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Comparative study of the essential oils and extracts of *Achillea fragrantissima* (Forssk.) Sch. Bip. and *Achillea santolina* L. (Asteraceae) from Egypt

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Essential oils obtained by steam distillation from the aerial parts of *Achillea fragrantissima* (Forssk.) Sch. Bip. and the flower, leaf and stem of *A. santolina* L. as well as their lipophilic constituents obtained by solvent extraction were analysed using GLC and GLC-mass spectrometry. Nineteen constituents in the essential oil of *A. fragrantissima* were identified, in addition to 41 compounds from its *n*-hexane-ether extract. The hydrodistilled oil and the solvent extract contain santolina alcohol, artemisia alcohol, artemisia ketone, *cis*-thujone and *trans*-thujone as major constituents. In *A. santolina* altogether 54 volatile components were detected. The major components were 1,8-cineole, fragranol, fragranyl acetate and terpin-4-ol. Furthermore, the essential oils and the *n*-hexane-ether extracts of the two plants were screened for their antimicrobial activity.

1. Introduction

The genus Achillea (Fam. Asteraceae) is represented by about 115 species in the temperate regions of the Northern hemisphere, mainly in North Africa, Southeast Europe, and Southwest Asia (Boulos 2002). Among this genus, A. fragrantissima (known by its Arabic name Qaysoom) and A. santolina (known locally by Beatheran) are represented in Egypt (Täckholm 1974). They are strongly fragrant perennial herbs and have been used by Bedouins as stomachic and anthelmintic (Boulos 1983). A. fragrantissima was found to be used as a hypoglycaemic drug (Yaniv et al. 1987). A. santolina is used treat toothache (Boulos 1983), chest disorders and as a tonic, carminative (Afsharypuor et al. 1996), insect repellent and in dysentery (Alkofahi et al. 1996). Previous investigations of these plants revealed the isolation and identification of some bioactive compounds (Haddad et al. 1960; Bohlmann and Jastrow, 1962; Shalaby and Steinegger, 1964; Shalaby et al. 1965; Khafagy et al. 1965; Khafagy et al. 1976; Khafagy et al. 1978; Ahmed et al. 1989; Seida et al. 1990; Mustafa et al. 1992; Balboul et al. 1997).

The chemical composition of the essential oil of *A. fra-grantissima* has been previously studied and the major constituents were reported to be santolina alcohol, artemisia ketone, *cis*-thujone and *trans*-thujone (Shalaby and Richter, 1964; El-Deeb 1985; Aboutabl et al. 1986a; 1986b; Fleisher and Fleisher 1993; Hifnawy et al. 2001).

The oil of *A. santolina* has not been thoroughly studied with modern analytical methods. Since 1970 only few reports concerning volatile components of this plant growing in Iran (Afsharypuor et al. 1996), Egypt and Turkey have been published (Khafagy and El Fatatry 1970; Brunke et al. 1986) providing the evidence for presence of different chemotypes. In these previous studies, the oils from Egypt and Turkey showed similar components of their monoterpene fractions with 1,8-cineole, camphor, *trans-* and *cis-*sabinene hydrate, linalool, pinocarvone and 4-terpineol as the main constituents. From the Egyptian plant oil, fragranol and its diasteroisomer grandisol were identified. However, the main oil components of *A. santolina* growing in Iran include caryophyllene oxide, *cis-*nerolidol, camphor, olic aldehyde and linalool.

In this paper, we present a detailed comparative study between the hydrodistilled oils as well as the *n*-hexane-ether extracts of *A. fragrantissima* and *A. santolina*. Moreover, their antimicrobial activities were assayed.

