

Ecdysteroids from *Rhaponticum uniflorum*

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Five phytoecdysteroids were isolated from the roots of *Rhaponticum uniflorum*. They were identified as ecdysterone, ajugasterone C, ajugasterone C-20,22-monoacetonide, ajugasterone C-2,3,20,22-diacetonide and 5-deoxykaladasterone-20,22-monoacetonide by means of spectroscopic data. This is the first report of ajugasterone C-2,3,20,22-diacetonide and 5-deoxykaladasterone-20,22-monoacetonide isolated from a natural source.

1. Introduction

In China, there are two species of *Rhaponticum* (Compositae). *Rhaponticum uniflorum* (L.) DC. is distributed in northern China. Its roots have been used as a Chinese traditional medicine for the treatment of fever and intoxications [1]. Since the 1980s many researchers have studied *Rhaponticum uniflorum* [2–3]. Phytoecdysteroids and other compounds were isolated from this plant [4–6]. We report here the isolation and structural elucidation of two new ecdysteroids, as well as the isolation of 3 known ecdysteroids, from the roots of *R. uniflorum*.

2. Investigations, results and discussion

The dried powdered roots of *R. uniflorum* were extracted with methanol at room temperature. The extract was fractionated successively by petrol, ethyl acetate and n-butanol. The ethyl acetate extract was further separated by silica gel chromatography to give two new ecdysteroids, ajugasterone C-2,3,20,22-diacetonide (**4**) and 5-deoxykaladasterone-20,22-monoacetonide (**5**), and one known ecdysteroid, ajugasterone C-20,22-monoacetonide (**3**) [7]. The n-butanol extract was also purified by silica gel chromatography to give two known ecdysteroids, ecdysterone (**1**) [5] and ajugasterone C (**2**) [5].

Compound **4**, C₃₃H₅₂O₇. The IR spectrum of compound **4** contained absorption bands of hydroxy groups (3431 cm⁻¹) and of a keto group conjugated with a double bond (1662 cm⁻¹). Its UV spectrum ($\lambda_{\max}^{\text{MeOH}}$ 243 nm) also confirmed the presence of a 7-en-6-keto group in the steroid nucleus. In the EI MS the peaks at *m/z* 379, 343, 325, 317 and 299 confirmed the existence of an ajugasterone C ring skeleton [8]. A strong peak at *m/z* 185 could be the result of the side chain ion derived from cleavage between C-17 and C-20. The ¹³C NMR spectrum (Table) showed that the compound is quite similar to ajugasterone C except that six more peaks corresponding to the ketal groups were observed (δ 108.09, 106.81, 26.76, 26.83, 29.04 and 30.87), and the C-2, C-3, C-20, C-22 signals had shifted to a lower field. The MS of compound **4** showed the peaks of the molecular ion with *m/z* 560. This is higher by 40 amu than the molecular weight of compound **3**. Accordingly, the structure of compound **4** is ajugasterone C-2,3,20,22-diacetonide.

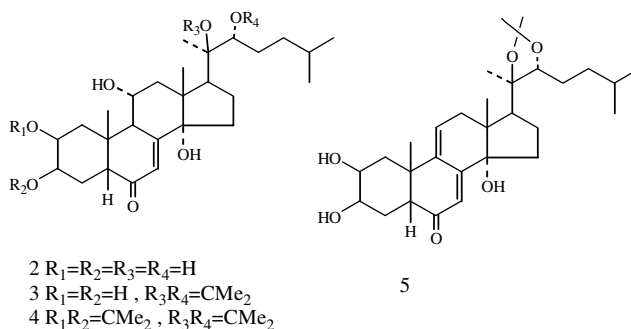
Ajugasterone C-2,3,20,22-diacetonide (**4**) is a native compound [9]. This follows from the fact that no acetone was used in the extraction and subsequent treatment of the extractive substances.

