57Fe NMR of Ferrocenes by 1H-**57Fe INEPT Techniques**

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Summary: 57Fe NMR spectra (11.66 MHz) of the sandwich complexes ferrocene (1), butylferrocene (2), and acetylferrocene (3) were measured at natural abundance of 57Fe by application of the polarization transfer technique (INEPT; 1H-*57Fe). In contrast to previous assumptions this technique works very well, although it has to be based on rather small (*∼*0.3*-*0.7 Hz) longrange scalar 57Fe*-*1H couplings across two bonds. Thus, the 57Fe NMR signal of ferrocene (*∼*150 mg in 2 mL of CDCl3/hexane (1:1)) was observed already after 4 transients, and after 400 transients (70 min of spectrometer time)* the ¹³C satellites $(^{1}J(^{57}Fe,^{13}C) = 4.7$ *Hz*) were *detected, allowing us to determine the isotope-induced chemical shift* $\frac{1}{4}$ ^{12/13}C(⁵⁷Fe) = -440 ppb at natural *abundance of the isotopes.*

After 50 years of intensive research, there is a huge family of complexes derived from ferrocene, FeCp₂, and the attraction of ferrocene chemistry has even increased in recent years, considering the role of ferrocenes in materials science.¹ Although the principal structural features of ferrocene derivatives are well-known, in particular in the solid state from X-ray structural analysis and in solution from ${}^{1}H$, ${}^{13}C$, and ${}^{31}P$ NMR (if appropriate) spectroscopy, more precise information on their electronic structures is still desirable. In this context, $57Fe$ NMR spectroscopy² can provide useful data, as has been shown for ferrocene derivatives, $2-5$ for CpFe half-sandwich compounds,6-⁹ and for *η*4-diene iron complexes.10,11 There is also increasing interest in applying 57Fe NMR to the study of biologically relevant iron-containing systems.^{12,13} Recently we have shown⁵

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that improvements in measuring 57Fe NMR signals of ferrocenes directly by single-pulse techniques at natural abundance promise a brighter future of 57Fe NMR as compared with the gloomy picture drawn originally.^{2-4,6-10} These improvements can be traced to better NMR instrumentation (e.g., higher field strengths, B_0 , and improved hardware such as frequency generation, amplifiers, and probe head design) and also to optimized experimental parameters.⁵ Here we report on the application of polarization transfer (PT) techniques $(1H-57Fe)$ to ferrocenes $(1-3)$.

The inherently unfavorable NMR properties of the ⁵⁷Fe nucleus ($I = \frac{1}{2}$; natural abundance 2.2%; receptivity with respect to $^{13}\mathrm{C}$ 4.19 \times $10^{-3})$ clearly indicate that direct observation of 57Fe NMR signals by single-pulse techniques will never be easy, even under the best experimental conditions. It had been shown already in the 1970s that 57Fe-enriched complexes can be studied by heteronuclear ${}^{13}C\{ {}^{57}Fe}$ double-resonance experiments.^{2,14} At natural abundance of ⁵⁷Fe, indirect detection via ${}^{1}H$ or ${}^{31}P^{2,8,9}$ appears to be now the most attractive alternative, which, however, requires a sizable scalar coupling between ⁵⁷Fe and ¹H or ³¹P, present only in complexes containing an Fe-H or Fe-P bond. In contrast, it appears that the inverse detection also works for $(Cp)Fe(CO)_2R$ with ^{2,3} $J(57Fe,{}^{1}H_R) \approx 1-2$ Hz.⁹ Another alternative would be the application of polarization transfer (PT) techniques $(^1H-^{57}Fe)$ such as INEPT15 or DEPT,16 leading to an improvement of the signal-to-noise ratio by a factor of ∼30 with respect to single pulse detection. However, it has been pointed out² that the application of PT techniques may not be feasible in the absence of Fe-H bonds, owing to the small magnitude of the coupling constants ^{*nJ*(57}Fe,¹H)

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Figure 1. 11.66 MHz ⁵⁷Fe NMR spectra (INEPT;¹² based on delays according to $^{2}J(5^{7}Fe,{}^{1}H) = 0.5$ Hz; measured at 23 ± 1 °C (assumed accuracy of temperature inside of the sample, controlled to 23 ± 0.2 °C outside of the sample)) of ferrocene, Cp₂Fe (saturated solution (∼150 mg) in CDCl₃/ hexane (∼1:1)). The delays in the INEPT pulse sequence were adjusted to $^2J(5^7Fe, ^1H) = 0.5 Hz$ (instead of the experimental value 0.4 ± 0.1 Hz) in order to compensate for pulse durations and dead times due to the hardware. Part A (refocused with optimized low-power ¹H composite pulse decoupling): acquisition time 3 s; repetition time 11 s; line broadening 0.2 Hz; 8 transients. The reduced intensity of the 57Fe NMR signal when compared with the spectrum in (B) indicates that some 57Fe magnetization is lost during the refocusing periods (0.22 s). Part B (basic pulse sequence; 1H nondecoupled): acquisition time 3 s; repetition time 8 s; line broadening 0.2 Hz; 8 transients.

