

Department of Pharmacology¹, School of Pharmaceutical Sciences, Shandong University; The affiliated Hospital of Shandong Medical College², Linyi, P.R. China

Connexin 43, a new therapeutic target for cardiovascular diseases

YU-NING SONG¹, HAO ZHANG¹, JUN-YI ZHAO^{1,2}, XIU-LI GUO¹

Received November 25, 2008, accepted December 29, 2008

*Dr. Xiu-Li Guo, Department of Pharmacology, School of Pharmaceutical Sciences, Shandong University, No. 44 WenHuaXi Road, Jinan 250012, P.R. China
guoxl@sdu.edu.cn*

Pharmazie 64: 291–295 (2009)

doi: 10.1691/ph.2009.8814

Junctional channels (JC) play essential roles in the normal function of the cardiovascular system, mediating the spread of the electrical impulse that triggers synchronized contraction of the cardiac chambers and contributing to the coordination of activities between cells of the arterial wall. In mammalian hearts, cells most prominently express JC built of Connexin40 (Cx40), Cx43 and Cx45, of which Cx43 is the predominant intercellular gap junction protein. Changes in cardiovascular Cx gene expression during development or in response to (patho)physiological signals are expected to be a crucial factor in normal cardiac development and functions, and several cardiac diseases, such as atrial fibrillation, hypertrophy, heart failure, atherosclerosis, etc. Although the underlying molecular mechanisms have not yet been elucidated, recent research has found a variety of novel potential therapies related to Cx43 that can help to learn more about the mechanism of those cardiovascular diseases and the signaling pathway.

1. Introduction

Gap junctions are specialized channels formed between the membranes of two adjacent cells. They permit the direct passage of small molecules from the cytosol of one cell to its neighbor, and thus form a system of cell-cell communication that exists alongside familiar secretion/receptor signaling. Because of the rich potential for regulation of junctional conductance, and directional and molecular gating, gap junctional communication plays a crucial role in many aspects of normal tissue physiology (Levin 2002).

Gap junction channels are located in the cell membrane. Each cell contributes a half channel (termed a connexon), with each half channel formed by the oligomerization of six protein subunits (connexins) in a hexameric torus. The connexons dock to provide direct cell-to-cell communication through channels, which are large enough for molecules of up to about 1000 MW to pass (Danesh-Meyer and Green 2008).

The connexins (Cx) are a multigene family of gap junction proteins, which has highly conserved transmembrane and extracellular regions. All connexins share a common architecture: four hydrophobic amino acid regions that are membrane-spanning domains, two extracellular loops and three cytoplasmic components; amino- and carboxy-terminal regions, and a cytoplasmic loop. Most cell types express several different connexin isoforms in a temporal-, spatial-, and differentiation-specific manner. As far as we know, 21 isoforms in humans have been described. Each connexin is characterized by its predicted molecular mass;

for example, connexin 43 (Cx43) has a predicted mass of 43 kDa. Small molecules less than 1 kDa, such as cAMP, IP₃, K⁺, and Ca²⁺ ions, are capable of traveling through gap junction channels. Cx43 has been identified in several tissues including heart muscle, central nervous system, renal tubules and bone, and provides communication between the intracellular and extracellular compartments. It is also involved in Ca²⁺ signaling and glutamate release in astrocytes, in transduction of cell survival signals in bone, and in promoting cell injury in response to metabolic inhibition in isolated ventricular myocytes, astrocytes and renal tubule cells (Hawat and Baroudi 2008). That's why alterations in the distribution and/or level of Cx expression can be found under many pathological conditions, especially cardiac diseases. This article reviews the physiological features of Cx43 and its complex role in the genesis, propagation, and potential therapeutics of cardiovascular diseases.

2. General features of connexin 43

In the connexins family, Cx26, Cx30, Cx36, Cx40, Cx43, Cx45 are the most common connexins in human body, and multiple Cxs are coexpressed in the heart. Three principal Cxs are found in cardiac myocytes, Cx43, Cx40 and Cx45. Cx43 predominates in the heart as a whole but is typically coexpressed in characteristic combinations and relative quantities with Cx40 and/or Cx45 in a chamber-related and myocyte type-specific manner (Despantez et al. 2007). The most ubiquitously expressed member of the

connexin family, Cx43, has a half-life of only 1~3 h (in cardiac myocytes), from synthesis to degradation by internal digestion within proteasomes or lysosomes (Hawat and Baroudi 2008). Cx43 gap junctions contribute to local metabolic homeostasis and synchronization of cellular activities by allowing bidirectional, intercellular movement of ions, metabolites and second messengers. Following injury, a Cx43 mediated bystander effect allows the propagation of cell death and survival-modulating signals that modulate the fate of the surrounding cells, subsequently causes many serious diseases (Herve et al. 2007).

