



# TWO TRIGLYCOSIDIC TRITERPENE ASTRAGALOSIDES FROM HAIRY ROOT CULTURES OF ASTRAGALUS MEMBRANACEUS\*

YU ZHOU, MASAO HIROTANI, HEKAI RUI and TSUTOMU FURUYA

School of Pharmaceutical Sciences, Kitasato University, Minato-ku, Tokyo 108, Japan

(Received 5 September 1994)

**Key Word Index**—Astragalus membranaceus; Leguminosae; hairy root cultures; Agrobacterium rhizogenes; triterpenoids; agroastragalosides III and IV.

**Abstract**—Two new triglycosidic triterpene astragalosides, named agroastragaloside III and agroastragaloside IV, were isolated from the hairy root cultures of *Astragalus membranaceus*. From their spectroscopic data, the structures of the two compounds were elucidated as  $3-O-\beta-(2',3'-\text{di-}O-\text{acetyl})$ -D-xylopyranosyl-6- $O-\beta$ -D-glucopyranosyl-25- $O-\beta$ -D-glucopyranosyl-25- $O-\beta$ -D-glucopyranosyl-25- $O-\beta$ -D-glucopyranosyl-cycloastragenol, respectively. Both are triterpenetridesmosides.

#### INTRODUCTION

In preceding papers [1, 2], we reported the isolation of nine triterpene oligoglycosides: acetylastragaloside I, astragaloside I–IV, agroastragaloside I, II, isoastragaloside I and 3-O- $\beta$ -D-xylopyranosyl-cycloastragenol from hairy root cultures of Astragalus membranaceus. We now describe the structural elucidation of an additional two triglycosidic triterpene oligoglycosides, agroastragaloside III (1) and agroastragaloside IV (2) from the same culture source.

#### RESULTS AND DISCUSSION

Compound 1, a new astragaloside derivative, named agroastragaloside III, analysed for C51H82O21 by HR-FAB-mass spectrometry. The IR spectrum of 1 showed the presence of a hydroxyl group (3425 cm<sup>-1</sup>) and an ester carbonyl group (1750 cm $^{-1}$ ). The base peak at m/z143 in the mass spectrum of 1 resulted from cleavage between C-17 and C-20 suggesting the presence of the partial structure A (a 25-hydroxy-20, 24-epoxy residue) as found in astragaloside IV [1]. In the <sup>1</sup>H NMR spectrum, 1 showed signals from a cyclopropane-methylene at  $\delta 0.18$ and 0.55 (each d, J = 4.2 Hz, H<sub>2</sub>-19), seven tertiary methyl groups at  $\delta$ 0.90, 1.25, 1.27, 1.33, 1.42, 1.66 and 1.76, and two acetyl methyl groups at  $\delta$ 1.97 and 2.03, respectively (Table 1). Furthermore, the <sup>1</sup>H NMR spectrum of 1 clearly showed three anomeric doublets at  $\delta 4.81$ (J = 7.6 Hz),  $\delta 4.91 (J = 7.2 \text{ Hz})$  and  $\delta 5.06 (J = 7.3 \text{ Hz})$  in the downfield region, indicative of the presence of three  $\beta$ linked sugars [3, 4]. This was supported by the <sup>13</sup>C NMR spectrum, which showed three anomeric carbon signals at  $\delta$ 99.0, 104.2 and 105.1. The <sup>13</sup>C NMR spectrum of 1 displayed a total of 51 carbon signals. Based on a DEPT experiment, <sup>1</sup>H-<sup>1</sup>H and <sup>13</sup>C-<sup>1</sup>H COSY spectra and comparison with <sup>13</sup>C NMR data of related astragalosides [1, 2], all signals could be assigned (Table 2). These data suggest that 1 contained one additional glucose moiety relative to that of astragaloside IV. In the <sup>1</sup>H NMR spectrum, signals at  $\delta$ 1.97 (3H, s) and 2.03 (3H, s), and  $^{13}$ C NMR signals at  $\delta$ 170.0 and 170.7, showed the presence of two acetoxyl groups in 1. The sites of attachment of the xylose and glucose moieties of 1 were determined by means of a HMBC spectrum to be at C-3, C-6 and C-25, respectively [5]. In the HMBC spectrum, the first anomeric proton signal at  $\delta 4.81$  (H-1') showed longrange correlation with the carbon at  $\delta$ 89.4 (C-3). Also the second and third anomeric proton signals at  $\delta 4.91$  (H-1") and 5.06 (H-1") showed long-range correlation with the carbons at  $\delta$ 79.4 (C-6) and 78.8 (C-25), respectively. Consequently, xylose and glucose residues should be attached to the hydroxyl groups at C-3, C-6 and C-25 of 1, respectively. Furthermore, the positions of the acetoxyl groups of 1 were elucidated from its HOHAHA spectrum. In this spectrum the effects of the anomeric proton ( $\delta 4.81$ ) of the xylopyranose were observed on all other protons after a long mixing time and clearly two acetoxyl groups were located at C-2' and C-3' of the xylose moiety. These data led us to the conclusion that 1 was  $3-O-\beta-(2',3'-di-O$ acetyl)-D-xylopyranosyl-6-O- $\beta$ -D-glucopyranosyl-25-O- $\beta$ -D-glucopyranosyl-cycloastragenol.