 $(n > 1)$ and the concomitant loss of ¹H and ⁵⁷Fe magnetization during the pulse sequence.

We show now for ferrocene $(1: \delta^{57}Fe)$ 1538.4, in CDCl₃ without hexane δ ⁽⁵⁷Fe) = +1541.7 ⁵), butylferrocene $(2; \delta^{57}Fe)$ 1570.8), and acetylferrocene $(3; \delta^{57}Fe)$ δ ⁽⁵⁷Fe) 1774.4) that this assumption was misleading. It has already been shown for $CpFe(CO)_2R$ that small coupling constants, in these particular cases $^{2,3}J^{(57}\text{Fe},^{1}\text{H}_{\text{R}})$, can be used for indirect detection of the ⁵⁷Fe resonance via ¹H NMR.⁹ Polarization transfer appears to work well for many ferrocene derivatives, and meaningful spectra can be acquired within a very short time (see Figures 1 and 2). Thus, the ⁵⁷Fe NMR signal of ferrocene (~150 mg in 2 mL of CDCl₃/hexane (1:1)) can easily be observed already after four or eight transients (spectrometer time <100 s). The same signal, complete with its ${}^{13}C$ satellites, was measured after 70 min (Figure 2), allowing us to determine ${}^{1}J(5^{7}Fe, {}^{13}C) =$ 4.7 Hz, in agreement with data from the 13C NMR spectra of ⁵⁷Fe-enriched ferrocene,¹⁴ and the previously unknown isotope-induced chemical shift ${}^{1}\Delta(1^{2/13}C(5^{7}Fe))$ $=$ -440 ppb at natural abundance of both isotopes. Considering the different nature of the $Fe-C$ bonds in ferrocene and of the $Fe-C_{Me}$ bond in $(Cp)Fe(CO)_2Me$, it is interesting to note that in the latter a similar value, $1\Delta(12/13C_{Me}(57Fe)) = -471$ ppb, has been measured for the η^1 -linked methyl group.⁹ The magnitude of the coupling constants $^{2}J(^{57}Fe,^{1}H)$ used for the calculation of the delays in the pulse sequences was assumed to be on the order of 0.3-0.7 Hz (best results were obtained for **1** with 0.5 Hz, for **2** with 0.3 Hz, and for **3**

Figure 2. 11.66 MHz ⁵⁷Fe NMR spectra (INEPT,¹² refocused and ¹H decoupled; based on $^{2}J(5^{7}Fe, 1H) = 0.5 Hz$; 23 \pm 1 °C (see legend of Figure 1)), measured for observation of 13C satellites and determination of the isotope-induced chemical shift ${}^{1}\Delta(1^{2/13}C(5^{7}Fe))$ (the same sample as used for spectra shown in Figure 1; result of 400 transients; ∼70 min of spectrometer time). For comparison, the $57Fe{^1H}$ single-pulse spectrum is shown on top, where the same time (acquisition time 1.5 s; 2800 transients, 30° pulses) has been used as for the INEPT spectrum.

with 0.7 Hz). The nondecoupled spectrum of ferrocene (**1**) recorded by INEPT (see Figure 1B) indicates that $\binom{2J^{57}\text{Fe},^{1}\text{H}}{\text{2}} \approx 0.4 \pm 0.1 \text{ Hz}$. The expected 10-line exists of the result of the pattern (the central line is canceled as the result of the pulse sequence) is not resolved; the separation of the nonresolved anti-phase multiplets is ∼1 Hz, and the magnitude of $^{2}J^{57}Fe,^{1}H$) should be slightly less than half of this value. The application of the DEPT pulse sequences¹⁵ (¹H decoupled or nondecoupled) was not successful. This could be the consequence of longer evolution times by a factor of ≥ 2 as compared to those of INEPT, which may cause serious loss of 1H and 57Fe magnetization. Attempts to measure the 57Fe NMR signal of bis(pentamethylcyclopentadienyl)iron, $FeCp*_{2}$ (**4**), by INEPT experiments within a short time, on the basis of $\frac{3J}{57}Fe$, ¹H), failed. It must be concluded that the magnitude of $3J(57Fe,1H)$ is less than 0.25 Hz (this means a sum of delays of \geq 2 s in the basic INEPT pulse sequence) and that loss of 1H magnetization prevents efficient polarization transfer. 57Fe NMR spectra of numerous other ferrocene derivatives, studied already by single-pulse techniques,^{3,5} have been remeasured successfully, using ${}^{1}H-{}^{57}Fe$ techniques.