3. Cardiovascular diseases related to Cx43

3.1. Arrhythmia

As we know, alterations in conduction velocity may contribute to cardiac arrhythmias. Propagation of electrical activity between myocytes in the heart requires gap junction channels, which contribute to coordinated conduction of the heartbeat. Two overlapping forms of remodeling of GJs and Cx expression may occur: (1) structural remodeling, which involves changes in the arrangement or organization of GJs, (2) remodeling of Cx expression, in which the amount and/or types of Cx expressed are altered (Severs et al. 2006). Howarth et al. (2007) suggested that altered expression of gap junction connexin proteins may partly underlie heart rhythm disturbances. Other research shows similar results (Kanagaratnam et al. 2002). Altered expression of connexin43 and its phosphorylation also contribute to the arrhythmogenic substrate, which induced through an inhibition of cell-to-cell coupling, in early stage heart failure of cardiomyopathic hamster (Sato et al. 2008).

Acquired adult heart disease with an arrhythmic tendency is marked by remodeling of GJs and Cx expression. With regard to alterations in Cx expression, the most widely documented change is downregulation of Cx43 in the ventricle of patients with end-stage congestive heart failure. The expression of Cx43 and Cx40 mRNA in the left ventricle of patients with end-stage heart failure due to idiopathic dilated cardiomyopathy (nonischemic heart disease, N-IHD), or ischemic heart disease, IHD) was observed. Results show that Cx43 mRNA is markedly downregulated, and the loss of Cx43 could, in some instances, exceed 90%. Meanwhile, Cx40 is strongly up-regulated (up to four or five times) in the ischemic category (Despantez et al. 2004). These changes occur regardless of the primary cause of the condition (e.g., idiopathic dilated cardiomyopathy, ischemic cardiomyopathy, valvular aortic stenosis or other etiologies) (Kostin et al. 2003; Yamada et al. 2003; Kostin et al. 2004). The reduction in ventricular Cx43 appears to develop progressively during the course of disease, as suggested by the pattern of change observed in pressure-overloaded hearts with valvular aortic stenosis classified according to ejection fraction and the finding of reduced Cx43 in the nonfailing ventricles of coronary artery bypass patients (Kostin et al. 2004). Synchronously, an increased expression of Cx45 and Cx40 has been reported in the failing human ventricle (Kanagaratnam et al. 2002; Yamada et al. 2003). Thus, the Cx co-expression patterns of the diseased human ventricle involve a generalized increase in the ratio of Cx45:Cx43 and, in ischemic heart disease, a localized increase in Cx40:Cx43.

3.2. Ischemic heart disease

A recent study showed that Cx43 is also localized in the inner membrane of cardiomyocyte mitochondria and as a new player in the pathophysiology of myocardial ischemia-reperfusion injury (Ruiz-Meana et al. 2008). Normal ventricular myocytes are extensively coupled by gap junctions and have the capacity to rapidly increase the amount of connexin within gap junction plaques to meet physiological demands for enhanced cell-cell communication. However, myocytes can also rapidly uncouple in response to injury or disease. Ischemic and non-ischemic heart disease is associated with changes in the expression of connexins and remodeling of gap junctions (Saffitz et al. 2007). In addition, Cx43 is also involved in the formation of reactive oxygen species, which are crucial to the signal transduction cascade of ischemic pre/post-conditioning's protection. One study shows that the heterozygous Cx43-deficiency abrogates the cardioprotection induced by ischemic preconditioning, although the 50% Cx43-deficiency mice do not exaggerate tissue damage under sustained ischemia (Schwanke et al. 2003). In another study, heterozygous connexin 43-deficient (Cx43+/-) mice were subjected to permanent coronary occlusion. Infarct size of area at risk was reduced significantly in Cx43 +/- mice compared with Cx43+/+ mice (Heusch et al. 2006).

