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# Electrochemical behavior of quinapril and its determination in pharmaceutical formulations by square-wave voltammetry at a mercury electrode

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The electrochemical behavior of the antihypertensive drug quinapril was investigated at a hanging mercury drop electrode using different voltammetric techniques such as cyclic voltammetry, squarewave voltammetry and chronoamperometry. A simple and sensitive square-wave voltammetric method for the electrochemical analysis of quinapril in its pharmaceutical formulations was developed and validated. The experimental and instrumental parameters affecting the peak current of quinapril were investigated. Various buffers such as Britton Robinson, borate and phosphate buffers at different pH values (3.0-11.0) were examined as supporting electrolyte. The optimum conditions were obtained using Britton Robinson buffer at pH 10.0 and frequency: 50 Hz, scan increment: 4 mV and pulse amplitude: 25 mV. A well-defined peak current was observed at the hanging mercury drop electrode at -1100 mV vs. Ag/AgCl reference electrode. This proposed method was validated by evaluating linearity, sensitivity, repeatability, accuracy, precision, selectivity, recovery, robustness and ruggedness. The linear calibration range was  $0.50-8.68 \,\mu g \, m L^{-1}$  (r = 0.9992). The detection and quantification limits of this method were 0.22 and 0.50  $\mu$ g mL<sup>-1</sup> and intra-day and inter-day precision were between 0.81– 4.32% (n = 7), respectively. The developed method was applied successfully for the determination of quinapril in its tablet dosage forms. The average amount of quinapril in tablets was found as  $20.26 \pm 0.12$  with RSD of 1.60% for 20 mg tablets and  $40.55 \pm 0.23$  with RSD of 1.52% for 40 mg tablets.

## 1. Introduction

Quinapril hydrochloride (QUI), 2-(2-{[1-(ethoxycarbonyl)-3-phenyl-propyl]amino}-1-oxopropyl)-1,2,3,4-tetra-hydro-

3-isoquinoline carboxylic acid monohydrochloride, is a nonpeptide, nonsulfhydryl angiotensin converting enzyme (ACE) inhibitor belonging to the third class of ACE inhibitors. The role of this kind of drugs is to inhibit the last step of the biosynthesis of angiotensin II, a potent vasoconstrictor, causing general vasodilatation. QUI is used for the treatment of mild to moderate hypertension and congestive heart failure, either alone or in conjunction with other drugs (Vertes and Haynie 1992; Forette et al. 1992; Cudina et al. 2006).



Several analytical methods have been reported for the determination of QUI in pharmaceutical formulations and biological fluids, including densitometry (Kowalczuk et al. 2003), spectrophotometry (Bonazzi et al. 1997; Kowalczuk et al. 2004), HPLC (Bonazzi et al. 1997; Abbara et al. 2002; Hengy and Most 1988; Gumieniczek and Hopkala 1998; Prieto et al. 2001; Kugler et al. 1995), gas chromatography - mass spectrometry (Ferry et al. 1987; Goto et al. 1992; Goto et al. 1992), capillary electrophoresis (Cudina et al. 2006; Gotti et al. 2000; Hillaert et al. 2001; Hillaert and Van den Bossche 2001; Hillaert et al. 2002; Prieto et al. 2002) and voltammetry (Prieto et al. 2003). The main problems encountered using chromatographic methods are either the need for derivatization or time-consuming extraction procedures. Since these methods have a slightly expensive instrumentation and high running costs, the use of simpler, faster, less expensive, but sensitive, and stability to on-line measurements (give the repeatable results), electrochemical methods can be an alternative.

The main objective of this study was to investigate the electrochemical behavior and reduction mechanism of QUI and its determination possibilities at a hanging mercury drop electrode (HMDE) using the particularly simple, rapid and sensitive technique of square-wave voltammetric (SWV) method. The experimental and instrumental parameters were optimized in order to improve sensitivity. The proposed voltammetric method was also applied to the quantitation of QUI in pharmaceutical formulations without the necessity of sample pre-treatment.

A voltammetric method concerning the reduction of QUI at the mercury electrode has been reported by Prieto et al. (2003). But in this study, electrochemical behavior and reduction mechanism of QUI were not investigated in detail and this method was not fully validated. In our study, reduction mechanism of QUI was investigated by cyclic voltammetry and the diffusion coefficient of QUI was calculated by chronoamperometry. Our method was fully validated and validity of the proposed method was also tested comparing the results to those of Prieto et al. (2003).

