-of-plane displacement and five-co-ordination of the iron atom in porphyrins.

However, if two equivalent weak-field axial ligands are bound, high-spin six-co-ordinated haems may be obtained where the iron atom may be found in the plane of the porphyrin.<sup>3</sup> There are several examples of high-spin six-co-ordinated iron(III) porphyrin complexes,<sup>3,4-6</sup> but the only well characterized analogous iron(II) complex is [ $\text{Fe}(\text{tpp})(\text{thf})_2$ ] ( $\text{tpp} = 5,10,15,20$ -tetraphenylporphyrinate, thf = tetrahydrofuran) which was reported to be stable only in the crystalline state.<sup>7,8</sup> In these six-co-ordinated complexes the iron atom is rigorously centred in the porphyrin plane<sup>3,8</sup> and the large high-spin ions are accommodated in the haem plane by porphyrin ring-core expansion with significant non-bonded repulsion between the axial ligands and the pyrrole N atoms of the porphyrinato core.<sup>3,8-10</sup>

Recently we have shown that six-co-ordinated high-spin complexes are easily formed when weak axial ligands such as  $\text{H}_2\text{O}$  or thf are bound to iron porphyrins encapsulated in detergent micelles.<sup>4-6,11</sup> A Mössbauer study of [ $\text{FeL}_2(\text{thf})_2$ ] [ $\text{L} = 3,7,12,17$ -tetramethyl-8,13-divinylporphyrin-2,18-dipropionate (dianion of protoporphyrin IX)] in frozen 5% aqueous solution showed isomer shift [0.91(2)  $\text{mm s}^{-1}$ ] and quadrupole splitting [2.49(2)  $\text{mm s}^{-1}$ ] values<sup>11</sup> similar to those of the high-spin six-co-ordinated tpp analogue.<sup>7,8</sup> A resonance-Raman

study of haem in aqueous thf showed an expanded porphyrin core with the high-spin  $\text{Fe}^{\text{II}}$  centred in the haem plane.<sup>12</sup> However, in the absence of the detergent (*e.g.* in aqueous thf) about 50% of the molecules are found to be in the form of aggregated species.<sup>11</sup> The detergent micelles stabilize monomeric species in aqueous solution as found earlier.<sup>4-6,13</sup> In view of the recent interest in the six-co-ordinated high-spin haems as haemoprotein models,<sup>3,7-12</sup> we report here the  $^1\text{H}$  n.m.r. and electronic spectra of the complex [ $\text{FeL}(\text{thf})_2$ ] in aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  solution. To our knowledge this is the first such study on a high-spin six-co-ordinated iron(II) haem in aqueous solution.