3.3. Heart hypoplasia

Although Cx43 is expressed in various cell types during embryonic development, mice with a global inactivation of Cx43 survive until birth but die in the perinatal period, which is due to an obstruction of the right ventricular outflow tract of the heart, as well as narrowing of the ventricular outlet region and hypertrophy of ventricular myocardium.

Congenital Diaphragmatic Hernia (CDH), one of the typical heart hypoplasia diseases, is one of the fatal diseases found in the newborn. The main symptom of CDH is heart hypoplasia and conotruncal defects as well as great vessel malformations that are likely related to disturbed neural crest developmental control. The latest study suggests that the main natural cause of CDH is the altered expression of Cx43, leading to the disturbance of GJ, which suggested that Cx43 might be a promising target of the CDH therapy (Levin 2002; Eckardt et al. 2006).

3.4. Vessel injury/repair

Endothelial dysfunction is thought to play a pivotal role in the initiation, progression, and occurrence of clinical complications of atherosclerosis. Maintenance of the functional integrity of the endothelial monolayer requires direct intercellular communication via gap junctions between individual cells. In mammalian vascular tissues, endothelial cells mainly express Cx37, Cx40, and Cx43, which is by far the predominant connexin (Chou et al. 2007). In human endothelial cells, different isoforms of connexins are subjected to different ways of regulation (Haussing et al. 2008). Studies showed that decreases in endothelial connexins occur in a variety of vascular disorders, closely related to atherosclerosis, such as hypertension and hyperlipidemia (Chou et al. 2007).

Tobacco abuse is still among the most important cardiovascular risk factors in modern society. Haussig et al. (2008) set an example to investigate whether sub-chronic nicotine exposure could induce endothelial dysfunction

and communication failure. Results showed that the levels of endothelial gap junction proteins Cx37 and Cx43 were significantly reduced. This indicated that nicotine induced functional intercellular communication failure in endothelial cells probably resulting from the down-regulation of Cx37 and Cx43 expression.

4. Novel potential therapies on cardiovascular disease with regulating connexin 43

Modulation of Cx43 expression has the potential to have widespread therapeutic applications to some cardiovascular disease. Although many mechanisms are far from clear, and moreover, the regulation of connexins and gap junctional conductance is very complex and differs between species, between cells from different tissues and between the connexin isoforms themselves, some novel potential therapies related to regulation of connexins have already been proposed as treatments of cardiovascular diseases.

4.1. Regulation of the phosphorylation of connexin 43

Phosphorylation of Cx43 subunits is known to be an important regulatory mechanism of Cx43 channels. Activation of protein kinases and protein phosphatases has been correlated with a reduction or increase of gap junction, since connexins could be phosphorylated by proteins kinases and dephosphorylated by protein phosphatases. The different phosphorylation form of connexins has different effects on gap junctional intercellular communication (GJIC). Suitable phosphorylation of connexin enhances GJIC, dephosphorylation or superphosphorylation of connexin restrains the GJIC. Alterations of phosphorylation state of Cx43 are related to cardiac function injury after hypoxia or reoxygenation. Detection of dephosphorylated Cx43 may serve as a diagnostic tool for examining ischemic heart disease (Matsushita et al. 2006; Wang et al. 2007).

Many enzymes, such as PKA, PKC, CK, MAPK, non-receptor tyrosine kinase, PP (PP1, PP2A, PP2B and PP2C), involved in the protein phosphorylation have effects on the regulation of Cx43 expression. Okadaic acid, a PP1/PP2 inhibitor, decreases ischemia-induced Cx43 dephosphorylation and the accumulation of non-phosphorylated Cx43 at the intercalated discs of myocytes in the whole heart (Jeyaraman et al. 2003). Thus, it shows a potent protection of cardiocytes under ischemia. Calyculin, which decreases ischemia-induced Cx43 dephosphorylation, showed similar results in the same experiment. It is concluded that isolated adult myocytes respond to ischemia in a manner similar to whole hearts and that ischemia-induced dephosphorylation of Cx43 is mediated, at least in part, by PP1-like phosphatase(s) (Jeyaraman et al. 2003; Ai and Pogwizd 2005).

