## **REVIEW**

Department of Pharmaceutical Technology<sup>1</sup>, Faculty of Health Sciences, Fernando Pessoa University, Porto; Institute of Biotechnology and Bioengineering<sup>2</sup>, Centre of Genetics and Biotechnology, University of Trás-os-Montes and Alto Douro (IBB/CGB-UTAD), Vila Real, Portugal

# **Nanoparticulate carriers (NPC) for oral pharmaceutics and nutraceutics**

C. M. LOPES<sup> $1,2$ </sup>, P. MARTINS-LOPES<sup>2</sup>, E. B. SOUTO  $1,2$ 

*Received October 20, 2009, accepted October 27, 2009*

*Prof. Dr. Eliana B. Souto, Department of Pharmaceutical Technology, Faculty of Health Sciences, Fernando Pessoa University, Rua Carlos da Maia, 296, P-4200-150 Porto, Portugal eliana@ufp.edu.pt*

*Pharmazie 65: 75–82 (2010) doi: 10.1691/ph.2010.9338*

The introduction of nanoparticulate carriers (NPC) in the pharmaceutic and nutraceutic fields has changed the definitions of disease management and treatment, diagnosis, as well as the supply food chain in the agri-food sector. NPC composed of synthetic polymers, proteins or polysaccharides gather interesting properties to be used for oral administration of pharmaceutics and nutraceutics. Oral administration remains the most convenient way of delivering drugs (e.g. peptides, proteins and nucleic acids) since these suffer similar metabolic pathways as food supply. Recent advances in biotechnology have produced highly potent new molecules however with low oral bioavailability. A suitable and promising approach to overcome their sensitivity to chemical and enzymatic hydrolysis as well as the poor cellular uptake, would be their entrapment within suitable gastrointestinal (GI) resistant NPC. Increasing attention has been paid to the potential use of NPC for peptides, proteins, antioxidants (carotenoids, omega fatty acids, coenzyme Q10), vitamins, probiotics, for oral administration. This review focuses on the most important materials to produce NPC for oral administration, and the most recent achievements in the production techniques and bioactives successfully delivered by these means.

## **1. Introduction**

Over the past few decades, new science disciplines and technologies, particularly bio- and nanotechnology, have deeply influenced the latest achievements in various areas including pharmaceutical and agri-food sectors. Several research lines have shown that the distribution profile of bioactive molecules (drugs and nutrients) can be modified at cell and tissue levels by their entrapment in submicron colloidal systems well-known as nanoparticulate carriers (NPC).

NPC are colloidal systems usually depicting a mean particle size below 1000 nm, being able to entrap bioactive molecules within their matrix and/or absorbed onto their surface (functionalized and/or coated). NPC have successfully been applied to overcome the physiological difficulties encountered in oral administration of bioactive molecules including pharmaceutically active compounds (peptides, proteins, genes, nucleic acids), nutraceutics, and probes/dyes usually applied for diagnostic purposes.

If directly coupled with the bioactive molecule (functionalized and/or coated) the NPC is solely used as a carrier system. Nevertheless, if NPC is produced from a bioactive raw-material the system itself, may reveal some biological activity, i.e. therapeutic function as its own carrier (Baran et al. 2002; Cascone et al. 2002; Duncan 2003; Kipp 2004).

NPC can be prepared from a variety of materials such as synthetic or semi-synthetic polymers, proteins, and also polysaccharides. The source materials may be of biological origin, such as albumin, gelatine, lactid acid, dextran, chitosan, or have "chemical" characteristics, e.g. synthetic polymers, carbon, silica, as well as metals (De Jong and Borm 2008).

Pharmazie **65** (2010) 2 75

To search for an appropriate NPC the main criteria to bear in mind upon the selection of a material include: (i) the required mean size and the inherent physicochemical properties of the loaded bioactive (e.g. aqueous solubility, chemical degradation) which influence the formulation stability and shelf life; (ii) the surface characteristics (e.g. charge and permeability) to improve their targeting; (iii) degree of biodegradability, biocompatibility and toxicity, (iv) drug release profile; and (v) antigenicity of the final product (Kreuter 1994). Based on these criteria, biodegradable raw-materials, with a limited life span as long as therapeutically needed, are considered an optimal option for NPC development.

The present review is focused on the most common rawmaterials used to produce NPC intended to deliver bioactives (pharmaceutics and/or nutraceutics) by oral administration. Several local or systemic applications may be foreseen providing successful examples of bioactives entrapped and delivered by means of NPC.

## **2. Nanoparticulate carriers for pharmaceutics and nutraceutics**

## *2.1. Polymeric-based NPC*

Polymeric-based NPC may be prepared from synthetic or semi-synthetic polymers, e.g. polyamines, poly(amino acids), polyesters, poly(alkyl cyanoacrykates), polyorthoesters (POE) and polyanhydrides (Linhardt 1989; Schwendeman et al. 1997). The bioactive can be entrapped (dissolved or dispersed) within the NPC matrix, adsorbed onto their surface, or even attached

in case a binding molecule is placed between the NPC surface and the bioactive. Furthermore, depending on the preparation procedure applied, two different structures may be obtained, i.e. nanospheres or nanocapsules. These will influence the NPC properties including their loading capacity, entrapment efficiency, yield of production, mean size, zeta potential, as well as the bioactive release profile. Nanospheres have a matrix-like structure, in which the bioactive is physically and uniformly dispersed or dissolved (Couvreur et al. 1995); whereas nanocapsules are composed of a polymeric shell and an inner oil core (Mainardes and Silva 2004). In these latter, the presence of oil leads to a vesicular structure in which the bioactive is confined to a cavity surrounded by a unique polymer membrane.

Polymeric-based NPC are of special interest from the pharmaceutical point of view due to their desirable properties such as biocompatibility, biodegradability, surface modification and functionalization facilities (Rawat et al. 2006). In addition, these carriers are more stable in the gastrointestinal (GI) tract than other colloidal carriers (e.g. liposomes), and can protect the entrapped bioactive molecules against GI tract environment. Moreover, modifying the type of polymer, well defined physicochemical characteristics may be accomplished according to the therapeutic needs, such as hydrophobicity, zeta potential, bioactive release profile (e.g. delayed, prolonged, triggered), and biological behaviour (e.g. targeting bioadhesion, improved cellular uptake) (Galindo-Rodriguez et al. 2005).

The main drawback of biodegradable NPC is their non-specific interaction either with cells and/or plasma proteins, leading to accumulation of the bioactive in non-target tissues. For a number of applications, surface-modified NPC have been developed to control their interactions with biologic milieu and therefore their biodistribution. In fact, it is possible to modify the NPC surface by adsorption or chemical grafting of hydrophilic molecules such as poly(ethylene glycol) (PEG) (Blummel et al. 2007; Ramachandran et al. 2008), polyethylene oxide (PEO) (Santander-Ortega et al. 2007; Tai et al. 2008), poloxamers (Petri et al. 2007; Kreuter and Gelperina 2008), poloxamine (Moghimi and Gray 1997), polysorbates (Ambruosi et al. 2006; Teeranachaideekul et al. 2008), and also phospholipids (des Rieux et al. 2006). In this context, PEG has received considerable attention as coating polymer. PEG-coated NPC could successfully avoid the mononuclear phagocyte system sequestration, thus increasing NPC blood circulation time in comparison to uncoated NPC (Gref et al. 1994; Gref 2002). These type of systems are well known as long-circulating particles (Kommareddy et al. 2005). They have been successfully produced for various pharmaceutical purposes such as control bioactive release profile, target particular organ/tissue, and deliver bioactive molecules like protein, peptides, and genes through a peroral route of administration (Langer 2000; Bhadra et al. 2002; Lee et al. 2005). The most suitable polymer may be chosen to achieve high entrapment efficiencies for a particular bioactive. Several methods are well described in the literature to prepare polymeric-based NPC, being usually selected according to the physicochemical characteristics of the bioactive to be entrapped (Montasser et al. 2000; Soppimath et al. 2001; Couvreur et al. 2002). Some polymers are less sensitive to processing conditions than others, due to their chemical characteristics, molecular weight and crystallinity (Lemoine et al. 1996).

