$^{13}\mathrm{C}{^{-119}}\mathrm{Sn}$ Correlation NMR in Solution and Solid-State CP/MAS NMR of Bis(tributylstannyl(IV)) O-5,6-Isopropylidene-L-(+)-ascorbate[†]

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Summary: Various NMR correlation techniques, ¹H-¹³C HMQC, ${}^{I}H^{-13}C$ HMBC, ${}^{1}H^{-119}Sn$ HMQC with gradient enhancement, and 13C-119Sn HMQC, enabled a full resonance assignment and the solution structure elucidation of bis(tributylstannyl(IV)) L-(+)-ascorbate (1) and O-5,6-isopropylidene-L-(+)-ascorbate (2) in deuteriochloroform. Double heteronuclear 13C-119Sn HMQC spectroscopy appeared indispensable for discriminating between ${}^{12}J({}^{13}C - O - {}^{119}Sn)$ and ${}^{3}J({}^{13}C = C - O - {}^{119}Sn)$ coupling pathways involving quaternary carbon atoms and provided useful information about the relative magnitudes of ${}^2J({}^{13}C-{}^{119}Sn)$ and ${}^3J({}^{13}C-{}^{119}Sn)$ coupling constants, as they turned out to have an uncommon coupling size sequence. While the tin atoms in compounds 1 and 2 are four-coordinate in solution, 117Sn CP/MAS NMR reveals that compound 2 has one four-coordinate and one fivecoordinate tin atom in the solid state.

Introduction

Organotin derivatives of bio-organic compounds have attracted growing interest because of their biological properties.1 This raises the issue of their detailed structural elucidation using multinuclear 2D NMR techniques in solution, as recently illustrated for a tin derivative of erythromycin A.2 Tin derivatives of another natural compound of interest, ascorbic acid, have been recently reported by Casas et al.,3 who investigated NMR spectra of dimethyltin(IV) and dibutyltin(IV) L-(+)-ascorbates in aqueous solutions and in the solid state.

This study reports the synthesis and structure determination in solution and solid state of two novel tincontaining derivatives of ascorbic acid, i.e., bis(tributylstannyl(IV)) L-(+)-ascorbate (1) and bis(tributylstannyl(IV)) O-5,6-isopropylidene L-(+)-ascorbate (2) (Scheme 1).

We present a full resonance and structure assignment strategy using 13C-119Sn correlation NMR as an essential tool in solution as well as ¹³C and ¹¹⁷Sn CP/MAS NMR for the solid state. Double heteronuclear ¹³C-¹¹⁹Sn correlation NMR, so far only described once by Berger and Mitchell,4 turned out necessary to discriminate between two pairs of ${}^2J({}^{13}C-O-{}^{119}Sn)$ and ${}^3J({}^{13}C=$ C-O-119Sn) coupling pathways involving quaternary carbon atoms. It enabled us to obtain interesting information with regards to the relative magnitudes of $^{2}J(^{13}C-^{119}Sn)$ and $^{3}J(^{13}C-^{119}Sn)$ coupling constants which are of relevance to structural organotin chemistry. ¹³C and ¹¹⁷Sn CP/MAS NMR experiments provide information on the coordination at tin in the solid state of compound 2, which, unfortunately, appeared unsuitable for X-ray diffraction analysis.

Experimental Section

Syntheses. Bis(tributylstannyl(IV)) L-(+)-Ascorbate (1). To Bu_3SnOCH_3 (1.85 g, 5.76 mmol)⁵ in 10 mL of dry ethanol in a 50 mL Schlenk tube was added L-(+)ascorbic acid (0.507 g, 2.88 mmol) in 20 mL of dry ethanol. The mixture was heated under argon atmosphere at 80 °C until the acid

Dedicated to Prof. Dr. K.-H. Thiele on the occasion of his 70th birthday

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was completely dissolved. Subsequently, the volume of the solution was reduced to 50% by distillation under normal pressure. The residual solvent was removed by distillation in vacuo. The product was a yellowish liquid that darkens upon standing at room temperature. Yield of isolated product: 1.92 g (88%). Anal. Found: C, 47.36; H, 7.78; Sn, 31.44. Calcd for $C_{30}H_{60}O_6Sn_2$: C, 47.78; H, 8.02; Sn, 31.48.

