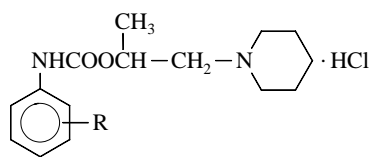


Table: Studied substances, their rate constants of alkaline hydrolysis and activation parameters

| Compd. | Temp. (°C) | k · 10 <sup>5</sup> (s <sup>-1</sup> · l · mol <sup>-1</sup> ) | t <sub>1/2</sub> (h) | n  | r     | F     | s       | a       | b       |  |   |  |
|--------|------------|--|----------------------|----|-------|-------|---------|---------|---------|--|---|--|
|        |            |  |                      |    |       |       |         |         |         | E <sub>A</sub> (kJ · mol <sup>-1</sup> ) | ΔH <sup>‡</sup> (kJ · mol <sup>-1</sup> ) | ΔS <sup>‡</sup> (J · K <sup>-1</sup> · mol <sup>-1</sup> ) |
| 1      | 60         | 3.297 ± 0.135  | 59                   | 11 | 0.990 | 462.8 | 0.00488 | 0.0029  | 0.00504 |  |   |  |
|        | 65         | 5.890 ± 0.244  | 33                   | 11 | 0.992 | 581.6 | 0.00819 | -0.0110 | 0.00898 |  |   |  |
|        | 70         | 9.612 ± 0.282  | 20                   | 10 | 0.997 | 1163  | 0.00780 | -0.0104 | 0.01465 |  |   |  |
|        | 75         | 14.21 ± 0.401  | 14                   | 10 | 0.997 | 1254  | 0.01111 | -0.0160 | 0.02165 |  |   |  |
| 2      | 60         | 3.118 ± 0.141  | 63                   | 7  | 0.995 | 488.7 | 0.00227 | -0.0016 | 0.00475 |  |   |  |
|        | 65         | 5.651 ± 0.115  | 35                   | 10 | 0.998 | 2420  | 0.00318 | -0.0026 | 0.00861 |  |   |  |
|        | 70         | 8.948 ± 0.275  | 22                   | 6  | 0.998 | 1060  | 0.00350 | -0.0036 | 0.01364 |  |   |  |
|        | 75         | 12.53 ± 0.502  | 16                   | 7  | 0.996 | 623.8 | 0.01708 | -0.0086 | 0.01910 |  |   |  |
| 3      | 60         | 4.509 ± 0.089  | 44                   | 12 | 0.998 | 2649  | 0.00685 | -0.0108 | 0.00675 |  |   |  |
|        | 65         | 7.691 ± 0.283  | 26                   | 16 | 0.991 | 735.4 | 0.01658 | -0.0187 | 0.01134 |  |   |  |
|        | 70         | 11.06 ± 0.433  | 18                   | 13 | 0.992 | 653.0 | 0.01114 | -0.0124 | 0.01632 |  |   |  |
|        | 75         | 15.73 ± 0.806  | 13                   | 11 | 0.988 | 380.4 | 0.02388 | -0.0241 | 0.02330 |  |   |  |
| 4      | 60         | 3.179 ± 0.121  | 64                   | 16 | 0.990 | 694.0 | 0.01055 | -0.0096 | 0.00467 |  |   |  |
|        | 65         | 3.820 ± 0.185  | 53                   | 12 | 0.987 | 383.1 | 0.01091 | 0.0053  | 0.00561 |  |   |  |
|        | 70         | 5.136 ± 0.214  | 38                   | 14 | 0.990 | 572.9 | 0.01297 | -0.0124 | 0.00795 |  |   |  |
|        | 75         | 6.369 ± 0.296  | 32                   | 8  | 0.994 | 462.7 | 0.00851 | -0.0022 | 0.00944 |  |   |  |



1: 2-OC<sub>6</sub>H<sub>13</sub>  
 2: 2-OC<sub>7</sub>H<sub>15</sub>  
 3: 3-OC<sub>7</sub>H<sub>15</sub>  
 4: 4-OC<sub>7</sub>H<sub>15</sub>

lic buffers at pH 7; basic media have an unfavourable influence on its stability in solution.