2. Investigations, results and discussion

The hydrodistillation of the fresh aerial parts of A. fragrantissima gave pale yellow oil with an aromatic fragrant odour. While the oils obtained from A. santolina are yellowish-green with an aromatic, fragrant and pleasant odour. The odour and flavour of the n-hexane-ether extracts were believed to be closer to that of those plant oils. Using GLC and GLC-MS, the compounds identified and their proportions are listed according to their retention indices on a OV-1 fused silica capillary column (Table 1). In A. fragrantissima, a total of 49 components were identified. The major compounds in the hydrodistilled oil were found to be cis-thujone and santolina alcohol with 29.48 and 18.29%, respectively. In the *n*-hexane-ether extract the percentages were 17.28 and 36.69, respectively. The oil was found to be rich in artemisia ketone (15.24%), transthujone (10.38%), trans-pinocarveol (6.83%) and yomogi alcohol (4.35%). These findings are in agreement with the previous reports of A. fragrantissima studies (Shalaby and

Compound		RI*	A. fragrantissin	A. fragrantissima		A. santolina		
			Essential oil	Hexane extract	Essential oil			Hexane extract
					Flower	Eaf	Stem	
1	Isopentyl acetate	876	_	_	tr	_	_	_
	Santolina triene	902	1.30	-	-	-	-	-
	α-Thujene	920	-	-	tr 70	tr	tr	tr
	α-Pinene Camphene	924 935	0.10	_	$\begin{array}{c} 0.70\\ 0.60 \end{array}$	0.35 0.29	0.15 0.26	tr 0.51
	Sabinene	955 959	_	_	tr	tr	0.20	1.82
	β-Pinene	962	_	_	tr		_	tr
	1,8-Dehydrocineole	974	-	-	tr	tr	-	_
	Yomogi alcohol	991	4.35	-	0.70	0.97	0.38	1.83
	<i>n</i> -Decane	993	-	-	tr	-	-	-
11	1	1004	_	-	0.50	0.47	tr	_
	<i>p</i> -Cymene 1,8-Cineole	1008 1014	- 1.84	0.50	0.40 3.00	1.90 1.93	0.42 0.69	0.48
	Limonene	1014	1.04	0.50	5.00 tr	1.95 tr	0.09	0.48 tr
	Santolina alcohol	1014	18.29	36.69	u —	u —	_	u —
	Bergamal	1038	_	0.41	_	_	_	-
	Artemisia ketone	1048	15.24	tr	1.66	0.87	0.40	0.51
18		1048	-	-	tr	1.22	0.98	-
19	5	1053	-	0.91	1.02	1.80	1.50	3.87
	<i>n</i> -Octanol	1055	-	1.51	_	-	-	-
	Artemisia alcohol	1074 1077	0.27	1.21	0.9	0.52	0.39 0.26	-
	Terpinolene trans-Sabinene hydrate	1077	_	- 0.41	tr tr	tr tr	0.20	-
	Linalool	1081	_	-	0.52	tr	_	_
	<i>cis</i> -Thujone	1090	29.48	17.28	-	-	_	-
	trans-Thujone	1098	10.83	4.71	_	_	-	_
27		1114	-	0.51	-	_	-	-
28		1120	6.83	0.32	-	-	-	-
	Camphor	1124	-	-	3.76	3.03	3.40	6.60
	<i>trans</i> -dihydro-α-Terpineol	1143 1143	-	0.31 0.11	-	_	-	-
31	<i>cis</i> -Chrysanthenol Borneol	1143	_	0.11	4.50	4.80	3.56	0.34
	Lavandulol	1153	2.22	0.42	tr	tr	0.18	-
	Artemisyl acetate	1153	0.72	0.63	_	_	_	-
35	Terpin-4-ol	1155	0.78	0.30	6.60	6.53	5.89	1.38
36	α-Terpineol	1165	-	0.91	0.34	0.30	0.12	1.24
37		1175	0.34	-	-	-	-	-
	cis-Piperitol	1178	_	_	tr	0.53	1.01	0.70
	Myrtenol Methyl chavicol	1178 1180	_	0.12	tr —	tr —	0.10	0.60
	trans-Piperitol	1180	_	0.12	tr	tr	tr	tr
	Fragranol	1196	_	_	11.84	13.22	18.69	8.78
43	<i>cis</i> -Sabinene hydrate acetate	1203	-	-	tr	_	_	_
	Carvone	1212	-	-	tr	_	-	-
	Carvotanacetone	1218	-	6.67	_	_	-	-
	trans-Myrtanol	1246	-	-	tr	-	-	-
	<i>cis</i> -Chrysanthenyl acetate	1250	-	1.41	tr tr	-	-	-
	<i>neo</i> -iso-3-Thujyl acetate Bornyl acetate	1251 1253	_	_	tr 0.30	0.17	0.15	0.53
	Hydroxy citronellal	1263	_	0.32	-		-	-
	Lavandulyl acetate	1266	_	_	tr	-	_	-
	Thymol	1267	-	-	0.90	tr	0.31	0.99
	trans-Sabinyl acetate	1275	1.77	-	-	-	-	-
	Carvacrol	1276	-	-	tr	0.69	-	-
	iso-Dihydro carveol acetate	1302	_	0.41	-	-	-	-
	Fragranyl acetate α-Terpinyl acetate	1335 1337	_	_	51.70 0.40	47.14 2.30	45.10 3.88	50.70 0.67
	Eugenol	1337	_	0.91	- 0.40	2.30	5.88 —	
	Neryl acetate	1352	_	-	tr	tr	tr	_
	Methyl p-anisate	1358	_	-	tr	_	_	_
61	trans-Myrtanol acetate	1368	-	_	0.51	0.51	0.59	0.81
62	(E) - α -Damascone	1377	_	-	tr	0.21	0.38	-
	cis-Jasmone	1388	-	tr	-	-	-	-
	Methyl eugenol	1367	-	tr 0.10	-	—	-	_
	<i>n</i> -Tetradecane (<i>E</i>)-Caryophyllene	1400 1411	_	0.10	0.21	0.30	0.33	0.29
00	(L)-Caryophynene	1411	-	_	0.21	0.50	0.55	0.23