Compound 2, also a new astragaloside derivative, named agroastragaloside IV, analysed for  $C_{49}H_{80}O_{20}$  by HR-FAB-mass spectrometry. The IR spectrum of 2 showed the presence of a hydroxyl group (3430 cm<sup>-1</sup>) and an ester carbonyl group (1755 cm<sup>-1</sup>). The base peak at m/z

<sup>\*</sup>Part 102 in the series 'Studies on Plant Tissue Cultures'. For Part 101 see ref. [2].

1408 Y. Zhou *et al.* 

143 in the mass spectrum resulted from cleavage between C-17 and C-20, again suggesting the presence of the partial structure A (a 25-hydroxy-20,24-epoxy residue) as found in 1. The <sup>1</sup>H NMR spectrum was very similar to that of 1 except for the absence of one acetyl methyl signal at  $\delta$ 1.97 (Table 1). The <sup>13</sup>C NMR spectrum was also closely similar to that of 1 except for the absence of the ester carbonyl carbon signal ( $\delta$ 170.7) and for the carbon signals owing to the xylose moiety. Furthermore, the <sup>1</sup>H NMR spectrum of 2 clearly showed three anomeric doublets at  $\delta 4.80 \ (J = 7.7 \ Hz), \ \delta 4.92 \ (J = 7.5 \ Hz)$  and 5.07 (J = 7.7 Hz) in the downfield region, indicative of the presence of three  $\beta$ -linked sugars, as in 1 [3, 4]. The presence of three  $\beta$ -linked sugars was supported by the <sup>13</sup>C NMR spectrum, which showed three anomeric carbon signals at  $\delta$ 99.0, 104.9 and 105.1. The <sup>13</sup>C NMR spectrum of 2 displayed a total of 49 carbon signals. Based on a DEPT experiment, <sup>1</sup>H-<sup>1</sup>H and <sup>13</sup>C-<sup>1</sup>H COSY spectra and comparison with <sup>13</sup>C NMR data of related astragalosides[1, 2], all signals could be assigned (Table 2). These data suggest that 1 was identical to 2, except for the presence of one less acetyl group. Furthermore, the sites of attachment of the xylose and glucose moieties of 2 were confirmed by means of a HMBC spectrum to be at C-3, C-6 and C-25, respectively [5]. The position of the acetoxy group in 2 was elucidated from its HOHAHA spectrum. In this spectrum, the effects of the anomeric proton ( $\delta 4.80$ ) of the xylopyranose were observed on all other protons after a long mixing time and clearly one acetyl group was located at C-2' of the xylose moiety. Therefore, the structure of 2 was determined to be 3-O- $\beta$ -(2'-O-acetyl)-D-xylopyranosyl-6-O- $\beta$ -D-glucopyranosyl-25-O-β-D-glucopyranosyl-cycloastra-

Kitagawa et al. [6] reported the structural elucidation of astragaloside VII (3) as 3-O- $\beta$ -D-xylopyranosyl-6-O- $\beta$ -D-glucopyranosyl-25-O- $\beta$ -D-glucopyranosyl-cycloastragenol isolated from an extract of Astragali Radix. It has quite a similar structure to the two new compounds agroastragaloside III (1) and agroastragaloside IV (2).