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## Experimental

The alkaline hydrolysis of the studied substances was carried out in aqueous-ethanolic NaOH (0.1 mol · l<sup>-1</sup>) in closed flasks tempered in an ultrathermostat at 60.0, 65.0, 70.0 and 75.0 ± 0.2 °C. The buffer solutions at pH 7.0, 8.0 and 9.0 possessed an amount of NaOH in the concentration of 0.0115 mol · l<sup>-1</sup>, 0.0291 mol · l<sup>-1</sup> and 0.0461 mol · l<sup>-1</sup>, respectively. The ionic strength of buffers was 0.1 mol · l<sup>-1</sup>. Hydrolysis was carried out at 70.0 ± 0.2 °C. The C<sub>2</sub>H<sub>5</sub>OH concentration was 50% (v/v). The concentration of the studied substances was 0.001 mol · l<sup>-1</sup>. The concentration of the corresponding alkyloxyaniline, produced by the reaction, was determined spectrophotometrically [7], (spectrophotometer Specol 11, Carl Zeiss Jena, Germany). Rate constants of alkaline hydrolysis were calculated using the kinetic equation of the second order. Values of E<sub>A</sub> were determined by the Arrhenius equation. Values for ΔH<sup>‡</sup> and ΔS<sup>‡</sup> were determined by the Eyring equation [8].

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## Effect of curing and storage on fluid-bed granules

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Aqueous colloidal polymer dispersions (latexes or pseudo-latexes) have been developed as an alternative to organic polymer solutions for the coating of solid dosage forms [1–3]. So films or coatings of water-insoluble polymers can be prepared without toxic organic solvents.

During coating of solid dosage forms with an aqueous polymer dispersion, coalescence of the colloidal polymer particles into a homogeneous film is often incomplete. To overcome this problem, it is recommended to expose the product to elevated temperatures (curing) [4, 5].

The objective of this work was to study the influence of storage time and temperature on granules, coated with a commercial ethylcellulose pseudolatex, Aquacoat<sup>®</sup>. Oven-curing of new coated theophylline granules at 70 °C decreased the released amount of drug of about a quarter

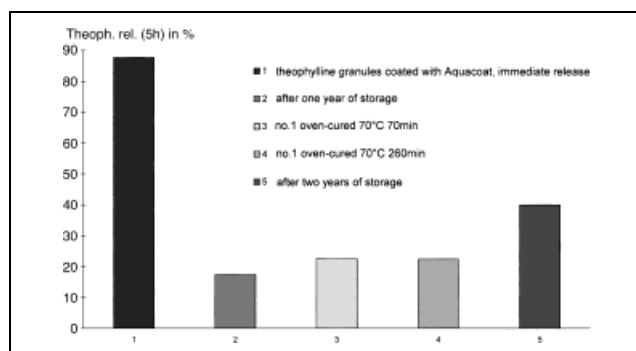


Fig. 1: Effect of storage and curing on theophylline release from coated granules

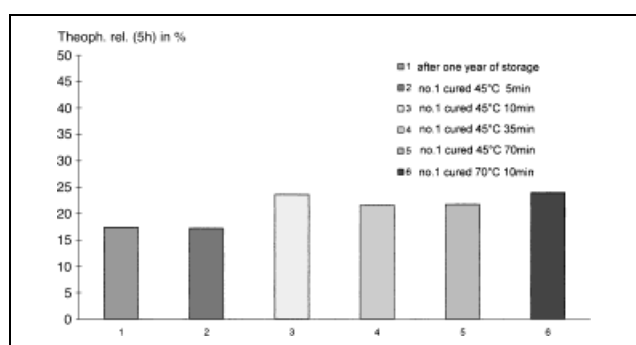


Fig. 2: Effect of storage and curing time on theophylline release from one year old coated granules

within five hours. The curing times of 70 min and 260 min caused no significant differences in drug release pattern.

