ORGANOMETALLICS

Volume 12, Number 3, March 1993

© Copyright 1993 American Chemical Society

Communications

Bioorganometallic Chemistry. 2. Synthesis and Structural Studies of the Reactions of a Nucleobase, 1-Methylcytosine, with a $(\eta^5$ -Pentamethylcyclopentadienyl)rhodium Aqua Complex

David P. Smith,[†] Marilyn M. Olmstead,[‡] Bruce C. Noll,[‡] Marcos F. Maestre,[†] and Richard H. Fish^{*,†}

Lawrence Berkeley Laboratory, University of California, Berkeley, California 94720, and Department of Chemistry, University of California, Davis, California 95616

Received December 23, 1992

Summary: The reactions of a nucleobase, 1-methylcytosine (MC), with a Cp*Rh aqua complex, $[(\eta^5 - Cp^*) Rh(H_2O)_2(OTf)_2J_x(2)$, provided two different complexes depending on the solvent media. Complex 3, $f(\eta^5-Cp^*)$ - $Rh(\eta^{1}(N3)-MC)(\eta^{2}(O2,N3)-MC)](OTf)_{2}$, was formed when acetone was used as the solvent; however, when complex **3** was recrystallized from water (pH 5.1) or when water was used as the reaction solvent, complex 4, trans- $[(\eta^{5} Cp^*$) $Rh(\eta^1(N3)-MC)(\mu-OH)J_2(OTf)_2$, was isolated as a crystalline solid. The structures of 3 and 4 were verified by ¹H NMR, FAB/MS, elemental analysis, and singlecrystal X-ray analysis. The structure of complex 3 showed one MC ligand bound via N3 and the other chelated via N3 and C=O2 (Rh-O2 = 2.251(6) Å). Inspection of several bond lengths of complex 4 indicates extensive intramolecular hydrogen bonding of the μ -OH groups with the exocyclic NH_2 (HO---HNH = 1.93(1) Å) and the 2-C==0 group (OH - - - O = C = 1.96(1) Å).

The reactions of inorganic metal complexes with DNA/ RNA nucleobases, nucleosides, nucleotides, and oligonucleotides have been extensively studied,¹ while few studies have been directed toward organometallic complexes with these biologically important ligands.² We have been investigating the coordination chemistry of the highly electrophilic (η^5 -pentamethylcyclopentadienyl)rhodium dicationic complex [Cp*Rh(S)₃]²⁺ (S = CH₃COCH₃, CH₃-

