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## 2-(Arylpropionylamino)- and 2-(arylacryloylamino)benzophenones: Farnesyltransferase inhibition and antimalarial activity

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Structural variation of the 2-acylamino moiety of some benzophenone farnesyltransferase inhibitors led to the *para*-trifluoromethylphenylpropionyl derivative with relatively low farnesyltransferase inhibition but considerable antimalarial activity and no cytotoxicity.

### 1. Introduction

Farnesyltransferase is involved in the post-translational modification of numerous proteins of which the majority has important functions in intracellular signal transduction. Farnesyltransferase catalyzes the transfer of a farnesyl residue from farnesyl pyrophosphate to the thiol of a cysteine side chain of the protein substrate. The cysteine residue is part of a characteristic carboxy-terminal consensus sequence, the so-called CAAX box (C, cysteine; A, amino acid with aliphatic side chain; X, serine or methionine) (Fu and Casey 1999; Wittinghofer and Waldmann 2000a, b; Bell 2000). Various inhibitors of the farnesyltransferase have been developed as potential cancer therapeutics. The compounds of several pharmaceutical companies are in advanced stages of clinical studies (Cox and Der 2002; Purcell and Donehower 2002).

In addition to mammals, farnesyltransferases were also identified in other eukaryotic organisms including pathogenic protozoa of the genera *Plasmodium* (Chakrabarti et al. 1998, 2002), *Trypanosoma* (Yokoyama et al. 1998, 2000; Bruckner et al. 2002), *Leishmania* (Bruckner et al. 2002) and *Toxoplasma* (Ibrahim et al. 2001). Therefore, inhibition of the farnesyltransferase has also been suggested as new strategy for the treatment of parasitic infections (Cox and Der 2002). The most important of these protozoa caused diseases is Malaria tropica caused by the infection with *Plasmodium falciparum*. Approximately 40% of the world population lives in areas with malaria risk, and 2 to 3 million people die each year from malaria. Because of the increasing spread of malaria parasites resistant to chloroquine and other commonly used antimalarials there is an urgent need for new therapeutics (Sachs and Malaney 2002; Ridley 2002).

We have developed a novel class of farnesyltransferase inhibitors based on a benzophenone scaffold (Schlitzer 2002). In course of our studies towards the establishment of structure activity relationships of this type of compounds we identified inhibitors **4a** and **5a** as promising lead structures especially because of their inhibition of yeast farnesyltransferase with IC<sub>50</sub>-values of 40 nM and

5 nM, respectively (Kettler et al. 2003). In addition, the phenylpropionic acid derivative **4a** displayed an interesting *in vitro* antimalarial activity with an IC<sub>50</sub> of 310 nM (Wiesner et al. 2003b). In the present study it was our objective to see how structural variations of phenylpropionic and the cinnamic acid residue at the 2-amino group of the benzophenone core influence farnesyltransferase inhibition as well as antimalarial activity.

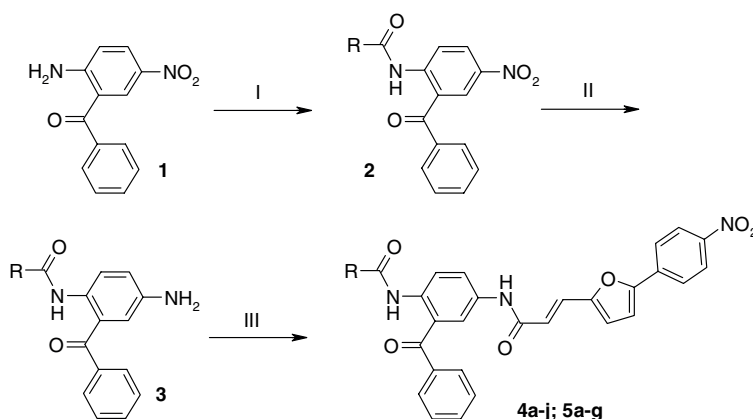
### 2. Investigations and results

Target compounds were prepared from the commercially available 2-amino-5-nitrobenzophenone **1**, which was first acylated at the 2-amino group by appropriate 3-arylpropionic acid chlorides and cinnamic acid chlorides, respectively (Scheme 1). Then, the 5-nitro group was reduced and the resulting amino function was acylated by 3-[5-(4-nitrophenyl)-2-furyl]acrylic acid chloride (Böhm et al. 2001). *Para*-nitro and trifluoromethyl cinnamic acid were obtained from the corresponding benzaldehydes by Knoevenagel condensation. *Para*-, *meta*- and *ortho*-trifluoromethylphenylpropionic acid were prepared by catalytic hydrogenation of the appropriate cinnamic acid derivatives which in turn were obtained from the aldehydes. 4-Nitrophenylpropionic acid was obtained by nitration of phenylpropionic acid (Moloney et al. 1999).