4.2. Regulation of C43 by stimulating some related receptors

Certain receptors also have an effect on the Cx43 expression when they are activated. A typical example is the α -1-adrenoceptor subtype, whose activation selectively up-regulates the expression of connexin 43 in rat cardiomyocytes (Salameh et al. 2008), while Cx40, Cx45 are not influenced. Furthermore, the up-regulation of Cx43 expression could be inhibited by α_{1A} -adrenoceptor agonists, such as prazosin (Rojas et al. 2008). However, the role of α_1 -adrenoceptor subtypes in the regulation of Cx43 remained unclear until now. The physiological meaning of

this regulation could be that in case of adaptation of the heart muscle to higher exercise (hypertrophy), the electrical communication is regulated independently, which might provide a possibility to separate between contractility adaptation and establishment of electrical pathways for conduction. However, at present, this just remains a hypothesis.

Another example is that the activation of M_3 receptor improves GJIC and structural remodeling of Cx43. The mRNA level of Cx43 could be recovered from down-regulation level under pathologic conditions, such as ischemia (Yu et al. 2007). M_3 receptor agonist treatment before ischemia occurrence could abrogate the down-regulation of Cx43 (Yue et al. 2006). Another report showed that Cx43 and M_3 receptor are co-located in the cardiac myocyte, which makes both M_3 and Cx43 potential therapeutic targets in cardiovascular diseases, such as arrhythmia (Zhang et al. 2006).

4.3. Actions of hormones

Stress impacting the heart and the renin-angiotensin-aldosterone system (RAAS) causes changes in cellular structure and electrophysiology. Modulation of intercellular coupling through gap junctions could lead to an alteration in conduction velocity and conduction block. Aldosterone (Ald) shows a complex influence on the Cx43 expression. A significant down-regulation of Cx43 in neonatal rat cardiac myocytes is induced when exposed to 10^{-4} M Ald, but a significant up-regulation of Cx43 is induced with 10^{-8} M Ald. These results suggest that Ald affects gap junction remodeling in a dose dependent manner by modulating Cx43 synthesis through different receptors, which results in a concomitant change of conduction velocity (Shyu et al. 2001). AngII facilitates Cx43 synthesis, up-regulates Cx43 expression and increases the number and size of gap junction profiles in cardiac cells via AT1 receptor and ERK and p38 signal pathway (Polontchouk et al. 2002). Therefore, an effective prevention of RAAS activation during development of gap junction remodeling, including angiotensin-converting enzyme inhibitors, angiotensin receptor blockers, aldosterone receptor antagonists, etc, might be a new therapeutic strategy for the treatment of arrhythmias (Murray et al. 2007).

T3, a thyroid hormone, up-regulates Cx43 and accelerates gap junction formation in cultured neonatal cardiac myocytes. Thyroid status not only modulates the mechanical function of cardiomyocytes but also cell-to-cell communication which is essential for myocardial electrical and metabolic synchronizations (Tribulova et al. 2004).

4.4. Anti-cancer drugs

Cx43 also plays a crucial role in the growth, differentiation and proliferation of tumor cells. Many anti-cancer drugs show a certain influence on the expression of Cx43 which has the potential to be used in the treatment of cardiovascular diseases.

Recent reports studied the ability of Coleusin Factor (CF, also named FSK88), a new anti-cancer drug, on regulating the Cx43 expression and GJIC level in rat osteosarcoma UMR106 cells. Results demonstrated that CF increased the mRNA and protein expression of Cx43 in a dose- and time-dependent manner and up-regulated the diminished GJIC level in UMR106 cells by dye transfer experiments. In addition, Cx43 distribution in the plasma membrane was also dramatically enhanced by CF treatment. These

results provide the first evidence that CF could regulate connexin and GJIC, indicating that Cx43 may be a target of CF which could be used to exert cardioprotection as well as its anti-tumor effects (Geng et al. 2007).

As₂O₃, an anti-cancer drug, regulates the endothelial gap junctions. Cx43 transcripts and gap junctions in human aortic endothelial cells (HAEC) are reduced with gap-junction communication attenuated by As₂O₃. The reduction of Cx43 involves both down-regulation at the transcriptional level and an increase of degradation. This indicates that gap-junction communication in the vascular endothelium is inhibited by As₂O₃, which might be useful for the therapy of vascular dysfunction (Chou et al. 2007).

