

Departments of General Chemistry<sup>1</sup> and Pharmacognosy<sup>2</sup>, University of Istanbul, Turkey; Department of Chemistry<sup>3</sup>, University of Peshawar, Pakistan; Department of Botany<sup>4</sup>, Government Postgraduate Jahan Zeb College, Saidu Sharif, Swat, Pakistan

## Norditerpenoid alkaloids from the roots of *Aconitum leave* Royle

A. ULUBELEN<sup>1</sup>, A. H. MERIÇLİ<sup>2</sup>, F. MERIÇLİ<sup>2</sup>, U. KOLAK<sup>1</sup>, M. ARFAN<sup>3</sup>, M. AHMAD<sup>3</sup> and H. AHMAD<sup>4</sup>

From the roots of *Aconitum leave* Royle growing wild in Northern Pakistan, Swat district, three new alkaloids, 8-methyllycaconitine (**1**), 14-demethyllycaconitine (**2**), *N*-deethyllycaconitine-*N*-aldehyde (**3**) have been isolated along with four known compounds, lappaconitine (**4**), lycaconitine (**5**), lapaconidine (**6**) and lycoctonine (**7**).

### 1. Introduction

*Aconitum* as well as *Delphinium* species have been used as medicinal agents for centuries, especially as cardio-tonic, febrifuges and sedatives [1, 2]. Lycaconitine (**5**) obtained from several *Aconitum* species was found to overcome multidrug resistance in cancer chemotherapy [3]. Intrategmental administration of methyllycaconitine was found to decrease both nicotine induced and food induced dopamine release in the Nucleus accumbens [4]. The affinity of lycaconitine for two neuronal nicotinic acetylcholine receptor subtypes was determined [5]. Like *Delphinium* species, in the old days *Aconitum* species were also used against lice and scorpions [6]. Alkaloids, although highly poisonous, were tested for cardiovascular (hypotensive and bradycardic) action [7, 8].

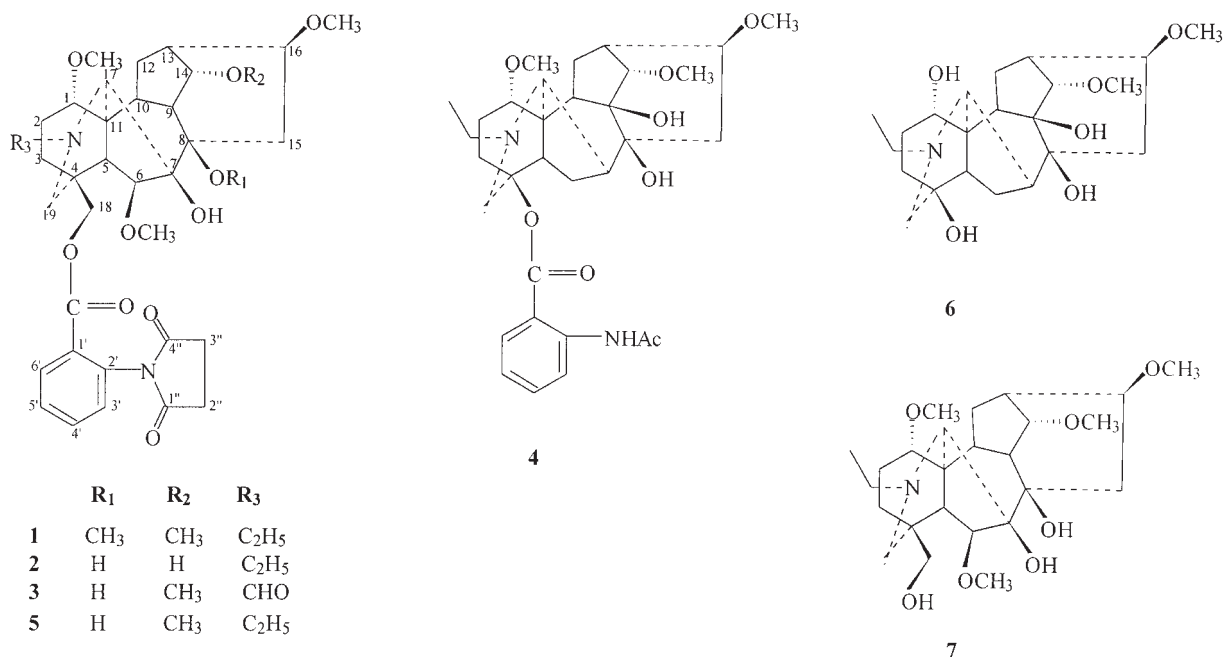
### 2. Investigations, results and discussion

Besides of our studies on *Delphinium* species [9–12], we now studied an *Aconitum* species, *A. leave* Royle also from Pakistan. In an earlier study, the presence of lappaconitine (**4**) was established in the plant and no other alkaloid was found [13].