 Table 1: Percentage composition of essential oils and n-hexane-ether extracts of Achillea fragrantissima (Forssk.) Sch. Bip. and A. santolina L. and their retention indices

Table 1: (continued)

Compound	RI*	A. fragrantissima		A. santolina			
		Essential oil	Hexane extract	Essential oil			Hexane extract
				Flower	Eaf	Stem	_
67 Lavandulyl isobutyrate	1425	_	_	tr	tr	tr	0.50
68 Massoia lactone	1442	_	tr	_	_	_	_
69 Germacrene D	1472	_	-	1.20	1.07	1.50	0.89
70 Bicyclogermacrene	1492	3.58	-	0.60	0.48	1.01	-
71 Butylated hydroxytoluene	1493	_	1.51	_	-	-	_
72 Lavandulyl-2-methylbutyrate	1506	_	-	1.22	0.50	0.71	_
73 β-Sesquiphellandrene	1510	0.58	_	_	-	-	-
74 Dendrolasin	1558	_	-	0.72	0.86	1.33	1.32
75 Spathulenol	1561	0.20	1.05	_	-	-	tr
76 Caryophyllene oxide	1576	-	-	0.66	0.56	0.31	3.90
77 10- <i>epi</i> -γ-eudesmol	1619	_	-	0.93	0.85	0.93	3.02
78 α-Muurolol	1630	-	-	1.23	1.25	1.18	4.25
79 β-Eudesmol	1633	0.14	-	_	-	-	-
80 Khusinol	1655	-	tr	_	-	-	_
81 Oplopanone	1695	-	tr	_	-	-	_
82 Palmitic acid methylester	1911	_	0.10	_	-	-	-
83 Palmitic acid	1948	-	3.96	_	-	-	_
84 Phytol	2100	_	1.22	_	-	-	_
85 Linoleic acid	2115	-	4.55	_	-	-	_
86 α-Santonine	2117	-	-	_	-	-	tr
87 Linolenic acid	2120	-	4.55	_	-	-	_
88 Methyloctacosane	2831	-	-	-	-	_	tr
89 Campesterol	3110	-	tr	-	-	-	-
90 Stigmasterol	3170	-	tr	-	-	-	-
91 β-Sitosterol	3220	-	tr	-	-	_	tr
92 Fucosterol	3280	-	tr	-	_	-	_
Total identified (%)		98.86	94.02	97.62	95.62	96.09	96.53

* RI = retention indices relative to $C_{9}-C_{28}$ *n*-alkanes on the OV-1 column, tr = traces (< 0.1), hex.extract = *n*-hexane-ether extract