Because of the reduction step involved in the synthesis according to Scheme 1, an alternative route had to be followed for the preparation of the nitro compounds **4k** and **5h** (Scheme 2). First, the 2-amino group of **1** was protected as trifluoroacetamide (**6**). After reduction of the 5-nitro group, the resulting amine **7** was acylated with 3-[5-(4-nitrophenyl)-2-furyl]acrylic acid chloride. After removal of the protective group from **8** the resulting intermediate **9** (Kettler et al. 2003) was acylated by 4-nitrophenylpropionic acid chloride and 4-nitrocinnamic acid chloride, respectively.

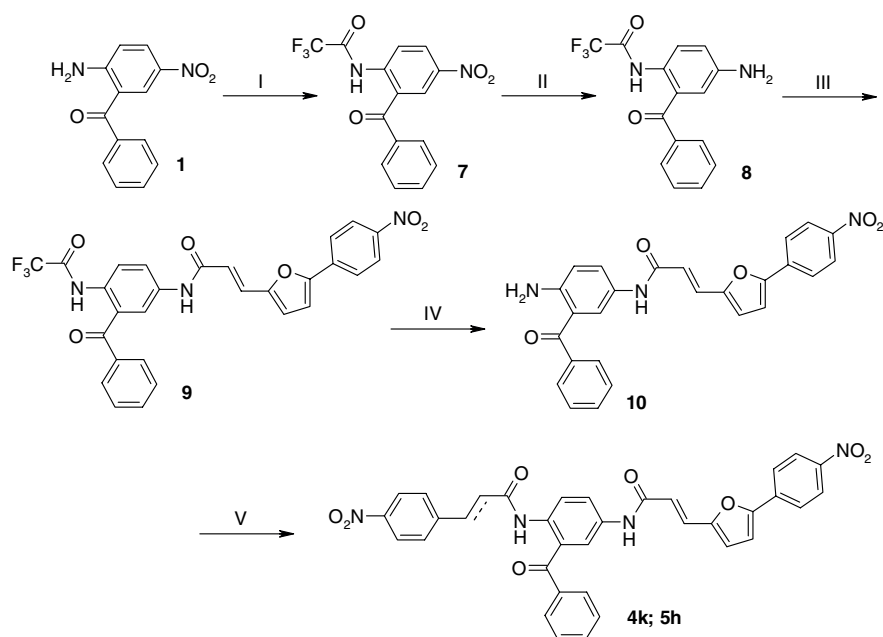
The farnesyltransferase inhibitory activity of the inhibitors was determined using the fluorescence enhancement assay as described by Pompliano et al. (1992). The assay employs yeast farnesyltransferase (FTase) fused to glutathione S-

Scheme 1



(I)  $R-CO-Cl$ , toluene/dioxane, reflux, 2h; (II)  $SnCl_2 \times 2 H_2O$ , EtOAc, reflux 2h; (III) 3-[5-(4-nitrophenyl)-2-furyl]acrylic acid chloride, toluene/dioxane, reflux, 2h

Scheme 2



(I) TFAA, DCM/pyridine, 0 °C, 2h; (II)  $SnCl_2 \times 2 H_2O$ , EtOAc, reflux 2h; (III) 3-[5-(4-nitrophenyl)-2-furyl]acrylic acid chloride, toluene/dioxane, reflux, 2h; (IV)  $K_2CO_3$ , dioxane/ $H_2O$ , reflux, 3h; (V)  $O_2N-C_6H_4-(CH_2)_2-COCl$  or  $O_2N-C_6H_4-CH=CH-COCl$ , toluene/dioxane, reflux, 2h

transferase at the N-terminus of the  $\beta$ -subunit (Del Villar et al. 1997). The heterologous expression of the farnesyltransferase genes from *P. falciparum* has not been achieved so far and, therefore, no recombinant enzyme is available for routine screening (Chakrabarti et al. 2002).