In the present study, the dried and powdered roots of the plant (1 kg) were extracted with n-hexane, and 8 g of an

oily residue was obtained. The remaining plant material was extracted with 90% EtOH. The ethanol extract was concentrated and acidified with 5% HCl and extracted with CHCl<sub>3</sub> to yield 30 g of non-alkaloidal residue. The acidic aqueous solution was basified with 10% NaOH solution and extracted with CHCl<sub>3</sub> to yield 26 g of a crude alkaloidal mixture. A part of the alkaloidal mixture (5 g) was separated by VLC (Vacuum Liquid Chromatography) eluting with petroleum ether, a gradient of CH<sub>2</sub>Cl<sub>2</sub> was added up to 100% followed by EtOH up to 10%. When the CH<sub>2</sub>Cl<sub>2</sub> reached to 50% a crystalline compound started to separate (frac. 9 to frac. 14), about 2 g of a single crystalline compound was obtained. The structure of the crystalline compound was established as lappaconitine (**4**) using spectral data, m.p., optical rotation and TLC comparison with authentic sample [14, 15]. Frac. 15–21, frac. 22–29, frac. 30–36 were alkaloidal mixtures. They were separated by a Chromatotron apparatus [16]. Fractions 15–21 yielded lycaconitine (**5**), 8-methyllycaconitine (**1**), 14-demethyllycaconitine (**2**), fractions 22–29 contained *N*-deethyllycaconitine-*N*-aldehyde (**3**), fractions 30–36 yielded lapaconidine (**6**) and lycoctonine (**7**). Compounds **1**, **2** and **3** were new, while **4**–**7** were known.

8-Methyllycaconitine (**1**) is an amorphous compound with an optical rotation  $[\alpha]_D^{30} = +19.5^\circ$  (c = 0.1, CHCl<sub>3</sub>). The molecular formula C<sub>37</sub>H<sub>50</sub>N<sub>2</sub>O<sub>10</sub> was calculated from



HRMS ( $m/z$  682.3464) and confirmed by  $^{13}\text{C}$  NMR and APT (Attached Proton Test) spectral data. The  $^1\text{H}$  NMR spectrum of **1** showed signals for aromatic protons at  $\delta$  8.02 (1H, dd,  $J = 8$  and 1.5 Hz, H-6'), 7.68 (1H, dt,  $J = 1.5, 8$  and 8 Hz), 7.54 (1H, dt,  $J = 1.5, 8$  and 8 Hz) (H-5' and H-4'), 7.27 (1H, dd,  $J = 1.5, 8$  Hz, H-3'), five methoxy groups were at  $\delta$  3.42, 3.40, 3.35, 3.32 and 3.24 (each 3H, s). A typical signal for succinimid  $\text{CH}_2\text{-CH}_2$  group was at  $\delta$  2.90 as a broad singlet (4H, s, 2'', 3''). A signal at  $\delta$  3.65 (1H, t,  $J = 4.5$  Hz, H-14) typical for methoxyl carrying C-14, a broad singlet signal at  $\delta$  3.92 indicated H-6 $\alpha$  in a methoxyl carrying C-6 position. Other signals were at  $\delta$  2.92 (1H, s, H-17), 1.12 (3H, t,  $J = 7$  Hz) (N- $\text{CH}_2\text{CH}_3$ ). The  $^{13}\text{C}$  NMR spectrum of **1** (Table) is in agreement with the suggested structure.

The UV and IR spectra of 14-demethyllycaconitine (**2**) were similar to those of compound (**1**). The HRMS of **2** indicated the molecular formula  $\text{C}_{35}\text{H}_{46}\text{N}_2\text{O}_{10}$  ( $m/z$  654.3155) which was correlated by its  $^{13}\text{C}$  NMR and APT spectra. The  $^1\text{H}$  NMR spectra of compounds **1** and **2** were quite similar except for the chemical shift of H-14 at  $\delta$  4.09 (1H, t,  $J = 4.5$  Hz, H-14) which indicated a hydroxy group at C-14 instead of a methoxyl. The only other difference was the presence of three methoxy groups in compound **2** at  $\delta$  3.36 (6H, s,  $2 \times \text{OMe}$ ), 3.26 (3H, s, OMe) versus five methoxyls of compound **1**. The aromatic protons and all the other signals were in their respective