Richter 1964; El-Deeb 1985; Aboutabl et al. 1986a; 1986b; Fleisher and Fleisher 1993; Hifnawy et al. 2001). We report the presence of *trans*-pinocarveol (6.83), artemisyl acetate (0.72%) and β -sesquiphellandrene (0.58%) for the first time in the oil of this plant. Solvent extraction has the advantage of recovering the higher molecular weight natural compounds. Forty-one compounds were identified in this extract. Carvotanacetone (6.67%), trans-thujone (4.71%), linolic acid (4.55%) and linolenic acid (4.55%) were major components in the *n*-hexane-ether extract. An interesting finding was the detections of 17 additional volatile components not previously mentioned in the literature e.g. bergamal, cis-sabinene hydrate, n-octanol, trans-sabinene hydrate, iso-3-thujanol, trans-dihydro-α-terpineol, cis-chrysanthenol, carvotan acetone, cis-chrysanthenyl acetate, hydroxy citronellal, iso-dihydrocarveol acetate, methyl eugenol, n-tetradecane, butylated hydroxytoulene, khusinol, oplopanone and phytol. Besides, palmitic acid methyl ester, linolenic acid as well as a traces from campesterol, stigmasterol, β -sitosterol and fucosterol were also detected. Various alkanols such as C24, C26, C28, and C30 with retention indices (RI) 2664, 2882, 3043 and 3278, respectively have been also identified. On the other hand, myrecne, nerol, namyl ketone, β-pinene, limonene and bornyl acetate previously reported as oil components (Shalaby and Richter 1964; El-Deeb 1985) could not be detected. We confirmed the presence of α -pinene, santolina triene, 1.8-cineole, artemisia alcohol, spathulenol and β -eudesmol as minor compounds identified by Hifnawy et al. (2001) as well as the presence of 2-methyl-1-propen-1-one, (2-methylbutyl)-cyclopropane, p-cymene, 2-methyl-1-propanol, cyclopropane carboxylic acid methyl ester, propanoic acid 2-methyl-3methylbutyl ester, butanoic acid 3-methyl-3-methyl butyl ester, 3-hexenyl butanoate (*E*), 3-hexenyl butanoate (*Z*), sabinyl alcohol (sabinol), sabinol isomer and γ -muurolene could not be detected in our sample.

Crushed flowers, leaves and stems of A. santolina were separately hydrodistilled. Fifty-four compounds (97.62%) were identified in the flower's oil whereas 43 (95.62%) and 37 (96.09%) compounds were characterized in the oils obtained from the leaves and stems, respectively. Fragranyl acetate, fragranol, lavandulyl isobutyrate, terpin-4ol and camphor were the main constituents in the investigated parts. The principle components in the n-hexaneether extract of the total aerial parts were fragranyl acetate (50.7%), fragranol (8.78%), camphor (6.6%), cis-sabinene hydrate (3.87%). Oxygenated sesquiterpenes present in the plant include α -muurolol (4.25%), caryophyllene oxide (3.9%), spathulenol, dendrolasin and 10-epi-y-eudesmol (3.02%). Another distinguishing feature appeared to be the presence of minor amounts of β -sitosterol, and methyloctacosane in the extract product, which were apparently absent in flower, leaf and stem distillates. In this context it should also be noted that we found no evidence for the presence of grandisol, cis-nerolidol, farnesol, 2-hydroxycyclopentadecanone, cis-a-santalol acetate and olic aldehyde in the volatile fractions previously reported in this plant (Khafagy and El Fatatry 1970; Brunke et al. 1986; Afsharypuor et al. 1996).

This divergence in the profiles of the volatile constituents of *A. fragrantissima* and *A. santolina* should be possibly attributed either due to geographical and ecological factors or an indication of the existence of different chemotypes of these plants.