4.5. Active compounds

There are some active compounds that regulate the expression of connexins, for instance, nitrofen. Cx43 mRNA was overexpressed in the hearts of nitrofen-exposed embryonal rats, which could be normalized by vitamin A. The underlying mechanism of action deserves further investigation (Gonzalez-Reyes et al. 2006).

Recent experiments investigated that heptanol, as a reversible gap junction inhibitor, could down-regulate the connexins, consequently impair gap junction. This down-regulation, which has little selectivity on connexins family, at least including Cx40, Cx43 and Cx45, etc., diminishes soon after removal of heptanol (Keevil et al. 2000).

Some antipsychotic drugs, such as thioridazine and its active metabolite, mesoridazine, have well-known cardiac conduction side-effects that have resulted in fatal or nearly fatal clinical consequences. The physiological mechanisms responsible for these cardiac side-effects are unclear. Matesic et al. (2006) observed the effect of thioridazine and mesoridazine on gap junction-mediated intercellular communication between cells that express the major cardiac gap junction subtype Cx43. In light of the fact that the concentrations of thioridazine and mesoridazine used in these experiments are in the range of those used clinically in patients, the results suggest that inhibition of gap junction intercellular communication may be one factor contributing to the cardiac side-effects observed in some patients taking these medications.

5. Summary

Together with the above-mentioned studies, alterations of intercellular communication through gap junctional connections are likely contributing factors to the occurrence of many cardiovascular diseases. Such alterations may be as simple as microfibrosis interrupting gap junctional communication without a change in connexin quantity or distribution or as complex as remodeling of the three different connexin isoforms (Cx40, Cx43, Cx45) to change the makeup of heteromeric/heterotypic gap junctions. Dramatic increase in the understanding of the molecular mechanisms of normal and abnormal cellular physiology led to the development of exciting new theories of cardiovascular disease. But there are still many things remaining unknown about Cx43 and a lot of research needs to be done. However, Cx43 certainly is a promising target in cardiovascular diseases, which will lead to novel therapy options in the future.

Acknowledgement: This work was funded by the Natural Science Foundation of China Grants (30672451).