Bis(tributyIstannyl(IV)) *O***-5,6-Isopropylidene-L-(+)-ascorbate (2).** Compound **2** was prepared analogously to compound **1** from 1.41 g (4.39 mmol) of Bu₃SnOCH₃ and *O***-5**,6-isopropylidene-L-(+)ascorbic acid (0.475 g, 2.20 mmol). A white solid was obtained. Yield of isolated product: 1.77 g (86%). Mp: 57-59 °C. Anal. Found: C, 49.79; H, 8.20; Sn, 30.23. Calcd for $C_{33}H_{64}O_6Sn_2$: C, 49.90; H, 8.12; Sn, 29.89.

NMR Measurements. The 1 H (360.13 MHz), 13 C (90.56 MHz), and 119 Sn (134.35 MHz) NMR spectra were recorded at 300 K from deuteriochloroform solutions on a Bruker AMX 360 spectrometer equipped with a 5 mm broadband probe and a 5 mm broadband inverse probe with z-shielding. 1 H and 13 C chemical shifts were referred to internal Si(CH₃)₄ ($\delta = 0.00$), and 119 Sn chemical shifts were referred to external neat Sn-(CH₃)₄ placed in a coaxial capillary ($\delta = 0.00$).

The 13 C (125.79 MHz) $^{-119}$ Sn (186.53 MHz) HMQC spectrum⁴ was recorded at 303 K on a Bruker AMX 500 instrument equipped with a Silicon Graphics SGIO2 computer and a BSMS digital lock, using a 13 C spectral width of 190 ppm and a 119 Sn spectral width of 12 ppm, with appropriate folding of the low-frequency 119 Sn resonance (SnB) along F_1 in order to improve digital resolution. Eight FIDs of 16K data points were recorded with 3840 scans and a recycling delay of 10 s. The delay for heteronuclear coupling evolution was set to 28 ms. 1 H decoupling was applied during acquisition. Processing consisted of zero-filling in F_1 to 32 K (no zero-filling in F_2), followed by a sine-bell multiplication and magnitude FT calculation.

The gradient enhanced $^1H-^{13}C$ HMQC, $^1H-^{13}C$ HMBC, and $^1H-^{119}Sn$ HMQC 2D spectra were recorded as explained previously. 6

¹³C (62.90 MHz) and ¹¹⁷Sn (89.13 MHz) CP/MAS NMR spectra of compound 2 were recorded on a Bruker DRX 250 spectrometer. Compound 2 was packed in a standard 4 mm ZrO₂ rotor. The ¹¹⁷Sn Hartmann-Hahn cross-polarization match was set with tetracyclohexyltin using a 1H 90° pulse of $5 \mu s$. Cross-polarization contact time and recycling delay were set to 2 ms and 4 s, respectively. Two spinning rates (5 and 9 kHz) were used to identify the isotropic chemical shift. The number of scans varied between 200 and 800. 117Sn chemical shifts were referred to tetracyclohexyltin ($\delta = -$ 97.35) taken as a secondary reference. The principal components of the $^{117}\mbox{Sn}$ shielding tensors were analyzed according to Herzfeld and Berger⁷ using WINFIT software.⁸ They are reported, following Haeberlen's notation,⁹ as the isotropic chemical shift $[\delta_{iso}]$ $-\sigma_{\rm iso} = -(\sigma_{11} + \sigma_{22} + \sigma_{33})/3$] (ppm), the anisotropy ($\zeta = \sigma_{33} - \sigma_{33} - \sigma_{33}$ $\sigma_{\rm iso}$), and the asymmetry ($\eta = |\sigma_{22} - \sigma_{11}|/|\sigma_{33} - \sigma_{\rm iso}|$), σ_{11} , σ_{22} , and σ_{33} being the three components of the shielding tensor expressed in its principal axis system with the following rule: $|\sigma_{33} - \sigma_{iso}| \geq |\sigma_{11} - \sigma_{iso}| \geq |\sigma_{22} - \sigma_{iso}|$. With this convention, ζ is signed and expressed in ppm.

Results and Discussion

The complete, final $^{117/119}$ Sn and 13 C resonance assignments for compounds **1** and **2**, including scalar coupling data, are given in Table 1.

