ORIGINAL ARTICLES

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A new alkaloid from *Narcissus serotinus* L.

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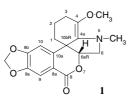
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A new alkaloid derivative of [2]benzopyrano-[3,4-c] indole, isomer of 3-epimacronine, 4-methoxy-5-methyl-1,2,3,5,6,6 α R-hexahydro-[1,3]dioxolo[4',5':6,7]isochromeno [3,4-*c*]indol-8-one, has been iso-lated from *Narcissus serotinus* L. (Amaryllidaceae) and its structure was elucidated by mass and spectral analysis.

1. Introduction

In Greece the genus *Narcissus* is represented by the species N. tazetta L., N. poeticus L. and N. serotinus L. (Tutin et al. 1980). An alkaloid of the pretazettine-type, an isomer of 3-epimacronine (Viladomat et al. 1990) which differs in the position of the methoxy-group and the double bond was isolated. The structure of the new compound is 4-methoxy-5-methyl-1,2,3,5,6,6αR-hexahydro-[1,3]dioxolo[4',5':6,7]isochromeno[3,4-c]indol-8-one (1). Most of the Amaryllidaceae alkaloids have biological and pharmacological properties and recent research in a series of alkaloids of this family and this genus demonstrated cytotoxic and antiviral activity (Furusawa et al. 1980; Gabrielsen et al. 1992; Weniger et al. 1995; Lopez et al. 2002). This is the first report of this alkaloid structure in the Amaryllidaceae family and provides scope for further investigation regarding a biological-pharmacological use.



2. Investigations, results and discussion

The proposed structure of this new alkaloid (1), was based on the following spectroscopic data obtained by ¹H NMR, ¹³C NMR, ¹H-¹H COSY, NOESY, ¹H-¹³C COSY, DEPT, EI-MS, CI-MS and IR.

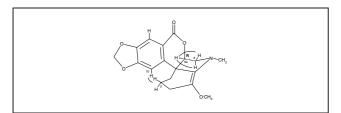


Fig.: Configuration of compound 1

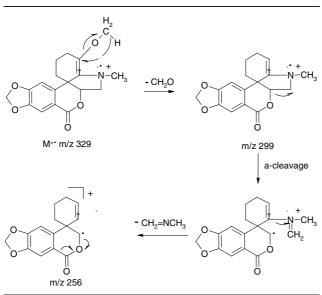
Better assignment was proposed by selective proton decoupling experiments, which showed that irradiation of H-6 α $(\delta 4.20, dd)$ induced simplification of the m signal at $\delta 1.80$ (H-6, H-6') to a dd. This reveals that after decoupling, H-6 is no longer coupled with H- 6α , giving two doublets and the signal of H-6 α (dd) turns to s. The ¹H-¹H COSY demonstrated a correlation between H-6 α (δ 4.20) and each of the H-6 protons (δ 1.80–2.30), as well as between H-3 and H-2 protons (δ 1.80–2.30). The configuration of C_{6 α} is R as can be shown based on the following considerations: $H_{6\alpha}$ appears at 4.20 ppm as a doublet of doublets ($J_{6\alpha, 6} = J_{6\alpha, 6'}$ = 6Hz), which suggests a conformation where $H_{6\alpha}$ is located at equal angles from the two neighbouring protons. On this ground, the configuration of $C_{6\alpha}$ becomes R, whereas proximity in space appears between one of the two protons on carbon 2 and H₁₀, as is shown by the reconstruction of the molecule using molecular models (Fig.). This proximity is evidenced by NOE Spectroscopy (NOESY), where correlation appears between H_{10} and the multiplet from 2.30 to 1.80 ppm where H_6 , H_2 and H_1 appear.

The ¹H-¹³C COSY spectrum demonstrated the following assignments: H-9 (7.65s) to C-9 (107.04); H-10 (7.25s) to C-10 (103.14); $-\text{OCH}_2\text{O}$ (6.10–6.08 dd) to OCH₂O (102.20); H-6\alpha (δ 4.20 dd) to C-6 α (74.86); OCH₃ (3.55 s) to OCH₃ (58.04) and H-3 (2.55–2.70 m) to C-3 (34.81). Finally, the carbon multiplicities were established by DEPT spectrum.