Farnesylpyrophosphate and the dansylated pentapeptide Ds-GlyCysValLeuSer were used as substrates. Upon farnesylation of the cysteine thiol, the dansyl residue is placed into a lipophilic environment. The resulting enhancement of fluorescence at 505 nm is used to monitor the enzyme reaction.

The farnesyltransferase inhibitory activities of compounds **4** and **5** are displayed in the Table. In the case of the phenylpropionyl derivatives **4** as well as with the cinnamoyl derivatives **5** all new compounds turned out to be considerably less active than the initial unsubstituted leads

**4a** and **5a**. Clear structure activity relationships cannot be delineated.

Compounds **4** and **5** were assayed for their inhibitory activity against intraerythrocytic forms of *P. falciparum* strain Dd2 using a semi-automated microdilution assay (Desjardins et al. 1979; Trager and Jensen 1976; Ancelin et al. 1998). The growth of the parasites was monitored through the incorporation of tritium labeled hypoxanthine. Comparability of different experiments was ensured by concurrent assay of standard compounds. The Dd2 strain used for the inhibition assays is resistant to several commonly used anti-malarial drugs (chloroquine, cycloguanil and pyrimethamine) (Table).

Activity of the cinnamic acid substituted derivatives **5** against the intraerythrocytic forms of *P. falciparum* was

Table: Farnesyltransferase inhibition and anti-malarial activity<sup>a, b</sup> of compounds 4 and 5

Compd.	R	IC <sub>50</sub> (nM) FTase	IC <sub>50</sub> (nM) <i>P. falciparum</i>	Compd.	R	IC <sub>50</sub> (nM) FTase	IC <sub>50</sub> (nM) <i>P. falciparum</i>
<b>4a</b>		40	310	<b>5a</b>		5	2300
<b>4b</b>		215	3250				
<b>4c</b>		435	1300	<b>5b</b>		136	21000
<b>4d</b>		411	440	<b>5c</b>		150	8300
<b>4e</b>		1378	61	<b>5d</b>		224	20000
<b>4f</b>		300	625				
<b>4g</b>		900	1080				
<b>4h</b>		646	440	<b>5e</b>		184	27000
<b>4i</b>		231	125	<b>5f</b>		343	42000
<b>4j</b>		900	165	<b>5g</b>		169	42000
<b>4k</b>		178	1400	<b>5h</b>		71	10000

<sup>a</sup> Activity was assayed by measuring radioactive hypoxanthine uptake by the multi-resistant *P. falciparum* strain Dd2. IC<sub>50</sub> values (nM) for standard antimalarials were: chloroquine, 170; pyrimethamine, 2500; cycloguanil, 2200; quinine, 380; lumefantrine, 30; artemisinin, 18

<sup>b</sup> Antimalarial activity of compounds **4a–k** has been published elsewhere (Wiesner et al. 2003b)

generally very low. This can be mainly attributed to the low solubility of this type of compounds in the culture medium which most probably prevents effective drug levels inside the parasite.

In case of the phenylpropionyl derivatives **4** antimalarial activity changes considerably with the substitution on the terminal phenyl residue. While the hydroxy- (**4b**), methoxy- (**4c**) and nitro- (**4k**) derivatives are markedly less active than the lead, the methyl- (**4d**) and the fluoro- (**4h**) derivatives are roughly equipotent to the lead **4a**. However, the chloro- (**4i**) and the bromo- (**4j**) derivatives are considerably more active while the trifluoromethyl derivative (**4e**) is the most active compound of this series dis-

playing an IC<sub>50</sub>-value of 61 nM which is already in an interesting range.

Shifting the trifluoromethyl group from the para- to the meta- (**4f**) or ortho- (**4g**) position resulted in an increasing reduction in activity.

The phenylpropionyl substituted derivatives were assayed for potential cytotoxic activity using the MTT-assay employing HL-60 cell line. In this test, the reduction of a tetrazolium bromide to the corresponding formazane is used to measure cell viability because the reaction occurs only in active mitochondria and therefore in living cells. Compounds were assayed in concentrations between 10<sup>-9</sup> to 10<sup>-4</sup> mol/l. Apart from low activity in the micromolar

range for the meta- and ortho-trifluoromethyl derivatives (**4f**, **g**), no cytotoxic activity was observed especially not for the most active antimalarial compound **4e**.