 (a) Tullius, T. D. In Metal-DNA Chemistry; Tullius, T. D., Ed.; ACS Symposium Series 402; American Chemical Society: Washington, DC 1989; Chapter 1, and references therein. (b) Barton, J.K. Comments Inorg. Chem. 1985, 3, 321 and references therein. (c) Pyle, A. M.; Barton, J. K. In Progress in Inorganic Chemistry: Bioinorganic Chemistry; Lippard, S. J., Ed.; Wiley: New York, 1990; Vol. 38, p 413, and references therein. (d) Marzilli, L. G. In Progress in Inorganic Chemistry; Lippard, S. J., Ed.; Wiley: New York, 1990; Vol. 38, p 413, and references therein. (d) Marzilli, L. G. In Progress in Inorganic Chemistry; Lippard, S. J., Ed.; Wiley: New York, 1977; Vol. 23, p 255, and references therein. (e) Howe-Grant, M. E.; Lippard, S. J. Met. Ions Biol. Syst. 1980, 11, 63 and references therein. (f) Raudaschl-Sieber, G.; Schollhorn, H.; Thewalt, U.; Lippert, B. J. Am. Chem. Soc. 1985, 107, 3591. (g) Reily, M. D.; Hambley, T. W.; Marzilli, L. G. J. Am. Chem. Soc. 1988, 110, 2999. (i) Alink, M. A.; Nakahara, H.; Hirano, T.; Inagaki, K.; Hakanishi, M.; Kidani, Y.; Reedijk, J. Inorg. Chem. 1991, 130, 1236 and references therein. (i) Reily, M. D.; Marzilli, L. G. J. Am. Chem. Soc. 1986, 108, 8299. (j) Qu, Y.; Farrell, N. J. Am. Chem. Soc. 1991, 113, 4851 and references therein. (k) Caradonna, J. P.; Lippard, S. J.; Gait, M. J.; Singh, M. J. Am. Chem. Soc. 1982, 104, 5793. (l) Marcelis, A. T. M.; den Hartog, J. H. J.; Reedijk, J. J. Am. Chem. Soc. 1982, 104, 2664. (m) Sherman, S. E.; Gibson, D.; Wang, A. H.-J.; Lippard, S. J. J. Am. Chem. Soc. 1988, 110, 7368. (n) Mukundan, S., Jr.; Xu, Y.; Zon, G.; Marzilli, L. G. J. Am. Chem. Soc. Dalton Trans. 1979, 596. (p) Rainen, L.; Howard, R. A.; Kimball, A. P.; Bear, J. L. Inorg. Chem. 1975, 14, 2752. (q) Torres, L. M.; Marzilli, L. G. J. Am. Chem. Soc. 1991, 113, 4678. (r) Scheller, K. H.; Mitchell, P. Bear, J. L. Inorg. Chem. 1975, 14, 2752. (q) Torres, L. M.; Marzilli, L. G. J. Am. Chem. Soc. 1991, 113, 4678. (r) Scheller, K. H.; Mitchell, P.; Bear, J. L. Inorg. Ch

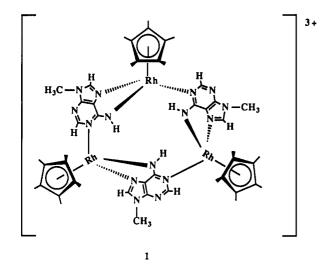
American Chemical Society: Washington, DC, 1995; Chapter 11, and references therein. (t) Hodgson, D. J. In Progress in Inorganic Chemistry; Lippard, S. J., Ed.; Wiley: New York, 1977; Vol. 23, p 211.
(2) (a) Toney, J. H.; Marks, T. J. J. Am. Chem. Soc. 1985, 107, 947.
(b) Kuo, L. Y.; Kanatzidis, M. G.; Marks, T. J. J. Am. Chem. Soc. 1987, 109, 7207. (c) Toney, J. H.; Brock, C. P.; Marks, T. J. Am. Chem. Soc. 1986, 108, 7263. (d) Kuo, L. Y.; Kanatzidis, M. G.; Sabat, M.; Tipton, A. L.; Marks, T. J. J. Am. Chem. Soc. 1991, 113, 9027.

^{*} To whom correspondence should be addressed. [†] University of California at Berkeley.

¹ University of California at Berkel ² University of California at Davis.

CN) with nitrogen heterocyclic compounds in organic solvents.³ Similar studies in aqueous solution with biologically important nitrogen ligands as mentioned above are of interest in view of the utility of Cp*Rh aqua complexes as anchors for single DNA molecules, in conjunction with surface microscopy techniques, for application to the human genome involving mapping and sequencing DNA bases,⁴ and as potential chemotherapeutics as well as reagents in biotechnology.^{1b,c}

Recently, we reported that 9-methyladenine formed an unusual and unprecedented cyclic trimer, $[(\eta^5-\text{Cp}^*)\text{Rh}-(\mu_2-\eta^1(N1):\eta^2(N6,N7)-9-\text{methyladenyl})]_3(\text{OTf})_3$ (1) upon reaction in water with a $(\eta^5$ -pentamethylcyclopentadienyl)rhodium aqua complex, 2, from pH 6 to 9.⁵ In that study, it was evident that the formation of the $\eta^2(\text{N6},\text{N7})$ five-



membered adenyl chelate occurred by a condensation reaction of the exocyclic NH_2 group with 2, a reactive Cp*Rh hydroxy species at pH 5–7.^{5,6} Similar cyclic trimer structures were also observed for adenosine,⁵ adenosine 3'-monophosphate, and the phosphate methyl ester of adenosine 5'-monophosphate in reactions with 2 in aqueous solution.⁷