The EI-MS showed a molecular ion at m/z 329 (50%) and CI-MS (CH₄) M + 1, m/z 330 (100%). The most important and characteristic fragments in EI-MS with their relative intensities are: 329 ([M⁺·], 50), 314 (8), 299 (40), 272 (60), 271 (35), 256 (68), 241 (86), 240 (90), 228 (23), 213 (30), 212 (18), 135 (30), 57 (100).

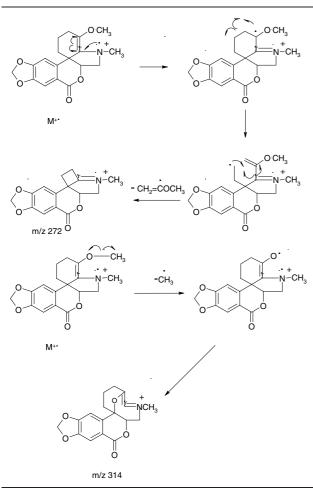
The fragment at m/z 299 is due to the single cleavage decomposition of the parent ion. The origin of the m/z 256 might have been the α -cleavage next to the nitrogen atom and subsequent loss of the fragment CH₂ = N-CH₃. Loss of CO₂ resulted in the formation of the fragment ion m/z 212 (Scheme 1). The fragment at m/z 314 originated after the loss of -CH₃ radical from M⁺. The low intensity (8%) of this fragment could be explained by the unfavorable stereochem-

Scheme 1



istry of the $-OCH_3$ group; it is known in the case of Amaryllidaceae alkaloids that minor changes in the stereochemistry are frequently sufficient to cause appreciable differences in the MS of many stereoisomers. This happens for tazettine and criwelline which differ only in the configuration of the $-OCH_3$ group, but this is sufficient to cause considerable variations in the MS of these alkaloids. Also, experiments with deuterium labeling of the > NCH₃ group demonstrated

Scheme 2



that only $-OCH_3$ was implicated in the $M^{+\bullet} -CH_3$ process (Duffield et al. 1965). Fragmentation after RDA process, followed by a hydrogen transfer to the nucleus with the double bond, constitutes a general process for these systems (Schoes et al. 1968), but in the case of this compound such type of fragmentation does not seem to appear. On the contrary, a proposed possible fragmentation way of $M^{+\bullet}$, results in the formation of ion m/z 272 after the loss of the fragment [$-CH_2COCH_3$] as radical (Scheme 2). Also the fragment at m/z 57 is given as base peak, due to its stability as positive ion and that seems to favor this type of fragmentation.

3. Experimental

3.1. Apparatus

The IR spectrum was obtained using a Paragon 500 Perkin Elmer spectrophotometer, ¹H-, ¹³C-, ¹H-¹H COSY, NOESY, ¹H-¹³C COSY, DEPT and irradiation spectra were obtained on the Bruker DRX-400 apparatus. EI-MS, CI-MS were recorded on a VG-Trio 2000 FISONS mass spectrometer.

3.2. Plant material

Plant material was collected from the hillsides at Drapano cape near Chania-Crete, at an altitude of 50 m and was identified by Dr. C. Fournaraki; a voucher specimen is deposited in the herbarium of the Mediterranean Agronomic Institute of Chania, No 3593.