From a general point of view, polymeric-based NPC have been developed mainly by three methods: dispersion of performed polymers, polymerization of monomers, and ionic gelation or coacervation of hydrophilic polymers. Nevertheless, several other techniques have emerged in the last years, being possible nowadays to obtain NPC by e.g. supercritical fluid technology (Reverchon and Adami 2006; Jensen et al. 2007; Moisan et al.

2008) and particle replication in non-wetting templates (PRINT) (Roland et al. 2005).

The pharmacokinetic behaviour of the bioactive compound (e.g. absorption, biodistribution pattern and elimination) is influenced by the polymeric composition of the NPC (e.g. hydrophobicity, surface charge and biodegradation profile), by any adjuvant substances included, and by the loaded bioactive (e.g. molecular weight, charge, localization in the nanosphere by adsorption or incorporation) (Reis et al. 2006). In addition, the nature of polymers influences both NPC size and the bioactive release profile. Although natural polymers generally give a relatively fast release, synthetic polymers enable controlled bioactive release over a longer period of time. Among the different classes of biodegradable polymers, the thermoplastic aliphatic poly(esters) like poly(lactide) (PLA), poly(glycolide) (PGA), and especially the copolymer of lactide and glycolide, poly(lactide-co-glycolide) (PLGA), poly( $\varepsilon$ -caprolactone) (PCL), are most commonly used for the NPC production. They have outstanding favourable properties, such as good biocompatibility, biodegradability (not requiring surgery for removal), and mechanical strength (Jain 2000). In addition, these polymers are easy to formulate into different carriers for a variety of bioactives e.g. vaccines, peptides, and proteins. They protect the loaded bioactive against degradation and control its site specific delivery. Due to their strongly hydrophobic nature, they are more efficient for the entrapment of hydrophobic rather than hydrophilic bioactive molecules.

Poly( $\varepsilon$ -caprolactone) (PCL) biodegradation suffers slow biodegradation, thus making this polymer a suitable candidate for long term delivery extending over a period of more than one year. This has led to its application in the preparation of different delivery systems in the form of microspheres, nanospheres and implants (Sinha et al. 2004). PCL also shows compatibility with a wide variety of polymers when it is used in the development of NPC (Gref et al. 2002; Lemarchand et al. 2003). It is promising to combine them with natural chitosan to develop amphiphilic copolymers which can be used to produce spherical or elliptical NPC suitable for bioactive delivery and targeting (Yu et al. 2006).

### *2.2. Protein-based NPC*

Relevant proteins for oral administration of pharmaceutics and nutraceutics include albumin and gelatine. Albumin is one of the major proteins of human plasma, being up to approximately 60% of the total plasma proteins. This protein shows solubilizing properties in dilute salt solutions and precipitates at relatively low temperatures. The thermal treatment at elevated temperatures (95–170  $\degree$ C) or the use of chemicals agents are on the basis of the preparation of albumin to form NPC (Gayathri 2003).

Albumin NPC have been prepared by coacervation and chemical cross-linkage with glutaraldehyde, and interferon  $\gamma$  was either adsorbed or entrapped in those albumin NPC (Segura et al. 2005). These NPC were able to load, mainly by electrostatic interactions, high amounts of interferon in its bioactive form. Other successful examples include albumin-based NPC for the delivery of DNA and oligonucleotides (Vogel et al. 2005). It was found that the presence of albumin during oligonucleotidesprotamine complexation led to remarkable improvements in particle stability. Furthermore, the introduction of albumin reduced considerably the cytotoxic side effects of the NPC compared with those made solely of oligonucleotides and protamine (Weyermann et al. 2005). After cellular uptake the oligonucleotides were distributed within the cytoplasm.

The use of albumin NPC to deliver oligonucleotides offers a number of advantages. This negatively charged system is able to load oligonucleotides, without requirements of positive

compounds. Moreover, albumin NPC promote the nuclear accumulation of the oligonucleotide, without additives destabilizing endosomal membrane (Arnedo et al. 2004). Furthermore, NPC are quickly removed from the circulation by opsonization with various serum components followed by phagocytosis. Thus, oligonucleotides entrapped into these NPC will most likely be protected for enough time to get intact into macrophages, where these systems would be able to promote the adequate intracellular uptake to exert their activity.

Cationic albumin coated PLA-PEGylated conjugated NPC, containing an active fragment analogue of arginine vasopressin, were developed by the double emulsion/solvent evaporation procedure (Lu et al. 2005; Xie et al. 2006) to promote the substance to penetrate through blood brain barrier via intravenous injection (Xie et al. 2006).

Gelatine is also a well-known protein widely used in medical applications. It is suitable to develop NPC since it exhibits biodegradability, weak antigen activity, and superior biocompatibility compared to other natural proteins, such as albumin (Lee et al. 2001). Gelatine is a natural macromolecule derived from collagen by prolonged boiling of animal bones and skin, being inexpensive, easily to cross-link and to be chemically modified, showing therefore a huge potential for the preparation of NPC.

One of the first approaches to develop gelatine-based NPC was reported to a system based on gelatine NPC entrapped in PLGA microspheres (Li et al. 1997). Gelatine NPC containing bovine serum albumin (BSA) as model protein were previously prepared, and then entrapped within the hydrophobic PLGA microspheres to create nanoparticle-microsphere assemblies. A phase separation method followed by a solvent extraction method was applied to produce such assemblies. Release experiments showed that this new system possesses sustained release characteristics for proteins, demonstrating as well the ability to protect proteins against integrity loss or denaturation. The loading of BSA in gelatine-based NPC was conducted using a modified w/o emulsion method, without recurring to emulsifiers (Li et al. 1998). The release of BSA from those particles was also observed to follow a diffusion-controlled release mechanism.

A coacervation process was applied to load DNA into gelatinebased NPC with the cell ligand transferrin covalently bound to the gelatine (Truong-Le et al. 1999). This system was developed for active gene delivery and targeting. After encapsulation within NPC, DNA was partially resistant to DNAse I and its biological integrity was successfully demonstrated. This gene delivery device also allowed the incorporation of lysosomolytic agents to reduce degradation of the DNA in the endosomal and lysosomal compartments of cells, improving the bioavailability of DNA molecules (Leong et al. 1998). Furthermore, NPC could be lyophilized for storage without loss of DNA bioactivity (Leong et al. 1998).

It was also demonstrated that gelatine-based NPC can easily be surface modified with sulfhydryl groups, involved in the covalent attachment of proteins, such as avidin (Coester et al. 2000). Avidin-conjugated gelatine NPC were pointed out as potential carrier system for peptide nucleic acid derivatives in antisense therapy (Coester et al. 2000; Balthasar et al. 2005).

Modified gelatine has also been synthesized and used to prepare NPC. Example are thiol-modified gelatine NPC prepared by desolvation, which have been employed to entrap DNA with greater transfection efficiency than regular gelatine NPC (Kommareddy and Amiji 2005). PEGylated gelatine NPC were also found to be more stable against DNAase degradation than regular gelatine NPC (Kaul and Amiji 2005), and also with higher transfection efficiencies and high desirable application for systemic delivery of gene material to solid tumours (Goldie and Mansoor 2005).

#### *2.3. Polysaccharide-based NPC*

Polysaccharides are diverse in their structure and properties since these compounds have a larger number of reactive groups, and a wide range of molecular weight, which contribute to their variable chemical structure (Liu et al. 2008). Polysaccharides can be divided into polyelectrolytes and non-polyelectrolytes. The polyelectrolytes group include positively charged polysaccharides (chitosan) and negatively charged polysaccharides (e.g. alginate, pectin, heparin, hyaluronic acid). The presence of chemically very different groups on the molecular chains allows them to be chemically and biochemically modified resulting in many types of polysaccharide derivates.