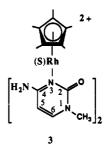
In order to ascertain the scope of this condensation reaction with nucleobases that have exocyclic NH₂ groups, we studied the reaction of the aqua complex 2, having an empirical formula of $[Cp*Rh(H_2O)_2(OTf)_2]_x$, with 1methylcytosine (MC). We found that two different Cp*Rh 1-methylcytosine complexes could be isolated depending on the solvent media used in the reaction.

(5) Smith, D. P.; Baralt, E.; Morales, B.; Olmstead, M. M.; Maestre, M. F.; Fish, R. H. J. Am. Chem. Soc. 1992, 114, 10647.

M. F.; Fish, K. H. J. AM. Chem. Soc. 1992, 114, 10647.
(6) (a) Kang, J. W.; Maitlis, P. M. J. Organomet. Chem. 1971, 127. (b) Espinet, P.; Bailey, P. M.; Maitlis, P. M. J. Chem. Soc., Dalton Trans. 1979, 1542. (c) Espinet, P.; Bailey, P. M.; Piraino, P.; Maitlis, P. M. Inorg. Chem. 1979, 18, 2706. (d) Nutton, A.; Bailey, P. M.; Braund, N. C.; Goodfellow, R. J.; Thompson, R. S.; Maitlis, P. M. J. Chem. Soc., Chem. Commun. 1980, 631. (e) Nutton, A.; Bailey, P. M.; Maitlis, P. M. J. Chem. Soc., Dalton Trans. 1981, 1997. (f) Nutton, A.; Maitlis, P. M. J. Chem. Soc., Dalton Trans. 1981, 2335. (g) Nutton, A.; Maitlis, P. M. J. Chem. Soc., Dalton Trans. 1981, 2339. (h) Kölle, U.; Kläui, W. Z. Naturforsch. 1991, 46B, 75.

(7) Smith, D. P.; Kohen, E.; Maestre, M. F.; Fish, R. H. Submitted for publication.

Reaction of 2 with 2 equiv of 1-methylcytosine in acetone for 36 h, followed by extraction with CH₂Cl₂, gave an orange precipitate (67%), complex 3, whose ¹H NMR spectrum in DMSO-d₆ revealed only one set of 1-methylcytosine and Cp* resonances and considerable downfield chemical shifts for H5 and H6 in comparison to free 1-methylcytosine ($\Delta\delta$ (H5) = 0.25 ppm, $\Delta\delta$ (H6) = 0.26 ppm). In



addition, two broad N(4)H₂ resonances were also shifted downfield upon coordination (7.70, 8.50 ppm compared to free 1-methylcytosine at 6.91 ppm). The FAB/MS data were consistent with a monomeric species (*m*-nitrobenzyl alcohol; *m*/*z* 512, [Cp*Rh(MC)(OTf)]) and provided no evidence for a dimer. The ¹H NMR data (one set of 1-methylcytosine signals for 3 down to -90 °C in CD₃OD) supports the solution structure of 3 as $[(\eta^5-Cp^*)Rh(\eta^1-(N^3)-MC)_2(S)](OTf)_2$, where a solvent molecule (S = H₂O, DMSO, CH₃OH) appears to take up the third coordination site on $(\eta^5-Cp^*)Rh.^8$

