ORGANOMETALLICS

Volume 12, Number 3, March 1993

0 Copmt 1993 **American Chemical Society**

Communications

Bioorganometallic Chemistry. 2. Synthesis and Structural Studies of the Reactions of a Nucleobase, 1 -Methylcytosine, with a $(n^5$ -Pentamethylcyclopentadienyl)rhodium Aqua **Complex**

David P. Smith,[†] Marilyn M. Olmstead,[†] Bruce C. Noll,[†] Marcos F. Maestre,[†] and Richard H. Fish^{*,†}

Lawrence Berkeley Laboratory, University of California, Berkeley, California 94720, and Department of Chemistry, University of California, Davis, California 95616

Received December 23, 1992

*Summary: The reactions of a nucleobase, l-methylcytosine (MC), with a Cp*Rh aqua complex,* $[(\eta^5 - Cp^*) Rh(H_2O)_2(OTf)_2I_x$ (2), provided two different complexes *depending on the solvent media. Complex 3,* $[(\eta^5 - Cp^*) Rh(\eta^2(N3)$ -MC $)(\eta^2(O2,N3)$ -MC $)(OTf)_{2}$, was formed when *acetone was used as the solvent; however, when complex 3 was recrystallized from water (pH 5.1) or when water was used as the reaction solvent, complex 4, trans-[(q5-* Cp^*)Rh(η ¹(N3)-MC)(μ -OH)]₂(OTf)₂, *was isolated as a crystalline solid. The structures of 3and 4 were verified by 'H NMR, FABIMS, elemental analysis, and singlecrystal X-ray analysis. The structure of complex 3 showed one MC ligand bound via N3 and the other chelated via N3 and C=02 (Rh-02 = 2.251(6) Å). Inspection of several bond lengths of complex 4indicates extensive intramolecular hydrogen bonding of the p-OH groups with the exocyclic NH₂ (HO---HNH = 1.93(1)* Å) *and the 2-C=0 group* $(OH--O=C = 1.96(1)$ *Å).*

The reactions of inorganic metal complexes with DNA/ RNA nucleobases, nucleosides, nucleotides, and oligonucleotides have been extensively studied,' while few studies have been directed toward organometallic complexes with these biologically important ligands.² We have been investigating the coordination chemistry of the highly electrophilic **(\$-pentamethylcyclopentadieny1)rhodium** dicationic complex $[Cp*Rh(S)₃]^{2+}$ (S = $CH₃COCH₃$, CH₃-

(1) (a) Tullius, T. D. In *Metal-DNA Chemistry;* Tullius, T. D., Ed.; ACS Symposium Series **402;** American Chemical Society: Washington, DC **1989;** Chapter **1,** and references therein. (b) Barton, J. K. *Comments Znorg. Chem.* **1985,3,321** and references therein. (c) Pyla, A. M.; Barton, J. K. In *Progress* in *Inorganic Chemistry:* Bioinorganic *Chemistry;* Lippard, S. J., Ed.; Wiley: New York, **1990;** Vol. **38,** p **413,** vd references therein. (d) Marzilli, L. G. In Progress in Inorganic Chemistry; Lippard,
S. J., Ed.; Wiley: New York, 1977; Vol. 23, p 255, and references therein.
(e) Howe-Grant, M. E.; Lippard, S. J. Met. Ions Biol. Syst. 1980, 11, 63 and references therein. *(0* Raudaechl-Sieber, **G.;** Schollhom, H.; Thewalt, U.; Lippert, B. J. *Am. Chem.* SOC. **1985,107, 3591.** (9) Reily, M. D.; Hambley, T. W.; Marzilli, L. *G.* J. *Am. Chem.* SOC. **1988,110,2999.** (h) **Alink,M.A.;Nakahara,H.;Hirano,T.;Inegaki,K.;Hakanishi,M.;Kidani,** Y.; Reedijk, J. *Inorg. Chem.* **1991, 30, 1236** and references therein. (i) Reily, M. D.; Marzilli, L. **G.** J. *Am. Chem. SOC.* **1986,108,8299.** (j) **Qu,** Y.; Farrell, N. J. *Am. Chem. SOC.* **1991,113,4851** and references therein. (k) Caradonna, J. P.; Lippard, S. J.; Gait, M. J.; Singh, M. J. *Am. Chem. Soc.* **1982,104,5793.** (1) Marcelis, A. T. M.; den Hartop, J. H. J.; Fbedijk, J. *J. Am. Chem.* **SOC. 1982,104,2664. (m)** Sherman, **S.** E.; Gibson, D.; Wang, A. H.-J.; Lippard, S. J. J. *Am. Chem.* SOC. **1988,120,7368.** (n) Mukundan, **S.,** Jr.; Xu, Y.; Zon, **G.;** Marzilli, L. *G.* J. *Am. Chem.* Soc. Wuxuunuun, 3., Jr., Xal, 1., Zoll, 3., Hadjiliadis, H. J. Chem. Soc., 1981, 113, 3021. (o) Pheumatikakis, G.; Hadjiliadis, H. J. Chem. Soc., Dalton Trans. 1979, 596. (p) Rainen, L.; Howard, R. A.; Kimball, A. P.; Bear, J. G. J. Amt. Chemi. Soc. 1991, 113, 4916. (17 Schuler, A. H., Winterland, P. S. Sigel, H. In Metal-DNA Chemistry; Tullius, T. D., Ed.; ACS Symposium Series 402;
R.; Prijs, B.; Sigel, H. J. Am. Chem. Soc. 1981, 103, 247. (s)