Polysaccharides are an important class of physiological materials with a very special properties like biocompatible, biodegradable, and protecting properties, which make them a promising choice to prepared NPC for bioactive entrapment and delivery (Sinha et al. 2004). In addition, most of natural polysaccharides have hydrophilic groups, such as hydroxyl, carboxyl and amino groups, enabling them to set up non-covalent bonds with biological tissue following bioadhesion (Lee et al. 2000). NPC prepared with bioadhesive polysaccharide (e.g. chitosan, alginate) could prolong the residence time, and therefore increase the absorption of the loaded bioactive.

Several studies have been published reporting the application of polysaccharides and their derivates as for bioactive delivery of several types of proteins (Li et al. 2007; Bayat et al. 2008; Rawat et al. 2008; Thirawong, Thongborisute et al. 2008).

Among various polysaccharides, chitosan and their derivates have early been used to prepare NPC. Chitosan, a non-branched polyamine of *D*-glucosamine and *N*-acetyl glucosamine molecules, is a naturally occurring linear polysaccharide, which is characterized for its biodegradable, non-toxic and biocompatible properties (Hirano 1996). Positively charged chitosan will bind to cell membranes and is reported to decrease the transepithelial electrical resistance and transiently opening tight conjunction between epithelial cells (Artursson et al. 1994). The safety of the chitosan, its mucoadhesive ability allowing to prolong residence time in the GI tract, and its ability to enhance absorption by increasing cellular permeability, have all been relevant properties to highlight its suitability for oral drug delivery (Bowman and Leong 2006). Due to its properties, chitosan allows the preparation of organic solvent free mucoadhesive NPC (Janes et al. 2001). Water soluble chitosan derivatives, easily dissolved in neutral aqueous medium, can also be used to produce NPC, aiming to avoid the potential cytotoxicity induced by acidic solution and, thus maintaining the bioactivity of the loaded molecules (Amidi et al. 2006; Sandri et al. 2006; Sandri et al. 2007; Chen et al. 2008).

Other polysaccharides used to prepare NPC, alone and in combination with other polysaccharides, include alginate (You and Peng 2004; Zahoor et al. 2005; Ahmad et al. 2006; Borges et al. 2007), dextran and dextran sulphate (Chauvierre et al. 2003; Bertholon et al. 2006; Tiyaboonchai and Limpeanchob 2007), heparin (Passirani et al. 1998; Labarre et al. 2005; Choi et al. 2006; Liu et al. 2007), amylose (Lalush et al. 2005), hialuronic acid (Chauvierre et al. 2003; Han et al. 2005), pectin (Chauvierre et al. 2003) and pullulan (Nishikawa et al. 1996; Akiyoshi et al. 1997; Akiyoshi et al. 2000; Kuroda et al. 2002; Na et al. 2003; Choi et al. 2006).

Liu et al. (2008) published a comprehensive review focusing on the mechanisms of polysaccharide-based NPC preparation. According to structural characteristics, polysaccharide-based NPC are prepared mainly by four different mechanisms, namely covalent cross-linking (Bodnar et al. 2005; Zhi et al. 2005; Liu et al. 2007), ionic crosslinking (Lu et al. 2006; Jain and Banerjee 2008; Tsai et al. 2008; Zhang et al. 2008), polyelectrolyte

**Pharmazie 65** (2010) 2 77

complexation (Lee et al. 2008), and self-assembly of hydrophobically modified polysaccharides (Jeong et al. 2006; Opanasopit et al. 2007; Yang et al. 2008). This latter method seems to be particularly suitable to develop NPC with targeting properties, e.g. polyester or polyakylcyanoacrylate NPC coated with PEG. To reveal an active targeting, specific ligands have to be attached to NPC surface to enable molecular recognition (Lemarchand et al. 2004). However, at the surface of PEGcoated NPC there are no reactive groups, which limits any chemical coupling of such ligands (Stella et al. 2000). Therefore, polysaccharide coatings have been considered as an alternative to the PEG coating because they have specific receptors in certain cells or tissues (Listinsky et al. 1998; Stahn and Zeisig 2000). In addition, several mucosal surfaces are good targets for polysaccharide recognition such as nasal, pulmonary and peroral. Lemarchand et al. (2004) reviewed different strategies to coat the surface of polymeric as well as inorganic NPC with polysaccharide and their medical application, mainly for imaging cancer approaches.

## **3. Uptake of oral pharmaceutics- and nutraceutics-loaded NPC**

Oral drug delivery is the preferable route for drug administration because it is non-invasive, avoids pain and discomfort associated with injections, and it decreases the contamination risk. It is also physiologically desirable, since the exogenous bioactive imitates their physiological pathway undergoing first hepatic bypass. Moreover, among the non-invasive routes of administration that have been evaluated for the delivery of bioactives, the oral route remains the most convenient, although it is not the most efficient for peptides and proteins due to their low absorption rate. The main reasons for the low oral bioavailability of such bioactives are related to chemical and conformational stability, cellular and luminal enzymatic degradation in the GI tract and poor intrinsic penetration of the intestinal membrane (Hamman et al. 2005; des Rieux et al. 2006; Morishita and Peppas 2006).

The development of an effective oral delivery system for proteins requires a comprehensive perception of their physicochemical properties, such as molecular weight, hydrophobicity, ionization coefficient and pH stability, as well as of the biological barriers that limit protein absorption through the GI tract (Mahato et al. 2003). Strategies to improve the oral bioavailability of bioactives have ranged from changing their physicochemical properties by modification of their lipophilicity and enzyme susceptibility, to add novel functionality using transport-carrier molecules that are recognized by endogenous transport-carrier systems in the GI tract and/or to their inclusion in specially adapted NPC (Morishita and Peppas 2006). Bioactive association with colloidal carriers, such as polymeric NPC, is one of several approaches proposed to improve their oral bioavailability. Such NPC are more stable in the GI tract than other colloidal carriers (e.g. liposomes), and can protect entrapped bioactives from GI environment. The use of different polymers allows the modulation of physicochemical and bioactive release properties and consequently the biological behaviour (Galindo-Rodriguez et al. 2005). Additionally, the NPC surface can be easily modified by adsorption or chemical grafting of certain hydrophilic molecules as previously mentioned. Moreover, their submicron size and their large specific surface area favour absorption compared to larger carriers. Consequently, it has already been widely shown that nanoencapsulation of bioactives protect them against the harsh environment of the GI tract and increases their transmucosal uptake and absorption (des Rieux et al. 2006).

Paracellular and transcellular routes have been explored to predict the intestinal absorption of proteins. Paracellular pathway is commonly pointed out as limited to protein absorption, due to the low surface area and tightness of the junctions of the intercellular spaces (Salamat-Miller and Johnston 2005). Strategies to modify the physicochemical properties of the intestinal wall and to modulate the tight junctions associated with the paracellular pathway have been mentioned (Salamat-Miller and Johnston 2005). A particular case has been reported to chitosan NPC because this natural polymer shows the special feature of adhering to mucous surface and transiently opening or widening the tight injunction between epithelial cells (Janes et al. 2001; Pan et al. 2002). Adhesive chitosan NPC were helpful in increasing the relative pharmacological bioavailability of insulin (Pan et al. 2002) and calcitonin (Prego et al. 2006) associated with the ability to weaken the intestinal barrier in a reversible way. A distinct advantage of proper particle size helped to increase the bioactive effects (Pan et al. 2002).

One of the advantages of NPC, when administered orally, is that they can be absorbed transcellularly, although in small quantities, not only through the membranous epithelial cells (M-cells) of the Peyer's patches in the Gut-Associated Lymphoid Tissue (GALT), but also through the much more numerous gut enterocytes (Hussain et al. 2001). The uptake of NPC carrying proteins by enterocytes is a limited but capable process (des Rieux et al. 2006). To improve the uptake of NPC, surface modifications (Vila et al. 2004; Bhattarai et al. 2006) and enhancing mucoadhesion (Tobio et al. 2000; Garcia-Fuentes et al. 2005) properties are usually explored with efficiency to promote the contact of proteins with the intestinal epithelium, increasing the concentration at the site of absorption. In the GI tract, the cationic NPC are favoured to bind to the mucous layer negatively charged, thus cationic polymers are selected as preferential mucoadhesive coating.