Table 1. ¹¹⁹Sn and ¹³C Chemical Shifts (ppm) and ⁿJ(¹¹⁹Sn-¹³C) Coupling Constants^a (values in Hz) for Compounds 1 and 2 in CDCl₃ and/or the Solid State

	1 (CDCl ₃)	2 (solid)	2 (CDCl ₃)
C1	174.7	178.5	174.7
	$^{3}J = 9$		$^{3}J = 9$
C2	122.5	120.4	122.8
	$^{3}J = 9$; $^{2}J = 23$		$^{3}J = 9$; $^{2}J = 23$
C3	155.3	164.3	155.2
	$^{3}J = 12; ^{2}J = 29$		$^3J = 12; ^2J = 26$
C4	77.9	75.4	77.1
	$^{3}J = 9$	or 74.3	$^{3}J = 9$
C5	72.1	74.3	76.1
		or 75.4	
C6	63.4	65.5	65.5
C7		115.0	109.3
C8, C9 b		$26.7 - 28.5^{e}$	26.2, 25.7
C1'c	$17.8 \ (^{1}J = 364)$	$17.1 - 21.2^f$	$18.0 \ (^{1}J = 365)$
$C1''^d$	$18.0 \ (^{1}J = 349)$	$17.1 - 21.2^f$	$18.0 \ (^{1}J = 350)$
$C2'^c$	$27.7 (^2J = 18)$	$26.7 - 28.5^{e}$	$27.8 \ (^2J = 18)$
$C2''^d$	$27.5 (^2J = 20)$	$26.7 - 28.5^{e}$	$27.6 \ (^2J = 21)$
C3'c	$26.9 (^3J = 64)$	$26.7 - 28.5^{e}$	$27.0 (^{3}J = 64)$
$C3''^d$	$27.0 (^{3}J = 66)$	$26.7 - 28.5^{e}$	$26.9 (^3J = 68)$
C4', C4"	13.5	12.6 - 14.4	13.5
¹¹⁹ Sn	123.6 (SnB)	112.0 (SnB)	118.6 (SnB)
2.1	152.5 (SnA)	18.1 (SnA)	146.0 (SnA)
	102.0 (51111)	10.1 (51111)	110.0 (51111)

 a $^nJ=^nJ(^{13}C-^{119}Sn)$ $(n=1,\ 2,\ or\ 3).$ b Diastereotopic methyl groups. c Bu $_3$ SnB. d Bu $_3$ SnA. e Overlapping ranges of ^{13}C signals for C8, C9, C2′, C2″, C3′, C3″. f Overlapping ranges of ^{13}C signals for C1′ and C1″.

The solution assignment, as explained below for **2**, is extrapolated to **1**, on the basis of their very similar NMR data, while the solid-state ¹³C assignment for **2** (**1** is a liquid) is based on a comparison of ¹³C data in solution and solid states.

The assignments are pairwise not evident for the olefinic quaternary carbons C2 and C3, the tertiary carbons C4 and C5, and the tin atoms SnA and SnB. Furthermore, the ¹³C assignment of the butyl carbons to the tin atom to which the butyl group is bound is likewise not straightforward. Compound 2 poses the question of the correct assignment of the ²J(¹³C-O- 119 Sn) and $^{3}J(^{13}C=C-O-^{119}Sn)$ coupling pathways for the quaternary carbons C2 and C3 to the tin atoms SnB and SnA to which they are respectively linked through a bridging oxygen atom. While it is well recognized 10 that usually ²J(¹³C-¹¹⁹Sn) coupling constants are smaller than ${}^{3}J({}^{13}\text{\r{C}}-{}^{119}\text{Sn})$ ones in aliphatic carbon chains, this rule is far from being applicable without risk to errors to the ${}^{2}J({}^{13}C-O-{}^{119}Sn)$ and ${}^{3}J({}^{13}C=C-O-{}^{119}Sn)$ coupling pathways.

Compound 2 displays two well-separated and narrow ¹¹⁹Sn resonances at 146.0 and 118.6 ppm. Also in the ¹³C spectrum, two sets of signals are associated with all carbons of the two Bu₃Sn groups (except for the methyl ones; Figure 1a).

The ¹H NMR signals of the butyl group protons are not sufficiently separated to be assigned by a 2D ¹H–¹³C HMQC spectrum. By contrast, the characteristic ¹H4 doublet, appearing at 4.32 ppm, well separated from the ¹H5 and ¹H6 multiplets around 4.0 ppm, allows the identification of the ¹³C4 resonance through their ¹H–¹³C HMQC cross-peak, the identification of the ¹³C5 resonance then following logically. The ¹³C6 resonance is easily identified from its different DEPT pattern.