References

- Ai X, Pogwizd SM (2005) Connexin 43 downregulation and dephosphorylation in nonischemic heart failure is associated with enhanced colocalized protein phosphatase type 2A. *Circ Res* 96: 54–63.
- Chou Y, Tsai CH, Ueng KC, Tian TY, Chen SC, Yeh HI (2007) Endothelial gap junctions are down-regulated by arsenic trioxide. *Eur J Pharmacol* 569: 29–36.
- Danesh-Meyer HV, Green CR (2008) Focus on molecules: connexin 43—Mind the gap. *Exp Eye Res* 6. [Epub ahead of print].
- Despantez T, Dupont E, Severs NJ, Weingart R (2007) Gap junction channels and cardiac impulse propagation. *J Membrane Biol* 218: 13–28.
- Eckardt D, Kirchhoff S, Kim JS, Degen J, Theis M, Ott T et al. (2006) Cardiomyocyte-restricted deletion of connexin 43 during mouse development. *J Mol Cell Cardiol* 41: 963–971.
- Geng S, Sun B, Liu S, Wang J (2007) Up-regulation of connexin 43 and gap junctional intercellular communication by Coleus Factor is associated with growth inhibition in rat osteosarcoma UMR106 cells. *Cell Biol Int* 31: 1420–1427.
- Gonzalez-Reyes S, Fernandez-Dumont V, Calonge WM, Martinez L, Tovar JA (2006) Expression of Connexin 43 in the hearts of rat embryos exposed to nitrofen and effects of vitamin A on it. *Pediatr Surg Int* 22: 61–65.
- Haussing S, Schubert A, Mohr FW, Dhein S (2008) Sub-chronic nicotine exposure induces intercellular communication failure and differential down-regulation of connexins in cultured human endothelial cells. *Atherosclerosis* 196: 210–218.
- Hawat G, Baroudi G (2008) Differential modulation of unapposed connexin in 43 hemichannel electrical conductance by protein kinase C isoforms. *Pflügers Arch* 456: 519–527.
- Herve JC, Derangeon M, Bahbouhi B, Mesnil M, Sarrouilhe D (2007) The connexin turnover, an important modulating factor of the level of cell-to-cell junctional communication: comparison with other integral membrane proteins. *J Membr Biol* 217: 21–33.
- Heusch G, Büchert A, Feldhaus S, Schulz R (2006) No loss of cardioprotection by postconditioning in connexin 43-deficient mice. *Basic Res Cardiol* 101: 354–356.
- Howarth FC, Nowotny N, Zilahi E, El Haj MA, Lei M (2007) Altered expression of gap junction connexin proteins may partly underlie heart rhythm disturbances in the streptozotocin-induced diabetic rat heart. *Mol Cell Biochem* 305: 145–151.
- Jeyaraman M, Tanguy S, Fandrich RR, Lukas A, Kardami E (2003) Ischemia-induced dephosphorylation of cardiomyocyte connexin-43 is reduced by okadaic acid and calyculin A but not fostriecin. *Mol Cell Biochem* 242: 129–134.
- Kanagaratnam P, Rothery S, Patel P, Severs NJ, Peters NS (2002) Relative expression of immunolocalized connexins 40 and 43 correlates with human atrial conduction properties. *J Am Coll Cardiol* 39: 116–123.
- Keevil VL, Huang CL, Chau PL, Sayeed RA, Vandenberg JI (2000) The effect of heptanol on the electrical and contractile function of the isolated, perfused rabbit heart. *Pflügers Arch* 440: 275–282.
- Kostin S, Rieger M, Dammer S, Hein S, Richter M, Klövekorn WP, et al. (2003) Gap junction remodeling and altered connexin 43 expression in the failing human heart. *Mol Cell Biochem* 242: 135–144.
- Kostin S, Dammer S, Hein S, Klövekorn WP, Bauer EP, Schaper J (2004) Connexin 43 expression and distribution in compensated and decompensated cardiac hypertrophy in patients with aortic stenosis. *Cardiovasc Res* 62: 426–436.
- Levin M (2002) Isolation and community: a review of the role of gap-junctional communication in embryonic patterning. *J Membr Biol* 185: 177–192.
- Matesic DF, Abifadel DN, Garcia EL, Jann MW (2006) Effect of thioridazine on gap junction intercellular communication in connexin 43-expressing cells. *Cell Biol Toxicol* 22: 257–268.
- Matsushita S, Kurihara H, Watanabe M, Okada T, Sakai T, Amano A (2006) Alterations of phosphorylation state of connexin 43 during hypoxia and reoxygenation are associated with cardiac function. *J Histochem Cytochem* 54: 343–353.
- Murray KT, Mace LC, Yang Z (2007) Nonantiarrhythmic drug therapy for atrial fibrillation. *Heart Rhythm* 4(3 suppl): s88–90.
- Polontchouk L, Ebel B, Jackels M, Dhein S (2002) Chronic effects of endothelin-1 and angiotensin II on gap junctions and intercellular communication in cardiac cells. *FASEB J* 16: 87–89.
- Rojas Gomez DM, Schulte JS, Mohr FW, Dhein S (2008) Alpha-1-adrenoceptor subtype selective regulation of connexin 43 expression in rat cardiomyocytes. *Naunyn Schmiedebergs Arch Pharmacol* 377: 77–85.
- Ruiz-Meana M, Rodriguez-Sinovas A, Cabestrero A, Boengler K, Heusch G, Garcia-Dorado D (2008) Mitochondrial connexin 43 as a new player in the pathophysiology of myocardial ischaemia-reperfusion injury. *Cardiovasc Res* 77: 325–333.
- Saffitz JE, Hames KY, Kanno S (2007) Remodeling of gap junctions in ischemic and nonischemic forms of heart disease. *J Membr Biol* 218: 65–71.

