National Laboratory of Applied Organic Chemistry, Lanzhou University, Gansu Province, P.R. China

Sesquiterpene polyol esters from Euonymus phellomana Loes

HONG WANG, LI YANG, XUAN TIAN and YAO-ZU CHEN

Three new (1-3) and six known (4-9) β -dihydroagarofuran sesquiterpene polyol esters were isolated from a methanol extract of the seed oil of *Euonymus phellomana* Loes. and their structures were established on the basis of spectral analysis, including 2D-NMR spectroscopy.

1. Introduction

Plants of the Celastraceae family have been the subject of continued and growing interest, due to the range of biological activities shown by many members of this family. Some of them have been used in folk medicine from ancient times as a stimulant and to invigorate the circulation of blood. In the last 30 years, various β -dihydroagarofuran sesquiterpene polyol esters have been isolated from Celastraceae. Some of these sesquiterpene polyol ersters exhibit insect antifeedant and/or insecticidal activities as well as antitumor activity [1, 2]. Recently, we have investigated the chemical constituents of Euonymus phellomana Loes. (Celastraceae). Euonymus phellomana Loes., a defoliate shrub mainly distributed in the west of China, has been used for the treatment of rheumatism and fractures as a folk medicine. From the methanol extract of the seed oil, three new (1-3) and six known (4–9) β -dihydroagarofuran sesquiterpenoids have been isolated. The structure of compounds 1-3 was established by 2D-NMR spectra.

2. Investigations, results and discussion

Repeated column chromatography of the methanol extract of seed oil of *Euonymus phellomana* Loes. yielded compounds 1-9. The structures of compounds 4-9 were de-



termined by spectral data as 1 β -hydroxy-2 β ,6 α ,9 β ,12-tetraacetoxy-8 β -benzoyloxy- β -dihydroagarofuran [3], 1 β -(β -) furancarboxy-2 β ,6 α -diacetoxy-4 α -hydroxy-9 α ,12-di(α -methyl) butanoyl- β -dihydroagarofuran [4], 1 β -benzoyloxy-2 β ,6 α -diacetoxy-4 α -hydroxy-9 α ,12-di(α -methyl)butanoyl- β -dihydroagarofuran [4], 1 β ,8 β -dibenzoyloxy-2 β -hydroxy-6 α ,9 β ,12-triacetoxy- β -dihydroagarofuran [5], 1 β , 8 β -dibenzoyloxy-2 β ,6 α ,9 β ,12-tetraacetoxy- β -dihydroagarofuran [4], and 1 β ,2 β -dibenzoyloxy-4 α -hydroxy-6 α ,9 α ,12-triacetoxy- β -dihydroagarofuran [4].

Compound 1 was analyzed for $C_{31}H_{42}O_{13}$ by spectral data. It showed on ester carbonyl band at 1745 cm^{-1} and a hydroxyl at 3420 cm^{-1} in IR. UV showed the presence of an acetate and a furan moiety. FABMS: m/z 623 (M + 1). The ¹H and ¹³C NMR spectrum revealed the presence of three acetoxy [8H: 2.09 (3H, s), 2.11 (3H, s), 2.27 (3 H, s); δC: 21.1, 21.2, 21.5, 169.4, 170.2, 170.5], one (β-)furancarboxy [δH: 6.72 (1H, d), 7.45 (1H, d), 8.01 (1 H, s); δC: 109.9, 118.7, 143.8, 148.9, 161.8] and one (a-methyl)butanoyl [8H: 0.68 (3H, t), 0.88 (3H, d), 1.26 (2 H, m), 1.98 (1 H, m); δC: 11.3, 15.6, 25.3, 40.7, 174.4] esters, and the MS showed peaks due to the loss of acetic acid, β -furoic acid and (α -methyl)butanoic acid. In addition, the ¹³C NMR spectrum showed signals attributed to three methyls (δ 29.3, 25.5, 25.0), three methylenes (δ 65.4, 41.9, 34.5), five oxygenated methines (δ 78.1, 69.9, 69.1, 68.2, 49.1) and four quaternary carbons (δ 91.1, 84.5, 69.7, 55.0). The ¹HNMR spectrum contained signals assignable to protons on the carbon atoms carrying four secondary ester groups, i.e. δ 6.11 (1 H, s, H-6), 5.58 (1 H, d, J = 3.4 Hz, H-1), 5.47 (1 H, d, J = 3.4 Hz, H-2),5.22 (1 H, d, J = 7.0 Hz, H-9) and a primary ester group, i.e. δ 4.99, 4.41 (2 H, ABq, J = 13.0 Hz, H-15a, b) on the basis of the multiplicity and the ¹H-¹HCOSY spectrum. These data suggested that 1 is a β -dihydroagarofuran sesquiterpene substituted with five ester groups.