An X-ray structural determination of compound 3 recrystallized from methanol, a weakly coordinating solvent, clearly shows, in the solid state, that one of the 1-methylcytosine ligands bonds via a four-membered-ring chelate, N3-Rh-O=C2 (Rh-N3a, 2.143(7) Å; Rh-O2a, 2.251(6) Å) with the other ligand bound through the expected N3 site (Rh-N3b, 2.126(8) Å) (Figure 1). Several structurally characterized examples of cytosine N3,O2 metal semichelates exist and exhibit longer M-O bond lengths (M = Cu, 2.76 Å; M = Cd, 2.64, 2.56, and 2.89 Å; M = Hg, 2.84 Å)⁹ in comparison to those observed in compound 3. The structural consequences of N3,O2 chelation, versus N3 coordination alone, are manifested in complex 3 by a longer C-O bond length (C2a-O2a, 1.264-(10) Å; C2b–O2b, 1.209(11) Å), a shorter C2–N3 bond length (C2a-N3a, 1.350(10) Å; C2b-N3b, 1.402(11) Å), a

^{(3) (}a) Fish, R. H.; Kim, H.-S.; Babin, J. E.; Adams, R. D. Organometallics 1988, 7, 2250. (b) Fish, R. H.; Baralt, E.; Kim, H.-S. Organometallics 1991, 10, 1965.

^{(4) (}a) Mapping and Sequencing the Human Genome; National Research Council Report; National Academy Press: Washington, DC, 1988. (b) Houseal, T. W.; Bustamante, C.; Stump, R. F.; Maestre, M. F. Biophys. J. 1989, 56, 507. (c) Zuccheri, G.; Smith, D. P.; Fish, R. H.; Maestre, M. F. Manuscript in preparation.

⁽⁸⁾ Complex 3, $[(\eta^{5}-Cp^{*})Rh(\eta^{1}(N3)-MC)(\eta^{2}(O_{2},N3)-MC)](O_{2}SCF_{3})_{2}$: In a Vacuum Atmospheres drybox, 200 mg of $[Cp^{*}Rh(H_{2}O)_{2}(OTf)_{2}]_{x}$ (0.36 mmol) and 91 mg of 1-methylcytosine (MC, 0.73 mmol) were slurried for 36 h in 20 mL of acetone. The solvent of the orange solution was stripped in vacuo, and the remaining orange solid was slurried in 20 mL of CH₂Cl₂ for 2 h and then filtered and dried to give 195 mg (67%) of complex 3. Analytically pure 3 was obtained by recrystallization from minimal methanol. ¹H NMR (DMSO-d₆, ppm): 8.52 (b, 1H, NH), 7.82 (d, J_{HH} = 7.4 Hz, 1 H, H6), 7.72 (b, 1H, NH), 5.84 (d, J_{HH} = 7.4 Hz, 1 H, H5), 3.28 (s, 3 H, Me), 1.71 (s, 15H, Cp*). FAB/MS (m-nitrobenzyl alcohol; m/z (relative intensity) 512.1 (52), [Cp*Rh(MC)(OTf)]; 387.0 (18), [Cp*Rh(OTf)]; 362.1 (100), [Cp*Rh(MC)]; 237.0 (30), [Cp*Rh-H]; 126.1 (40), [MC + H]. Anal. Calcd for RhO₆S₂F₆N₆C₂₂H₂₉: C, 33.60; [$(\eta^{5}$ -Cp*)Rh($\eta^{1}(N3)$ -MC)($\eta^{2}(N3,O2)$ -MC)](O₃SCF₃)₂1.5MeOH were obtained from methanol/Et₂O solution at -30 °C under an inert atmosphere. Crystal data: Mo K $\alpha(\lambda = 0.710 73 Å)$; T = 130 K; space group P2₁/n; Z = 4; a = 9.096(3) Å, b = 27.396(7) Å, c = 13.952(5) Å, β $= 99.14(3)^{\circ}; 0^{\circ} > 2\theta > 50^{\circ}; 4160$ observed reflections ($F > 4.0\sigma(F)$); R =0.0717 and $R_w = 0.0745$. To satisfactorily refine the structure, one of the two methanol molecules present in the crystal was modeled at 100% site occupancy and the second at 50% occupancy. The structure was solved by the Patterson method using SHELXTL PLUS.