## 2. Investigations, results and discussion

The SWV method was used in this study because of its adequate sensitivity and the rapidity of the measurements. The peak current intensity, shape and characteristics of all voltammograms were strongly dependent on various electrolytes and pH of the medium. Thus, the effect of pH on the peak current  $(i_p)$  and the reduction potential  $(E_p)$  of QUI was investigated over a range between 3.0 and 11.0. In acidic solutions, no reduction peak of QUI was observed. QUI exhibited a cathodic peak in Britton Robinson buffer at pH 6.0-11.0, but good peak shape was observed at pH 8.0-11.0 (Fig. 1). Peak current of QUI was increased with the increase of pH (Fig. 2). Maximum peak current and good peak shape were obtained in BR buffer at pH 10.0. In addition, BR, phosphate and borate buffers at pH 10.0 were investigated for the analytical determination of OUI (Fig. 3). The best results such as the maximum peak current, a well-defined cathodic peak, suitable sensitivity and repeatability were obtained with BR buffer at pH 10.0. Therefore, 0.04 M BR buffer at pH 10.0 was chosen as a supporting electrolyte for the rest of the present work.

The SWV response of the accumulated drug markedly depends on the instrumental parameters. Different parameters influencing peak current intensity and shape of the SWV peak of QUI were studied in order to improve sensitivity. Thus, frequency (f), scan increment ( $\Delta E$ ) and pulse amplitude ( $E_{SW}$ ) were investigated for 3.38 µg · mL<sup>-1</sup> of QUI solution in BR buffer at pH 10.0 in the proposed method.

f was varied from 40 to 120 Hz using  $\Delta E$  of 5 mV and  $E_{SW}$  of 25 mV. A linear relationship was obtained between



Fig. 1: Effect of BR buffer at different pH on the SWV peak of QUI  $(3.38 \ \mu g \cdot mL^{-1})$  at  $f = 50 \ Hz$ ,  $\Delta E = 5 \ mV$  and  $E_{SW} = 25 \ mV$ . a) pH = 10.0, b) pH = 11.0, c) pH = 8.0, d) pH = 9.0



Fig. 2: Effect of BR buffer at different pH on the SWV peak of QUI  $(3.38 \ \mu g \cdot mL^{-1})$  at  $f = 50 \ Hz$ ,  $\Delta E = 5 \ mV$  and  $E_{SW} = 25 \ mV$ 



Fig. 3: Effect of different buffer at pH = 10.0 on the SWV peak of QUI  $(3.38 \ \mu g \cdot mL^{-1})$  at f = 50 Hz,  $\Delta E = 5 \ mV$  and  $E_{SW} = 25 \ mV$ . a) BR, b) Phosphate, c) Borate buffers

the peak current and the f up to 100 Hz. The best peak definition was found at f = 50 Hz, which is used in this study.

At f of 50 Hz and  $E_{SW}$  of 25 mV,  $\Delta E$  was varied from 2 to 6 mV. The peak current increased to 4 mV and then it decreased; because the reduction peak of QUI was broad. Hence,  $\Delta E$  of 4 mV was chosen for the determination of QUI.

At f of 50 Hz and  $\Delta E$  of 4 mV,  $E_{SW}$  varied from 10 to 40 mV. Peak current of QUI was increased with the increase of  $E_{SW}$ , but the narrowest square-wave peak was obtained at 25 mV, and this was chosen as optimal value for analytical determination.

Consequently, the highest peak current values with the best peak definition were found as f = 50 Hz,  $\Delta E = 4$  mV and  $E_{SW} = 25$  mV, and these values were used for further measurements of QUI. Under these optimal experimental and instrumental conditions, a well-defined SWV peak of QUI was observed at -1100 mV in the proposed method (Fig. 4).

The reversibility of the reduction process of QUI was investigated by using cyclic voltammetry. The cyclic voltammogram of  $10.11 \,\mu\text{g} \cdot \text{mL}^{-1}$  of QUI in BR buffer at



Fig. 4: SW voltammograms of QUI at different concentrations using the optimum conditions. a) Supporting electrolyte, b) 1.48, c) 3.38, d) 4.76, e) 6.10, f) 7.41 and g) 8.68 μg · mL<sup>-1</sup> of QUI

pH 10.0 at HMDE exhibited a single well-defined peak at -1100 mV (Fig. 5). This peak may be attributed to the carbonyl group of the amide bound, activated by the neighboring nitrogen, might undergo reduction to alcohol, taking two electrons and two protons (Prieto et al. 2003).

No oxidation peak was observed on the reverse scan, indicating the irreversible nature of the electrode process (Fig. 5). The peak potential shifted to a more negative value on the increase of the scan rate, confirming the irreversible nature of the reduction process (Bard and Faulkner 1980).

