ORIGINAL ARTICLES

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Synthesis of some new antianemics I. Iron pullulan complexes of pharmaceutical interest

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First experiments were performed in order to obtain a polynuclear iron(III) complex with pullulan, which could be applied in therapy of sideropenic anemia. Complete synthesis of the complex can be realized with oligomers of depolymerized pullulan at an average molar mass $8000 - 10000$ g · mol⁻¹. Maximum iron contents of 51.4 mg · cm⁻³ is effected by pullulan depolymerizates $(M_w \approx 9000 \text{ g} \cdot \text{mol}^{-1}$, $[\eta] = 0.085 \text{ dl} \cdot \text{g}^{-1}$ at 130 °C for 240 min, at pH = 10.5 and at an initial mass ratio Fe/ligand of 1 : 3. The effected concentrations and complex stability respond to requirements of pharmacological application of parenteral antianemic preparations.

1. Introduction

The task of creating new, therapeutically active iron(III) complexes remains a challenge, because iron deficient anemia is wide-spread in the world [1]. Numerous preparations based on organic and inorganic compounds of biand trivalent iron are used today for the prevention and therapy of sideropenic anemia [2]. Iron-carbohydrate complexes (i.e. iron-dextran) are preferred as the standard in modern iron therapy [3, 4]. Synthesis of these complexes with native polysaccharides is impossible. Our experience with iron dextran complex and similar preparations shows that only when oligosaccharides have limited molar mass, complexation can occur. Therefore, depolymerization yielding oligomers of well-known degree of polymerization is mandatory. Synthesis and complex stability are directly influenced by the ligands' degree of polymerization. Usable reaction conditions (pH, T, t, P) are important for iron contents in the preparation and for its toxicity [5]. The possibility of different types of carbohydrate ligands

to form water-soluble complexes with Fe(OH)O in alkaline solution shows that there is no strict choice of ligands. Application of hydrogenated pullulan as a ligand is not known. Pullulan is a water-soluble polysaccharide very similar to dextran in its composition [6, 7]. Pullulan is an amorphous polymer with a linear flexible chain, the main axis of which consists of α -D-glycose units. The structural unit, which is regularly repeated, is maltotriose $(\alpha$ -1,4-triglycoside) linked by α -1,6-glycosidic bonds. The interest in this polysaccharide is considerable because of its good physicochemical qualities, good degradability and wide opportunities for industrial and pharmaceutical applications [8, 9].

Pullulan has all conditions to form water-soluble complexes with iron. That's why pullulan and its hydrogenated oligomers were examined as ligands for the synthesis of polynuclear antianemic complexes. Since the stability of a complex is directly affected by the characteristics of the applied ligand, the conditions of preparation and characterization of pullulan oligomers as well as the conditions of complex synthesis are optimized.

2. Investigations, results and discussion

Preliminary researches of complex synthesis with different samples of native pullulan (NP-1 and NP-2) of high molar mass showed negative results. Neither with prolonged time of reaction nor under more drastic reactive conditions in an autoclave, the complex was formed. Therefore, the starting components were depolymerized to oligomers of adequate polymerization degree (P_w) under different reactive conditions (pH, T and t). For investigating the influence of the initial molar mass of pullulan on the depolymerization rate, the progress of depolymerization of two samples was followed, the chosen molar masses 220570 and $328000 \text{ g} \cdot \text{mol}^{-1}$. The influence of pullulan concentration on its depolymerization rate in solutions with constant HCl concentration was verified for the concentrations 6.3 and 7.1%. Conditions of depolymerization, according to previous investigations [10], were chosen depending on ligand qualities and initial M_w . A temperature of 80– 85 C and a pH value of 2.0 proved to be optimal for the depolymerization. Under more drastic conditions of depolymerization (higher temperature and lower pH values), higher contents of mono- and oligosaccharides with lower M_w (below 1000 g · mol⁻¹) appeared. These saccharides inevitably have to be removed from depolymerizates before complexation with Fe(OH)O, because a very alkaline environment under high temperature and pressure causes their decomposition to toxic products [11]. On the other hand, if the average molar mass of the applied polysac-

charide is too high, the product will be a gel, not practical for any kind of application [12].

