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# Cytotoxic activity of nepetin, a flavonoid from *Eupatorium ballotaefolium* HBK

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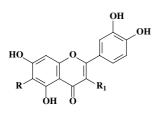
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The present study evaluated the cytotoxic activity of nepetin and quercetin-3-*O*-glucoside, compounds isolated from the aerial parts of *Eupatorium ballotaefolium*. The antimitotic activity was determined as the ability to inhibit sea urchin eggs development and five tumor cells lines growth. Moreover, the activities of these compounds were compared to quercetin in the same models. Nepetin inhibited the proliferation of the five tumor cell lines, once quercetin-3-*O*-glucoside did not present any activity even at the highest tested concentration and quercetin only inhibited proliferation of the B16 cell line. On the sea urchin assay, nepetin and quercetin induced a dose-dependent inhibition on egg development, while quercetin-3-*O*-glucoside did not modify normal egg cleavage, even at the highest tested concentration (100  $\mu$ g/ml).

## 1. Introduction

The genus Eupatorium (Asteraceae) is constituted by a taxonomically complex group of nearly 600 species distributed mainly in the tropical regions of the Americas (Maia et al. 2002). Several plants of this genus are widely used in folk medicine in different parts of the world due to their astringent, antirheumatic, antimicrobial, hepatoprotective, disinfectant and analgesic properties. They are used in the treatment of fever, headache, stomach ulcer, diarrhea and malaria (Lang et al. 2001; El-Seedi et al. 2002). Concomitantly, extracts of *Eupatorium* species have been shown several pharmacological activities, including hepatotoxic, cytotoxic, antiinflammatory, antioxidant and antigonorrhoeal effects (Mongelli et al. 2000; Kaushal et al. 2001). In fact, a number of bioactive compounds have been isolated of Eupatorium (Habtemarian 1998; El-Seedi et al. 2002). Thus, the genus seems to be a promising resource of new drugs and value-added products.



(1) R = OMe, R<sub>1</sub> = H
(2) R = H, R<sub>1</sub> = O-glucosyl
(3) R = H, R<sub>1</sub> = OH

In the course of our continuous search for cytotoxic agents from natural sources we have investigated extracts of plants from the Northeastern Brazilian flora. Since the chloroform fraction of the aerial parts of *E. ballotaefolium* showed cytotoxic activity, this was used as the rationale for the current study to identify compounds responsible for this effect. Herein we describe the isolation of nepetin (1) and quecertin-3-*O*-glucoside (2), as well as the study of their cytotoxic activity on tumor cell lines and on sea urchin eggs. The related compound, quercetin (3), previously isolated from *Lippia sidoides* (Costa et al. 2001) has been included in the biological tests for comparison purposes, since the two isolated compounds were quite similar to this well-known flavonoid (Middleton Jr et al. 2000).

### 2. Investigations, results and discussion

Nepetin (1) inhibited the proliferation of five tumor cell lines (Table 1), once quercetin-3-*O*-glucoside (2) did not present any activity even at the highest concentration tested. Quercetin (3) only inhibited the proliferation of B16 cell line with an IC<sub>50</sub> of 8.54  $\mu$ g/ml.

The study of alterations in sea urchin egg development is a suitable model for detecting cytotoxic, antimitotic and teratogenic activities of new compounds (Jacobs et al. 1981). Nepetin (1) and quercetin (3) induced a dose-dependent inhibition on egg development, while quercetin-3-O-glucoside did not modify normal egg cleavage, even at the highest tested concentration (100 µg/ml). The IC<sub>50</sub> values are presented in Table 2.