Scan rate studies were carried out to assess whether the processes at the HMDE electrode was under diffusion or adsorption control. The effect of the potential scan rate between 20 and 1000 mV  $\cdot$  s<sup>-1</sup> on the peak current and the potential of QUI were evaluated. A 57 mV negative shift in the peak potential confirmed the irreversibility of the reduction process. When the scan rate was varied from 20 to 800 mV  $\cdot$  s<sup>-1</sup> in 8.68 µg  $\cdot$  mL<sup>-1</sup> solution of QUI, a linear dependence of the peak intensity i<sub>p</sub> (µA) upon the square root of the scan rate v<sup>1/2</sup> (mV  $\cdot$  s<sup>-1</sup>) was found, demonstrating a diffusional behavior (ip (µA) = 0.0224 v<sup>1/2</sup> (mV  $\cdot$  s<sup>-1</sup>) – 0.0258, r = 0.9920, n = 6) (Bard and Faulkner 1980; Laviron et al. 1980).

A plot of log  $i_p$  (logarithm of peak current) versus log v (logarithm of scan rate) gave a straight line with a slope of 0.5316, very close to the theoretical value of 0.5, which is expressed for an ideal reaction the diffusion controlled electrode process (Laviron et al. 1980). The



Fig. 5: Cyclic voltammogram of 10.11  $\mu$ g · mL<sup>-1</sup> QUI solution at HMDE

following equation was obtained (scan rate range =  $20-800 \text{ mV} \cdot \text{s}^{-1}$ ):

$$\begin{split} \log i_p \left( \mu A \right) &= 0.5316 \log \nu \left( mV \cdot s^{-1} \right) - 1.7703 \,, \\ r &= 0.9885 \, (n = 6) \end{split}$$

The experimental Cottrell slope was determined from the chronoamperometric  $i_p$  versus  $t^{-1/2}$  plot. The diffusion coefficient was calculated from Cottrell Equation (Baranski et al. 1985). The constant potential applied was slightly more cathodic than the cyclic voltammetric  $E_p$  from -1200 to -950 mV. A HMDE with a surface area of 0.0199 cm<sup>2</sup> was employed. The diffusion coefficient was calculated as  $7.71 \times 10^{-6} \pm 2.41 \times 10^{-7}$  (n = 7).

Validation of the proposed SWV method for assay of QUI was examined via stability, linearity, sensitivity, precision, accuracy, recovery, selectivity, robustness, and ruggedness (ICH 2005; USP 2006; Ermer 2001; Taverniers et al. 2004; Ghoneim et al. 2006; Adhoum and Monser 2005; Hammam et al. 2006; Nouws et al. 2006).

The standard stock of QUI was kept in the dark at +4 °C. The stability of QUI stock solution (1000 µg · mL<sup>-1</sup>) was tested for two months and QUI in methanol solution was stable for at least two months period. Under the optimum conditions, the stability of 100.00 µg · mL<sup>-1</sup> of QUI solution in BR buffer at pH 10.0 was evaluated for 52 h period by the proposed method (Table 1). This solution was kept in the dark at +4 °C. Repetition of sample analysis after 30 h did not show any significant changes in the peak potential and peak current of QUI. Nevertheless, 100.0 µg · mL<sup>-1</sup> of QUI standard solution was prepared daily with BR at pH 10.0.

Under the optimum conditions, SWV voltammograms recorded with increasing amounts of QUI. As shown in

Table 1: Stability data of QUI standard solution  $(100.00 \ \mu g \cdot mL^{-1})$ 

	Peak current (nA)	Peak potential (mV)
Standard QUI 1 h after 2 h after 3 h after 4 h after 5 h after 23 h after	$\begin{array}{c} 409.80 \pm 6.37 \\ 411.83 \pm 4.55 \\ 415.11 \pm 3.38 \\ 409.67 \pm 5.00 \\ 411.19 \pm 4.35 \\ 406.45 \pm 4.96 \\ 412.05 \pm 6.07 \end{array}$	-1100 -1100 -1100 -1096 -1100 -1100 -1108
30 h after 48 h after 50 h after 52 h after	$\begin{array}{c} 412.10 \pm 5.40 \\ 370.65 \pm 2.31 \\ 347.35 \pm 4.86 \\ 308.10 \pm 6.12 \end{array}$	-1100 -1112 -1108 -1100

Table 2: Analytical parameters for voltammetric determination of QUI using developed SWV method (n = 8)