Peyer's patches are specific structures dispersed through the lymphoid nodules of the intestinal mucosa called O-MALT (Organized Associated Lymphoid Mucosa). They are rich in M cells, cells specialized for antigen sampling but also a potential portal for oral delivery of peptides and proteins since they possess high transcytotic capacity, and are able to transport a broad range of materials (Clark et al. 2001; Florence 2005). Uptake by M-cells have demonstrated to result in enhanced physiologic action of proteins after oral administration of NPC (Pinto-Alphandary et al. 2003; Borges et al. 2006). Furthermore, adsorptive endocytosis seems to occur through clathrin coated pits and vesicles, fluid phase endocytosis and phagocytosis (Qaddoumi et al. 2003). NPC size, electrical charge and surface hydrophobicity, are the main factors that influence their uptake by Peyer's patches. It is well accepted that hydrophobic, negatively charged protein-loaded NPC smaller that  $1 \mu m$  potentially show the best absorption rate (Jung et al. 2000; Yoo and Park 2004), however no generalities must be taken because other condition can rule the final absorption process.

Bioactive natural compounds, such as vitamins, antimicrobials, antioxidants, probiotics, bioactive peptides, can be incorporated into food matrices to developed innovative functional food, known as nutraceutic products. In the latest years, nutraceutics have been gathering attention from the scientific community, food manufacturers and consumers. Even though the involvement of these compounds in physiological functions is not yet fully understood, it is widely accepted that their addition to food matrices may have physiological benefits or disease preventing properties (Elliott and Ong 2002). One example of these benefits is the antihypertensive effect of dietary peptides derived from milk protein, mediated by angiotensin converting enzyme inhibition (Groziak and Miller 2000).

Considering oral administration, the most important key point for the effectiveness of a nutraceutic is keeping its bioavailability through dietary supplementation depending on several factors. The development of nutraceutic oral formulations is a great challenge mainly due to insufficient gastric residence time, low permeability and/or poor solubility within the gut, as well as instability under environment conditions encountered in food processing (temperature, oxygen, light) or in the GI tract (pH, enzymes, presence of other nutrients) (Bell 2001). Many approaches have been explored and even applied to protect and deliver bioactive molecules by oral route, such as chemical modifications, coupling agents, hydrogels and polymer-based NPC.

Although nanotechnology applications for pharmaceutical systems are well investigated and documented in which various systems have been developed for intelligent, modulated, and selective delivery of drugs to specific areas, the use of NPC in food sector are new emergent, but they are predicted to grow rapidly in the near future.

In order to facilitate the most favourable nanotechnological responses, the effective delivery of nutraceutics delivered by means of NPC will require appropriate food formulation and manufacturer process able to maintain the bioactive form fully functional until the time of consumption and deliver this integrate form to the physiological target (Chen et al. 2006).

Due to their submicron-meter size NPC seems to be promising to improve the bioavailability of nutraceutics, especially poorly soluble and high molecular weight bioactives, such as natural antioxidants, functional lipids (e.g. carotenoids, phytosterol, omega 3 fatty acids), proteins and numerous other compounds that are widely used as active ingredients in various food products (Chen et al. 2006). In the same way of oral pharmaceutical administration, in food matrices the NPC can dramatically prolong bioactive residence time in the GI tract by decreasing the influence of intestinal clearance mechanisms and increasing the specific surface available to interact with the biological support (Kawashim 2001; Arbos et al. 2002). In addition, NPC can penetrate deeply into tissues through fine capillaries, cross the epithelial lining fenestration and are generally taken up efficiently by cells (Desai et al. 1996, 1997; Lamprecht et al. 2004; des Rieux et al. 2006), thus allowing the nutraceutics delivery to target sites in the body. Nutrient digestion or absorption may increase or decrease depending on the structural, chemical, and physical states of NPC and the nutrients bound within them.

Some strategies are being explored to improve nutraceutics bioavailability. One of these approaches include the surface coating of NPC with a specific protein, which can modify the adhesive properties of NPC and their behaviour in the GI tract. In fact, some proteins (e.g. lectin) are able to bind specifically to sugar-residue-bearing sites located at the surface of epithelial cells (Goldstein et al. 1980). The application of this strategy is reported by covalent binding of lectin to polyvinylmethylether maleic anhydride NPC resulting in the decrease of terminal elimination rate in the GI mucosa, while BSA-coated NPC demonstrated high ability to adhere to the stomach mucosa (Arbos et al. 2002). Protein-coated surfaces can also be used to uptake the bioactive molecule by specific target-cell population (Goppert and Muller 2005) and protection of sensitive nutraceutics for the GI environment, e.g.  $\beta$ -lactoglobulin coated chitosan NPC (Chen and Subirade 2005). Due to its mucoadhesive properties, chitosan NPC should then adhere to the intestinal wall to facilitate the absorption of the nutraceutic. In another recent study (Jang and Lee 2008), the results clearly demonstrate the stability of chitosan NPC for L-ascorbic acid during the heat food processing and the possibility to enhance its antioxidant activity due to the prolonged release of L-ascorbic acid from chitosan NPC.

Pharmazie **65** (2010) 2 79

Other example of successful applications of NPC for nutraceutics is their use to deliver coenzyme Q10 (CoQ10), which is an antioxidant agent with well-established pharmacological activities. However, CoQ10 is only marketed as a nutritional supplement without any claim of therapeutic activity probably due to its poor physicochemical and biopharmaceutical properties leading to its low oral bioavailability. Ankola et al. (2007) documented the potential of NPC in improving the therapeutic value of molecules like CoQ10, facilitating its use as first line therapeutic agent for prophylaxis and therapy by overcoming the problems associated with its delivery in physiological conditions. These authors developed biodegradable NPC based on PLGA, using the quaternary ammonium salt didodecyldimethylammonium bromide as a stabilizer.

There are many other nutraceutics that are poorly absorbed, such as vitamin B12, vitamin K2, vitamin E and many phytonutrients, especially the polyphenols and terpenes, the latter of which include the carotenoids, chromonols, lemonoids and saponins, in which NPC seems to have great potential to enhance the bioavailability of those compounds and to generate new nutraceutic products.

Some nutraceutics-loaded NPC (e.g. vitamins, antimicrobials, antioxidants, etc) are commercially available (Chaudhry et al. 2008). Examples of these products include different Nanoceuticals<sup>TM</sup> from RBC Life Sciences<sup>®</sup> Inc. USA, Nano Calcium/Magnesium from Mag-I-Cal.com USA and nanoselenium-enriched Nanotea from Shenzhen Become Industry & Trade Co., Ltd., China. Another example is Nutri-Nano<sup>TM</sup> CoQ-10 from Solgar, USA. The product Novasol® from Aquanova® Germany contained nano-structured supplements based on "Nano-Sized Self-assembled Liquid Structures (NSSL)" from NutraLease Ltd. Israel, and NanoClustersTM delivery device for food products from RBC Life Sciences® Inc. USA. BioDelivery Sciences International has introduced their Bioral<sup>TM</sup> nanocochleate nutrient delivery system for micronutrients and antioxidants. The nanocochleates, with approximately 50 nm in size, are based on a phosphatidylserine carrier derived from soya bean.