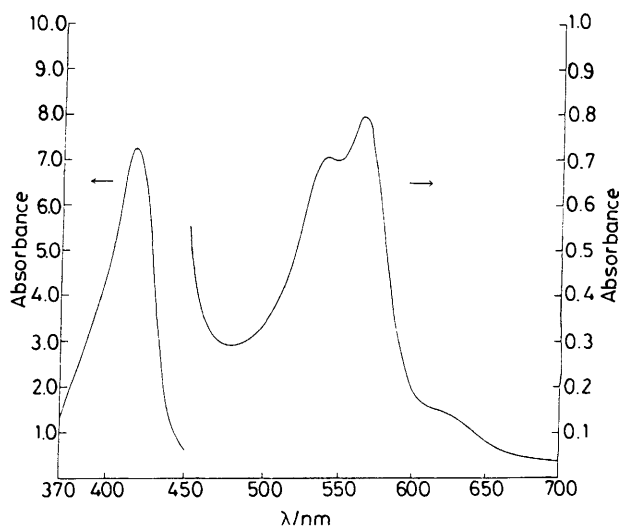
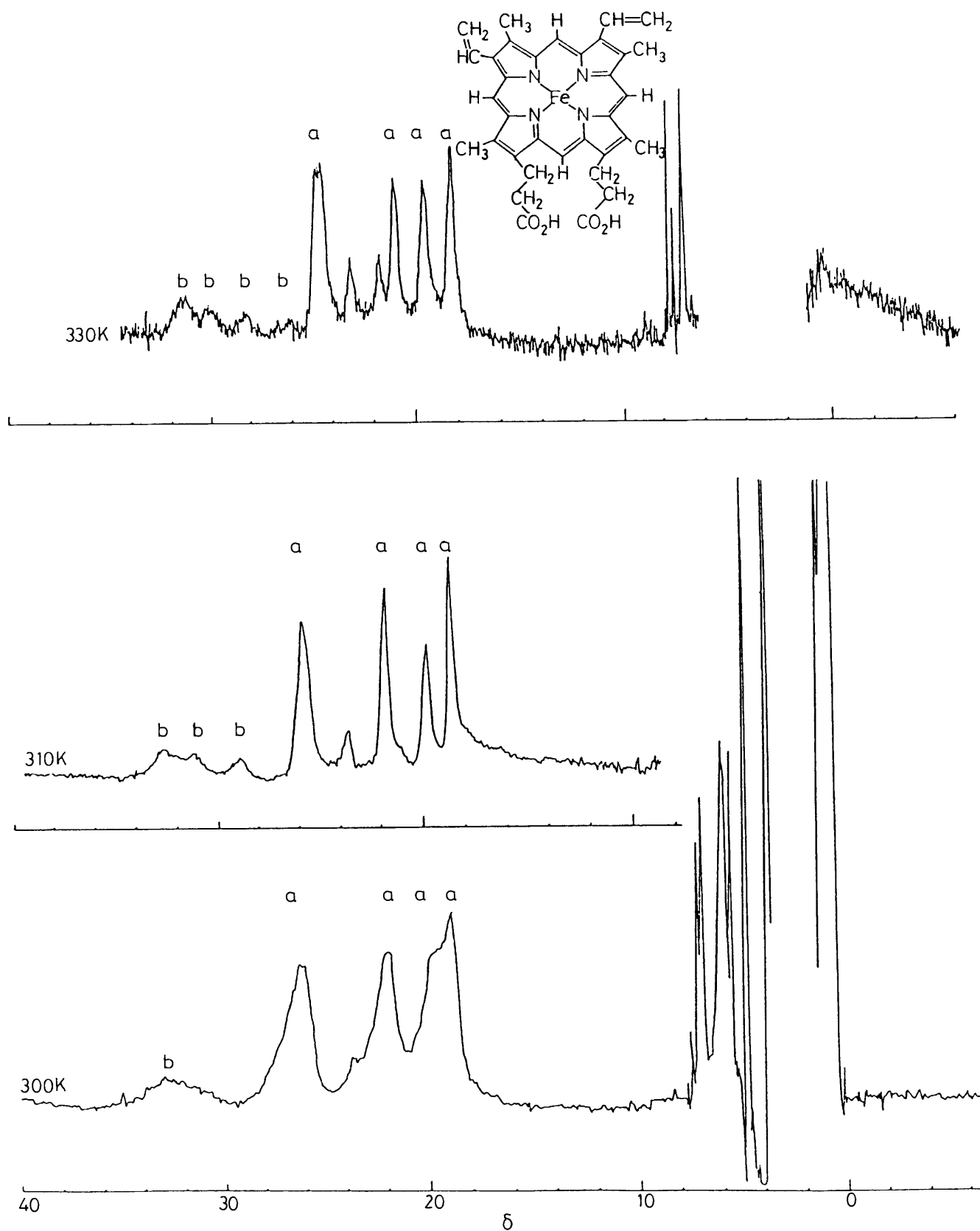


Figure 1. Electronic spectrum of  $6.5 \times 10^{-5} \text{ mol dm}^{-3}$  [ $\text{FeL}(\text{thf})_2$ ] encapsulated in 5% aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  micelles at 298 K and pH 10.0



**Figure 2.** Proton n.m.r. spectra (500 MHz) of  $[\text{FeL}(\text{thf})_2]$  at 330 (top) and 300 K (bottom) in 5% aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  solution. Assignments: a, haem  $\text{CH}_3$ ; b, meso-H

## Experimental

Iron(III) protoporphyrin IX chloride (haemin chloride) was purchased from Sigma Chemicals,  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  from E. Merck Co. A solution of the iron(III) complex (0.01–1.0  $\text{mmol dm}^{-3}$ ) in 5% aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  micelles<sup>4–6</sup> at pH 10.0 was prepared by dissolving it in an alkaline, thoroughly deoxygenated,  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  solution. The solution was allowed to stand for about 2 h at 40–50 °C and then to equilibrate with excess of tetrahydrofuran for another 3 h prior to reduction of the complex by sodium dithionite ( $\text{Na}_2\text{S}_2\text{O}_4$ ).

All operations were carried out under a nitrogen atmosphere and all reagents (including water– $\text{D}_2\text{O}$ ) were deoxygenated. A minimum amount of a saturated aqueous solution of  $\text{Na}_2\text{S}_2\text{O}_4$  was used to reduce the iron(III) complex. For the optical spectrum, a *ca.*  $10^{-5}$   $\text{mol dm}^{-3}$  solution of the iron(III) porphyrinate was taken in a quartz cuvette fitted with a rubber septum, and the reduction was carried out by injecting the dithionite solution, with gentle shaking, until a red solution was obtained. The sample for n.m.r. studies was prepared by reducing a *ca.* 1  $\text{mmol dm}^{-3}$  solution of the iron(III) porphyrinate in an n.m.r. tube fitted with a rubber septum which was previously flushed with nitrogen gas.

Experimental details for the optical and n.m.r. spectral measurements are as reported previously.<sup>13</sup> To get a good signal-to-noise ratio in the n.m.r. spectra, about 4 000–5 000 transients were required over a spectral width of *ca.* 45 kHz with 8K data points. In order to observe the paramagnetically shifted porphyrin protons, the micelle- and the water-proton signals needed to be saturated. A microprogram for multi-frequency irradiation was used to irradiate all these proton signals.