^{(9) (}a) Cu: Szalda, D. J.; Kistenmacher, T. J. Acta Crystallogr. 1977, B33, 865.
(b) Cd: Aoki, K.; Saenger, W. J. Inorg. Biochem. 1984, 20, 225.
(c) Hg: Authier-Martin, M.; Beauchamp, A. L. Can. J. Chem. 1977, 55, 1213.

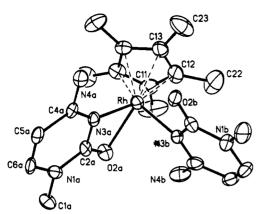
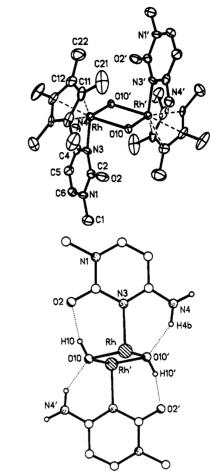


Figure 1. Molecular structure of 3, $[(\eta^5-Cp^*)Rh(\eta^1(N3)-MC)-MC)]$ $(\eta^2(O2,N3)-MC)](OTf)_2 \cdot 1.5 MeOH$, with atoms shown as 50% ellipsoids. Only the cation is shown for clarity. Selected bond lengths (Å) and angles (deg): Rh-N3a, 2.143 (7); Rh-N3b, 2.126 (8); Rh-O2a, 2.251 (6); N3a-Rh-N3b, 90.0 (3); N3a-Rh-O2a, 61.0 (2); O2a-Rh-N3b, 88.0 (2).

shorter C4-N4 bond length (N4a-C4a, 1.308 Å; N4b-C4b, 1.349(13) Å), and a smaller O2-C2-N3 bond angle (O2a-C2a-N3a, 117.1(7)°; O2b-C2b-N3b, 121.6(8)°). The N4a---O2b through-space distance (3.175 Å) precludes any intramolecular hydrogen bonding between these sites.

Upon recrystallization of complex 3 from H_2O (pH 5.1), a new complex, 4, is formed. Complex 4 can also be isolated by the dropwise addition of a deoxygenated, aqueous solution of 1 equiv of 1-methylcytosine to an aqueous solution of 2 that was adjusted to pH 5-6 by addition of NaOH. The ¹H NMR spectrum of 4 in DMSO- d_6 showed upfield chemical shifts for H5 (5.27 ppm) and H6 (7.45 ppm) of 0.32 and 0.09 ppm, respectively, in comparison to free 1-methylcytosine, while one set of exocyclic NH₂ protons at 7.18 ppm was shifted downfield by 0.27 ppm. The other exocyclic NH₂ signal was not observed and is apparently broadened into the base line. FAB/MS verified the dimeric nature of 4 (m-nitrobenzyl alcohol; m/z 766, $[(Cp*Rh)_2(MC)(\mu-OH)](OTf); m/z 659, [Cp*Rh(\mu-OH)]_2$ (OTf), while the single-crystal X-ray analysis confirmed the trans stereochemistry and the extensive intramolecular hydrogen bonding of the bridging hydroxyl groups with the C=O and the exocyclic NH_2 groups (Figure 2).¹⁰ This hydrogen-bonding network creates a hydrophobic environment around the metal centers and appears to be the reason that 4 is insoluble in H_2O at pH 5.1.

The core structure of 4 is quite similar to cis^{-3a} and trans-[$(\eta^5$ -Cp*)Rh $(\eta^1$ -(N))(μ -OH)] $_2^{2+11}$ dimeric complexes with heteroaromatic nitrogen ligands (cis stereochemistry,