#### **4. Conclusions**

There is a current need identify novel strategies to overcome limitations of oral delivery of sensitive bioactives being developed by biotechnology. A recently emerging scientific field – nanotechnology - works on a small scale to produce nanoparticulate carriers (NPC), but its implications in the pharmaceutical and agri-food markets are becoming massive. Besides the chemical nature of NPC, these submicron meter systems show promising features not only for pharmaceutical and cosmetics application, but also to develop innovate functional food. Delivery of therapeutic proteins/peptides and other nutraceutics have being receiving a considerable amount of attention worldwide, but several limitations to oral administration of such sensitive bioactives may still be pointed out. Intestinal membrane permeability, size, intestinal and hepatic metabolism and bioactive solubility are the major barriers playing an important role in decreasing bioavailability after oral administration. A number of approaches have been used to overcome these limitations. Nonconventional polymeric, protein and polysaccharide NPC have been developed which can be structurally modified to increase their targeting efficiency and delivery of entrapped bioactives in their matrix. Hydrophilic bioactives usually depict poor membrane permeability which may be overcome by increasing their membrane partitioning characteristics and their affinity to the mediated NPC. Likewise, surfacing NPC with mucoadhesive polymers seem to be an interesting strategy for the oral delivery

of sensitive bioactives enhancing their GI stability and bioavailability.

#### **References**

- Ahmad Z, Pandey R, Sharma S, Khuller GK (2006) Alginate nanoparticles as antituberculosis drug carriers: formulation development, pharmacokinetics and therapeutic potential. Indian J Chest Dis Allied Sci 48: 171–176.
- Akiyoshi K, Deguchi S, Tajima H, Nishikawa T, Sunamoto J (1997) Microscopic structure and thermoresponsiveness of a hydrogel nanoparticle by self-assembly of a hydrophobized polysaccharide. Macromolecules 30: 857–861.
- Akiyoshi K, Kang EC, Kurumada S, Sunamoto J, Principi T, Winnik FM (2000) Controlled association od amphiphilic polymers in water: thermosensitive nanoparticles formed by self-assembly of hydrophobically modified pullulans and poly(N-isopropyllacrylamide). Macromolecules 33: 3244–3249.
- Ambruosi A, Gelperina S, Khalansky A, Tanski S, Theisen A, Kreuter J (2006) Influence of surfactants, polymer and doxorubicin loading on the anti-tumour effect of poly(butyl cyanoacrylate) nanoparticles in a rat glioma model. J Microencapsul 23: 582–592.
- Amidi M, Romeijn SG, Borchard G, Junginger HE, Hennink WE, Jiskoot W (2006) Preparation and characterization of protein-loaded N-trimethyl chitosan nanoparticles as nasal delivery system. J Control Release 111: 107–116.
- Ankola DD, Viswanad B, Bhardwaj V, Ramarao P, Kumar MN (2007) Development of potent oral nanoparticulate formulation of coenzyme Q10 for treatment of hypertension: can the simple nutritional supplements be used as first line therapeutic agents for prophylaxis/therapy? Eur J Pharm Biopharm 67: 361–369.
- Arbos P, Arangoa MA, Campanero MA, Irache JM (2002) Quantification of the bioadhesive properties of protein-coated PVM/MA nanoparticles. Int J Pharm 242: 129–136.
- Arnedo A, Irache JM, Merodio M, Espuelas Millan MS (2004) Albumin nanoparticles improved the stability, nuclear accumulation and anticytomegaloviral activity of a phosphodiester oligonucleotide. J Control Release 94: 217–227.
- Artursson P, Lindmark T, Davis SS, Illum L (1994) Effect of Chitosan on the Permeability of Monolayers of Intestinal Epithelial-Cells (Caco-2). Pharm Res 11: 1358–1361.
- Balthasar S, Michaelis K, Dinauer N, von Briesen H, Kreuter J, Langer K (2005) Preparation and characterisation of antibody modified gelatin nanoparticles as drug carrier system for uptake in lymphocytes. Biomaterials 26: 2723–2732.
- Baran ET, Ozer N, Hasirci V (2002) *In vivo* half life of nanoencapsulated L-asparaginase. J Mater Sci Mater Med 13: 1113–1121.
- Bayat A, Larijani B, Ahmadian S, Junginger HE, Rafiee-Tehrani M (2008) Preparation and characterization of insulin nanoparticles using chitosan and its quaternized derivatives. Nanomedicine 4: 115–120.
- Bell LN (2001) Stability testing of nutraceuticals and functional food. In: R.E. C Wildman (Ed.), Handbook of nutraceuticals and functional food. New York: CRC Press: 501–516.
- Bertholon I, Vauthier C, Labarre D (2006) Complement activation by core-shell poly(isobutylcyanoacrylate)-polysaccharide nanoparticles: influences of surface morphology, length, and type of polysaccharide. Pharm Res 23: 1313–1323.
- Bhadra D, Bhadra S, Jain P, Jain NK (2002) Pegnology: a review of PEGylated systems. Pharmazie 57: 5–29.
- Bhattarai N, Ramay HR, Chou S-H, Zhang M (2006) Chitosan and lactic acid-grafted chitosan nanoparticles as carriers for prolonged drug delivery. Int J Nanomed 1: 181–187.
- Blummel J, Perschmann N, Aydin D, Drinjakovic J, Surrey T, Lopez-Garcia M, Kessler H, Spatz JP (2007) Protein repellent properties of covalently attached PEG coatings on nanostructured SiO(2)–based interfaces. Biomaterials 28: 4739–4747.
- Bodnar M, Hartmann JF, Borbely J (2005) Preparation and characterization of chitosan-based nanoparticles. Biomacromolecules 6: 2521–2527.
- Borges O, Cordeiro-da-Silva A, Romeijn SG, Amidi M, de Sousa A, Borchard G, Junginger HE (2006) Uptake studies in rat Peyer's patches, cytotoxicity and release studies of alginate coated chitosan nanoparticles for mucosal vaccination. J Control Release 114: 348–358.
- Borges O, Tavares J, de Sousa A, Borchard G, Junginger HE, Cordeiro-da-Silva A (2007) Evaluation of the immune response following a short oral vaccination schedule with hepatitis B antigen encapsu-

lated into alginate-coated chitosan nanoparticles. Eur J Pharm Sci 32: 278–290.