	SWV Method
Regression equation* Correlation coefficient (r)	y = 124.9965x - 13.8369 0.9992
Standard error of slope Standard error of intercept	0.81 2.91
Linearity range ( $\mu g \cdot mL^{-1}$ ) Number of data points	0.50-8.68
Limit of detection (LOD) $(\mu q \cdot mI^{-1})$	0.22
Limit of quantitation (LOQ) ( $\mu$ g · mL <sup>-1</sup> )	0.50

 $^{*}$  y = bx + a; x = concentration (µg  $\cdot$  mL^{-1}), y = peak current (nA), a = intercept, b = slope

	Intra-day	Intra-day			Inter-day		
Added $(\mu g \cdot mL^{-1})$	$\begin{array}{c} Found^a \\ (\mu g \cdot m L^{-1}) \end{array}$	Precision RSD %	Accuracy <sup>b</sup> (Bias %)	$\begin{matrix} Found^a \\ (\mu g \cdot m L^{-1}) \end{matrix}$	Precision RSD %	Accuracy <sup>b</sup> (Bias %)	
0.99	$1.02 \pm 0.01$	2.67	3.03	$1.00\pm0.02$	4.32	1.01	
3.38	$3.41\pm0.02$	1.09	0.89	$3.39\pm0.02$	1.79	0.30	
7.41	$7.44\pm0.02$	0.81	0.40	$7.42\pm0.03$	0.96	0.13	

Table 3: Evaluation of precision and accuracy of the proposed method for the determination of QUI (n = 7)

 $^a$  Found  $=\bar{x}=$  mean  $\pm$  standard error, RSD = Relative standard deviation,

<sup>b</sup> Accuracy = [(Found - Added)/Added]  $\times$  100

Fig. 4, peak currents increased linearly with increasing amounts of QUI. The calibration graphs of the peak current versus concentration were found to be linear over the range of  $0.50-8.68 \,\mu g \cdot m L^{-1}$ . The linearity was checked by preparing standard solutions for 9 different concentrations. The calibration curve is described by the following regression equation:

$$i_p = 124.9965 \text{ C} - 13.8369$$
,  $r = 0.9992 (n = 8)$ 

where  $i_p$  is the SWV peak current (nA) and C is the QUI concentration ( $\mu g \cdot m L^{-1}$ ), r is the correlation coefficient. Statistical evaluation of the regression lines regarding the standard error of the intercept and standard error of the slope and analytical characteristic of the proposed method are given in Table 2.

The sensitivity of the developed method was checked in terms of limit of detection (LOD) and limit of quantitation (LOQ) value. The LOD is defined as the lowest concentration of an analyte in a sample can be detected. LOD may be calculated according the formula:

## LOD = 3.3 (SD/S)

where SD is the standard deviation of y-intercepts of regression lines and S is the mean slope of the calibration curves. The calculated LOD value for the proposed method was  $0.22 \ \mu g \cdot mL^{-1}$ .

The LOQ is defined as the lowest concentration of an analyte in a sample which can be determined quantitatively with an acceptable level of accuracy and precision under the optimum conditions of the method. The LOQ value of this method was found to be  $0.50 \ \mu g \cdot mL^{-1}$  (RSD = 5.88%) for the developed method (n = 8). These LOD and LOQ data indicated that the proposed method could be considered sensitive.

The precision of this method was investigated with respect to repeatability and intermediate precision. In order to measure repeatability of the voltammetric instrument, 12 consecutive measurements were made with the same standard solution 3.38  $\mu g \cdot m L^{-1}$  of QUI under the optimum condition on the same day. The mean of measured peak potential and peak current were found to be 1102.67  $\pm$  0.75 with RSD of 0.24% and 412.22  $\pm$  3.46 with RSD of 2.90%, respectively. These results confirmed the high repeatability and precision of the proposed method for the QUI analysis.

The intra- and inter-day precision of this method were evaluated at three different concentration levels of QUI (0.99, 3.38 and 7.41  $\mu$ g · mL<sup>-1</sup>) in the linear range were used. The intra- and inter-day precision was studied on the same day and in 7 different days over a period one week. The RSD values of intra-day and inter-day precision of this method were in the range of 0.81–4.32% (Table 3). This indicates the high precision of the proposed method.

The accuracy of the proposed method was verified by calculating the percentage relative error (bias %) at three concen-

trations. The intra- and inter-day accuracy were carried out as mentioned in the precision section. The results obtained from intra- and inter-day accuracy of this developed method were found to be between 0.13 and 3.03% (Table 3).