The antitumor potential of flavonoids has been extensively discussed in the literature. Edwards et al. (1979) concluded

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Substances	CEM µg/ml	HL-60 µg/ml	HCT-8 µg/ml	MCF-7 µg/ml	B-16 µg/ml
Doxorubicin	0.021	0.016	0.014	0.110	0.032
	0.016-0.026	0.013-0.019	0.011-0.018	0.097-0.116	0.024 - 0.045
(1)	8.24	6.42	7.18	14.18	9.42
	6.34-10.66	5.48-7.52	5.66-9.12	13.29-15.12	8.96-9.90
(2)	>25	>25	>25	>25	>25
(3)	>25	>25	>25	>25	8.24 (5.86-12.44)

Table 1: Antiproliferative effects of nepetin (1), quercetin-3-O-glucoside (2) and quercetin (3) on tumoral cell lines

Data are presented as IC<sub>50</sub> values and 95% confidence interval obtained by non-linear regression for leukemias (HL-60 and CEM), breast (MCF-7), colon (HCT-8) and skin (B-16) cancer cells from 3 independent experiments.

 Table 2: Antimitotic activity of nepetin (1), quercetin-3-O-glucoside (2) and quecetin (3) on sea urchin (Lytechinus variegatus) eggs development

Substances	1 <sup>st</sup> cleavage	3 <sup>rd</sup> cleavage	Blastulae
	μg/ml	μg/ml	μg/ml
Doxorubicin	6.3	0.3	0.5
	4.3–9.1	0.2-0.7	0.3–1.1
(1)	12.02	3.15	3.82
	9.00–16.06	2.08–4.78	2.69–5.44
(2) (3)	>100.0 94.74 85.19–105.4	>100.0 54.33 47.59-62.03	>100.0 44.19 25.05-77.95

Data are presented as IC\_{50} values and 95% confidence interval obtained by non-linear regression for first and third cleavages, and blastulae from 3 independent experiments.

that flavonoids were not a promising group of antitumor compounds. On the other hand, Beutler et al. (1996) showed that, in fact, there are some structure-activity requirements for flavonoid cytotoxicity, at least when this activity is related to the inhibition of tubulin polimerization. According to several authors, the presence of a 3-methoxyl substitution increases the cytotoxicity of flavonoids (Beutler et al. 1996; Middleton Jr et al. 2000; Costa-Lotufo et al. 2003). Costa-Lotufo et al. (2003) described the cytotoxicity of kaempferol and isokaempferide using the same assays. These two flavonoids differ only by the presence of a methoxyl group on C-3 in isokaempferide instead of a hydroxyl group present in many flavon-3-ols like kaempferol, while neptin possesses a methoxyl group on C-6. Isokaempferide was at least three times more active than kaempferol against tumor cell lines, corroborating this theory at least when the cytotoxicity is determined on tumor cell lines. In the sea urchin assay, kaempferol and isokaempferide presented the same activity. The activity of nepetin (1) on tumor cell lines was stronger than that of kaempferol, but was weaker compared to isokaempferide and, in the sea urchin assay, nepetin was as active as kaempferol and isokaempferide. Thus, the presented data suggest that the presence of a methoxyl group on C-6 could also be related to an increased cytotoxicity of flavonoids.

## 3. Experimental

#### 3.1. Extraction and isolation

Aerial parts of *E. ballotaefolium* were collected from their natural habitat on Meruoca Mountain, State of Ceará, in July 2001. The botanical material was authenticated by Prof. Edson P. Nunes of the Departamento de Biologia. A voucher specimen (# 27646) has been deposited at the Herbarium Prisco Bezerra (EAC), Universidade Federal do Ceará. The air-dried and powdered aerial parts of *E. ballotaefolium* (1.2 kg) were exhaustively extracted with *n*-hexane and EtOH at room temperature to give 21.6 g and 50.3 g of the respective extracts, after evaporation of the solvents under reduced pressure. The EtOH extract was fractionated over Si gel using CHCl<sub>3</sub>, EtOAc and MeOH. The CHCl<sub>3</sub> fraction was repeatedly chromatographed over Si gel, eluting with *n*-hexane and *n*-hexane-EtOAc (100:  $0 \rightarrow 0$ : 100) to yield nepetin (1, 469.4 mg). The EtOAc fraction was chromatographed on a Silica gel column eluted with CHCl<sub>3</sub>, CHCl<sub>3</sub>, EtOAC (100:0 $\rightarrow$ 0:100), and MeOH to yield quercetin-3-*O*-glucoside (2, 27.8 mg). The structures of the compounds 1 and 2 were established by spectroscopic analysis and comparison with literature data (Wenkert and Gottlieb 1977; Agrawal 1989).