references therein. (t) Hodgson, D.J.In Progress in Inorganic Chemistry;
Lippard, S. J., Ed.; Wiley: New York, 1977; Vol. 23, p 211.
(2) (a) Toney, J. H.; Marks, T. J. J. Am. Chem. Soc. 1985, 107, 947.
(b) Kuo, L. Y.; Kana

^{*} To whom correspondence should be addressed. ⁺University of California at Berkeley.

University of California at Davis.

CN) with nitrogen heterocyclic compounds in organic solvents.3 Similar studies in aqueous solution with biologically important nitrogen ligands **as** mentioned above are of interest in view of the utility of Cp*Rh aqua complexes **as** anchors for single DNA molecules, in conjunction with surface microscopy techniques, for application to the human genome involving mapping and sequencing DNA bases,⁴ and as potential chemotherapeutics as well as reagents in biotechnology.^{1b,c}

Recently, we reported that 9-methyladenine formed an unusual and unprecedented cyclic trimer, $((n^5$ -Cp^{*})Rh- $(\mu_2 - \eta^1(N1))\eta^2(N6, N7)$ -9-methyladenyl)]₃(OTf)₃ (1) upon reaction in water with a $(\eta^5$ -pentamethylcyclopentadienyl)rhodium aqua complex, **2,** from pH 6 to 9.5 In that study, it was evident that the formation of the $n^2(N6, N7)$ five-

membered adenyl chelate occurred by a condensation reaction of the exocyclic NH2 group with **2,** a reactive Cp*Rh hydroxy species at pH 5-7.5*6 Similar cyclic trimer structures were also observed for adenosine,⁵ adenosine 3'-monophosphate, and the phosphate methyl ester of adenosine 5'-monophosphate in reactions with **2** in aqueous solution. 7

In order to ascertain the scope of this condensation reaction with nucleobases that have exocyclic $NH₂$ groups, we studied the reaction of the aqua complex **2,** having an empirical formula of $[Cp*Rh(H_2O)_2(OTf)_2]_x$, with 1methylcytosine (MC). We found that two different Cp*Rh 1-methylcytosine complexes could be isolated depending on the solvent media used in the reaction.

(5) Smith, D. P.; Baralt, E.; Morales, B.; Olmstead, M. M.; Maestre, M. F.; Fish, R. H. *J. Am. Chem. Soc.* 1992, *114*, 10647.

(6) (a) Kang, J. W.; Maitlis, P. M. J. Organomet. Chem. 1971, 127. (b)
Espinet, P.; Bailey, P. M.; Maitlis, P. M. J. Chem. Soc., Dalton Trans.
1979, 1542. (c) Espinet, P.; Bailey, P. M.; Piraino, P.; Maitlis, P. M.
Inorg. J. Chem. SOC., Dalton *Trans.* 1981,1997. *(0* Nutton, **A.;** Maitlis, P. M. J. *Chem. Soc.,* Dalton *Trans.* 1981,2335. (g) Nutton, A.; Maitlia, P. M. J. Chem. SOC., Dalton Trans. 1981, 2339. (h) K611e, U.; Kliiui, W. *2.* Naturforsch. 1991, 46B, 75.