**Table:**  $^{13}\text{C}$  NMR assignments of 8-methyllycaconitine (**1**), 14-demethyllycaconitine (**2**) and N-deethyl-lycaconitine-N-aldehyde (**3**)

C	1	2	3
1	83.8 d	83.4 d	83.4 d
2	25.9 t	25.9 t	25.8 t
3	33.5 t	33.6 t	33.5 t
4	37.4 s	37.4 s	37.3 s
5	43.2 d	43.6 d	43.3 d
6	90.7 d	90.6 d	90.7 d
7	88.4 s	87.9 s	88.2 s
8	80.1 s	77.4 s	76.8 s
9	49.9 d	49.8 d	49.8 d
10	38.0 d	37.9 d	38.0 d
11	50.9 s	50.6 s	50.8 s
12	28.6 t	28.6 t	28.8 t
13	46.0 d	45.9 d	46.2 d
14	83.8 d	75.8 d	83.6 d
15	33.1 t	33.8 t	32.8 t
16	82.5 d	83.0 d	82.8 d
17	64.4 d	65.0 d	64.6 d
18	69.5 t	69.5 t	69.4 t
19	52.3 t	52.4 t	52.0 t
20	50.6 t	51.0 t	—
21	13.9 q	14.0 q	182.3 d
Ar-CO	164.1 s	164.1 s	164.2 s
1'	126.8 s	126.7 s	126.8 s
2'	132.8 s	132.8 s	132.6 s
3'	129.4 d	129.4 d	130.0 d
4'	133.6 d	133.6 d	134.0 d
5'	131.1 d	131.3 d	131.4 d
6'	130.0 d	129.9 d	130.2 d
1''	176.5 s	175.8 s	175.6 s
2''	31.9 t	32.0 t	32.8 t
3''	33.5 t	33.6 t	34.0 t
4''	175.0 s	174.8 s	175.6 s
OMe-1	57.7 q	58.0 q	58.0 q
OMe-6	55.7 q	56.0 q	56.2 q
OMe-8	58.0 q	—	58.3 q
OMe-14	58.1 q	—	58.3 q
OMe-16	56.2 q	56.3 q	56.0 q

places. The  $^{13}\text{C}$  NMR (Table) clearly showed that compound **2** is 14-demethyllycaconitine.

The  $^1\text{H}$  NMR spectrum of N-deethyllycaconitine-N-aldehyde (**3**) showed the presence of an aldehyde group at  $\delta$  9.85 (1H, s), and the lack of the methyl triplet at around  $\delta$  1.10 suggested that the aldehyde group should be situated at N as N-CHO instead of an ethyl group as N- $\text{CH}_2\text{CH}_3$ . Four methoxy groups and all other signals were similar to those of compounds **1** and **2**. The  $^{13}\text{C}$  NMR (Table) correlated to the given structure. The molecular formula  $\text{C}_{35}\text{H}_{44}\text{N}_2\text{O}_{11}$  was calculated from its HRMS ( $m/z$  668.2947).

### 3. Experimental

#### 3.1. Equipment

UV spectra were recorded in MeOH on a Shimadzu UV 1601 spectrophotometer. IR spectra were recorded in  $\text{CHCl}_3$  on a Perkin-Elmer 983 spectrophotometer. Optical rotations were determined in Opt. Act. Ltd. AA-5 polarimeter.  $^1\text{H}$  NMR (200 MHz) and  $^{13}\text{C}$  NMR (50 MHz) spectra were recorded on a Bruker AC-200 instrument. EIMS and HRMS were recorded on a ZapSpec instrument.

#### 3.2. Plant material

The roots (1 kg dry wt) of *Aconitum leave* Royle were collected and identified by one of us (H.A.) in Swat district of Northern Pakistan, at an elevation 1200 m, in August 2000. A voucher specimen is deposited in the Herbarium of Department of Botany, Jahan Zeb College, Saidu Sharif, Swat, Pakistan (RA-01).