- Salameh A, Krautblatter S, Baessler S, Karl S, Rojas Gomez D, Dhein S et al. (2008) Signal transduction and transcriptional control of cardiac connexin 43 up-regulation after alpha-1-adrenoceptor stimulation. *J Pharmacol Exp Ther* 326: 315–322.
- Sato T, Ohkusa T, Honjo H, Suzuki S, Yoshida MA, Ishiguro YS, et al. (2008) Altered expression of connexin 43 contributes to the arrhythmogenic substrate during the development of heart failure in cardiomyopathic hamster. *Am J Physiol Heart Circ Physiol* 294: 1164–1173.
- Schwanke U, Li X, Schulz R, Heusch G (2003) No ischemic preconditioning in heterozygous connexin 43-deficient mice—a further in vivo study. *Basic Res Cardiol* 98: 181–182.
- Severs NJ, Dupont E, Thomas N, Kaba R, Rothery S, Jain R et al. (2006) Alterations in cardiac connexin expression in cardiomyopathies. *Adv Cardiol* 42: 228–242.
- Shyu KG, Chen CC, Wang BW, Kuan P (2001) Angiotensin II receptor antagonist blocks the expression of connexin 43 induced by cyclical mechanical stretch in cultured neonatal rat cardiac myocytes. *J Mol Cell Cardio* 33: 691–698.
- Tribulova N, Shneyvays V, Mamedova LK, Moshel S, Zinman T, Shainberg A et al. (2004) Enhanced connexin-43 and α -sarcomeric actin expression in cultured heart myocytes exposed to triiodo-L-thyronine. *J Mol Histol* 35: 463–470.
- Wang R, Zhang C, Ruan Y, Liu N, Wang L (2007) Changes in phosphorylation of connexin 43 in rats during acute myocardial hypoxia and effect of antiarrhythmic peptide on the phosphorylation. *J Huazhong Univ Sci Technol Med Sci* 27: 241–244.
- Yamada KA, Rogers JG, Sundset R, Steinberg TH, Saffitz JE (2003) Up-regulation of connexin 45 in heart failure. *J Cardiovasc Electrophysiol* 14: 1205–1212.
- Yu H, Yue P, Zhang Y, Zhao W, Xing Y, Huo Y et al. (2007) Activation of cardiac M₃ receptor influences connexin 43 expression. *Chin Pharmacol Bull* 23: 711–715.
- Yue P, Lü YJ, Yang BF (2006) Advances in the study of cardiac M₃ receptor as a novel target of antiarrhythmic drugs. *Yao Xue Xue Bao* 41: 702–705.
- Zhang Y, Yue P, Xiao J, Yu HY, Pan ZW, Lin DH et al. (2006) Integration between M₃ muscarinic acetylcholine receptor and connexin 43 as antiarrhythmic targets in rat ventricular myocardium. *Yao Xue Xue Bao* 41: 395–400.

ERRATUM

Key Laboratory of Organism Functional Factors of the Changbai Mountain (Yanbian University)¹, Ministry of Education, Yanji; College of Pharmacy², Yanbian University, Yanji; College of Medicine³, Jilin, P.R. China

Evaluation of anticonvulsant activity of QUAN-0806 in various murine experimental seizure models

LI-PING GUAN^{1,2}, DONG-HAI ZHAO³, ZHE JIANG², HU-RI PIAO², ZHE-SHAN QUAN^{1,2}

Unfortunately, in the contribution by Li-Ping Guan et al. published in *PHARMAZIE* 2009, 64: 248–251 (doi: 10.1691/ph.2009.8738) the species of experimental animals was misprinted. In fact, Kunming mice were used instead of C57B/6 mice, as has erroneously been stated. Thus, the respective paragraph should read as follows:

4. Experimental

4.1 Animals

Kunming mice of either sex (obtained from the Laboratory of Animal Research, College of Pharmacy, Yanbian University) weighing from 18 to 25 g were used in these studies. The ratio of male and female mice in the control and drug-treated groups was kept same to avoid variation in re-

sponses due to sex differences. The animals were housed in plastic cages and were maintained on a 12:12 h light-dark (7:00 A.M. to 7:00 P.M.) schedule in a temperature-controlled ($21 \pm 2^\circ\text{C}$) animal room and relative humidity (about 40–50%). The animals had free access to standard rodent chow and water and were acclimatized to their environment for one week prior to experimentation. The animals were randomly distributed into different groups. Each experimental group comprised of a minimum of 10 animals. Mice were moved from the animal care facility to the testing laboratory room; each animal was caged separately after recording its body weight and was randomized to receive the treatments according to a random number table. Each mouse was used only once. The MES test and the rotarod test were carried out by the standard described in the Antiepileptic Drug Development Program (ADD), Epilepsy Branch, National Institutes of Health, Bethesda, MD, U.S.A. (Hester 1979a; Hester 1980b). All animal procedures conform to the Provision and General Recommendations of Chinese Experimental Animal Administration Legislation.