2.1. Depolymerization of native pullulan

Pullulan NP-1 ($M_w = 220570 \text{ g} \cdot \text{mol}^{-1}$) was depolymerized by hydrolysis in HCl/water. Depolymerization was followed by viscosimetry [13]. Conditions that allow application of this method are steadiness of pullulan in acidic water solution at 25° C (depolymerization would be interrupted by cooling), and the constancy of ratio between pullulan intrinsic viscosity in water and hydrochloric acid (important for calculation of average weight molar mass). The average number at molar mass, polydispersity (M_w/M_n) , and contents of reducing groups (RG) characterize some depolymerizates (Table 1).

According to previous experiences, the time of depolymerization at 80 \degree C and pH 2.0 with pullulan NP-1 is limited to 90–150 min. In this interval depolymerizates with M_w between 21000 and 13000 g mol⁻¹ are obtained (Table 1).

Similar M_w values with relatively small deviation were
also obtained with NP-2 pullulan $(M_w =$ also obtained with NP-2 pullulan $(M_w =$ $328000 \text{ g} \cdot \text{mol}^{-1}$) under the same conditions. This shows that the depolymerization velocity does not depend on initial pullulan molar mass in this interval. Therefore, in the second series, analogously to complexes when low molar dextran is used $(M_w = 5000 \text{ g} \cdot \text{mol}^{-1})$, for the depolymerization of NP-2 pullulan higher temperature and longer time intervals (0–180 min) were applied at the same pH value. With this sample, in the interval of 130–180 min (Table 2), depolymerizates with M_w under 10000 g · mol⁻¹ could be obtained.

Table 1: Characteristic parameters of native pullulan NP-1 and its oligomers, obtained by depolymerization with HCl (pH 2, 80° C, $90-150$ min)

Sample $(c = 6.3\%)$	t (min)	η_r	[n] $(dl \cdot g^{-1})$	P_{w}	M_w $(g \cdot mol^{-1})$
$NP-1$				1362	220570
$Dep-1$	90	1.500	0.147	125	20313
$Dep-2$	100	1.481	0.142	119	19291
$Dep-3$	110	1.462	0.138	114	18485
Dep-4	120	1.423	0.129	103	16715
$Dep-5$	130	1.385	0.118	90	14633
Dep-6	140	1.365	0.113	85	13718
Dep-7	150	1.356	0.111	82	13357

The influence of pullulan concentration on depolymerization showed slight variations within the limits of experimental error. Thus, in the observed time interval concentrations could be neglected. Depolymerization increases considerably the content of reducing saccharides. Experiments have shown that unfavorable ligand qualities for the outcome of complex preparation arise precisely from their reducing activity. In preparation with higher contents of RG, sedimentation occurs. Decreasing complex stability increases toxicity. Therefore, the reducing activity of the obtained pullulan oligomers has been decreased by NaBH4. In accordance with kinetic laws, hydrogenation of pullulan oligomers is a pseudo first order reaction [14].

2.2. Synthesis of polynuclear iron (III) complexes with pullulan

On the basis of numerous data [4, 5], which refer to the synthesis of iron(III) complexes with different carbohydrates, it can be concluded that there are no fundamental differences among many quoted procedures.

The influence of molar masses of ligands on creating of complexes was investigated. Five depolymerizates with the ligands masses from 4000 to 15000 g · mol⁻¹ were used (Table 3). According to published data about dextran complexes [16], we expected a positive outcome of complex synthesis.