#### 3.3. Extraction and isolation

The powdered plant material was first extracted with n-hexane yielding 8 g of an oily residue, then extracted with 90% EtOH. The concentrated EtOH extract was acidified with 5% HCl and extracted with  $\text{CHCl}_3$ . The remaining aqueous EtOH was basified with 10% NaOH and extracted with  $\text{CHCl}_3$  to yield 26 g of a crude alkaloidal mixture. A part of this mixture (5 g) was fractionated by VLC using a basic alumina column (100 g,  $\text{Al}_2\text{O}_3$  EM 1085) eluting with petroleum ether, and gradients  $\text{CH}_2\text{Cl}_2$  up to 100% and EtOH 10%. Fractions 1–8 were oily and discarded, fractions 9–14 (2 g) (petroleum ether-dichloromethane, 1:1) yielded lappaconitine (**4**) (2 g). Other fractions were mixtures of alkaloids and could be separated on Si gel plates of a Chromatotron apparatus. Eluting with  $\text{CH}_2\text{Cl}_2$  and a gradient of EtOH starting from 1% and by a slow increase up to 10%. Fractions 15–21 (1.3 g) yielded lycaconitine (**5**, 50 mg), 8-methyllycaconitine (**1**, 12 mg), 14-demethyllycaconitine (**2**, 8 mg), fractions 22–29 (270 mg) yielded N-deethyllycaconitine-N-aldehyde (**3**, 7 mg), fractions 30–36 (1.6 g) yielded lapaconidine (**6**, 17.3 mg) and lycocotinine (**7**, 12 mg). The known compounds were identified by comparing their  $^1\text{H}$  and  $^{13}\text{C}$  NMR data to those of authentic compounds and co-TLC behavior with standards.

##### 3.3.1. 8-Methyllycaconitine (**1**)

$\alpha_D^{25} = +19.5^\circ$  ( $c = 0.1$ ); UV  $\lambda_{\text{max}}^{\text{MeOH}}$  (log  $\epsilon$ ) nm: 278 (2.10), 230 (4.00); IR  $\nu_{\text{max}}^{\text{CHCl}_3}$   $\text{cm}^{-1}$ : 3460 (OH), 1710 (C=O), 1600, 1580, 1515 (arom. C–H), 1070 (C–O);  $^1\text{H}$  NMR (see text);  $^{13}\text{C}$  NMR (Table); EIMS (rel. int) at  $m/z$  682 [ $\text{M}$ ] $^+$  (2), 667 [ $\text{M-Me}$ ] $^+$  (7), 637 [ $\text{M-Me-OMe}$ ] $^+$  (50), 623 (18), 584 [ $\text{M-C}_4\text{H}_4\text{NO}_2$ ] $^+$  (8), 480 [ $584\text{-C}_7\text{H}_4\text{O}$ ] $^+$  (7), 464 [ $584\text{-C}_7\text{H}_4\text{O}_2$ ] $^+$  (35), 406 (12), 358 (15), 202 [ $\text{C}_{11}\text{H}_8\text{NO}_3$ ] $^+$  (100), 174 (30), 101 (35), 84 (65), 71 (53). HRMS: 682.3467  $\text{C}_{37}\text{H}_{50}\text{N}_2\text{O}_{10}$  (calcd. 682.3464).

##### 3.3.2. 14-Demethyllycaconitine (**2**)

$\alpha_D^{25} = +20.0^\circ$  ( $c = 0.1$ ); UV  $\lambda_{\text{max}}^{\text{MeOH}}$  (log  $\epsilon$ ) nm: 278 (2.05), 225 (4.10); IR  $\nu_{\text{max}}^{\text{CHCl}_3}$   $\text{cm}^{-1}$ : 3450 (OH), 1715 (C=O), 1605, 1580, 1510 (arom. C–H), 1075 (C–O), 880, 790, 657;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$ : 8.08 (1H, dd,  $J = 1.5$  and 8.0 Hz, H-6'), 7.70 (1H, dt,  $J = 1.5, 8, 8$  Hz) 7.56 (1H, dt,  $J = 1.5, 8, 8$  Hz) (H-4' and H-5'), 7.28 (1H, dd,  $J = 1.5$  and 8 Hz, H-3'). The  $^{13}\text{C}$  NMR spectrum is in agreement with the suggested structure; EIMS (rel. int) at  $m/z$ : 654 [ $\text{M}$ ] $^+$  (12), 623 [ $\text{M-OMe}$ ] $^+$  (7), 452 [ $\text{M-C}_{11}\text{H}_8\text{NO}_3$ ] $^+$  (55), 436 [ $\text{M-C}_{11}\text{H}_8\text{NO}_4$ ] $^+$  (20), 284 (15), 256 (20), 202 [ $\text{C}_{11}\text{H}_8\text{NO}_3$ ] $^+$  (60), 97 (33), 83 (40), 71 (72). HRMS: 654.3152  $\text{C}_{35}\text{H}_{46}\text{N}_2\text{O}_{10}$  (calcd. 654.3155).