(7) Smith, D. P.; Kohen, E.; Maestre, M. F.; Fish, R. H. Submitted for publication.

Reaction of **2** with 2 equiv of 1-methylcytosine in acetone for 36 h, followed by extraction with CH_2Cl_2 , gave an orange precipitate (67 *5%* **1,** complex 3, whose 'H NMR spectrum in DMSO- d_6 revealed only one set of 1-methylcytosine and Cp* resonances and considerable downfield chemical shifts for **H6** and H6 in comparison to free l-methylcytosine $(\Delta \delta(H5) = 0.25$ ppm, $\Delta \delta(H6) = 0.26$ ppm). In

addition, two broad N(4)Hz resonances were **also** shifted downfield upon coordination (7.70,8.50 ppm compared to free 1-methylcytosine at 6.91 ppm). The FAB/MS data were consistent with a monomeric species (m-nitrobenzyl alcohol; m/z 512, $[Cp*Rh(MC)(OTf)]$ and provided no evidence for a dimer. The 'H NMR data (one set of 1-methylcytosine signals for 3 down to **-90** "C in CDsOD) supports the solution structure of 3 as $[(n^5 \text{-}Cp^*) \text{R}h(n^1 \text{-}Cp^*)]$ (N^3) -MC)₂(S)](OTf)₂, where a solvent molecule $(S = H_2O)$, DMSO, CH₃OH) appears to take up the third coordination site on $(n^5$ -Cp^{*})Rh.⁸

An X-ray structural determination of compound 3 recrystallized from methanol, a weakly coordinating solvent, clearly shows, *in the* solid *state,* that one of the 1-methylcytosine ligands bonds via a four-membered-ring chelate, N3-Rh-O= $C2$ (Rh-N3a, 2.143(7) Å; Rh-O2a, 2.251(6) **A)** with the other ligand bound through the expected N3 site (Rh-N3b, 2.126(8) **A)** (Figure 1). Several structurally characterized examples of cytosine N3,02 metal semichelates exist and exhibit longer M-O bond lengths ($M = Cu$, 2.76 Å; $M = Cd$, 2.64, 2.56, and 2.89 Å; $M = Hg$, 2.84 Å)⁹ in comparison to those observed in compound 3. The structural consequences of N3,02 chelation, versus N3 coordination alone, are manifested in complex 3 by a longer C-0 bond length (C2a-O2a, 1.264 (10) **A;** C2b-O2b, 1.209(11) **A),** a shorter C2-N3 bond length (C2a-N3a, 1.350(10) A; C2b-N3b, 1.402(11) A), a

^{(3) (}a) Fish, R. H.; Kim, H.-S.; Babin, J. E.; Adams, R. D. Organometallics **1988**, 7, 2250. (b) Fish, R. H.; Baralt, E.; Kim, H.-S.
Organometallics **1991**, 10, 1965.

⁽⁴⁾ (a) Mapping **and** Sequencing the *Human* Genome; National Research Council Report, National Academy Press: Washington, DC, 1988. (b) Houseal, T. W.; Bustamante, C.; Stump, R. F.; Maestre, M. F. Biophys. J. 1989, 56, 507. (c) Zuccheri, G.; Smith, D. P.; Fish, R. H.; Maestre, **M.** F. Manuscript in preparation.