To study the accuracy of the proposed method and to check the interference from excipients used in the dosage forms, recovery experiments were carried out using the standard addition method. This study was performed by addition of known amounts of QUI ( $0.96 - 4.61 \,\mu\text{g} \cdot \text{mL}^{-1}$ ) to known concentrations of the tablets and the mixtures were analyzed by the proposed method. The recovery results were calculated using the regression equation of the standard addition method. The recoveries of QUI were found to be from 100.44 to 100.96% (Table 4). These data showed that excipients presented in tablet dosage forms did not interfere with the analysis of OUI.

The mean regression equation of standard addition method was found to be

$$y = 130.2825 C + 401.6324$$
,  $r = 0.9989$ 

 $i_p$  is the SWV peak current (nA) and C is the added QUI concentration ( $\mu g \cdot m L^{-1}$ ), r is the correlation coefficient. There was no difference between the slopes of the two methods with calibration curve and standard addition methods. These values showed that excipients from tablet dosage forms did not significantly interfere and the developed method could be considered selective. The calibration curve method, which is easier and quicker than the standard addition method, was used in quantitative analysis of QUI.

The selectivity of the developed method was tested by analysis of  $3.38 \ \mu g \cdot m L^{-1}$  of QUI standard solution and pharmaceutical formulation as tablet solution containing  $3.38 \ \mu g \cdot m L^{-1}$  of QUI. Comparison of the recorded voltammograms obtained from both solutions showed that the peak potential and peak current of QUI did not change (Fig. 6). Therefore, no significant excipient interference was observed and this SWV method could successfully be applied to the analysis of QUI in the presence of tablet excipients. Consequently, the proposed method could be considered selective.

The effect of small variables such as pH (9.9–10.1) and concentration of supporting electrolyte (0.03–0.05 M) were evaluated for 3.38  $\mu g \cdot m L^{-1}$  of QUI. Only one parameter was changed in each experiment. The obtained mean percent-

Table 4: Recovery data of the developed method for the analysis of OUI (n = 6)

$\begin{array}{l} Added \\ (\mu g \cdot m L^{-1}) \end{array}$	$\begin{array}{l} Found \\ (\mu g \cdot m L^{-1}) \end{array}$	Recovery (%)	RSD of Recovery (%)
0.96 1.90 3.27 4.61	$\begin{array}{c} 0.97 \pm 0.01 \\ 1.92 \pm 0.02 \\ 3.29 \pm 0.01 \\ 4.63 \pm 0.01 \end{array}$	$\begin{array}{c} 100.96 \pm 0.17 \\ 100.88 \pm 0.83 \\ 100.65 \pm 0.34 \\ 100.44 \pm 0.20 \end{array}$	0.28 1.43 0.58 0.34

Found =  $\bar{x}$  = mean  $\pm$  standard error, RSD % = Relative standard deviation



Fig. 6: The SW voltammograms of QUI (3.38 µg · mL<sup>-1</sup>) a) Standard solution, b) Tablet solution of QUI at optimum conditions

Table 5: Robustness data of SWV method (n = 7)

	Found $(\mu g \cdot mL^{-1})$	RSD %	Recovery (%)
Standard $(3.38 \ \mu\text{g} \cdot \text{mL}^{-1})$ pH 9.9 pH 10.1 Buffer Molarity (0.03 M) Buffer Molarity (0.05 M)	$\begin{array}{c} 3.40 \pm 0.02 \\ 3.48 \pm 0.02 \\ 3.44 \pm 0.02 \\ 3.44 \pm 0.02 \\ 3.40 \pm 0.02 \end{array}$	1.08 1.28 1.25 1.25 1.49	$\begin{array}{c} 100.53 \pm 0.41 \\ 103.05 \pm 0.50 \\ 101.72 \pm 0.48 \\ 101.70 \pm 0.48 \\ 100.72 \pm 0.57 \end{array}$
Friedman analysis: $p = 0.072 > p = 0.05$			

 $\bar{\mathbf{x}} = \text{Mean} \pm \text{standard error}, \text{RSD} = \text{Relative standard deviation}$ 

Table 6: Ruggedness of the proposed method (Added of QUI 3.38  $\mu g \cdot m L^{-1}) \ (n=7)$ 

1. Analyst found $(\mu g \cdot mL^{-1})$	2. Analyst found $(\mu g \cdot mL^{-1})$
$ar{\mathbf{x}} = 3.40 \pm 0.02$ SD = 0.04 RSD % = 1.08	$\bar{\mathbf{x}} = 3.41 \pm 0.02$ SD = 0.05 RSD % = 1.57
$\begin{array}{l} t_c = 0.42,  t_t = 2.18,  p > 0.05 \\ F_c = 1.56,  F_t = 4.28,  p > 0.05 \end{array}$	