## Results and Discussion

The optical spectrum of  $[\text{FeL}(\text{thf})_2]$  in 5%  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  aqueous solution is shown in Figure 1. This spectrum is clearly different ( $\lambda_{\text{max}}$  at 415, 540, and 564 nm) from those of high-spin ( $S = 2$ ) five-co-ordinated or intermediate-spin ( $S = 1$ ) four-co-ordinated species in the same detergent solution.<sup>13</sup> Gradual addition of thf to the four-co-ordinated  $[\text{FeL}]$  in  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  solution, which shows a split Soret band,<sup>13</sup> led to a sharp band at 415 nm when excess of thf was added. The spectrum in Figure 1 is similar to that found for six-co-ordinated haems<sup>14</sup> indicating co-ordination of two molecules of thf to  $[\text{FeL}]$ . Optical spectral study at various concentrations showed no evidence for aggregation<sup>15</sup> of the haem in 5% aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$ . Thus the species in solution is a monomeric six-co-ordinated haem with thf as the axial ligands.

The  $^1\text{H}$  n.m.r. (500 MHz) spectra of  $[\text{FeL}(\text{thf})_2]$  in 5% aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  at various temperatures are shown in Figure 2. Though the room-temperature (298 K) spectrum is broad, four sharp well resolved lines are observed at > 300 K. These spectra agree quite well with those reported for deoxy-haemoglobin<sup>16</sup> and other high-spin ( $S = 2$ ) haems.<sup>13</sup> However, the downfield shift of the proton resonances of the six-co-ordinated species  $[\text{FeL}(\text{thf})_2]$  is larger than that reported previously for other high-spin ( $S = 2$ ) haems. Assignment of the various resonances is based on the relative intensity and multiplicity and by comparison with the assignments for high-spin iron(II) haems<sup>17</sup> and haemoglobin.<sup>16,18</sup>

The porphyrin proton resonances in the n.m.r. spectrum of  $[\text{FeL}(\text{thf})_2]$  in aqueous  $\text{NMe}_3(\text{C}_{16}\text{H}_{33})\text{Br}$  are much broader than those observed for high-spin haems in organic solvents<sup>17</sup> and are similar to those observed for haemoglobin.<sup>16–18</sup> A

possible cause of such line broadening for haems encapsulated in a hydrophobic cavity was discussed recently.<sup>13</sup> As in the case of high-spin five-co-ordinated species,<sup>13</sup> the isotropic shift of the methyl and *meso* protons shows a linear temperature dependence in the range 310–330 K indicating that only one spin state is populated in this range.<sup>13</sup>

There are three significant differences in the n.m.r. spectra of the five- and six-co-ordinated high-spin iron(II) haem complexes. First, the spread of the haem methyl resonances for the six-co-ordinated complex  $[\text{FeL}(\text{thf})_2]$  is much smaller (*ca.* 6.5 p.p.m.) than that found for the five-co-ordinated high-spin species (*ca.* 16 p.p.m.) studied under identical conditions.<sup>13</sup> The smaller spread of the haem methyl signals for the former may be due to a smaller haem in-plane asymmetry.<sup>13,16–18</sup> Secondly, the average shift of the four haem methyl resonances has a larger downfield bias for the six-co-ordinated species (*ca.* 21 p.p.m.) than that of the five-co-ordinated haem (*ca.* 17 p.p.m.).<sup>13</sup> A similar dependence of the methyl proton resonances on co-ordination number was found for the iron(III) porphyrinates,<sup>19</sup> where the larger downfield shifts for the six-co-ordinated complex were attributed to the added effect of  $\pi$ -spin transfer on a dominant  $\sigma$ -spin transfer mechanism.<sup>19</sup> Thirdly, the broad and weak resonances near 30 p.p.m. downfield, resolved as four broad and weak peaks at 320 K (Figure 2), are likely to be the *meso*-H signals.\* For the five-co-ordinated high-spin haem the *meso*-H peaks appear at  $\delta$  0–6.<sup>17</sup> The large downfield bias of the *meso*-H resonances for the high-spin six-co-ordinated haems studied here agrees quite well with the similar observation for haemin complexes.<sup>19</sup>

It is interesting that for the haem complexes where the iron atom lies in the plane of the porphyrin (*viz.* four-co-ordinated intermediate-spin,<sup>13,20</sup> six-co-ordinated low-spin,<sup>13</sup> and six-co-ordinated high-spin) the downfield shift of the *meso* protons is larger than that of the methyl-proton resonances. However, for the high-spin five-co-ordinated haems where the iron atom is out of the porphyrin plane the downfield shift of the methyl-proton resonances is larger than that of the *meso*-proton resonances.<sup>17</sup>

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\* Assigned by comparison with the proton n.m.r. spectra of various iron-(II) and -(III) haems reported previously.<sup>13,17–20</sup>

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