- Bowman K, Leong KW (2006) Chitosan nanoparticles for oral drug and gene delivery. Int J Nanomed 1: 117–128.
- Cascone MG, Lazzeri L, Carmignani C, Zhu Z (2002) Gelatin nanoparticles produced by a simple W/O emulsion as delivery system for methotrexate. J Mater Sci Mater Med 13: 523–526.
- Chaudhry Q, Scotter M, Blackburn J, Ross B, Boxall A, Castle L, Aitken R, Watkins R (2008) Applications and implications of nanotechnologies for the food sector. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 25: 241–258.
- Chauvierre C, Labarre D, Couvreur P, Vauthier C (2003) Novel polysaccharide-decorated poly(isobutyl cyanoacrylate) nanoparticles. Pharm Res 20: 1786–1793.
- Chen F, Zhang ZR, Yuan F, Qin X, Wang M, Huang Y (2008) *In vitro* and *in vivo* study of N-trimethyl chitosan nanoparticles for oral protein delivery. Int J Pharm 349: 226–233.
- Chen LY, Remondetto GE, Subirade M (2006) Food protein–based materials as nutraceutical delivery systems. Trends Food Sci Technol 17: 272–283.
- Chen LY, Subirade M (2005) Chitosan/beta-lactoglobulin core-shell nanoparticles as nutraceutical carriers. Biomaterials 26: 6041–6053.
- Choi SH, Lee JH, Choi SM, Park TG (2006) Thermally reversible pluronic/heparin nanocapsules exhibiting 1000-fold volume transition. Langmuir 22: 1758–1762.
- Clark MA, Jepson MA, Hirst BH (2001) Exploiting M cells for drug and vaccine delivery. Adv Drug Deliv Rev 50: 81–106.
- Coester C, Kreuter J, von Briesen H, Langer K (2000) Preparation of avidinlabelled gelatin nanoparticles as carriers for biotinylated peptide nucleic acid (PNA). Int J Pharm 196: 147–149.
- Couvreur P, Barratt G, Fattal E, Legrand P, Vauthier C (2002) Nanocapsule technology: a review. Crit Rev Ther Drug Carrier Syst 19: 99–134.
- Couvreur P, Dubernet C, Puisieux F (1995) Controlled Drug-Delivery with Nanoparticles-Current Possibilities and Future-Trends. Eur J Pharm Biopharm 41: 2–13.
- De Jong WH, Borm PJ (2008) Drug delivery and nanoparticles:applications and hazards. Int J Nanomed 3: 133–149.
- des Rieux A, Fievez V, Garinot M, Schneider Y-J, Preat V (2006a) Nanoparticles as potential oral delivery systems of proteins and vaccines: A mechanistic approach. J Control Release 116: 1–27.
- Desai MP, Labhasetwar V, Amidon GL, Levy RJ (1996) Gastrointestinal uptake of biodegradable microparticles: effect of particle size. Pharm Res 13: 1838–1845.
- Desai MP, Labhasetwar V, Walter E, Levy RJ, Amidon GL (1997) The mechanism of uptake of biodegradable microparticles in Caco-2 cells is size dependent. Pharm Res 14: 1568–1573.
- Duncan R (2003) The dawning era of polymer therapeutics. Nat Rev Drug Discov 2: 347–360.
- Elliott R, Ong TJ (2002) Science, medicine, and the future-Nutritional genomics. Br Med J 324: 1438–1442.
- Florence AT (2005) Nanoparticle uptake by the oral route: Fulfilling its potential? Drug Discov Tod: Technologies 2: 75–81.
- Galindo-Rodriguez SA, Allémann E, Fassi H, Doelker E (2005) Polymeric nanoparticles for oral delivery of drugs and vacines: A critical evaluation of *in vivo* studies. Crit Rev Ther Drug Car Sys 22: 419–463.
- Garcia-Fuentes M, Torres D, Alonso MJ (2005) New surface-modified lipid nanoparticles as delivery vehicles for salmon calcitonin. Int J Pharm 296: 122–132.
- Gayathri VP (2003) Biopolymer albumin for diagnosis and in drug delivery. Drug Development Research 58: 219–247.
- Goldie K, Mansoor A (2005) Tumor-Targeted Gene Delivery Using Poly(Ethylene Glycol)-Modified Gelatin Nanoparticles: *In Vitro* and *in Vivo* Studies. Pharm Res 22: 951–961.
- Goldstein IJ, Hughes RC, Monsigny M, Osawa T, Sharon N (1980) What should be called lectin? Nature 285: 1438–1442.
- Goppert TM, Muller RH (2005) Adsorption kinetics of plasma proteins on solid lipid nanoparticles for drug targeting. Int J Pharm 302: 172–186.
- Gref R (2002) Surface engineered nanoparticles as drug carriers. In: Baraton, MT, Editor. Synthesis, functionalization and surface treatment of nanoparticles. American Scientific Publishers: 233–256.
- Gref R, Minamitake Y, Peracchia MT, Trubetskoy V, Torchilin V, Langer R (1994) Biodegradable long-circulating polymeric nanospheres. Science 263: 1600–1603.
- Gref R, Rodrigues J, Couvreur J (2002) Polysaccharides grafted with polyester: novel amphiphilic copolymers for biomedical application Macromolecules 35: 9861–9867.