 $\bar{\mathbf{x}} =$  Mean  $\pm$  standard error, SD = Standard deviation, RSD % = Relative standard deviation

 $t_{\rm c}=$  calculated t value,  $t_{\rm t}=$  tabulated t value,  $F_{\rm c}=$  calculated F value,  $F_{\rm t}=$  tabulated F value

age recoveries and RSD % value based on the average of 7 replicate measurements were not significantly affected within the studied ranges of variations in the procedural operational conditions (Table 5). The statistical comparison was done with Friedman analysis and no difference was found between analysis results (p = 0.072 > p = 0.05). Consequently, this SWV was reliable for the analysis of QUI and the proposed method could be considered robust.

Two analysts analyzed  $3.38 \ \mu g \cdot m L^{-1}$  of QUI standard solution with the proposed method using the same instrument under the same optimized conditions at different days. The obtained results were found to reproducible, thus this developed method could be considered rugged with results of RSD % value of 1.08 and 1.57% for first and second analysts, respectively (Table 6). The statistical comparison of the results was done with the t- and F-tests ( $t_c = 0.42$  and  $F_c = 1.56$ , p > 0.05). The results show no statistical differences between different analysts.

In order to evaluate the applicability of the proposed method, a commercial tablet formulation of Acuitel<sup>®</sup> Tablets at two different dosage forms as 20 mg and 40 mg containing QUI was studied. The QUI content of tablets was determined by the calibration curve method (Table 7). As shown in Table 7, the excipients presented in tablet dosage forms did not interfere with the analysis of QUI.

To test the reliability of the developed method, QUI tablets were also analyzed with a reported method (Prieto et al. 2003). The t- and F-tests were carried out on the data to statistically examine the validity of the obtained results (Table 7). Since the calculated t- and F-values did not exceed the theoretical values, which verified there was no significant difference between the proposed and reported methods.

The SWV method described here is sensitive, accurate, rapid, reliable and simple to perform and a low cost quantitative method, thus suitable for analysis of QUI in pharmaceutical formulations. Preparation of the sample is easy and no separation and extraction procedure is required. Therefore, the presented method can be recommended for routine analysis of QUI in quality control laboratories.

## 3. Experimental

### 3.1. Apparatus

All experiments were performed using a BAS 100 B/W (Bioanalytical System, USA) electrochemical analyzer. A three electrode system consisted of HMDE as working electrode; an Ag/AgCl with saturated KCl as reference electrode and a platinum wire as counter electrode were used. The peak heights were automatically or manually measured using the "tangent fit" capability of the instrument. All measurements were performed at room temperature. All pH measurements were made with a Mettler Orion Model 420A digital pH meter calibrated with standard buffers.

#### 3.2. Chemicals and reagents

QUI was kindly provided from Dr. Reddy's Laboratories (Hyderabad, India) and it was used without further purification. Acuitel Tablets  $^{\text{R}}$  (20 mg

Table 7: Application of the proposed and reported methods to the analysis of commercial pharmaceutical formulations (Acuitel<sup>®</sup> Tablets) of QUI (n = 7)

Labeled claim (mg)	Proposed method	Proposed method		Comparison method (Prieto et al. 2003)	
	20.00	40.00	20.00	40.00	
Found (mg)	$\bar{\mathbf{x}} = 20.26 \pm 0.12$	$\bar{\mathbf{x}} = 40.55 \pm 0.23$	$\bar{\mathbf{x}} = 20.02 \pm 0.14$	$\bar{\mathbf{x}} = 39.95 \pm 0.29$	
RSD %	1.60	1.52	1.83	1.94	
t <sub>c</sub> value	1.31	1.61			
t <sub>t</sub> value	2.18	2.18			
F <sub>c</sub> value	1.34	1.58			
F <sub>t</sub> value	4.28	4.28			
Recovery %	$\bar{\mathbf{x}} = 101.30 \pm 0.61$	$\bar{\mathbf{x}} = 101.38 \pm 0.58$	$\bar{\mathbf{x}} = 100.08 \pm 0.69$	$\bar{\mathbf{x}} = 99.88 \pm 0.73$	
RSD % of recovery	1.60	1.52	1.83	1.94	

 $\bar{x}=\text{Mean}\pm$  standard error, RSD % = Relative standard deviation

 $t_c = \text{calculated t value, } t_t = \text{tabulated t value, } F_c = \text{calculated F value, } F_t = \text{tabulated F value}$ 

and 40 mg QUI per tablet) were kindly supplied by Pfizer A. Ş. (Istanbul, Turkey). All chemicals for preparation of buffers were used analytical reagent grade (Merck or Sigma).