 (8) Complex 3, $[(\eta^5$ -Cp^{*})Rh(η^1 (N3)-MC)(η^2 (O2₁N3)-MC)1 (O₃SCF₃)₂: In a Vacuum Atmospheres drybox, 200 mg of $[Cp*Rh(H₂O)₂(OTf)₂$ (0.36 mmol) and 91 mg of 1-methylcytosine (MC, 0.73 mmol) were slurried for 36 h in 20 mL of acetone. The solvent of the orange solution was stripped in vacuo, and the remaining orange solid was slurried in 20 mL
of CH₂Cl₂ for 2 h and then filtered and dried to give 195 mg (67%) of complex 3. Analytically pure 3 was obtained by recrystallization from minimal methanol. ¹H NMR (DMSO-d₆, ppm): 8.52 (b, 1H, NH), 7.82 H, H5), 3.28 **(8,** 3 H, **Me),** 1.71 **(8,** 15H, Cp*). FAB/MS (m-nitrobenzyl **(d,** *JHH* * 7.4 Hz, 1 H, H6), 7.72 (b, lH, NH), 5.84 (d, *Jw* 7.4 Hz, 1 alcohol; *m/z* (relative intensity) 512.1 (52), [Cp*Rh(MC)(OTf)]; 387.0 aronoi; *m/z* (telative intensity) 512.1 (32), [Cp*Rh(MC)]; 237.0 (30), [Cp*Rh -
(18), [Cp*Rh(OTf)]; 362.1 (100), [Cp*Rh(MC)]; 237.0 (30), [Cp*Rh -
H]; 126.1 (40), [MC + H]. Anal. Calcd for RhO₈S₂F₈N₆C₂₂H₂₉: C, of $[(\eta^5$ -Cp^{*})Rh(η^1 (N3)-MC)(η^2 (N3,O2)-MC)](O₃SCF₃)₂-1.5MeOH were obtained from methanol/Et₂O solution at -30 ^oC under an inert atmosphere. Crystal data: Mo K $\alpha(\lambda = 0.71073 \text{ A})$; $T = 130 \text{ K}$; space
group $P2_1/n$; $Z = 4$; $a = 9.096(3) \text{ A}$, $b = 27.396(7) \text{ A}$, $c = 13.952(5) \text{ A}$, $\beta = 99.14(3)^o$; $0^o > 2\theta > 50^o$; 4160 observed reflections $(F$ = 99.14(3)°; 0° > 2*8* > 50°; 4160 observed reflections (*F* > 4.0 $\sigma(F)$; *R* = 0.0717 and *R_w* = 0.0745. To satisfactorily refine the structure, one of the two methanol molecules present in the crystal was modeled occupancy and the second at 50% occupancy. The structure **waa** solved by the Patterson method using SHELXTL PLUS.

^{(9) (}a) Cu: Szalda, D. J.; Kietenmacher, T. J. Acta Crystallogr. 1977, B33, 865. (b) Cd: Aoki, K.; Saenger, W. J. *Inorg. Biochem.* 1984, 20, 225. (c) Hg: Authier-Martin, M.; Beauchamp, A. L. **Can.** J. Chem. 1977,66, 1213.

Figure 1. Molecular structure of 3, $[(\eta^5$ -Cp^{*})Rh $(\eta^1$ (N3)-MC)- $(\eta^2(O2, N3)$ -MC)](OTf)₂-1.5MeOH, with atoms shown as 50% ellipsoids. Only the cation is shown for clarity. Selected bond lengths (Å) and angles (deg): Rh-N3a, 2.143 (7); Rh-N3b, 2.126 *(8);* Rh-OQa, 2.251 (6); N3a-Rh-NSb, 90.0 (3); N3a-Rh-Oaa, 61.0 (2); 02a-Rh-N3b, 88.0 (2).

shorter C4-N4 bond length (N4a-C4a, 1.308A; N4b-C4b, 1.349(13) **A),** and a smaller 02-C2-N3 bond angle (02a-C2a-N3a, 117.1(7)°; O2b-C2b-N3b, 121.6(8)°). The N4a---02b through-space distance (3.175 **A)** precludes any intramolecular hydrogen bonding between these sites.

Upon recrystallization of complex 3 from H_2O (pH 5.1), anew complex, **4,** is formed. Complex **4** can **also** be isolated by the dropwise addition of a deoxygenated, aqueous solution of 1 equiv of l-methylcytosine to an aqueous solution of **2** that was adjusted to pH *5-6* by addition of NaOH. The ¹H NMR spectrum of 4 in DMSO- d_6 showed upfield chemical **shifta** for H5 (5.27 ppm) and H6 (7.45 ppm) of 0.32 and 0.09 ppm, respectively, in comparison to free 1-methylcytosine, while one set of exocyclic NH₂ protons at 7.18 ppm was shifted downfield by 0.27 ppm. The other exocyclic NH₂ signal was not observed and is apparently broadened into the **base** line. FABIMS verified the dimeric nature of **4** (m-nitrobenzyl alcohol; *mlz* 766, $[(Cp*Rh)₂(MC)(\mu-OH)](OTf); m/z 659, [Cp*Rh(\mu-OH)]₂$ -(OTf), while the single-crystal X-ray analysis confirmed the trans stereochemistry and the extensive intramolecular hydrogen bonding of the bridging hydroxyl groups with the C= O and the exocyclic NH₂ groups (Figure 2).¹⁰ This hydrogen-bonding network creates a hydrophobic environment around the metal centers and appears to be the reason that 4 is insoluble in H_2O at pH 5.1.