## **REVIEW**

- Groziak SM, Miller GD (2000) Natural bioactive substances in milk and colostrum: effects on the arterial blood pressure system. Br J Nutr 84 (Suppl 1): S119–125.
- Hamman JH, Enslin GM, Kotze AF (2005) Oral delivery of peptide drugs: barriers and developments. BioDrugs 19: 165–177.
- Han SK, Lee JH, Kim D, Cho SH, Yuk SH (2005) Hydrophilized poly(lactide-co-glycolide) nanoparticles with core/shell structure for protein delivery. Sci Technol Adv Mat 6: 468–474.
- Hirano S (1996) Chitin biotechnology applications. Biotechnol Annu Rev 2: 237–258.
- Hussain N, Jaitley V, Florence AT (2001) Recent advances in the understanding of uptake of microparticulates across the gastrointestinal lymphatics. Adv Drug Deliv Rev 50: 107–142.
- Jain D, Banerjee R (2008) Comparison of ciprofloxacin hydrochlorideloaded protein, lipid, and chitosan nanoparticles for drug delivery. J Biomed Mater Res B Appl Biomater 86: 105–112.
- Jain RA (2000) The manufacturing techniques of various drug loaded biodegradable poly(lactide-co-glycolide) (PLGA) devices. Biomaterials 21: 2475–2490.
- Janes KA, Calvo P, Alonso MJ (2001) Polysaccharide colloidal particles as delivery systems for macromolecules. Adv Drug Deliv Rev 47: 83–97.
- Janes KA, Fresneau MP, Marazuela A, Fabra A, Alonso MJ (2001) Chitosan nanoparticles as delivery systems for doxorubicin. J Control Release 73: 255–267.
- Jang KI, Lee HG (2008) Stability of chitosan nanoparticles for L-ascorbic acid during heat treatment in aqueous solution. J Agric Food Chem 56: 1936–1941.
- Jensen H, Bremholm M, Nielsen RP, Joensen KD, Pedersen JS, Birkedal H, Chen YS, Almer J, Sogaard EG, Iversen SB, Iversen BB (2007) In situ high–energy synchrotron radiation study of sol–gel nanoparticle formation in supercritical fluids. Angew Chem Int Ed Engl 46: 1113–1116.
- Jeong YI, Kim SH, Jung TY, Kim IY, Kang SS, Jin YH, Ryu HH, Sun HS, Jin S, Kim KK, Ahn KY, Jung S (2006) Polyion complex micelles composed of all–trans retinoic acid and poly (ethylene glycol)–grafted–chitosan. J Pharm Sci 95: 2348–2360.
- Jung T, Kamm W, Breitenbach A, Kaiserling E, Xiao JX, Kissel T (2000) Biodegradable nanoparticles for oral delivery of peptides: is there a role for polymers to affect mucosal uptake? Eur J Pharm Biopharm 50: 147–160.
- Kaul G, Amiji M (2005) Cellular interactions and *in vitro* DNA transfection studies with poly(ethylene glycol)–modified gelatin nanoparticles. J Pharm Sci 94: 184–198.
- Kawashim Y (2001) Nanoparticulate system for improved drug delivery. Adv Drug Deliv Rev 47: 1–2.
- Kipp JE (2004) The role of solid nanoparticle technology in the parenteral delivery of poorly water–soluble drugs. Int J Pharm 284: 109–122.
- Kommareddy S, Amiji M (2005) Preparation and evaluation of thiol– modified gelatin nanoparticles for intracellular DNA delivery in response to glutathione. Bioconj Chem 16: 1423–1432.
- Kommareddy S, Tiwari SB, Amiji MM (2005) Long–circulating polymeric nanovectors for tumor–selective gene delivery. Technol Cancer Res Treat 4: 615–625.
- Kreuter J (1994) Drug Targeting with Nanoparticles. Eur J Drug Met Pharm 19: 253–256.
- Kreuter J, Gelperina S (2008) Use of nanoparticles for cerebral cancer. Tumori 94: 271–277.
- Kuroda K, Fujimoto K, Sunamotto J (2002) Hierarchical self–assembly of hydrophobically modified pullulan in water: gelation by networks of nanoparticles. Langmuir 18: 3244–3249.
- Labarre D, Vauthier C, Chauvierre C, Petri B, Muller R, Chehimi MM (2005) Interactions of blood proteins with poly(isobutylcyanoacrylate) nanoparticles decorated with a polysaccharidic brush. Biomaterials 26: 5075–5084.
- Lalush I, Bar H, Zakaria I, Eichler S, Shimoni E (2005) Utilization of amylose–lipid complexes as molecular nanocapsules for conjugated linoleic Acid. Biomacromolecules 6: 121–130.
- Lamprecht A, Saumet JL, Roux J, Benoit JP (2004) Lipid nanocarriers as drug delivery system for ibuprofen in pain treatment. Int J Pharm 278: 407–414.
- Langer R (2000) Biomaterials in drug delivery and tissue engineering: one laboratory's experience. Acc Chem Res 33: 94–101.
- Lee CH, Jung KY, Choi JG, Kang YC (2005) Nano–sizedY(2)O(3): Eu phosphor particles prepared by spray pyrolysis. Mat Sc Engineering B: Sol State Mat Adv Technol 116: 59–63.
- **Pharmazie 65** (2010) 2 81
- Lee CH, Singla A, Lee Y (2001) Biomedical applications of collagen. Int J Pharm 221: 1–22.
- Lee JW, Park JH, Robinson JR (2000) Bioadhesive–based dosage forms: the next generation. J Pharm Sci 89: 850–866.
- Lee PW, Peng SF, Su CJ, Mi FL, Chen HL, Wei MC, Lin HJ, Sung HW (2008) The use of biodegradable polymeric nanoparticles in combination with a low–pressure gene gun for transdermal DNA delivery. Biomaterials 29: 742–751.
- Lemarchand C, Couvreur P, Besnard M, Costantini D, Gref R (2003) Novel polyester–polysaccharide nanoparticles. Pharm Res 20: 1284–1292.
- Lemarchand C, Gref R, Couvreur P (2004) Polysaccharide–decorated nanoparticles. Eur J Pharm Biopharm 58: 327–341.
- Lemoine D, Francois C, Kedzierewicz F, Preat V, Hoffman M, Maincent P (1996) Stability study of nanoparticles of poly(epsilon–caprolactone), poly(D,L–lactide) and poly(D,L–lactide–co–glycolide). Biomaterials 17: 2191–2197.
- Leong KW, Mao HQ, Truong–Le VL, Roy K, Walsh SM, August JT (1998) DNA–polycation nanospheres as non–viral gene delivery vehicles. J Control Release 53: 183–193.
- Li JK, Wang N, Wu XS (1997) A novel biodegradable system based on gelatin nanoparticles and poly(lactic–co–glycolic acid) microspheres for protein and peptide drug delivery. J Pharm Sci 86: 891–895.
- Li JK, Wang N, Wu XS (1998) Gelatin nanoencapsulation of protein/peptide drugs using an emulsifier–free emulsion method. J Microencapsul 15: 163–172.
- Li YY, Chen XG, Liu CS, Cha DS, Park HJ, Lee CM (2007) Effect of the molecular mass and degree of substitution of oleoylchitosan on the structure, rheological properties, and formation of nanoparticles. J Agr Food Chem 55: 4842–4847.
- Linhardt RJ (1989) Biodegradable polymers for controlled release of drugs. In Controlled Release of Drugs. M Rosoff, Editor, VCH Publishers: New York: 53–95.
- Listinsky JJ, Siegal GP, Listinsky CM (1998) Alpha–L–fucose: a potentially critical molecule in pathologic processes including neoplasia. Am J Clin Pathol 110: 425–440.
- Liu H, Chen B, Mao Z, Gao CY (2007) Chitosan nanoparticles for loading of toothpaste actives and adhesion on tooth analogs. J Appl Polym Sci 106: 4248–4256.
- Liu Z, Jiao Y, Liu F, Zhang Z (2007) Heparin/chitosan nanoparticle carriers prepared by polyelectrolyte complexation. J Biomed Mater Res A 83: 806–812.
- Liu Z, Jiao Y, Wang Y, Zhou C, Zhang Z (2008) Polysaccharides–based nanoparticles as drug delivery systems. Adv Drug Deliv Rev 60: 1650–1662.
- Lu B, Xiong SB, Yang H, Yin XD, Zhao RB (2006) Mitoxantrone–loaded BSA nanospheres and chitosan nanospheres for local injection against breast cancer and its lymph node metastases. I: Formulation and *in vitro* characterization. Int J Pharm 307: 168–174.
- Lu W, Tan YZ, Hu KL, Jiang XG (2005) Cationic albumin conjugated pegylated nanoparticle with its transcytosis ability and little toxicity against blood-brain barrier. Int J Pharm 295: 247–260.
- Mahato RI, Narang AS, Thoma L, Miller DD (2003) Emerging trends in oral delivery of peptide and protein drugs. Crit Rev Ther Drug Carrier Syst 20: 153–214.
- Mainardes RM, Silva LP (2004) Drug delivery systems: past, present, and future. Curr Drug Targets 5: 449–455.
- Moghimi SM, Gray T (1997) A single dose of intravenously injected poloxamine-coated long-circulating particles triggers macrophage clearance of subsequent doses in rats. Clin Sci (Lond) 93: 371–379.
- Moisan S, Marty JD, Cansell F, Aymonier C (2008) Preparation of functional hybrid palladium nanoparticles using supercritical fluids: a novel approach to detach the growth and functionalization steps. Chem Commun (Camb): 1428–1430.
- Montasser I, Briancon S, Lieto J, Fessi H (2000) Methods of obtaining and formation mechanisms of polymer nanoparticles J Pharm Belg 55: 155–167.
- Morishita M, Peppas NA (2006) Is the oral route possible for peptide and protein drug delivery? Drug Discov Tod 11: 905–910.
- Na K, Lee ES, Bae YH (2003) Adriamycin loaded pullulan acetate/sulfonamide conjugate nanoparticles responding to tumor pH: pH-dependent cell interaction, internalization and cytotoxicity *in vitro*. J Control Release 87: 3–13.
- Nishikawa T, Aklyoshi K, Sunamotto J (1996) Macromolecular complexation between bovine serum albumin and the self-assembled hydrogel nanoparticles of hydrophobized polysaccharides. J Am Chem Soc 118: 6110–6115.