The positions of the ester groups were unambiguously established by HMBC experiments. The carbonyl ¹³C signal at δ 161.8 showed long range correlation with the ¹H signals at δ 5.22 (H-9). Thus, the furoyl group must be located on C-9. The long range couplings of the carbonyl signals at 170.5, 170.2 and 169.4 with the proton signals at δ 5.47 (H-2), 5.58 (H-1) and 6.11 (H-6) and the acetyl methyl signals at δ 2.09, 2.11 and 2.27 indicated the presence of acetyl groups on C-2, C-1 and C-6. The last carbonyl ¹³C signals at δ 174.4 were correlated with the ¹H signals at δ 4.99, 4.41 (H-15a, b), 0.88 (H-16), 1.26 (H-18) and 1.98 (H-19), respectively. Thus, the isobutyryl groups had to be situated on C-15. Therefore, a planar structure for **1** was established.

The relative stereochemistry of **1** was confirmed via the NOESY difference spectra. In NOESY there are correlation between H-1, H-2/H-3ax ($J_{1,2} = 3.4$), showing H-1ax,

H(J _{Hz})	1	2	3	4	7	8
1	5.58 d (3.4)	5.65 d (3.4)	5.95 s	4.34 dd (8.5, 3.5)	5.79 d (4.0)	5.83 d (4.0)
2	5.47 d (3.4)	5.51 d (3.4)	5.94 t	5.23 td (3.5, 3.5, 2.7)	4.29 m	5.51 dt (4.0, 4.0, 2.5)
3	2.02 m	2.20 m	2.27 m	2.31	2.43 m	2.52 ddd (15.2, 6.2, 4.0)
		1.99 m	2.10 m	1.29	1.89 m	
4				2.29	2.34 m	2.37 brp (7.7)
6	6.11 s	6.13 s	6.25 s	6.30 s	6.88 s	6.09 s
7	2.23 m	2.18 m	2.24 s	2.59 d (4.0)	2.58 d (3.5)	2.59 d (4.0)
8	2.57 dd	2.59 m	2.63 m	5.72 dd (4.0, 5.6)	5.66 dd (3.5, 5.5)	5.65 dd (4.0, 5.7)
	2.53 dd	2.55 m				
9	5.22 d (7.0)	5.33 d (8.0)	5.53 d (7.1)	5.63 d (5.6)	5.54 d(5.5)	5.54 d(5.7)
12	1.47 s	1.48 s	1.58 s	1.42 s	1.59 s	1.60 s
13	1.47 s	1.49 s	1.57 s	1.58 s	1.46 s	1.47 s
14	1.55 s	1.56 s	1.61 s	1.15 d (7.7)	1.22 d (7.0)	1.15 d (7.7)
15a	4.99 d (13.0)	5.00 d (13.0)	5.42 d (12.7)	5.11 d (12.5)	5.59 d (13.0)	5.47 d (13.5)
15b	4.41 d (13.0)	4.44 d (13.0)	4.69 d (12.7)	4.77 d (12.5)	4.68 d (13.0)	4.60 d (13.5)

Table 1: ¹H NMR data of compounds 1–4, 7 and 8 (δ in CDCl₃, J(Hz))*

* **2**: AcO × 3[2.09 (s, 3 H), 2.12 (s, 3 H), 2.28 (s, 3 H)]; BzO[7.43~7.47 (t, 2 H), 7.56~7.60 (t, 1 H), 8.02~8.04 (d, 2 H)]; α -MeBu[0.79 (d, J = 8.0, 3 H), 0.54 (t, 3 H), 1.15 (m, 1 H), 0.91 (m, 1 H), 1.87 (m, 1 H)]. **3**: AcO × 2[2.14 (s, 3 H), 2.34 (s, 3 H)]; BzO × 3[7.12~7.16 (t, 2 H), 7.30~7.36 (m, 5 H), 7.45~7.57 (m, 4 H), 7.74~7.76 (d, 2 H)].

H-2eq; H-6: δ 6.11(s) indicated angle of 7 β , 6 should be 90° [6, 7]. Cross signals of H-1ax and δ 8.01, 6.72(-OFu) determined α substituted ester group at C-9.

Considering the above data for 1 the structure was elucidated to be 1β , 2β , 6α -triacetoxy- 4α -hydroxy- 9α -(β -)furancarboxy-15-(α -methyl)butyroyloxy- β -dihydroagarofuran.

Compound 2 was found to be spectroscopically similar to 1. FAB MS m/z: 633 (M + 1). Its spectral data (Tables 1 and 2) suggested the presence of three acetate esters, one benzyl ester and one isobutyrate ester and the 1 β ,2 β ,6 α ,9 α ,15-quinquesterified- β -dihydroagarofuran parent. Close comparison of the ¹H and ¹³C NMR spectra of 1 and 2 revealed the benzoyl at C-9 of 2 replacing a furoylate of 1. In the same way as in 1 (HMBC experiment), the positions of the three acetate esters were shown to be situated at C-1, C-2 and C-6, the benzoyl esters at C-9, and the one isobutyrate ester at C-11, respectively.