h

Figure 2. (a, upper view) Molecular structure of 4, trans- $[(\eta^5-Cp^*)Rh(\eta^1(N3)-MC)(\mu-OH)]_2(O_3SCF_3)_2$, with atoms as 50% ellipsoids. Selected bond length (Å) and angles (deg): Rh-N3, 2.181(5); Rh-O10, 2.138(5); Rh-O10', 2.118(5); C(2)-O(2), 1.232(10); C(4)-N(4), 1.32(9); H4b-O10', 1.93(1); H10-O2, 1.96(1); N3-Rh-O10, 87.9(2); Rh-O10-Rh', 101.2(2); N(4)-C(4)-N(3), 117.7(6); O(2)-C(2)-N(3), 123.1(6); N4-H4-O10'149(1); O10-H10-O2, 155(1). (b, lower view) Structure of the $[Rh(\mu-OH)(MC)]_2$ core emphasizing the intramolecular hydrogen bonding.

N = quinoline, 5; trans stereochemistry, N = pyridine, 6); however, some slight differences are apparent. The planar $Rh_2(\mu-OH)_2$ fragment of 4 shows a wider O-Rh-O angle of 78.8(2)° in comparison to 5 (75.1(1), 74.8(1)°) and 6(76.8-(2)°). As well, a smaller Rh–O–Rh angle (101.2(2)°) for 4 was observed in comparison to 104.7(1) and 105.0(1)° for 5 and 103.1(2)° for 6. This results in a closer Rh-Rh through-space distance of 3.290(2) Å for 4 in comparison to 3.322(1) Å for 5 and 3.308(1) Å for 6.

This slight deformation is presumably caused by the unusual binding of the 1-methylcytosine in which the (Rh-OH)₂ core acts as a covalent electrophile, a hydrogen bond donor, and a hydrogen bond acceptor. This system is a rare example of a complex in which a μ -hydroxy ligand exhibits simultaneous hydrogen bond donor and acceptor capabilities.¹¹ Although other metal complexes of 1methylcytosine have shown extensive intermolecular

⁽¹⁰⁾ Complex 4, $[(\eta^5-Cp^*Rh)(\eta^1(N3)-MC)(\mu-OH)]_2(O_3SCF_3)_2-4H_2O:$ Complex 3 (43 mg) was dissolved in 5 mL of deoxygenated water, and the solution was stirred for 36 h, resulting in precipitation of a yellow-orange solid. The volume of the reaction mixture was reduced to ca. 2 mL, and the orange precipitate (23 mg, 81%) was collected by filtration and dried for 18 h under vacuum. ¹H NMR (DMSO- d_6 , ppm): 7.45 (H6, d, $J_{\rm HH}$ = 7.4 Hz, 1 H); 7.18 (NH, b, 1H), 5.27 (H5, d, J_{HH} = 7.4 Hz, 1 H); 3.18 (CH₃, s, 3 H); 1.78 (Cp*, s, 15H). FAB/MS (*m*-nitrobenzyl alcohol; m/z (relative intensity)): 766 (10), [(Cp*Rh(μ -OH))₂(MC)(OTf) - H₂O]; 659 (23), $\begin{array}{l} (Cp^*Rh(\mu-OH))_2(OTf); (Cp^*Rh(\mu-OH))_2(VRC)(O1f) &= H_2O]; (59 (23), \\ [(Cp^*Rh(MC)(OTf)]; (641 (13), [(Cp^*Rh(MC)])_2(OTf) &= H_2O]; (512 (20), [(Cp^*Rh(MC))]; (362 (82), [Cp^*Rh(MC)]; (237 (25), [Cp^*Rh - H]; 149 (100), [OTf]; 126 (30), [MC + H]. Anal. Calcd for Rh_2O_{14}S_2F_{04}C_{32}H_{54}: C, 33.99; H, 4.81; N, 7.43. Found: C, 33.53; H, \\ (Charlie H)_{14} & (Charlie H)_{14} &$ 4.40; N, 7.82. Orange parallelepipeds of 4 were obtained from water at room temperature. Crystal data: Mo K α ($\lambda = 0.710$ 73 Å); T = 130 K; space group $P2_1/n$; Z = 2; a = 8.077(2) Å, b = 19.541(4) Å, c = 13.508(3)A, $\beta = 106.50(3)^{\circ}$; 2484 observed reflections (F > 4.0 σ (F)); R = 0.0486 and $R_{\rm w} = 0.0480$. The structure was solved by direct methods using SHELXTL PLUS. Hydrogen atoms bonded to carbon were located from a difference map and subse equently refined using a riding model with C-H = 0.96 Å and isotropic U values equal to 1.2 times the equivalent isotropic U of