#### 3.3. Standard solutions

The stock solution of QUI ( $1000 \ \mu g \cdot mL^{-1}$ ) was prepared in MeOH and kept in the dark and at +4 °C. Standard solutions of QUI were prepared daily by appropriate dilution of the stock solution with (BR) buffer at pH 10.0. Different buffers as supporting electrolytes, namely BR, phosphate and borate buffers were used. The investigated buffers were prepared in Milli-Q water. The pH of the solutions was adjusted with 0.1 M HCl or 0.1 M NaOH.

#### 3.4. General analytical procedure

The supporting electrolyte (2 mL), containing BR buffer at pH 10.0, was pipetted into colored electrochemical cell. Dissolved oxygen was removed from the solution by a purified nitrogen gas stream through the cell for at least 10 min. After the voltammogram of supporting electrolyte was recorded, then aliquots standard solutions of QUI were added. Nitrogen was pressed through the solution for 30 s and the square-wave voltammograms of these solutions were recorded using a new mercury drop. The studied potential range was from -400 to -1800 mV versus Ag/AgCl. SWV optimum conditions were as follows: (f) = 50 Hz, ( $\Delta E$ ) = 4 mV and ( $E_{SW}$ ) = 25 mV.

#### 3.5. Tablet solutions and procedure

QUI determination was also performed in commercially available two different tablet dosage forms Acuitel<sup>®</sup> Tablets. The amount of QUI present in each tablet was 20 or 40 mg. Ten tablets were weighed accurately and finely powdered and mixed. A portion of the powder equivalent to the average weight of one tablet was transferred into a 50 mL volumetric flask and 50 mL of methanol was added. The content of the flask was sonicated for 15 min to provide dissolution and then completed to volume with methanol. This solution was centrifuged for 15 min at 5000 rpm to separate out the insoluble excipients. An aliquot of clear supernatant was taken and diluted BR buffer at pH 10.0 to achieve the desired concentration. Then the procedure was completed as mentioned under Section 3.4. The drug content per tablet was determined by using the calibration curve method.

#### References

- Abbara C, Aymard G, Hinh S, Diquet B (2002) Simultaneous determination of quinapril and its active metabolite quinaprilat in human plasma using high-performance liquid chromatography with ultraviolet detection. J Chromatogr B Anal Technolog Biomed Life Sci 766: 199–207.
- Adhoum N, Monser L (2005) Determination of trimebutine in pharmaceuticals by differential pulse voltammetry at a glassy carbon electrode. J Pharm Biomed Anal 38: 619–623.
- Baranski AS, Fawcett WR, Gilbert CM (1985) Use of microelectrodes for the rapid determination of the number of electrons involved in an electrode reaction. Anal Chem 57: 166–170.
- Bard AJ, Faulkner LR (1980) Electrochemical Methods, Fundamentals and Applications; Wiley, New York p. 529.
- Bonazzi D, Gotti R, Andrisano V, Cavrini V (1997) Analysis of ACE inhibitors in pharmaceutical dosage forms by derivative UV spectroscopy and liquid chromatography (HPLC). J Pharm Biomed Anal 16: 431– 438.
- Cudina O, Jankovic I, Comor M, Vladimirov S (2006) Interaction of quinapril anion with cationic surfactant micelles of cetyltrimethylammonium bromide. J Colloid Interface Sci 301: 692–696.
- Ermer J (2001) Validation in pharmaceutical analysis. Part I: An integrated approach. J Pharm Biomed Anal 24: 755–767.
- Ferry JJ, Horvath AM, Easton-Taylor M, Toothaker RD, Colburn WA (1987) Determination of quinapril and its active metabolite in humanplasma and urine by gas-chromatography with electron-capture detection. J Chromatogr B 421: 187–191.
- Forette B, Koen R, Vicaut E (1992) Efficacy and safety of quinapril in the elderly hypertensive patient. Amer Heart J 123: 1426–1432.
- Ghoneim EM, El-Desoky HS, Ghoneim MM (2006) Adsorptive cathodic stripping voltammetric assay of the estrogen drug ethinylestradiol in pharmaceutical formulation and human plasma at a mercury electrode. J Pharm Biomed Anal 40: 255–261.