Under normal conditions of storage (room temperature and humidity) the released amount of theophylline from coated granules within five hours was after one year about five times less than the released amount of new coated theophylline granules. After two years of storage (same conditions), surprisingly the released amount of drug within five hours was half of the release of the new coated granules (Fig. 1). This effect may be caused by aging of the material.

As can be seen in Fig. 2, oven-curing of the one year old granules did not show an effect by 45 °C and 5 min. By 45 °C and 10, 35 and 70 min, the theophylline release was slightly increased as well as at 70 °C and 10 min.

## Experimental

A powder mixture of Emcoel<sup>®</sup> 50M (microcrystalline cellulose, Mendell Co. Inc., Carmel, NY, USA), theophylline (China), lactose hydrous (Sheffield Products, Norwich, NY, USA) and Ac-Di-Sol<sup>®</sup> (croscarmellose sodium, FMC Corporation, Newark, DE, USA) was used to produce granules in a fluid-bed rotor granulator (GPCG1, Glatt Air Techniques, Ramsey, NJ, USA) while continuously spraying a binder solution of 2.5% Plasdone<sup>®</sup> K-29/32 (polyvinylpyrrolidone, ISP Technologies Inc., Wayne, NJ, USA). After drying in the rotor granulator to a humidity of 2–4% the granules over 0.85 mm and under 1.7 mm were coated in the same granulator with a mixture of Aquacoat<sup>®</sup> (ethylcellulose aqueous dispersion, FMC Corporation, Newark, DE, USA) and 30% dibutyl sebacate (Sigma Chemical Co., St. Louis, MO, USA). The thickness of the layer was 4 mg/cm<sup>2</sup>.

The dissolution test was made by USP basket-apparatus at 37 °C and 50 rpm. The medium was 900 ml of water. Samples were measured by UV spectrophotometry (Beckmann DU-70, Beckmann Inc., Fullerton, CA, USA).

The moisture content was determined using an automated gravimetric moisture content analyzer (Computrac, Max-50, Arizona Instruments, Tempe, AZ, USA).

The content uniformity was examined and the USP requirements were met for all batches.

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## Effect of adhesive and drug reservoir on *in vitro* transdermal delivery of nicotine

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Transdermal delivery of drugs from polymeric devices has been around for over a decade and significant advances have been made in this field [1]. A complete design of a transdermal device or patch incorporates a drug-loaded matrix or reservoir totally covered by a rate-controlling membrane. The entire skin-contacting surface of the patch is coated with an adhesive [2]. Accordingly the topical permeability of a drug depends on the rate of release from its formulated product including drug reservoir, membrane and adhesive. The purpose of this investigation was to evaluate the influence of adhesive and drug reservoir on *in vitro* permeability of nicotine as a model drug. Transdermal delivery of nicotine into the systemic circulation has been shown to facilitate tobacco withdrawal by mitigating or even preventing abstinence symptoms and diminishing craving [3, 4]. Both natural rubber and silicone are used as the base of adhesive to determine the permeation of nicotine *in vitro*. A polysulfone hydrophobic membrane is employed as the rate-controlling membrane because of the good wetting of the membrane by the aqueous fluid beneath it [5]. In the drug reservoir of the *in vitro* diffusion cell, nicotine was incorporated into the hydrogel formulations including Carbopol polymer (polyacrylic acid), methylcellulose (MC) and hydroxypropyl cellulose (HPC).

Various nicotine loading amounts are incorporated into pH 7.0 McIlvaine buffer for the *in vitro* permeation study through a polysulfone membrane combined with natural rubber or silicone adhesive. The profile of nicotine flux and the permeability coefficient is shown in the Table. There is an ascendant trend for nicotine to increase its flux from the nicotine concentration of 3 to 9% either through a natural rubber adhesive or through a silicone adhesive. After the calculation of the correlation coefficient between the drug concentration of 3 to 9% and flux, a linear relationship ( $r = 0.999$  for natural rubber;  $r = 0.998$  for silicone adhesive) was observed. No significant difference was found between the flux from drug re-