Table 1. <sup>1</sup>H NMR spectral data of compounds 1 and 2 (400 MHz,  $\delta$ , pyridine- $d_5$ )

Н	1	2		
3α	3.65 dd	3.41 dd		
	(11.6, 4.4)	(11.5, 4.2)		
5α	1.88 m	1.89 d		
		(8.1)		
6β	3.78 ddd	3.80 ddd		
	(8.5, 8.5, 4.0)	(7.8, 7.8, 3.5)		
16α	4.88 m	4.87 m		
17α	2.42 d	2.42 d		
	(7.2)	(7.7)		
H <sub>3</sub> -18	1.33 s	1.33 s		
19a	0.18 d	0.16 d		
	(4.2)	(4.1)		
19b	0.55 d	0.54 d		
	(4.2)	(4.1)		
H <sub>3</sub> -21	1.25 s	1.26 s		
22a	1.61 m	1.60 m		
22b	2.79 dd	2.80 dd		
	(19.8, 11.5)	(19.6, 11.5)		
24	3.91 m	3.89 m		
H <sub>3</sub> -26	1.42 s	1.42 s		
H <sub>3</sub> -27	1.66 s	1.66 s		
H <sub>3</sub> -28	1.76 s	1.82 s		
H <sub>3</sub> -29	1.27 s	1.29 s		
H <sub>3</sub> -30	0.90 s	0.91 s		
1'(xylose)	4.81 d	4.80 d		
	(7.6)	(7.7)		
1"(6-C-glucose)	4.91 d	4.92 d		
	(7.2)	(7.5)		
1"'(25-C-glucose)	5.06 d	5.07 d		
	(7.3)	(7.7)		
Ac	1.97 s	2.04 s		
Ac	2.03 s			

### EXPERIMENTAL

Mps: uncorr. <sup>1</sup>H and <sup>13</sup>C NMR: 400 and 100 MHz. FAB- and EI-MS: 20 eV. Specific rotations: MeOH. CC: silica gel (C-200).

Table 2. <sup>13</sup>C NMR data of compounds 1–3 (100 MHz,  $\delta$ , pyridine- $d_s$ )

	C	1	2	3*
Aglycone	1	32.1	32.2	
	2	29.9	30.1	
	3	89.4	89.0	88.6
	4	42.4	42.4	
	5	52.5	52.6	52.4
	6	79.4	79.4	79.1
	7	34.4	34.4	
	8	45.7	45.6	
	9	21.0	21.3	
	10	28.8	29.0	
	11	26.3	26.3	
	12	33.6	33.6	
	13	45.4	45.4	
	14	46.3	46.3	
	15	45.7	45.7	72.5
	16	73.7	73.7	73.5
	17	58.2	58.1	58.1
	18	21.4	21.4	
	19 20	29.0 87.4	28.8	07.3
	20	28.0	87.3 27.9	87.2
	22	35.2		
	23	26.2	35.2 26.2	
	23 24	82.2	82.2	82.2
	24 25	82.2 78.8		82.2 78.6
	25 26	23.1	78.7 23.1	70.0
	27	25.1	25.8	
	28	28.4	28.4	
	29	16.7	16.7	
	30	20.0	19.9	
2 O R D Vulanuranagul	1'	104.2	104.9	107.3
3- <i>O</i> -β-D-Xylopyranosyl moiety	2′	73.3	75.8	75.3
	3'	77.0	76.4	77.8
	4'	71.5	72.0	71.4
	5′	66.9	67.3	66.8
6-O-β-D-Glucopyranosyl	1"	105.1	105.1	104.8
moiety	2"	75.7	75.8	75.5
morety	3"	79.2	79.2	78.7
	4"	72.0	71.5	72.0
	5"	78.7	78.7	77.8
	6"	63.2	63.2	62.8
25- <i>O</i> -β-D-Glucopyranosy	1′′′	99.0	99.0	98.8
moiety	2'''	75.3	75.3	75.1
	3'''	78.3	78.3	78.4
	4'''	71.5	71.5	71.5
	5'''	78.1	78.2	77.8
	6′′′	62.9	62.9	62.8
Acetoxyl group		170.0	170.2	
J D 1 mg		170.7		
		21.0	21.2	

<sup>\*</sup>Data for compound 3 (astagaloside VII) from ref. [6].

Culture of hairy roots. Methods of isolation and culture of hairy roots of A. membranaceus were performed according to ref. [1]. Roots were subcultured every 4 weeks on modified Ganborg B5 medium [7] containing 30 g sucrose, 45 g CaCl<sub>2</sub>·2H<sub>2</sub>O, 38 g MgSO<sub>4</sub>·7H<sub>2</sub>O in 11H<sub>2</sub>O

at 25° in the dark at 60 rpm on a rotary shaker. Large-scale culture was carried out using 500 ml conical flask containing 250 ml of the medium described above.