## **REVIEW**

- Opanasopit P, Ngawhirunpat T, Rojanarata T, Choochottiros C, Chirachanchai S (2007) Camptothecin-incorporating N-phthaloylchitosan-g-mPEG self-assembly micellar system: Effect of degree of deacetylation. Col Surf B Bioint 60: 117–124.
- Pan Y, Li YJ, Zhao HY, Zheng JM, Xu H, Wei G, Hao JS, Cui FD (2002a) Bioadhesive polysaccharide in protein delivery system: chitosan nanoparticles improve the intestinal absorption of insulin *in vivo*. Int J Pharm 249: 139–147.
- Pan Y, Zheng J-M, Zhao H-Y, LIi Y-J, Xu H, Wei G (2002b) Relationship between drug effects and particle size of insulin-loaded bioadhesive microspheres. Acta Pharmacol Sin 23: 1051–1056.
- Passirani C, Barratt G, Devissaguet JP, Labarre D (1998) Interactions of nanoparticles bearing heparin or dextran covalently bound to poly(methyl methacrylate) with the complement system. Life Sci 62: 775–785.
- Petri B, Bootz A, Khalansky A, Hekmatara T, Muller R, Uhl R, Kreuter J, Gelperina S (2007) Chemotherapy of brain tumour using doxorubicin bound to surfactant-coated poly(butyl cyanoacrylate) nanoparticles: revisiting the role of surfactants. J Control Release 117: 51–58.
- Pinto-Alphandary H, Aboubakar M, Jaillard D, Couvreur P, Vauthier C (2003) Visualization of insulin-loaded nanocapsules: *in vitro* and *in vivo* studies after oral administration to rats. Pharm Res 20: 1071–1084.
- Prego C, Torres D, Alonso MJ (2006) Chitosan Nanocapsules as Carriers for Oral Peptide Delivery: Effect of Chitosan Molecular Weight and Type of Salt on the *In Vitro* Behaviour and *In Vivo* Effectiveness. J Nanosci Nanotechnol 6: 2921–2928.
- Qaddoumi MG, Gukasyan HJ, Davda J, Labhasetwar V, Kim KJ, Lee VH (2003) Clathrin and caveolin-1 expression in primary pigmented rabbit conjunctival epithelial cells: role in PLGA nanoparticle endocytosis. Mol Vis 9: 559–568.
- Ramachandran R, Paul W, Sharma CP (2009) Synthesis and characterization of PEGylated calcium phosphate nanoparticles for oral insulin delivery. J Biomed Mater Res B Appl Biomater 88: 41–48.
- Rawat M, Singh D, Saraf S (2008) Development and *in vitro* evaluation of alginate gel-encapsulated, chitosan-coated ceramic nanocores for oral delivery of enzyme. Drug Dev Ind Pharm 34: 181–188.
- Rawat M, Singh D, Saraf S, Saraf S (2006) Nanocarriers: Promising vehicle for bioactive drugs. Biol Pharm Bol 29: 1790–1798.
- Reis CP, Neufeld R, Ribeiro A, Veiga F (2006) Nanoencapsulation i: Methods for preparation of drug loaded polymeric nanoparticles. Nanomedicine 2: 8–21.
- Reverchon E, Adami R (2006) Nanomaterials and supercritical fluids. J Supercrit Fluids 37: 1–22.
- Roland JP, Maynor BW, Euliss LE, Exner AE, Denison GM, DeSimone JM (2005) Direct fabrication and harvesting of monodisperse, shape-specific nanobiomaterials. J Am Chem Soc 127: 10096–10100.
- Salamat-Miller N, Johnston TP (2005) Current strategies used to enhance the paracellular transport of therapeutic polypeptides across the intestinal epithelium. Int J Pharm 294: 201–216.
- Sandri G, Bonferoni MC, Rossi S, Ferrari F, Gibin S, Zambito Y, Di Colo G, Caramella C (2007) Nanoparticles based on N-trimethylchitosan: evaluation of absorption properties using *in vitro* (Caco-2 cells) and *ex vivo* (excised rat jejunum) models. Eur J Pharm Biopharm 65: 68–77.
- Sandri G, Poggi P, Bonferoni MC, Rossi S, Ferrari F, Caramella C (2006) Histological evaluation of buccal penetration enhancement properties of chitosan and trimethyl chitosan. J Pharm Pharmacol 58: 1327–1336.
- Santander-Ortega MJ, Bastos-Gonzalez D, Ortega-Vinuesa JL (2007) Electrophoretic mobility and colloidal stability of PLGA particles coated with IgG. Col Surf B Bioint 60: 80–88.
- Schwendeman S, Costantino HR, Gupta RK, Langer R (1997) Peptide, protein and vaccine delivery from implantable polymeric systems: Processes and challenges. in Controlled Drug Delivery: Challenges and Strategies. K Park, Editor, American Chemical Society: Washington D.C.: 229–267.
- Segura S, Espuelas S, Renedo MJ, Irache JM (2005) Potential of Albumin Nanoparticles as Carriers for Interferon Gamma. Drug Devel Ind Pharm 31: 271–280.
- Sinha VR, Bansal K, Kaushik R, Kumria R, Trehan A (2004) Poly-epsiloncaprolactone microspheres and nanospheres: an overview. Int J Pharm 278: 1–23.
- Soppimath KS, Aminabhavi TM, Kulkarni AR, Rudzinski WE (2001) Biodegradable polymeric nanoparticles as drug delivery devices. J Control Release 70: 1–20.
- Stahn R, Zeisig R (2000) Cell adhesion inhibition by glycoliposomes: effects of vesicle diameter and ligand density. Tumour Biol 21: 176–186.
- Stella B, Arpicco S, Peracchia MT, Desmaele D, Hoebeke J, Renoir M, D'Angelo J, Cattel L, Couvreur P (2000) Design of folic acid-conjugated nanoparticles for drug targeting. J Pharm Sci 89: 1452–1464.
- Tai YC, McGuire J, Neff JA (2008) Nisin antimicrobial activity and structural characteristics at hydrophobic surfaces coated with the PEO-PPO-PEO triblock surfactant Pluronic F108. J Colloid Interf Sci 322: 104–111.
- Teeranachaideekul V, Junyaprasert VB, Souto EB, Muller RH (2008) Development of ascorbyl palmitate nanocrystals applying the nanosuspension technology. Int J Pharm 354: 227–234.
- Thirawong N, Thongborisute J, Takeuchi H, Sriamornsak P (2008) Improved intestinal absorption of calcitonin by mucoadhesive delivery of novel pectin-liposome nanocomplexes. J Control Release 125: 236–245.
- Tiyaboonchai W, Limpeanchob N (2007) Formulation and characterization of amphotericin B-chitosan-dextran sulfate nanoparticles. Int J Pharm 329: 142–149.
- Tobio M, Sanchez A, Vila A, Soriano I, Evora C, Vila-Jato JL, Alonso MJ (2000) The role of PEG on the stability in digestive fluids and *in vivo* fate of PEG-PLA nanoparticles following oral administration. Col Suf B Bioint 18: 315–323.
- Truong-Le VL, Walsh SM, Schweibert E, Mao H-Q, Guggino WB, August JT, Leong KW (1999) Gene Transfer by DNA-Gelatin Nanospheres. Arch Biochem Biophys 361: 47–56.
- Tsai ML, Bai SW, Chen RH (2008) Cavitation effects versus stretch effects resulted in different size and polydispersity of ionotropic gelation chitosan-sodium tripolyphosphate nanoparticle. Carbohydr Polym 71: 448–457.
- Vila A, Gill H, McCallion O, Alonso MJ (2004) Transport of PLA-PEG particles across the nasal mucosa: effect of particle size and PEG coating density. J Control Release 98: 231–244.
- Vogel V, Lochmann D, Weyermann J, Mayer G, Tziatzios C, van den Broek JA, Haase W, Wouters D, Schubert US, Kreuter J (2005) Oligonucleotideprotamine-albumin nanoparticles: preparation, physical properties, and intracellular distribution. J Control Release 103: 99–111.
- Weyermann J, Lochmann D, Georgens C, Zimmer A (2005) Albuminprotamine-oligonucleotide-nanoparticles as a new antisense delivery system. Part 2: cellular uptake and effect. Eur J Pharm Biopharm 59: 431–438.
- Xie Y-L, Lu W, Jiang X-G (2006) Improvement of cationic albumin conjugated pegylated nanoparticles holding NC-1900, a vasopressin fragment analog, in memory deficits induced by scopolamine in mice. Behav Brain Res 173: 76–84.
- Yang XD, Zhang QQ, Wang YS, Chen H, Zhang HZ, Gao FP, Liu LR (2008) Self-aggregated nanoparticles from methoxy poly(ethylene glycol)-modified chitosan: Synthesis; characterization; aggregation and methotrexate release *in vitro*. Col Surf B Bioint 61: 125–131.
- Yoo HS, Park TG (2004) Biodegradable nanoparticles containing proteinfatty acid complexes for oral delivery of salmon calcitonin. J Pharm Sci 93: 488–495.
- You JO, Peng CA (2004) Calcium-alginate nanoparticles formed by reverse microemulsion as gene carriers. Macromol Symp 219: 147–153.
- Yu H, Wang W, Chen X, Deng C, Jing X (2006) Synthesis and characterization of the biodegradable polycaprolactone-graft-chitosan amphiphilic copolymers. Biopolymers 83: 233–242.
- Zahoor A, Sharma S, Khuller GK (2005) Inhalable alginate nanoparticles as antitubercular drug carriers against experimental tuberculosis. Int J Antimicrob Agents 26: 298–303.
- Zhang YY, Yang Y, Tang KS, Hu X, Zou GL (2008) Physicochemical characterization and antioxidant activity of quercetin-loaded chitosan nanoparticles. J Appl Polym Sci 107: 891–897.
- Zhi JF, Wang Y, Luo G (2005) Adsorption of diuretic furosemide onto chitosan nanoparticles prepared with a water-in-oil nanoemulsion system. React Funct Polym 65: 249–257.