Chemical shifts *δ*(57Fe) of many ferrocene derivatives can be determined relatively quickly by INEPT experiments. Considering that an instrument with only moderate field strength $(B_0 = 8.46 \text{ T})$ was used, the studies can certainly be extended to less concentrated samples, even to reaction solutions, by using higher field strengths

*B*⁰ (up to 21 T) which have become increasingly available nowadays. The successful extension of the INEPT experiments to 187Os NMR can be envisaged, opening the access to an even less developed area² when compared with $57Fe$ NMR. The measurements of $57Fe$ NMR data may help to develop further concepts regarding structure and reactivity, 17 to understand other data coming, for example, from electrochemistry or $57Fe$ Mössbauer spectroscopy of ferrocenes,¹⁸ and to stimulate theoretical work on optimizing the geometry of transition-metal complexes and the calculation of metal nuclear magnetic shieldings.¹⁹

Experimental Section

The model compounds studied were commercially available and were used without further purification. The sample quality plays an important role in all polarization transfer (PT) experiments. Therefore, the line widths in the 1H NMR spectra should be checked; highly resolved ¹H NMR spectra indicate that loss of 1H magnetization during the initial steps of the PT pulse sequence may not be serious for PT efficiency. If there are broadened signals in the ¹H NMR spectra for those ¹H nuclei which will be used for PT, the great loss of 1H magnetization causes seriously reduced PT efficiency. Samples were prepared in 10 mm (o.d.) tubes by filtration of hexane/ $CDCl₃$ or $CHCl₃/CDCl₃$ solutions through silica under an atmosphere of Ar in order to remove paramagnetic impurities.

 $57Fe$ NMR spectra (see also Figures $1-3$ for further details) were recorded at 23 \pm 0.2 °C (control outside of the sample) using a Bruker Avance 360 instrument, equipped with a lowfrequency probe head, tunable from 6.8 to 16.8 MHz, for which the ¹H (90[°]) pulse duration was 32 μ s. The duration of the 90[°] pulse for 57Fe NMR (∼40 *µ*s) was calibrated first by measuring the 73 Ge NMR signal of GeCl₄ in CDCl₃ at 12.56 MHz, assuming that the value of the 73Ge 90° pulse would be similar for 57 Fe (11.66 MHz). The best homogeneity of B_0 was adjusted by optimizing the 1H NMR signals for each sample. Chemical shifts δ ⁽⁵⁷Fe) are given relative to δ ⁽⁵⁷Fe; Fe(CO)₅, neat) 0 with $E^{(57}Fe) = 3.237798$ MHz.

A problem in measuring 57Fe NMR spectra concerns the temperature dependence of the 57Fe NMR signals. This is

Figure 3. 11.66 MHz ⁵⁷Fe NMR spectrum (INEPT,¹²) refocused with optimized low-power 1H composite pulse decoupling; 23 ± 1 °C) of acetylferrocene, CpFe(C₅H₄C(O)-Me) (450 mg in 2 mL of $CHCl₃/CDCl₃$), after 20 h of measurement time. There are several overlapping 57Fe NMR signals as a result of the temperature dependence of ⁵⁷Fe chemical shifts. The line width $h_{1/2}$ after 32 scans was ∼0.5 Hz; the *δ*(57Fe) value was 1774.4.

shown in the case of acetylferrocene (**3**) for a spectrum recorded within 24 h (Figure 3). Even by using optimized low-power ¹H composite pulse decoupling (CPD), temperature gradients in the sample develop slowly and give rise to several overlapping 57Fe NMR signals (the temperature control of the spectrometer is switched on to keep the temperature constant (± 0.2) K), outside of the sample). On the basis of the present results, it can be assumed that the observation of identical line widths of 57Fe NMR signals of ferrocenes detected by single-pulse techniques with or without ¹H decoupling is due to temperature-induced chemical shifts which broaden the lines after long measurement times.

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