Extraction and separation procedures. After 4-weeks culture (143 flasks), roots (7.14 kg fr. wt) were harvested with nylon mesh and dried at  $60^{\circ}$  for 7 days. Dried roots (549 g) were refluxed ( $\times$ 3) with 70% EtOH. After filtration, the extracts was combined and the solvent evapd under red. pres. The residue was partitioned between *n*-BuOH and H<sub>2</sub>O, and the *n*-BuOH fr. evapd to dryness (crude saponin fr. 30.8 g). The crude saponin fr. was subjected to Diaion HP 20 CC and eluted successively with 20, 50, 80 and 100% MeOH. After removal of solvent, the 100% MeOH eluate (12.3 g) was subjected to CC over silica gel (ca 1 kg Wako gel C-200) and eluted with a CHCl<sub>3</sub>-MeOH solvent system to yield 7 frs (frs A-G).

Agroastragaloside III (1). Fr. E (0.8 g) was purified by HPLC-1 and the fr. containing the peak at  $R_t$  16.5 mins was collected. Further purification of the collected fr. was achieved by repeated HPLC-1 and 1 was isolated from the fr. containing the peak at  $R_t$  16.5 min. Compound 1 (41.1 mg), needles. Mp 191–193° (from MeOH).  $[\alpha]_D^{25} + 5.9$  (MeOH; c 2.74). FAB-MS m/z:1053 [M + Na]<sup>+</sup>. HR-FAB-MS:  $C_{51}H_{82}O_{21}Na$  (required 1053.5248, [M + Na]<sup>+</sup> at m/z 1053.5277). EIMS (direct inlet) 20 eV, m/z (rel. int.): 472 (9), 454 (21), 395 (10), 271 (10), 187 (16), 143 (100), 125 (78). IR  $v_{max}^{RB}$  cm<sup>-1</sup>: 3425 (OH), 1750 (CO<sub>2</sub>), 1250, 1035. <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 2.

Agroastragaloside IV (2). Fr. E (0.8 g) was purified by HPLC-1 and the fraction containing the peak at  $R_t$  12.5 mins was collected. Further purification of the collected fr. was achieved by repeated HPLC-2 and 2 was isolated from the fr. containing the peak at  $R_t$  12.1 min. Compound 2 (16.4 mg), needles. Mp 187–189° (from MeOH).  $[\alpha]_D^{2.5} + 13.9$  (MeOH; c 0.75). FAB-MS m/z: 1011 [M + Na]<sup>+</sup>. HR-FAB-MS:  $C_{4.9}H_{80}O_{20}Na$  (required 1011.5140, [M + Na]<sup>+</sup> at m/z 1011.5167). EIMS (direct inlet) 20 eV, m/z (rel. int.): 472 (2), 454 (11), 395 (10), 311 (5), 270 (9), 143 (100), 125 (30). IR  $v_{max}^{KBT}$  cm<sup>-1</sup>: 3430 (OH), 1755 (COO). <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 2.

HPLC analysis. Analysis and isolation of astragaloside derivatives in system HPLC-1: column (19 mm  $\times$  150 mm) packed with μ-Bondasphere  $5\mu C_{18}$ -100 Å solvent 82.5% MeOH in  $\rm H_2O$ , flow rate 6 ml min $^{-1}$ ; HPLC-2: column (19 mm  $\times$  150 mm) packed with μ-Bondasphere  $5\mu$   $C_{18}$ -100 Å solvent 80% MeOH in  $\rm H_2O$ , flow rate 6 ml min $^{-1}$ . Absorbance and differential refraction detection.

Acknowledgement—We are indebeted to the members of the Analytical Centre of this University for NMR and MS.

## REFERENCES

- 1. Hirotani, M., Zhou, Y., Rui, H. and Furuya, T. (1994) *Phytochemistry* 36, 665.
- Hirotani, M., Zhou, Y., Rui, H. and Furuya, T. (1994) Phytochemistry 37, 1403.

1410 Y. Zhou et al.

3. Kitagawa, I., Wang, H. K., Saito, M., Takagi, A. and Yoshikawa, M. (1983) Chem. Pharm. Bull. 31, 698.

- 4. Wang, H. K., He, K., Ji, L., Tezuka, Y., Kikuchi, T. and Kitagawa, I. (1989) Chem. Pharm. Bull. 37, 2041.
- 5. Summers, M. F., Marzilli, L. G. and Bax, A. (1986)
- J. Am. Chem. Soc. 108, 4285.
- Kitagawa, I., Wang, H. K., Saito, M. and Yoshikawa, M. (1983) Chem. Pharm. Bull. 31, 716.
- 7. Gamborg, O. L., Miller, R. A. and Ojima, K. (1968) Exp. Cell Res. **50**, 151.