### 3. Discussion

Although the newly prepared cinnamoyl derivatives **5b–g** displayed fair farnesyltransferase activity their unexpected low solubility prevented any relevant antimalarial activity. Therefore, this particular substructure was no longer considered for further studies.

With respect to antimalarial activity structure activity relationships observed with the phenylpropionic acid derivatives **4** closely resemble those observed in a series of structurally closely related inhibitors having an arylacetic acid substructure at 2-amino group of the benzophenone core (Wiesner et al. 2003a). In both cases, good activity was found with the chloro- and the bromo-substituent while the para-trifluoromethyl derivatives are the most active compounds in the series. Correlation between farnesyltransferase inhibitory and antimalarial activity is generally bad (Fig.) but one has to keep in mind that the activity against an isolated enzyme is compared with the activity against an intracellular living complete organism. Furthermore, since farnesyltransferase of *P. falciparum* is not available for routine screening, FTase of a different species (yeast) has to be used. Although amino acid sequences regarding the enzymes active site are quite similar, there are some differences.

Our general experience is that good farnesyltransferase inhibitors not necessarily need to be active antimalarial agents (Wiesner et al. 2003c). From the present set of data this conclusion has to be modified in an important aspect. The bromo and the trifluoromethyl compound show that relatively weak farnesyltransferase inhibitors can also be considerably active as antimalarials. This argues strongly for the possibility of the development of specific farnesyltransferase inhibitors with relative specificity against *P. falciparum*. One could argue that the activity of those compounds might originate from another mechanism than farnesyltransferase inhibition. However, we have shown with structurally closely related compounds that they in fact inhibit the protein farnesylation in *P. falciparum* cultures (Wiesner et al. 2004).

In addition it is important to note that the most active antimalarial compound **4e** displayed no measurable cytotoxic activity in the MTT-assay.

In conclusion, the results presented encourage further development of benzophenone-based farnesyltransferase inhibitors as potential antimalarial agents.

### 4. Experimental

#### 4.1. Preparation

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Jeol Eclipse 400 and a Jeol Eclipse 500 spectrometer. Mass spectra were obtained with a PE Biosystems API 2000. IR spectra were recorded on a JASCO FT/IR-410 Fourier Transform Infrared Spectrometer. Microanalyses were obtained from an elemental vario el and were within ± 0.4% of the calculated values. Melting points were obtained with a Reichert Austria microscope and are uncorrected. Liquid chromatography was carried out using silica gel 60 from ICM Silitech. The preparation of compounds **4a** and **5a** has been described elsewhere (Kettler et al. 2003)

#### 4.1.1. General procedure 1: Activation of various acids as acid chlorides and reaction with aromatic amines

The various carboxylic acids were dissolved in toluene and 0.1 mL SOCl<sub>2</sub> per mmol acid was added. The mixture was heated under reflux for 2 h and the volatiles were evaporated *in vacuo*. The resulting acyl chlorides were dissolved in toluene or dioxane (approx. 10 mL) and added to a solution of the appropriate aromatic amine in hot toluene (approx. 50 mL). The mixtures were heated under reflux for 2 h. Then, the solvent was removed *in vacuo* and the crude products were purified by recrystallisation from ethanol.

#### 4.1.2. General procedure 2: Reduction of aromatic nitro compounds

Aromatic nitro compounds **2** were dissolved in EtOAc (50–100 mL) and SnCl<sub>2</sub> × 2 H<sub>2</sub>O (1.125 g per mmol nitro compound) was added. The mixture was heated under reflux for 2 h. Then, NaHCO<sub>3</sub>-solution was added until pH 7–8 was reached and the organic layer was separated. The aqueous layer was extracted two times with EtOAc. The combined organic layers were washed with brine and dried over MgSO<sub>4</sub>. Then, the solvent was removed *in vacuo* to obtain the crude products.

#### 4.1.3. Compounds **2a–o**

The compounds **2a–o** were prepared from 2-amino-5-nitrobenzophenone **1** and the appropriate 3-arylpropionic acid chloride or cinnamic acid chloride according to general procedure 1.