The core structure of 4 is quite similar to cis^{-3a} and $trans\{-\frac{(n^5-Cp^*)Rh(n^1-(N))(\mu\cdot OH)}{2^{2+11}}$ dimeric complexes with heteroaromatic nitrogen ligands (cis stereochemistry,

a

b

Figure 2. (a, upper view) Molecular structure of **4,** *trans-* $[(\eta^5$ -Cp^{*})Rh($\eta^1(N3)$ -MC)(μ -OH)]₂(O₃SCF₃)₂, with atoms as *50%* ellipsoids. Selected bond length **(A) and** anglee **(deg):** Rh-N3,2.181(6); Rh-O10,2.138(5); Rh-OlO', 2.118(6); C(2)- 0(2), 1.232(10); C(4)-N(4), 1.32(9); H4b-O1O', 1.93(1); H1& 02,1.96(1); N3-Rh-010,87.9(2); Rh-Ol&Rh', 101.2(2); **N(4)-** $C(4)-N(3), 117.7(6); O(2)-C(2)-N(3), 123.1(6); N4-H4-O10'$ 149(1); 010-H10-02, 155(1). (b, lower view) Structure of the $[Rh(\mu\text{-}OH)(MC)]_2$ core emphasizing the intramolecular hydrogen bonding.

N = quinoline, **5;** trans stereochemistry, N = pyridine, **6);** however, some slight differences are apparent. **The** planar Rhz(p-OH)z fragment of **4** shows a wider **0-Rh-0** angle of $78.8(2)$ ^o in comparison to $5(75.1(1),74.8(1)$ ^o) and $6(76.8 (2)^\circ$). As well, a smaller Rh-O-Rh angle $(101.2(2)^\circ)$ for **⁴**was observed in comparison to 104.7(1) and 105.0(1)0 for 5 and $103.1(2)$ ^o for 6. This results in a closer Rh-Rh through-space distance of 3.290(2) **A** for **4** in comparison to 3.322(1) **A** for **5** and 3.308(1) **A** for **6.**

This slight deformation is presumably caused by the unusual binding of the 1-methylcytosine in which the **(Rh**-OH12 core *act8* **08** *a covalent electrophile, a hydrogen bond donor, and a hydrogen bond acceptor.* **This system is a** rare example of a complex in which a μ -hydroxy ligand exhibits simultaneous hydrogen bond donor and acceptor capabilities." Although other metal complexes of **1** methylcytosine have shown extensive *intermolecular*

 (10) Complex **4,** $[(\eta^5$ -Cp*Rh $)(\eta^1(N3)$ -MC $)(\mu$ -OH $)]_2(O_3SCF_3)_2$ -4H₂O: Complex **3 (43** *mg)* waa diseolved in **5 mL** of deoxygenated water, and the solution wan **stirred** for **36** h, resulting in precipitation of a yellow-orange solid. The volume of the reaction **mixture** waa reduced *to* **ca. 2** mL, and the orange precipitate (23 mg, 81%) was collected by filtration and dried
for 18 h under vacuum. 'H NMR (DMSO-d₆, ppm): 7.45 (H6, d, J_{HH} = dono)
7.4 Hz, 1 H); 7.18 (NH, b, 1H), 5.27 (H5, d, J_{HH} = 7.4 Hz, 1 H); 3.18 **s,3** H); **1.78** (Cp*,s, **1SH). FAB/MS** (m-nitrobenzyl alcohol; *m/z* (relative intensity)): **766 (lo),** [(Cp*Rh(p-OH))z(MC)(OM - HzOI; **659 (23),** mensity): $(16P+Rh(\mu\cdot OH))_2(M\cdot U)(1) - H_2O_1; 639 (23),$
 $[(Cp+Rh(\mu\cdot OH))_2(OTf)]$; $641 (13), [(Cp+Rh(\mu\cdot OH))_2(OTf)-H_2O]$; 512
 $(20), [Cp+Rh(MC)(OTf)]$; 362 (82), $[Cp+Rh(MC)]$; 237 (25), $[Cp+Rh-
H]$; 149 (100), [OTf]; 126 (30), $[MC + H]$. Anal. Calcd 4.40; N, 7.82. Orange parallelepipeds of 4 were obtained from water at
room temperature. Crystal data: Mo Ka ($\lambda = 0.71073$ Å); $T = 130$ K;
space group $P2_1/n$; $Z = 2$; $a = 8.077(2)$ Å, $b = 19.541(4)$ Å, $c = 13.508(3)$
Å, *R.* = **0.0480.** Thestmcturewanwlved **bydirectmethodausingSHELXTL** PLUS. Hydrogen atoms bonded to carbon were located from a difference map and subsequently refined using a riding model with $C-H = 0.96$ Å and isotropic *0* values **equal** *to* **1.2 times** the equivalent isotropic *U* of