Compound **5** was identified as 25-deoxy-9(11)-dehydro-20-hydroxyecdysone-20,22-mono acetonide based on the following evidence. Its UV spectrum in MeOH showed a strong maximum at λ_{\max} 287 nm; such a maximum value is

Table: ¹³C NMR spectral data of compounds **4–5** (100 MHz, 4 in CDCl₃, 5 in CD₃OD)

C	4	5
1	39.98	43.19
2	72.64	68.05
3	71.59	67.81
4	36.35	37.22
5	52.09	51.07
6	203.29	203.18
7	122.13	119.24
8	161.14	154.24
9	41.88	141.14
10	38.58	37.13
11	68.09	132.01
12	42.39	38.31
13	47.14	47.36
14	84.42	84.93
15	31.65	31.22
16	21.16	22.35
17	48.79	50.01
18	17.70	17.74
19	23.59	27.08
20	83.90	83.16
21	21.84	22.11
22	81.57	82.23
23	27.12	27.48
24	36.38	37.62
25	28.22	28.70
26	22.45	22.76
27	22.58	22.88
Ketal group		
–O–C–O–	108.09	107.22
	106.81	
CH ₃	26.76	29.39
CH ₃	26.83	29.68
CH ₃	29.04	
CH ₃	30.87	

characteristic of a conjugated dienone and was previously found for kaladasterone [10, 11] and 5-deoxykaladasterone [12]. The mass spectrum gave a [M]⁺ at *m/z* 502. The



presence of fragments at m/z 361, 343 (361–18) and 325 (343–18) is characteristic of an ecdysteroid bearing a 20,22 diol and no 25-OH [10]. ^1H NMR spectroscopy (1D and 2D-COSY experiments) allowed us to assign the 25-deoxy structure based on the chemical shifts and the doublet pattern of 26-Me and 27-Me. The $\Delta^{9(11)}$ structure was established from the disappearance of the 9-H signal and the appearance of a new olefinic proton signal at δ 6.22 coupled with three signals 2.73 ($J = 2$ Hz), 2.43 ($J = 6.5$ Hz) and 7-H signal by a small long-range coupling (observed with a 2D-COSY experiment). The observed downfield shifts of the 19-Me, 1-H and the small upfield shifts of 2-H, 3-H and 7-H are in accordance with a $\Delta^{9(11)}$ structure. The ^{13}C NMR spectrum (Table) showed that the compound is quite similar to 5-deoxykaladasterone [11] except that three more peaks corresponding to the ketal group were observed (δ 107.22, 29.39, 29.68), and the C-20, C-22 signals had shifted to a lower field. Thus, the structure of compound **5** was determined as 5-deoxykaladasterone-20,22-monoacetone.

3. Experimental

3.1. Equipment

M.p.s.: Kofler-microscope (Reichert) uncorr. Optical rotation: Polarimeter 241 (Perkin Elmer) solvent CHCl_3 or MeOH. IR-spectra were recorded on a Nicolet-5DX IR spectrometer. ^1H and ^{13}C NMR spectra were recorded with a Bruker AM-400, solvent CDCl_3 , CD_3OD , $\text{C}_5\text{D}_5\text{N}$, using TMS as internal standard. EIMS and FABMS were determined on a ZAB-HS mass spectrometer.

3.2. Plant material

The roots of *R. uniflorum* were purchased from the company of Chinese medicinal materials in province Gansu, China and were identified by Prof. Yin-Shou Zhou, Lanzhou Medical College. A voucher specimen No. 98001 was deposited in the Herbarium of the Department of Pharmacy, Lanzhou Medical College, Lanzhou, P.R. China.

3.3. Extraction and isolation

The air-dried roots of *R. uniflorum* (3 kg) were exhaustively extracted with MeOH at room temperature, the extract was concentrated under reduced pressure. The resulting residue was suspended in H_2O , extracted with petrol, EtOAc and *n*-BuOH, respectively. The EtOAc extract (50 g) was subjected to CC on a silica gel column (1000 g, 200–300 mesh) using petrol/EtOAc gradient as eluent to give fractions I–V according to TLC analysis. Compound **4** (48 mg) was obtained from fraction II (8:1), compound **5** (26 mg) from fraction IV (4:1) and compound **3** (57 mg) from fraction V (1:1), and purified by rechromatography on silica gel followed by recrystallization. The *n*-BuOH extract (40 g) was also subjected to CC on a silica gel column using $\text{CHCl}_3/\text{MeOH}$ gradient as eluent to give fractions I–IV. Compound **2** (31 mg) was obtained from fraction I (10:1), compound **1** (62 mg) from fraction II (8:1), and purified by rechromatography on silica gel followed by recrystallization.