Data in Table 3 show that high molar pullulan does not lead to successful results. It is more convenient to use depolymerizates of $6000-10000$ g · mol⁻¹. That confirms an earlier assumption that the average molar mass of the ligand has a limiting value which secures binding with Fe(OH)O particles. In favor of that are results with depolymerizates Dep-7 and Dep-17 with masses higher or smaller than necessary. They do not clarify the reaction mixture at all (i.e. dissolve solid forms of Fe(OH)O), not even under prolonged or more drastic conditions (autoclave). In the progress of the synthesis we have observed a second expected phenomenon. Boiling the solution under atmospheric pressure does not lead to dissolution of Fe(OH)O and changing of the color of the reaction system (e.g. Dep-7 and Dep-15). Under more drastic conditions (prolonged time, higher temperature and pressure), depending on pH (9.5–11.5), complexes are formed only partially $({\sim}60\%$ of dissolved Fe(OH)O) with iron contents of 5.9 mg \cdot cm⁻³ in the soluble fraction (Dep-7). It is evident that at higher pH values the synthesis should afford higher iron contents in the complex. At the same time, with dissolution of Fe(OH)O the color change from red into distinctly gray-brown shades. This might be the

Table 2: Characteristic parameters of native pullulan NP-2 and its oligomers, obtained by depolymerization with HCl (pH 2, 85° C, 0-180 min)

Sample $(c = 7.1\%)$	(min)	$[\eta]$ $(dl \cdot g^{-1})$	M_n $(g \cdot mol^{-1})$	P_{w}	M_{w} $(g \cdot mol^{-1})$	M_w/M_n	
$NP-2$				2025	328000		
Dep-8		0.233	29074	249	40396	1.389	
Dep-9	10	0.209	26002	212	34346	1.321	
$Dep-10$	30	0.184	14301	175	28399	1.986	
$Dep-11$	50	0.157	10769	138	22410	2.081	
$Dep-12$	80	0.131	8993	106	17104	1.902	
$Dep-13$	100	0.106	7929	77	12469	1.573	
$Dep-14$	130	0.091	5378	61	9929	1.846	
$Dep-15$	140	0.085	4767	55	8968	1.881	
$Dep-16$	150	0.071	4724	42	6856	1.451	
$Dep-17$	180	0.050	2672	25	4062	1.520	

Ligand	$t_{\rm DEP.}$ (min)	M_w $(g \cdot mol^{-1})$	pH	T $(^{\circ}C)$	t _{SYNT.} (min)	$+T^*$ (^{0}C)	$+\mathbf{t}^*_{\text{SYNT.}}$ (min)	Synth. result	Fe content $(mg \cdot cm^{-3})$
$NP-2$	$\overline{0}$	328000	10.5	80	120	140	120		
$NP-1$	θ	220570	10.5	70	120	140	90		
Dep-7	150	13357	10.5	100	120				
Dep-7	150	13357	9.5	70	120	120	90		
Dep-7	150	13357	10.5	70	120	120	90	30%	3.2
Dep-7	150	13357	11.5	70	120	120	90	60%	5.9
$Dep-15$	140	8968	10.5	100	120	-		40%	4.8
$Dep-15$	140	8968	10.5	80	120	120	90	100%	31.2
$Dep-16$	150	6856	10.5	70	120	100	180	80%	11.3
$Dep-16$	150	6856	10.5	80	120	120	60	60%	5.1
$Dep-16$	150	6856	10.5	80	60	105	60	40%	4.5
$Dep-17$	180	4062	10.5	80	120	100	90		

Table 3: Basic characteristics of pullulan oligomers and reaction conditions for complex synthesis

T* and t* relate to the synthesis of the complexes in an autoclave, at 200 kPa

result of alkaline degradation of the ligand in the progress of synthesis, which results in the presence colloidal dispersed reduced iron [5]. At complexes obtained with depolymerizates of $M_w = 6856 \text{ g} \cdot \text{mol}^{-1}$ (Dep-16), during the first 24 hours after synthesis visible sedimentation of the solid phase occurs. That indicates insufficient stability of the complex, but with achieving a little bit higher iron contents than in the previous case. The uncompleted synthesis ($\sim 80\%$ of dissolved Fe(OH)O), is probably due to inadequate conditions, in accordance to the ligands molar mass. Complete synthesis of complex, i.e. obtainment of a clear dark-red reaction mixture, was accomplished with depolymerizates Dep-15 and iron contents of $31.2 \text{ mg} \cdot \text{cm}^{-3}$.