Thus, the structure of **2** is $1\beta,2\beta,6\alpha$ -triacetoxy- 4α -hydroxy- 9α -benzoyloxy-15-(α -methyl)butyroyloxy- β -dihydroagarofuran.

Compound 3 was found to be spectroscopically very similar to 2. Its spectral data (Table 1 and 2) suggested the presence of two acetate esters and three benzoyl esters

Table 2: ¹³C NMR shifts of compounds 1-3 (δ in CDCl₃)*

С	1	2	3
1	69.9	69.9	69.3
2	68.2	68.1	69.3
3	41.9	41.9	42.1
4	69.7	69.7	69.6
5	91.1	91.1	91.2
6	78.1	78.1	78.1
7	49.1	49.1	49.2
8	34.5	34.5	34.7
9	69.1	69.5	71.2
10	55.0	55.1	55.3
11	84.5	84.6	84.7
12	25.0	25.0	25.8
13	25.5	25.7	25.3
14	29.3	29.3	29.3
15	65.4	65.4	66.3

* 2: AcO × 3[21.1, 21.2, 21.5, 169.4, 170.2, 170.5]; BzO [128.3, 129.1, 130.2, 133.5, 165.2]; α -MeBu [11.1, 15.6, 25.3, 40.6, 174.2]. 3: AcO × 2[21.3, 21.5, 170.3, 70.8]; BzO × 3[127.9(2C), 128.0(2C), 128.4(2C), 128.4(2C), 129.4(1C), 129.8(2C), 129.9(2C), 132.4(1C), 133.4(1C), 165.5(1C), 165.0(2C)]

and the $1\beta,2\beta,6\alpha,9,15$ -quinquesterified- β -dihydroagarofuran parent. Close comparison of the ¹H and ¹³C NMR spectra of **2** and **3** clearly revealed the two benzoyl at C-1, C-2 of **3** replacing two acetate of **1** and an acetate at C-15 of **3** replacing an isobutyrate of **1**. In the same way as in **2** (HMBC experiment), the positions of two acetate esters were shown to be situated at C-6 and C-15, the three benzoyl esters at C-1, C-2 and C-9, respectively. In the NOESY spectrum of **3**, the presence of cross signals between H-6ax and δ 8.08 (at C-9) showed that the stereochemistry of the substituted ester group at C-9 was β in compound **3**.

Thus, the structure of **3** is $1\beta_2\beta_9\beta$ -tribenzoyloxy- 4α -hy-droxy- 6α , 15-diacetoxy- β -dihydroagarofuran.

3. Experimental

3.1. Equipment

MP: uncorr. $[\alpha]_D$: JASCO-20C. MS: VGZAB-HS and VG-Auto Spec-3000. ¹H and ¹³C NMR, NOESY, HMQC and HMBC spectra: 400 MHz spectrometer, (Bruker AM-400) solvent, CDCl₃, int. Standard, TMS. CC: silica gel (200–300 mesh). Eluent, petrol-EtoAc (10:1–1:1). TLC: precoated silica gel (Merck 60F-254) plates. Reverse phase CC: RP-18, Merck short column, eluent, H₂O-MeOH (1:4).

3.2. Plant material

Plant material was collected from Wen county, Gansu province P.R. China in August 1996 and identified by Prof. Jizhou Sun, Department of Biology, Lanzhou University where a voucher is deposited.

3.3. Extraction and isolation

Air-dried and pulverized seeds (2.5 kg) of *Euonymus phellomana* Loes., were extracted with petrol at room temperature for a week. Removal of the solvent under reduced pressure afforded an oil. The oil was extracted two times with H₂O-MeOH (1:10). A portion (50.0 g) of this extract was chromatographed on a silica gel column using Petrol-EtoAc (10:1, 8:1, ..., 1:1, EtoAc), as eluent to give 65 fractions. 52 highly polar fractions were combined, on the basis of TLC, divided it into 3 groups, which were chromatographed on a RP-18 short column using H₂O-MeOH (1:4) as eluent to yield compounds 1 (6.4 mg), 2 (14.7 mg), 3 (14.2 mg), 4 (3.9 mg), 5 (300.0 mg), 6 (50.0 mg), 7 (10.5 mg), 8 (5.6 mg) and 9 (21.2 mg).