3.4.1. Ecdysterone (1)

White needles, m.p. 236–238 °C (EtOAc/MeOH); $[\alpha]_D^{20} +62.0$ (c 0.9, MeOH); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 242; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3479, 3289, 3226, 2959, 2875, 1658, 1640, 1468, 1447, 1384, 1229, 1145, 1053, 920, 878, 688; FABMS: 503 $[\text{M} + \text{Na}]^+$, 487 $[\text{M} + \text{Li}]^+$; EIMS m/z (%): 462 $[\text{M} - \text{H}_2\text{O}]^+$ (2), 444 $[\text{M} - 2\text{H}_2\text{O}]^+$ (5), 426 $[\text{M} - 3\text{H}_2\text{O}]^+$ (16), 408 $[\text{M} - 4\text{H}_2\text{O}]^+$ (3), 363 $[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2]^+$ (58), 345 $[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2 - \text{H}_2\text{O}]^+$ (100), 327 $[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2 - 2\text{H}_2\text{O}]^+$ (35), 161 $[\text{C}_8\text{H}_{17}\text{O}_3]^+$ (7), 143 $[\text{C}_8\text{H}_{17}\text{O}_3 - \text{H}_2\text{O}]^+$ (15), 125 $[\text{C}_8\text{H}_{17}\text{O}_3 - 2\text{H}_2\text{O}]^+$ (17), 107 $[\text{C}_8\text{H}_{17}\text{O}_3 - 3\text{H}_2\text{O}]^+$, 117 $[\text{C}_6\text{H}_{13}\text{O}_2]^+$ (4), 99 $[\text{C}_6\text{H}_{13}\text{O}_2 - \text{H}_2\text{O}]^+$ (42), 80 $[\text{C}_6\text{H}_{13}\text{O}_2 - 2\text{H}_2\text{O}]^+$ (32); ^1H NMR (400 MHz, $\text{C}_5\text{D}_5\text{N}$) δ : 1.01 (3H, s, 19-H), 1.21 (3H, s, 18-H), 1.36 (6H, s, 26, 27-H), 1.58 (3H, s, 21-H), 3.01 (2H, m, 5,17-H), 3.02 (1H, m, 5-H), 3.58 (1H, m, 9-H), 3.77 (1H, dd, 22-H), 3.90 (2H, m, 2,3-H), 6.26 (1H, d, $J = 1.8$ Hz, 7-H).

3.4.2. Ajugasterone C (2)

White powder, m.p. 196–198 °C (EtOAc/MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 240. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3430, 1658, 1641, 1440, 1384, 1059, 976. FABMS m/z : 503 $[\text{M} + \text{Na}]^+$, 487 $[\text{M} + \text{Li}]^+$. EIMS m/z : 462 $[\text{M} - \text{H}_2\text{O}]^+$, 444 $[\text{M} - 2\text{H}_2\text{O}]^+$, 426 $[\text{M} - 3\text{H}_2\text{O}]^+$, 379 $[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2]^+$, 361 $[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2 - \text{H}_2\text{O}]^+$, 343

$[\text{M} - \text{C}_6\text{H}_{13}\text{O}_2 - 2\text{H}_2\text{O}]^+$, 145 $[\text{C}_8\text{H}_{17}\text{O}_2]^+$, 127 $[\text{C}_8\text{H}_{17}\text{O}_2 - \text{H}_2\text{O}]^+$, 109 $[\text{C}_8\text{H}_{17}\text{O}_2 - 2\text{H}_2\text{O}]^+$, 101 $[\text{C}_6\text{H}_{13}\text{O}]^+$, 83 $[\text{C}_6\text{H}_{13}\text{O} - \text{H}_2\text{O}]^+$; ^1H NMR (CD_3OD) δ : 5.80 (1H, brs, 7-H), 4.05 (2H, m, 2,3-H), 3.58 (2H, m, 9, 22-H), 3.24 (1H, m, 11-H), 2.57 (1H, dd, 5-H), 1.24 (3H, s, 21-H), 1.05 (3H, s, 18-H), 0.93 (6H, d, $J = 7$ Hz, 26, 27-H), 0.85 (3H, s, 19-H).