Regarding that complex synthesis can be realized only under drastic conditions, analogously to the synthesis of dextran complex with iron [17], in further experiment the synthesis was conducted in an autoclave at 200 kPa and at $100-130$ °C. Since the molar mass of the used ligand had been determined earlier, and for finding the optimum of molar mass, we used two more depolymerizates Dep-13 and Dep-20 (Table 4).

Complete reaction could be achieved with pullulan oligomers in masses of $8000 - 10000$ g · mol⁻¹. Maximum iron contents of 51.4 mg per unit of preparation volume were reached by depolymerizates Dep-15. The result of synthesis with other depolymerizates is in accordance with previous facts, establishing the basis to obtain complexes with higher iron contents per volume. Possibility and result of iron pullulan complex synthesis were confirmed by FTIR spectroscopic studies [18].

2.3. Preliminary toxicological examinations of iron pullulan complex

For the pharmacological test, the iron complex with Dep-15 ligands (Table 4) was standardized on $50 \text{ mg } \text{Fe/cm}^3$, in 0.9% sodium chloride solution, at $pH = 6.5$ and the mass ratio Fe/ligand 1:2.84. The preparation was tested for acute toxicity (LD_{50}) using white mice of both sexes, 18–26 g. Application was conducted intravenously (dose of 900–1900 mg \cdot kg⁻¹). None of the applicated doses showed mortality within 10 days. Thus, the lethal dose could not be determined. Application of higher doses was not realizable because of the application volume, which could not be increased in consideration of standardized preparation. It can be concluded that the max. applicable dose is 1900 mg \cdot kg⁻¹ at the iron concentration built-in the complex of 50 mg \cdot cm⁻³.

The effected concentrations, complex stability, and acute toxicity respond to requirements of a pharmacological application of parenteral antianemic preparations. Our results are preliminary, but they contribute to our knowledge concerning the preparation of optimum ligands for successful synthesis and for conditions of synthesis, not only of the examined iron complex with pullulan but of similar polysaccharides. Research should be conducted to standardize partially depolymerized carbohydrates and to improve their physicochemical and pharmacobiological characteristics.

3. Experimental

3.1. Materials

Native pullulan (NP) was produced by biosynthesis with our own species (CH1, Pharmaceutical and Chemical Industry "Zdravlje", Leskovac-Yu), using the fungus Aureobasidium pullulans. Two samples of pullulan were

Table 4: Reaction conditions and characteristic parameters of complex synthesis with different pullulan oligomers of NP-1 and NP-2

Ligand	M_{w} $(g \cdot mol^{-1})$	M_n $(g \cdot mol^{-1})$	[n] $(dl \cdot g^{-1})$	RG $(\%)$	T $(^{\circ}C)$	t_{SYNT} (min)	pH	Synth. result	Fe content $(mg \cdot cm^{-3})$
Dep-7	13357	7004	0.110	5.72	100	180	11.0		15.9
$Dep-13$	12469	7929	0.106	6.34	100	180	11.0		16.2
$Dep-14$	9929	5378	0.091	7.59	120	150	10.5		26.3
$Dep-14$	9929	5378	0.091	7.59	120	180	10.5	$\mathrm{+}$	28.4
$Dep-14$	9929	5378	0.091	7.59	130	240	10.5	$\mathrm{+}$	28.6
$Dep-15$	8968	4767	0.085	10.74	130	210	10.5	\pm	38.8
$Dep-15$	8968	4767	0.085	10.74	130	240	10.5	$\mathrm{+}$	51.4
$Dep-20$	8040	5020	0.079	7.01	105	120	10.5	$^+$	28.5
$Dep-16$	6856	4724	0.071	12.29	100	180	10.5		21.3