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Table 2: Antimicrobial activity of Achillea fragrantissima and A. santolina hydrodistilled oils and n-hexane-ether extracts

Material*	Diameter of inhibition zone						
	Gram – ve bac	teria	Gram + ve bacteri	Fungus			
	E. coli	P. aeruginosa	S. aureus	B. subtilis	C. albicans		
Hydrodistilled oil of A. fragrantissima.	10	15	15	30	30		
Hydrodistilled oil of <i>A. fragrantissima</i> . Extract of <i>A. fragrantissima</i> ^{**}	17	19	-	19	15		
Hydrodistilled oil of A. santolina	8	30	13	21	27		
Extract of A. santolina**	18	17	-	17	13		
Ciprofloxacin (25 µg)	20	10	25	22	-		
Nystatin (25 µg)	-	-	-	-	25		

* = All assays consisted of 50 µl of a test solution, consists of 20 mg residue in 1 ml DMF and 25µg/50 µl of standard antimicrobial in DMF.

** = n-hexane-ether extract, - = no inhibition.

The susceptibility of various microorganisms to the inhibitory effect of the oils and extracts is represented in Table 2. The results show that: the hydrodistilled oils of *A. fragrantissima* and *A. santolina* exhibited a pronounced antifungal activity however; the *n*-hexane-ether extracts showed moderate effect. Hydrodistilled oil of both plants showed a range of activity against some human pathogens such as *B. subtilis* and *P. aeruginosa*. However, *S. aureus* was completely resistant against the *n*-hexane-ether extracts of *A. fragrantissima* and *A. santolina* which were active against *E. coli, P. aeruginosa* and *B. subtilis*.

3. Experimental

3.1. Plant material

Flowering aerial parts of *A. fragrantissima* (Forssk.) Sch. Bip. were collected from wild plants growing in the Sinai desert (vicinity of Sader Hetan) in April 2000, while those of *A. santolina* L. were collected from a wild population growing in the vicinity of Alexandria province, Egypt in April 2000. The identity of the plants have been kindly verified by Prof. Dr. N. El-Hadidi, Faculty of Science, Cairo University. Voucher specimens were deposited in the Herbarium of Pharmacognosy Department, Faculty of Pharmacy, Zagazig University.

3.2. Sample preparation

The fresh aerial parts of *A. fragrantissima* as well as flowers, leaves and stems of *A. santolina* were separately subjected to hydrodistillation. Fourhours produced oils with 1, 0.1, 0.12 and 0.04%, respectively. The aerial parts of the two plants were extracted by percolation with a mixture of *n*-hexane-ether (1:1, v/v) and the solvents were removed subsequently under reduced pressure. The yield was 1.3 and 0.9% for *A. fragrantissima* and *A. santolina*, respectively.

3.3. Analysis

The constituents of the volatile oils obtained by steam distillation and of the *n*-hexane-ether extracts were analysed by GLC and GLC-MS as reported (El-Shazly et al. 2002a). Compounds were identified by comparison of their retention indices (C_9 to C_{24} *n*-alkane mixture) and mass spectra with those reported in the literature (El-Shazly 1999; El-Shazly et al. 2002a; 2002b; Adams 1995; Asres et al. 1998; Engel et al. 1998; Masada 1976; Ryhage and Sydow 1963; Sydow 1963; Goad and Akihisa 1997).

3.4. Antimicrobial screening

The hydrodistilled oils and the *n*-hexane-ether extracts of *A. fragrantissima* and *A. santolina* were prepared for antimicrobial screening by dissolving 20 mg (oil or extract) in 1 ml dimethylformamide (DMF) and 50 µl was applied (equivalent to 1 mg). The activity of the solutions was screened by the cup-plate agar diffusion method (Woods and Washington 1995). The available microorganisms (Table 2) were *Staphylococcus aureus*, *Bacillus subtilis* (Gram positive bacteria); *Escherichia coli*, *Pseudomonas aerugino-sa* (Gram negative bacteria) and *Candida albicans* (fungus). The selected microorganisms were obtained from the stock cultures of the Department of Microbiology, Faculty of Pharmacy, Zagazig University. As positive controls, 25 µg of ciprofloxacin in DMF was used as standard antibiotic for bacteria while for fungi nystatin (in DMF) was used at the same concentration. The plates were incubated overnight at 37 °C for bacteria and 30 °C for fungi. The zones of inhibition were observed and recorded (in mm) as shown in Table 2.

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