the bonded **carbon.** The **amino** and hydroxy hydrogens were **alro** located of 0.94(2) and 0.92(2) Å, respectively, and fixed isotropic U 's but were otherwise not constrained. A difference map was featureless.

⁽¹¹⁾ Lahoz, F. G.; Carmona, D.; Oro, L. A.; Lamata, M. P.; Puebla, M.
P.; Foces-Foces, C.; Cano, F. H. J. Organomet. Chem. 1986, 316, 221.
(12) Liu, F.; Roesky, H. R.; Schmidt, H.-G.; Noltemeyer, M. Orga*nometallic8* **1992,** *11,* **2986.**

hydrogen bonding,13 we are unaware of a system which recognizes the bonding capabilities of this ligand so readily in an intramolecular fashion. In addition, the formation Of **4** provides some evidence for the structure of the reactive aqua complex **2** that corroborates the FAB/MS data, obtained previously, indicating a dimeric $[Cp*Rh(\mu-OH)]_2$ cationic complex (pH dependent). 5

Condensation reactions between the exocyclic $NH₂$ of 1-methylcytosine and **M-OH** centers to form either fourmembered-ring chelates $(\eta^2(N3,N4); Cp_2Mo^{2+},Pt^IV)$ or μ -1methylcytosyl complexes (Pt^{II}) are well documented.^{2d,13a,13c} However, we were not able to induce a similar condensation reaction between the exocyclic $NH₂$ and the μ -OH group with 4. Mild thermolysis (70 \degree C for 16 h) of 4 in DMSO- d_6 solutions results in overall decomposition with no evidence of a condensation reaction. The fact that we observe no apparent condensation reaction of the exocyclic $NH₂$ group with the bridging OH group could be indicative of the pronounced stability of the extensive intramolecular hydrogen-bonding regime shown in **4** or simply a manifeatation of the instability of a four-membered-ring chelate. Thermolysis of 3 under the same conditions gave a similar result, while heating 3 in acetone- d_6 at 50 °C for 24 h showed no reaction.

In future publications, we will report on the reactivity of **2** with other nucleobases, nucleotides, and sequencespecific oligonucleotides **as** well **as** ita utility **as** a tether, simultaneously bonding to both glass or electrode surfaces and sequence-specific dfgonucleotide-single DNA molecules, for eventual application to sequence and map the human genome.^{4c}

Acknowledgment. The studies at LBL were generously supported by Laboratory Directed Research and Development Funds aqd the Department of Energy under Contract No. DE-AC03-76SF00098. We thank Dr. Mina J. Bissell of LBL for her support of this project.

Supplementary Material Available: Tables of crystaldata, atomic coordinates and isotropic dieplacement coefficienta, bond lengths, bond angles, and anisotropic displacement coefficienta for 3 and 4 (18 pages). Ordering information is given on any current masthead page.

OM920820N

⁽¹³⁾ (a) Faggiani, *R;* **Lippert, B.; Lock, C.** J. **L.; Speranzini, R. A.** *J. Am. Chem. SOC.* **1981,103,1111. (b) Beyerle-Pfnur, R.; Schollhorn, H.; Thewalt, U.; Lippert, B.** *J. Chem. SOC., Chem. Commun.* **1985,1510. (c)** Schollhorn, H.; Beyerle-Pfnur, R.; Thewalt, U.; Lippert, B. J. Am. Chem. *SOC.* **1986,108,3680. (d) Lippert, B. In** *Progress in Inorganic Chemistry;* **Lippard,** S. J., **Ed.; Wiley: New York, 1989; Vol. 37, p 1, and references therein.**