used. Different pullulan fractions were characterized by the light-scattering method on molar mass: NP-1 $(M_w = 220570 \text{ g} \cdot \text{mol}^{-1})$ and NP-2 $(M_w = 328000 \text{ g} \cdot \text{mol}^{-1}).$

3.2. Methods

The amorphous β_2 -form of Fe(OH)O, as the only applicable form for this synthesis, was prepared by sedimentation from ferric chloride solution using sodium carbonate at room temperature [15]. The precipitate, after removing electrolytes, was directly used for the synthesis of the complex. This form of exceptional quality of Fe(OH)O was used because this gel gives complexes with hydrogenated low molar dextran and dextran carboxylic acid [3].

Native pullulan was depolymerized by HCl, pH 2, 80-85 °C, 0-180 min. For obtaining fractions with as much as possible a narrow distribution of molar masses, all depolymerizates were fractionally precipitated by ethanol, in order to remove ligands of low molar masses from fractions, which were prepared for the reaction with iron. Average weight molar masses (M_w) of the pullulan oligomers were determined by the viscosimetric method based on Khun-Mark-Houwink's equation, adapted for pullulan by Kato [19]:

$$
[\eta] = 1.91 \cdot 10^{-2} \cdot M_w^{0.67} \quad (in cm^3 \cdot g^{-1})
$$
 (1)

Average number molar masses (M_n) and the contents of RG were determined by Somogyis method [20]. Viscosimetric measurements were performed at 25 °C using Ostwalds viscosimeter [21]. Values of specific and intrinsic viscosity [h]were calculated by Schultz-Blaschke's equation.

The oligomers were reduced with NaBH₄ at 40 °C [22]. The quantity of NaBH₄ (96% activity) necessary for hydrogenation (Q), which depends of RG contents and initial ligands mass (m), was calculated by an empiric formula:

$$
Q (mg) = 0.95 \cdot 10^{-3} \cdot RG (\%) \cdot m (g)
$$
 (2)

The excess of unreacted NaBH4 was destroyed by HCl and by further treatment of the depolymerization solution on 70° C.

Solutions both of depolymerized pullulan and complex were purified and deionized by acidic and alkaline ion-exchangers (Amberlit IR-120 and IRA- 410). The iron content in the preparations was determined complexometrically. Chlorides in water solutions both of ligands and complexes

were determined by potentiometric titration with 0.1 M AgNO₃.
Polynuclear β-Fe(OH)O and its complexes with pullulan were synthesized according to Müller [5]. Methods of complex preparation involve neutralization of $FeCl₃$ solution with an alkaline solution of modified pullulan. The synthesis was realized either at boiling temperature under atmospheric pressure or in an autoclave at 200 kPa until 140 °C, at a mass ratio Fe/ ligand 1 : 3, and different reaction conditions (pH, T, t). Success of synthesis was defined as complete dissolution of the Fe(OH)O gel and as changing of the light-brown color of the initially homogeneous reaction mixture into dark-red, characteristic for polynuclear iron complexes with carbohydrates.

The test of systematic acute toxicity was executed by the Litchfield-Wilcoxon method [23], on white mice of both sexes, 18–26 g. To groups of 6 mice the following doses were successively applied: 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800 and 1900 mg · kg⁻¹ intravenously. During ten days the percentage of expiring was followed. These tests were conducted in the biomedical laboratory, Sector of applied research and development, ''Zdravlje" Leskovac.

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Received January 29, 2001 Goran S. Nikolic Accepted October 12, 2001 Faculty of Technology

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