3.4.3. Ajugasterone C-20,22-monoacetone (3)

White needles, m.p. 248–250 °C (EtOAc/MeOH); $[\alpha]_D^{20} +60.8$ (c 1.3, MeOH); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 237. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 1658, 1468, 1377, 1039. FABMS m/z : 543 $[\text{M} + \text{Na}]^+$, 527 $[\text{M} + \text{Li}]^+$; EIMS m/z : 520 $[\text{M}]^+$, 505 $[\text{M} - \text{CH}_3]^+$, 502 $[\text{M} - \text{H}_2\text{O}]^+$, 487 $[\text{M} - \text{CH}_3 - \text{H}_2\text{O}]^+$, 469 $[\text{M} - \text{CH}_3 - 2\text{H}_2\text{O}]^+$, 462, 444, 427, 379, 361, 344, 325, 317, 299, 266, 185, 127, 109, 59. ^1H NMR (CD_3OD) δ : 5.79 (1H, d, $J = 1.8$ Hz, H-7), 3.99 (1H, m, H-2), 3.94 (1H, m, H-3), 4.08 (1H, m, H-11), 3.65 (1H, m, H-22), 3.13 (1H, m, H-9), 0.91 (6H, d, $J = 3.6$ Hz, H-26, 27), 0.79, 1.04, 1.17, (each 3H, s, H-18, 19, 21), 1.30, 1.38 (each 3H, s, ketal group Me \times 2).

3.4.4. Ajugasterone C-2,3,20,22-diacetone (4)

White needles, m.p. 134–136 °C (Petrol/EtOAc); $[\alpha]_D^{20} +58.2$ (c 1.0, MeOH); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 243. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3431, 1662, 1459, 1056. EIMS m/z : 560 $[\text{M}]^+$, 545 $[\text{M} - \text{CH}_3]^+$, 542 $[\text{M} - \text{H}_2\text{O}]^+$, 502, 484, 460, 419, 379, 361, 357, 325, 317, 306, 299, 185, 142, 127. ^1H NMR (400 MHz, CDCl_3) δ : 5.84 (1H, d, $J = 2.4$ Hz, H-7), 4.52 (1H, m, H-2), 4.29 (1H, m, H-3), 4.12 (1H, m, H-11), 3.60 (1H, m, H-22), 2.83 (1H, m, H-9), 0.90 (6H, d, $J = 6.5$ Hz, H-26, 27), 0.78, 1.04, 1.16 (each 3H, s, H-18, 19, 21), 1.32, 1.33, 1.41, 1.49 (each 3H, s, ketal group Me \times 4). ^{13}C NMR: see Table.

3.4.5. 5-Deoxykaladasterone-20,22-monoacetone (5)

White powder, m.p. 235–237 °C (EtOAc/MeOH); $[\alpha]_D^{20} +54.8$ (c 0.8, MeOH); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 287. IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 1665. FABMS m/z : 525 $[\text{M} + \text{Na}]^+$, 509 $[\text{M} + \text{Li}]^+$; EIMS m/z : 487 $[\text{M} - 15]^+$, 469 $[\text{M} - \text{CH}_3 - \text{H}_2\text{O}]^+$, 444, 426, 402, 379, 361, 343, 325, 326, 317, 301, 299, 211, 185. ^1H NMR (400 MHz, CD_3OD) δ : 3.69 (1H, m, H-2), 3.76 (1H, m, H-3), 2.40 (1H, m, H-5), 5.66 (1H, brs, H-7), 6.22 (1H, dd, $J = 6.4$, 2 Hz, H-11), 2.73 (1H, m, $J = 2$ Hz, H-12), 2.43 (1H, m, $J = 6.5$ Hz, H-12), 2.45 (1H, m, H-17), 0.81, 1.06, 1.16 (each 3H, s, H-18, 19, 21), 0.88, 0.89 (each 3H, d, $J = 6.7$ Hz, H-26, 27), 1.28, 1.35 (each 3H, s, ketal group Me \times 2); ^{13}C NMR: see Table.

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