#### 3.2. Cytotoxicity assays

Nepetin (1), quercetin-3-*O*-glucoside (2) and quercetin (3) were tested for cytotoxic activity on five tumoral cell lines: B-16 (murine skin cancer), HCT-8 (human colon cancer), MCF-7 (human breast cancer) CEM and HL-60 (leukemia cancer) cell lines (Children's Mercy Hospital, Kansas City, MO, USA) using the 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2*H*-tetrazolium bromide assay (Mosmann 1983).

The sea urchin assay was performed essentially as described by Costa-Lotufo et al. (2002).

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#### References

Agrawal PK (1989) Carbon-13 NMR of Flavonoids, Elsevier Science Publishers B. V., Amsterdam, The Netherlands, 564p.

- Beutler JA, Hamel E, Vlietinck AJ, Haemers A, Rajan P, Roitman JN, Cardellina II JH, Boyd MR (1996) Structure-activity requirements for flavone cytotoxicity and binding to tubulin. J Med Chem 41: 2333– 2338.
- Costa SMO, Lemos TLG, Pessoa ODL, Pessoa C, Montenegro RC, Braz-Filho R (2001) Chemical constituents from *Lippia sidoides* and cytotoxic activity. J Nat Prod 64: 792–795.
- Costa-Lotufo LV, Jimenez PC, Wilke DV, Leal LKAM, Cunha GMA, Silveira ER, Canuto KM, Viana GSB, Moraes MEA, Moraes MO, Pessoa C (2003) Antiproliferative effects of several compounds isolated from *Amburana cearensis* A. C. Smith. Z Naturforsch 58c: 675–680.
- Edwards JM, Raffauf RF, LeQuesne PW (1979) Antineoplastic activity and cytotoxicity of flavones, isoflavones, and flavonones. J Nat Prod 42: 85–91.
- El-Seedi HR, Ohara T, Sata N, Nishiyama S (2002) Antimicrobial diterpenoids from *Eupatorium glutinosum* (Asteraceae). J Ethnopharmacol 81: 293–296.
- Habtemarian S (1998) Cistifolin, an integrin-dependent cell adhesion blocker from the anti-rheumatic herbal drug, gravel root (rhizome of *Eupatorium purpureum*). Planta Med 64: 683–685.
- Jacobs RS, White S, Wilson L (1981) Selective compounds derived from marine organisms: effects on cell division in fertilized sea urchin eggs. Fed Proc 40: 26–29.
- Kaushal V, Dawra RK, Sharma OP, Kurade NP (2001) Hepatotoxicity in rat induced by partially purified toxins from *Eupatorium adenophorum* (Ageratina adenophora). Toxicon 39: 615–619.
- Lang G, Passreiter CM, Medinila B, Castilo JJ, Witte L (2001) Non-toxic pyrrolizidine alkaloids from *Eupatorium semialatum*. Biochem Syst Ecol 29: 143–147.
- Maia JGS, Zoghbi MGB, Andrade HA, Silva MHL, Luz AIR, Silva JD (2002) Essential oils composition of *Eupatorium* species growing wild in the Amazon. Biochem Syst Ecol 30: 1071–1077.
- Middleton Jr E, Kandaswani C, Theoharides TC (2000) The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. Pharmacol Rev 52: 673–751.
- Mongelli E, Pampuro S, Coussio J, Salomon H, Ciccia G (2000) Cytotoxic and DNA interation activities of extracts from medicinal plants used in Argentina. J Ethnopharmacol 71: 145–151.
- Mosmann T (1983) Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Meth 16: 55–63.
- Wenkert E, Gottlieb HE (1977) Carbon-13 nuclear magnetic resonance spectroscopy of flavonoid and isoflavonoid compounds. Phytochemistry 16, 1811–1816.