##### 3.3.3. N-Deethyllycaconitine-N-aldehyde (**3**)

$\alpha_D^{25} = +22.5^\circ$  ( $c = 0.1$ ); UV  $\lambda_{\text{max}}^{\text{MeOH}}$  (log  $\epsilon$ ) nm: 280 (2.10), 230 (4.10); IR  $\nu_{\text{max}}^{\text{CHCl}_3}$   $\text{cm}^{-1}$ : 3450 (OH), 2850, 1730 (CHO), 1710 (C=O), 1605, 1580, 1520 (arom. C–H), 1080 (C–O);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$ : 8.05 (1H, dd,

$J = 1.8, 8 \text{ Hz}$ ), 7.68 (1H, dt,  $J = 1.8, 8, 8 \text{ Hz}$ ), 7.54 (1H, dt,  $J = 1.8, 8, 8 \text{ Hz}$ ), 7.28 (1H, dd, 1.8, 8 Hz) for H-6', H-5', H-4', H-3' respectively, four methoxy groups at  $\delta$  3.44, 3.42 (each 3H, s), 3.36 (6H, s,  $2 \times \text{OMe}$ ). The  $^{13}\text{C}$  NMR signals (Table) are in agreement with the suggested structure; EIMS (rel. int) at  $m/z$  668  $[\text{M}]^+$  (3), 653  $[\text{M-Me}]^+$  (5), 622  $[\text{M-Me-OMe}]^+$  (5), 593  $[\text{622-CHO}]^+$  (12), 535 (18), 466  $[\text{M-C}_{11}\text{H}_8\text{NO}_3]^+$  (30), 401 (65), 355 (80), 341 (92), 202 (100), 97 (28), 84 (45), 71 (80). HRMS: 668.2946  $\text{C}_{35}\text{H}_{44}\text{N}_2\text{O}_{11}$  (calcd. 668.2947).

Acknowledgements: Turkish authors thank to the Research Fund of the University of Istanbul No Ö-1026, Dr. M. Arfan thanks to University of Peshawar for the Research grant given for this study.

#### References

- Benn, M. H.; Jacyno, J. M.; Pelletier, S. W. (Ed.): *Alkaloids: Chemical and Biological Perspectives*, vol. 1, p. 153, John Wiley and sons, New York 1983
- Bisset, N. G.: *J. Ethnopharmacol.* **4**, 247 (1981)
- Kim, D. K.; Kwon, H. Y.; Lee, K. R.; Zee, O. I.: *Arch. Pharmacol. Res.* **21**, 344 (1998)
- Schilstrom, B.; Svensson, H.M.; Svensson, T. H.; Nomikos, G. G.: *Neuroscience* **85**, 1005 (1998)
- Jacyno, J. M.; Harwood, J. S.; Lin, N.H.; Cambell, J.E.; Sullivan, J. P.; Holladay, M. W.: *J. Nat. Prod.* **59**, 707 (1996)
- Gunther, R. T.: *The Greek Herbal of Dioscorides*, p. 316–317, Hafner Publishing company, London, New York 1968
- Chiao, H.; Pelletier, S. W.; Desai, H. K.; Rabagay, N. R.; Caldwell, R. W.: *Eur. J. Pharmacol.* **283**, 103 (1995)
- Desai, H. K.; Hart, B. P.; Caldwell, R.W.; Huang, J.; Pelletier, S. W.: *J. Nat. Prod.* **61**, 743 (1998)
- Ulubelen, A.; Arfan, M.; Sönmez, U.; Meriçli, A. H.; Meriçli, F.: *Phytochemistry* **47**, 1147 (1998)
- Ulubelen, A.; Arfan, M.; Sönmez, U.; Meriçli, A.H.; Meriçli, F.: *Phytochemistry* **48**, 385 (1998)
- Ulubelen, A.; Desai, H. K.; Teng, Q.; Meriçli, A. H.; Meriçli, F.; Kolak, U.; Arfan, M.; Lee, C. K.; Pelletier, S. W.: *Heterocycles* **51**, 1843 (1999)
- Ulubelen, A.; Meriçli, A. H.; Meriçli, F.; Kolak, U.; Arfan, M. Ahmad, M.; Ahmad, H.: *Heterocycles* **53**, 2279 (2000)
- Handa, V.; Handa, K. L.: *Planta Med.* **30**, 177 (1964)
- Jiang, S.-H.; Zhu, Y.-L.; Zhu, R.-H.: *Acta Pharm. Sinica* **17**, 282 (1982)
- Pelletier, S. W.; Mody, N. V.; Sawhney, R. S.: *Can. J. Chem.* **57**, 1652 (1979)
- Desai, H. K.; Trumbull, E. R.; Pelletier, S. W.: *J. Chromatogr.* **366**, 439 (1986)

Received September 18, 2001

Accepted November 13, 2001

Ayhan Ulubelen  
Faculty of Pharmacy  
University of Istanbul  
Beyazıt 34452  
Istanbul  
Turkey  
aulubelen@yahoo.com