3.3. Extraction and isolation

500 g of the air dried aerial parts and bulbs collected in October 2001 were crushed and extracted with 3 × 500 ml MeOH. The extract was filtered, concentrated and partitioned between a 2% HCl aqueous phase solution and CHCl3. The aqueous phase was made basically with NaHCO3 and re-extracted with CHCl3 affording 42 mg of a residue, after concentration under vacuum. The residue was subjected twice to silica gel 60 (0.04-0.063 mm) flash column chromatography (1 × 20 cm), CH₂Cl₂/MeOH 9.5:0.5, approxim. 5 ml/min t_R of the substance 120–260 ml; detection of the elutes by TLC (SiO₂, CH₂Cl₂/MeOH/NH₄OH 9.5:0.5:0.1, Rf 0.50; CH₂Cl₂/i-PrOH/ NH4OH 7:3:0.1, Rf 0.30; CH2Cl2/MeOH/NH4OH 9:1:0.1, Rf 0.80; Dragendorff reagent). After a second purification with the same chromatographic conditions the selected fractions yielded 14 mg of pure product. 4-Methoxy-5-methyl-1,2,3,5,6,6 α R-hexahydro-[1,3]dioxolo[4',5':6,7]isochromeno[3,4-c]indol-8-one (1): IR (KBr): 1721 (lactone C=0), 1448, 1416, 1257, 1163, 1105 cm⁻¹. ¹H NMR (CDCI₃, TMS): $\delta = 1.80-2.30$ (6 H, m, H-1, H-2, H-6), 2.41 (3 H, s, CH₃N <), 2.70–2.55 (2 H, m, H-3), 3.55 (3 H, s, $-OCH_3$), 4.20 (1 H, dd, $J_{6a,6} = J_{6a,6'} = 6$ Hz, H-6a), 6.08 (1 H, d, $J_{gem} = 1.2 \text{ Hz}$, $-\text{OCH}_2\text{O}-$), 6.10 (1H, d, $J_{gem} = 1.2 \text{ Hz}$, $-\text{OCH}_2\text{O}-$), 7.25 (1H, s, H-10) 7.65 (1H, s, H-9). ¹³C NMR (CDCl₃, TMS): $\delta = 29.34$ $(C\text{-}1), \ 31.26 \ (C\text{-}2), \ 34.81 \ (C\text{-}3), \ 42.09 \ (CH_3N <), \ 53.39 \ (C\text{-}6), \ 58.04$ (CH₃O-), 74.86 (C-6a), 102.20 (-OCH₂O-), 103.14 (C-10), 107.04 (C-9), 111.40 (C-10b), 116.33 (C-4a), 135.13 (C-4), 148.08 (C-8a), 152.37 (C-10a), 153.51 (C-9a), 161.43 (C-9b), 185.86 (C-8).

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References

- Gabrielsen B, Monath TP, Huggins JW, Kefauver DF, Pettit GR, Groszek G, Hollingshead M, Kirsi JJ, Shannon WM, Schubert EM (1992) Antiviral (RNA) activity of selected Amaryllidaceae isoquinolino constituents and synthesis of related substances. J Nat Products 55: 1569–1581.
- Duffield A, Aplin R, Budzikiewics H, Djerassi C, Murphy C, Wildman W (1965) Mass spectrometry in structural and stereochemical problems. A study of the fragmentation of some Amaryllidaceae alkaloids. J Am Chem Soc 87: 4902–4912.
- Furusawa E, Irie H, Combs D, Wildman WC (1980) Therapeutic activity of pretazettine on Raucher leukemia: comparison with related Amaryllidaceae alkaloids. Chemotherapy 26: 36–45.
- Lopez S, Bastida J, Viladomat F, Codina C (2002) Acetylcholinesterase inhibitory activity of some Amaryllidaceae alkaloids and *Narcissus* extracts. Life Sci 71: 2521–2529.
- Schoes H, Smith D, Burlingame A, Jeffs P, Dopke W (1968) Mass spectra of Amaryllidaceae alkaloids. The lycorenine series. Tetrahedron 24: 2825–2837.
- Tutin T, Heywood B, Burges N, Moore D, Valentine D, Walters S, Webb FD (1980) Amaryllidaceae in Flora Europaea, Cambridge University Press, p. 79, volume 5.
- Viladomat F, Bastida J, Tribo G, Codina C, Rubiralta M (1990) Alkaloids from *Narcissus bicolor*. Phytochemistry 29: 1307–1310.
- Weniger B, Italiano L, Beck JP, Bastida J, Bergomon S, Codina C, Lobstein A, Anton R (1995) Cytotoxic activity of Amaryllidaceae alkaloids. Planta Med 61: 77–79.