- Ghoneim EM, El-Attar MA, Hammam E, Khashaba PY (2007) Stripping voltammetric quantification of the anti-diabetic drug glipizide in bulk form and pharmaceutical formulation. J Pharm Biomed Anal 43: 1465–1469.
- Goto N, Kamata T, Ikegami K (1992) Trace analysis of quinapril and its active metabolite, quinaprilat, in human plasma and urine by gas-chromatography negative-ion chemical ionization mass-spectrometry. J Chromatogr B 578: 195–201.
- Goto N, Sato T, Shigetoshi M, Ikegami K (1992) Determination of dioxopiperazine metabolites of quinapril in biological-fluids by gas-chromatography mass-spectrometry. J Chromatogr B 578: 203–206.
- Gotti R, Andrisano V, Cavrini V, Bertucci C, Furlanetto S (2000) Analysis of ACE-inhibitors by CE using alkylsulfonic additives. J Pharm Biomed Anal 22: 423–431.
- Gumieniczek A, Hopkala H (1998) High-performance liquid chromatographic assay of quinapril in tablets. Pharm Acta Helv 73: 183–185.
  Hammam E, El-Attar MA, Beltagi AM (2006) Voltammetric studies on the
- Hammam E, El-Attar MA, Beltagi AM (2006) Voltammetric studies on the antibiotic drug cefoperazone quantification and pharmacokinetic studies. J Pharm Biomed Anal 42: 523–527.
- Hengy H, Most M (1988) Determination of the new ace-inhibitor quinapril and its active metabolite quinaprilate in plasma and urine by high-performance liquid-chromatography and pre-column labeling for fluorescentdetection. J Liq Chromatogr 11: 517–530.
- Hillaert S, De Grauwe K, Van den Bossche W (2001) Simultaneous determination of hydrochlorothiazide and several inhibitors of angiotensinconverting enzyme by capillary electrophoresis. J. Chromatography A 924: 439–449.
- Hillaert S, Van den Bossche W (2001) The quantitative determination of several inhibitors of the angiotensin-converting enzyme by CE. J Pharm Biomed Anal 25: 775–783.
- Hillaert S, Vander Heyden Y, Van den Bossche W (2002) Optimisation by experimental design of a capillary electrophoretic method for the separation of several inhibitors of angiotensin-converting enzyme using alkylsulphonates. J Chromatogr A 978: 231–242.
- ICH Harmonised Tripartite Guideline, International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use, "Validation of Analytical Procedures: Text and Methodology Q2 (R1)", (2005).
- Kowalczuk D, Hopkala H, Pietras R (2003) Simultaneous densitometric determination of quinapril and hydrochlorothiazide in the combination tablets. JPC-J. Planar Chromatogr – Modern TLC 16: 196–200.
- Kowalczuk D, Hopkala H (2004) Application of derivative spectrophotometry for simultaneous determination of quinapril and hydrochlorothiazide in the combination tablets. J AOAC Intern 87: 847–851.
- Kugler R, Olson SC, Smith D (1995) Determination of quinapril and quinaprilat by high-performance liquid-chromatography with radiochemical detection, coupled to liquid scintillation-counting spectrometry. J Chromatogr B 666: 360–367.
- Laviron E, Roullier L, Degrand C (1980) A multilayer model for the study of space distributed redox modified electrodes: Part II. Theory and application of linear potential sweep voltammetry for a simple reaction. J Electroanal Chem 112: 11–23.
- Nouws HPA, Matos CD, Barros AA, Rodrigues JA (2006) Electroanalytical determination of paroxetine in pharmaceuticals. J Pharm Biomed Anal 42: 341–346.
- Prieto JA, Alonso RM, Jimenez RM (2001) Solid-phase extraction and high-performance liquid chromatography applied to the determination of quinapril and its metabolite quinaprilat in urine. J Chromatogr Sci 39: 153–159.
- Prieto JA, Alonso RM, Jimenez RM (2002) Determination of the angiotensin-converting enzyme inhibitor quinapril and its metabolite quinaprilat in pharmaceuticals and urine by capillary zone electrophoresis and solidphase extraction. Electrophoresis 23: 102–109.
- Prieto JA, Alonso RM, Jimenez RM (2003) Square wave voltammetric determination of the angiotensin-converting enzyme inhibitors cilazapril, quinapril and ramipril in pharmaceutical formulations. Farmaco 58: 343–350.
- Taverniers I, Loose DM, Bockstaele EV (2004) Trends in quality in the analytical laboratory. II. Analytical method validation and quality assurance. Trends Anal Chem 23: 535–552.
- The United States Pharmacopoeia, The National Formulary, USP 29, NF 24, USP Convention, Rockville MD, 2006, 3050–3053.
- Vertes V, Haynie R (1992) Comparative pharmacokinetics of captopril, enalapril, and quinapril. Am J Cardiol 69: C8–C16.