3.4. 1β , 2β , 6α -Triacetoxy- 4α -hydroxy- 9α -(β -)furancarboxy-15-(α -methyl) butyroyloxy- β -dihydroagarofuran (1)

White needles crystals (petroleum ether/Me₂CO); m.p.: 154–156, $C_{31}H_{42}O_{13}$; $[\alpha]_D^{20} + 22.8^{\circ}$ (C = 0.30, MeOH); IR, ν_{max}^{KBr} (cm⁻¹): 3420, 1746, 1621, 1571, 1241, 1155, 1095; UV, λ_{max}^{MeOH} (nm): 206, 232; EIMS, *m/z* (%): 607 (M⁺-CH₃, 3), 562 (M⁺-HOAc, 4) 502 (M⁺-2HOAc, 30), 95 (Fu⁺, 100), 85 (C₄H₉CO⁺, 37), 57 (C₄H₉⁺, 46), 43 (Ac⁺, 2); FABMS *m/z*: 623 (M + 1); ¹HNMR and ¹³C NMR see Table 1 and Table 2.

3.5. 1β , 2β , 6α -Triacetoxy- 4α -hydroxy- 9α -benzoyloxy-15-(α -methyl) butyroyloxy- β -dihydroagarofuran (2)

White needles crystals; mp.: $170-172 \,^{\circ}C$, $C_{33}H_{44}O_{12}$; $[\alpha]_{20}^{20} + 39.7^{\circ}$ (C = 0.23, MeOH); IR, v_{max}^{KBr} (cm⁻¹): 3530, 1745, 1709, 1606, 1584, 1459, 1388, 1369, 940, 867; UV, $\lambda_{max}^{\text{MeOH}}$ (nm): 231, 275, 281; EIMS, m/z (%): 617 (M⁺-CH₃, 1), 572 (M⁺-HOAc, 2), 530 (572 + H-OAc, 2), 512 (M⁺ -2 HOAc, 56), 452 (512-HOAc, 1), 105 (C₆H₅CO⁺, 100), 85 (C₄H₉CO⁺, 20), 77 (C₆H₅⁺, 30), 57 (C₄H₉⁺, 20); FABMS m/z: 633 (M + 1); ¹H and ¹³C NMR see Table 1 and Table 2.

3.6. 1β,2β,9β-Tribenzoyloxy-4a-hydroxy-6a,15-diacetoxy-β-dihydroagarofuran (3)

Colorless oil; $C_{40}H_{42}O_{12}$; $[\alpha]_D^{20}$ +81.3° (C = 0.48, MeOH); IR, ν_{max}^{KBr} (cm⁻¹): 3556, 1729, 1604, 1454, 1388, 1368, 1276, 1240, 1152 ; UV, λ_{max}^{MeOh} (nm): 231, 275, 281; EIMS, *m*/*z* (%): 594 (M⁺-2 HOAc, 2), 532 M_{max} (nn), 23, 27, 26, 187, 187, m_2 (b), 57, (n = 2 Hore, 2, 35) (46), 428 (2), 397 (4), 337 (5), 228 (4), 202 (22), 166 (12), 105 (C₆H₅CO⁺, 100), 77 (C₆H₅⁺, 48), 45 (3); FABMS m/z; 715 (M + 1); ¹H and ¹³C NMR see Table 1 and Table 2.

Acknowledgements: We would like to thank Prof. Jizhou Sun, Department of Biology, Lanzhou University for the identification of the plant and the National Laboratory of Applied Organic Chemistry and Analysis Center Lanzhou University, P.R. China for measuring NMR and MS, respectively.

This work was supported by National Laboratory of Applied Organic Chemistry of Lanzhou University of China.

References

- 1 Reimar, B.; Wagner, H.: Pytochemistry 17, 1821 (1978)
- 2 Yoshihisa, T.; Syunji, O.; Kimiko, N.; Toshiakj, T.: J. Nat. Prod. 56, 815 (1993)
- 3 Zsuzsanna, R.; Andras, P.; Istvan, P.; Gyula, A.; Alajos, K.: J. Chem. Soc. Perkin. Trans I, 1079 (1989)
- 4 Zsuzsanna, R.; Istvan, P.: J. Chem. Soc. Perkin. Trans I, 1089 (1989)
- 5 Axel, R.; Hartmut, T.; Herbert, B.: Z. Naturforsch., Teil B, 31, 607 (1976)
- 6 Cecil, R. S. Jr.; Roger, W. M.; David. W.; William, K. R.; Nancy, E.;
- Jon, C.: J. Org. Chem. **41**, 3264 (1976) 7 Wakabayashi, N.; Wu, W. J.; Waters, R. M.; Redfern, R. E.; Mills, G. D. Jr.; Albert, B. D.; William, R. L.; Andrzejewski, D.: J. Nat. Prod., 51, 537 (1988)

Received September 5, 2000 Accepted October 13, 2000

Prof. Xuan Tian Department of Chemistry Lanzhou University 73000, P.R. China xuant@lzu.edu.cn