the bonded carbon. The amino and hydroxy hydrogens were also located on the difference map and restrained to have N-H and O-H bond lengths of 0.94(2) and 0.92(2) Å, respectively, and fixed isotropic U's but were otherwise not constrained. A difference map was featureless

⁽¹¹⁾ Lahoz, F. G.; Carmona, D.; Oro, L. A.; Lamata, M. P.; Puebla, M.
P.; Foces-Foces, C.; Cano, F. H. J. Organomet. Chem. 1986, 316, 221.
(12) Liu, F.; Roesky, H. R.; Schmidt, H.-G.; Noltemeyer, M. Orga-

nometallics 1992, 11, 2965.

hydrogen bonding,¹³ we are unaware of a system which recognizes the bonding capabilities of this ligand so readily in an *intra*molecular fashion. In addition, the formation of 4 provides some evidence for the structure of the reactive aqua complex 2 that corroborates the FAB/MS data, obtained previously, indicating a dimeric [Cp*Rh(μ -OH)]₂ cationic complex (pH dependent).⁵

Condensation reactions between the exocyclic NH_2 of 1-methylcytosine and M–OH centers to form either fourmembered-ring chelates ($\eta^2(N3,N4)$; Cp_2Mo^{2+} , Pt^{IV}) or μ -1methylcytosyl complexes (Pt^{II}) are well documented.^{2d,13a,13c} However, we were not able to induce a similar condensation reaction between the exocyclic NH_2 and the μ -OH group with 4. Mild thermolysis (70 °C for 16 h) of 4 in DMSO-d₆ solutions results in overall decomposition with no evidence of a condensation reaction. The fact that we observe no apparent condensation reaction of the exocyclic NH_2 group with the bridging OH group could be indicative of the pronounced stability of the extensive intramolecular hydrogen-bonding regime shown in 4 or simply a manifestation of the instability of a four-membered-ring chelate. Thermolysis of 3 under the same conditions gave a similar result, while heating 3 in acetone- d_6 at 50 °C for 24 h showed no reaction.

In future publications, we will report on the reactivity of 2 with other nucleobases, nucleotides, and sequencespecific oligonucleotides as well as its utility as a tether, simultaneously bonding to both glass or electrode surfaces and sequence-specific ofigonucleotide—single DNA molecules, for eventual application to sequence and map the human genome.^{4c}

Acknowledgment. The studies at LBL were generously supported by Laboratory Directed Research and Development Funds and the Department of Energy under Contract No. DE-ACO3-76SF00098. We thank Dr. Mina J. Bissell of LBL for her support of this project.

Supplementary Material Available: Tables of crystal data, atomic coordinates and isotropic displacement coefficients, bond lengths, bond angles, and anisotropic displacement coefficients for 3 and 4 (18 pages). Ordering information is given on any current masthead page.

OM920820N

^{(13) (}a) Faggiani, R.; Lippert, B.; Lock, C. J. L.; Speranzini, R. A. J. Am. Chem. Soc. 1981, 103, 1111. (b) Beyerle-Pfnur, R.; Schollhorn, H.; Thewalt, U.; Lippert, B. J. Chem. Soc., Chem. Commun. 1985, 1510. (c) Schollhorn, H.; Beyerle-Pfnur, R.; Thewalt, U.; Lippert, B. J. Am. Chem. Soc. 1986, 108, 3680. (d) Lippert, B. In Progress in Inorganic Chemistry; Lippard, S. J., Ed.; Wiley: New York, 1989; Vol. 37, p 1, and references therein.