##### 4.1.3.1. *N*-(2-Benzoyl-4-nitrophenyl)-3-(4-hydroxyphenyl)propionic acid amide (**2a**)

Yield 60%. <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 2.76 (t, <sup>3</sup>J = 8.0 Hz, 2 H, CH<sub>2</sub>), 2.99 (t, <sup>3</sup>J = 8.0 Hz, 2 H, CH<sub>2</sub>), 4.85 (s, 1 H, OH), 6.71 (m, 2 H, Ar-H), 7.10 (m, 2 H, Ar-H), 7.56 (m, 2 H, Ar-H), 7.69 (m, 3 H, Ar-H), 8.39 (dd, <sup>3</sup>J = 8.8 Hz, <sup>4</sup>J = 2.0 Hz, 1 H, Ar-H), 8.43 (d, <sup>4</sup>J = 2.0 Hz, 1 H, Ar-H), 8.89 (d, <sup>3</sup>J = 8.8 Hz, Ar-H), 11.00 (s, 1 H, NH).

##### 4.1.3.2. *N*-(2-Benzoyl-4-nitrophenyl)-3-(4-methoxyphenyl)propionic acid amide (**2b**)

Yield 57%. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ (ppm) = 2.76 (t, <sup>3</sup>J = 7.5 Hz, 2 H, CH<sub>2</sub>), 3.03 (t, <sup>3</sup>J = 7.5 Hz, 2 H, CH<sub>2</sub>), 3.71 (s, 3 H, OCH<sub>3</sub>), 6.78 (m, 2 H, Ar-H),

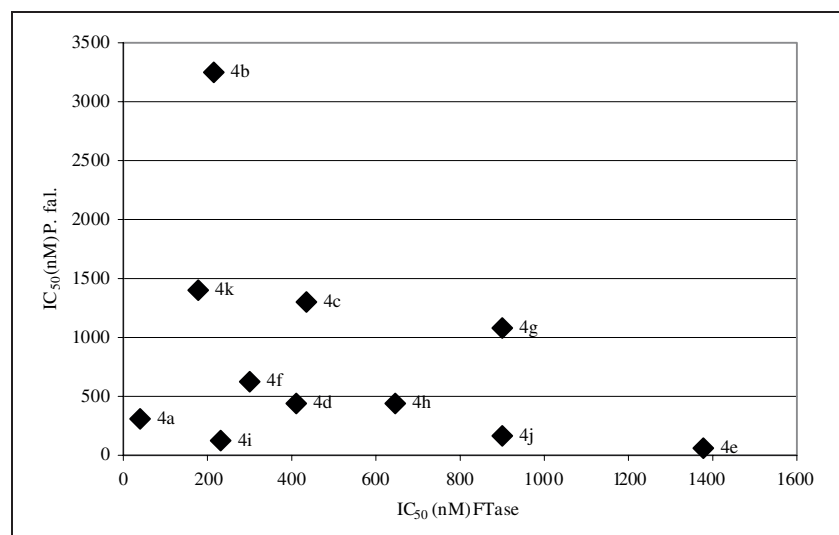


Fig.: Correlation between farnesyltransferase inhibition and antimalarial activity for compounds **4a–k**











4.5. *In vitro* measurement of *P. falciparum* parasite growth inhibition

Compounds were tested by a semiautomated microdilution assay against intraerythrocytic forms of *P. falciparum* (Desjardins et al. 1979). The *P. falciparum* strain Dd2 was cultivated by a modification of the method described by Trager and Jensen (1976). The culture medium consisted of RPMI 1640 supplemented with 10% human type 0<sup>+</sup> serum and 25 mM HEPES. Human type 0<sup>+</sup> erythrocytes served as host cells. The cultures were kept at 37 °C in an atmosphere of 5% O<sub>2</sub>, 3% CO<sub>2</sub>, and 92% N<sub>2</sub>.

Drug testing was carried out in 96-well microtiter plates. The compounds were dissolved in DMSO (10 mM) and prediluted in complete culture medium (final DMSO concentrations ≤ 1%) (In order to avoid a loss of lipophilic test compounds by adsorbance to the plastic material used for the assay, complete culture medium containing erythrocytes was used to dilute the DMSO stock solutions). Infected erythrocytes (200 µL per well, with 2% hematocrit and 0.4% parasitemia, predominantly ring-stage parasites) were incubated in duplicate with a serial dilution of the drugs for 48 h (Ancelin 1998). After the addition of 0.8 µCi [<sup>3</sup>H]-hypoxanthine in 50 µL medium per well, the plates were further incubated for 24 h. Cells were collected on glass fiber filters with a cell harvester (Micromate 196, Packard) and incorporated radioactivity measured using a β-counter (Matrix 9600, Packard).

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