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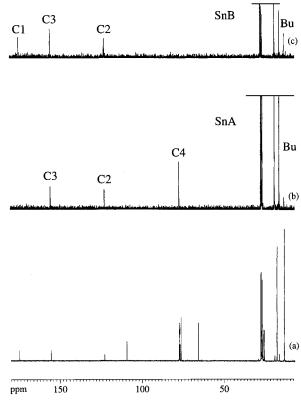


Figure 1. (a) ¹³C NMR spectrum of compound 2. (b) Crosssection, parallel to the ¹³C axis, of its 2D ¹³C-¹¹⁹Sn HMQC spectrum at the ¹¹⁹Sn resonance frequency of tin atom SnA (146.0 ppm) in F₁. (c) Cross-section of the 2D ¹³C-¹¹⁹Sn HMQC spectrum at the 119Sn resonance frequency of tin atom SnB (118.6 ppm).

The application of a 2D ¹H-¹¹⁹Sn heteronuclear correlation^{6,11} allows the ¹¹⁹Sn resonances to be identified. Thus the high-frequency 119Sn resonance at 146.0 ppm displays an intense correlation cross-peak with the ¹H4 resonance and a weaker one with the ¹H5 resonance; by contrast the low-frequency 119Sn resonance at 118.6 ppm displays only a weak cross-peak with the ¹H4 resonance and none with the ¹H5 one. This assigns the ¹¹⁹Sn resonance at 146.0 ppm to the tin atom SnA bound to carbon C3, the one at 118.6 ppm to the SnB tin atom bound to carbon C2, in line with a prediction resulting from the fact that the ¹¹⁹Sn NMR resonance is high-frequency shifted for the tin atom bound to the oxygen of the more acidic starting acid (p $K_A(C3OH)$ = 4.03; p $K_A(C2OH) = 11.3$). The ¹¹⁹Sn resonance assignment being achieved, the ¹³C resonances of the butyl groups are then easily assigned to their respective tin atoms through their ${}^{n}J(^{119}Sn-^{13}C)$ coupling satellites, which are well visible at the feet of the ¹¹⁹Sn resonances in the standard ¹¹⁹Sn spectrum.

Identification of the 13C2 and 13C3 resonances is possible from the 2D $^{1}H-^{13}C$ HMBC spectrum. The ^{13}C resonance at 155.2 ppm displays ¹H-¹³C HMBC crosspeaks with the ¹H4 and ¹H5 resonances, while, by contrast, that at 122.8 ppm correlates solely with the ¹H4 resonance. The ¹³C resonances at 155.2 and 122.8 ppm are therefore assigned to the C3 and C2 carbon

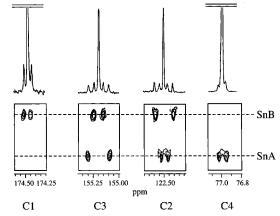


Figure 2. Expansions, at the resonance frequencies of the ¹³C resonances of carbons C1, C3, C2, and C4 along F₂, of the 2D contour plot of the ¹³C-¹¹⁹Sn HMQC spectrum of compound 2; the ¹¹⁹Sn resonance chemical shift scales along F₁ are not given because the signal SnB does not appear at its actual chemical shift, having been folded into F₁ (see Experimental Section).

atoms, respectively, in accordance with ¹³C literature data¹² on β , γ -olefinic ketones. The ¹³C2 and ¹³C3 resonances both display two pairs of unresolved ⁿJ-(13C-119/117Sn) satellites amounting to ca. 9 and 25 Hz, arising all from ${}^{2}J({}^{13}C-O-{}^{119/117}Sn)$ and ${}^{3}J({}^{13}C=C-O-{}^{119/117}Sn)$ ^{119/117}Sn) coupling pathways involving both SnA and SnB atoms.

None of the 2D correlation techniques used above enables one to unambiguously discriminate between the ²J and ³J couplings, unless empirical assumptions are formulated as to their relative sizes. 10 This issue can be addressed most efficiently by a 13C-119Sn double heteronuclear HMQC experiment. 4 In addition, one such single experiment enables one to check the selfconsistency of the ¹³C and ¹¹⁹Sn assignments proposed above from several 2D experiments.

Although it is customary to use the most sensitive nucleus for detection in a X-Y correlation, 13 we decided to detect ¹³C nuclei for the following reasons: (i) ¹³C resonances are commonly much sharper than those of ¹¹⁹Sn, while we wanted to detect quite small coupling constants; (ii) the ¹¹⁹Sn spectral window is narrower, in the present case, than the ¹³C one, and, moreover, a folding can be used to minimize the 2D data matrix at a given digital resolution in the F₁ (119Sn) dimension, since only two 119Sn resonances are present. The preparation delay was set to the compromise ${}^{n}J({}^{13}C-$ ¹¹⁹Sn) coupling value of 18 Hz.

Figure 1b,c shows the two-row cross-sections of the 2D ¹³C-¹¹⁹Sn HMQC spectrum at both ¹¹⁹Sn resonance frequencies, confirming the above assignments of both the SnA/SnB and the C2/C3 atom pairs. Figure 2 shows an expansion of the contour plots obtained in the 2D ¹³C-¹¹⁹Sn HMQC spectrum for the ¹³C2 and ¹³C3 resonances of interest, as well as for the 13C1 and 13C4 ones, the only ones of the ascorbate moiety to show ¹³C-¹¹⁹Sn HMQC correlations.

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The cross-peak pairs associated with the 119SnB resonance show that the ¹³C2-¹¹⁹SnB correlation doublet exhibits a larger scalar coupling splitting than the ¹³C3⁻¹¹⁹SnB doublet, which unambiguously demonstrates that the ²J(¹³C2-O-¹¹⁹SnB) coupling constant is larger than the ${}^{3}J({}^{13}C3=C2-O-{}^{119}SnB)$ one. Analogously, the cross-peak pairs associated with the 119SnA resonance show that the ¹³C3-¹¹⁹SnA correlation doublet exhibits a larger scalar coupling splitting than the ¹³C2-¹¹⁹SnA doublet, which again unambiguously demonstrates that the ²J(¹³C3-O-¹¹⁹SnA) coupling constant is larger than the ${}^{3}J({}^{13}C2=C3-O-{}^{119}SnA)$ one. This uncommon coupling size sequence¹⁰ for ²J(¹³C-¹¹⁹Sn) and ${}^{3}J({}^{13}C - {}^{119}Sn)$ coupling constants is worth stressing, because it can be of prime importance in resonance assignments as well as solution structure elucidations of organotin compounds in general.

The ^{117}Sn CP/MAS NMR spectrum of compound **2** (SnB $\delta_{iso}=112.0,\,\sigma_{11}=\sigma_{22}=-95.2$ and $\sigma_{33}=-145.7,\,\eta=0,\,\zeta=33.7;$ SnA $\delta_{iso}=18.1,\,\sigma_{11}=\sigma_{22}=74.5$ and $\sigma_{33}=-203.2,\,\eta=0,\,\zeta=185.1)$ reveals that SnB is four-coordinate, while SnA is five-coordinate in the solid state. This can be explained by an intermolecular coordination involving the carbonyl oxygen and the C3–O–SnA tin atom, as illustrated in Scheme 2.

Further evidence for the existence of such an intermolecular coordination is found in the ¹³C chemical shift differences between the solution and solid states which are largest at the level of the C3 (respectively 155.2 vs 164.3 ppm) and C=O (respectively 174.7 vs 178.5 ppm) carbon atoms. An alternative explanation, where the O1 carbonyl oxygen would coordinate the SnB atom, either intra- or intermolecularly, cannot explain satisfactorily the dramatic ¹³C chemical shift change at C3.

Conclusion

The solution-state structure of compounds $\bf 2$ and $\bf 1$ was unambiguously elucidated by various 2D correlation techniques. Especially the $^{13}C-^{119}Sn$ HMQC experiment

Scheme 2

proved to be indispensable for discriminating between ${}^2J(^{13}C^{-119}Sn)$ and ${}^3J(^{13}C^{-119}Sn)$ coupling pathways involving quaternary carbon atoms, revealing that the sequence ${}^3J(^{13}C^{-119}Sn) > {}^2J(^{13}C^{-119}Sn)$ usually observed in aliphatic chains 10 is not applicable to ${}^2J(^{13}C^{-119}Sn)$ and ${}^3J(^{13}C=C^{-119}Sn)$ coupling pathways.

From the five-coordination of only one tin atom, together with the ¹³C chemical shift differences between solution and solid states, an intermolecular coordination between SnA and the carbonyl oxygen is proposed for compound **2** in the solid state.

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Supporting Information Available: ¹¹⁷Sn CP/MAS NMR spectrum of **2**. This material is available free of charge via the Internet at http://pubs.acs.org.

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