

LONGIPINANE DERIVATIVES FROM STEVIA VISCIDA

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Abstract—Two new longipinane derivatives were isolated from the roots of *Stevia viscida*. The structures were deduced as longipinan- 9α ,15-diangeloyloxy-1-one and longipinan- 9α -angeloyloxy-15-tigloyloxy-1-one on the basis of spectral evidence.

INTRODUCTION

Studies of nearly 50 Stevia species have shown that longipinene derivatives are frequent in the genus [1, 2]. In continuation of our search for longipinene derivatives from Stevia [3–6] we studied the chemical constituents of the roots of S. viscida HBK. Chromatography of the hexane extracts yielded the new longipinane derivatives longipinan- 9α , 15-diangeloyloxy-1-one (1) and longipinan- 9α -angeloyloxy-15-tigloyloxy-1-one (2).

Compound 1, isolated as an oil, showed $[\alpha]_D - 26^\circ$ and IR absorptions at 1700 and 1640 cm⁻¹ (unsaturated ester groups). The ¹H NMR spectrum (Table 1) showed typical signals for a longipinane skeleton [3, 7] and for two angelate groups [8]. A triplet at $\delta 5.07 (J = 3.4 \text{ Hz})$ is assigned to H-9, as in related esters [7], while an AB system at $\delta 3.92$ and 3.81 ($J_{AB} = 11$ Hz), owing to a methylene group bearing an ester, is assigned to methylene-15. The α -orientation of this group is evident after comparison of the ¹H NMR spectral data of 1 with those of the C-15 functionalized esters 3 and 4, isolated from S. potrerensis [9] and S. elatior [10],† respectively. The stereochemistry of the methyl group at C-3 and of the angelate group at C-9 follows from the 13C NMR chemical shifts of C-3 (δ 27.1) and of C-10 (δ 46.5), respectively (Table 2), which are similar to those of triacetate 5 [7] (C-3, δ 26.8; C-10, δ 45.0) and different at C-3 for the epimeric triacetate 6 [11] (C-3, δ 32.4).

Compound 2 showed $[\alpha]_D - 33^\circ$ and IR absorptions at 1607 and 1650 cm⁻¹. Most of the ¹H and ¹³C NMR signals were very similar to those of diangelate 1 (Tables 1 and 2). The vinylic signals at $\delta 6.85$ (1H) and 6.10 (1H) together with the methyl group signals between $\delta 2.00$ and

Table 1. ¹H NMR spectral data of 1 and 2 (200 MHz, CDCl₃, coupling constants in Hz in parentheses)

Н	1	2
2α	2.55 dd (8.4, 19.9)	2.55 dd (8.3, 18.4)
2β	2.10 dd (6.0, 18.5)	2.12 dd (5.7, 18.5)
3	2.35 m	2.35 m
4	2.18 d (5.5)	2.18 d (5.5)
5	2.00 s	1.98 s
7	1.85 m	1.83 m
7'	1.30 m	1.29 m
9	5.07 t (3.4)	5.08 t (3.1)
11	3.03 d (5.4)	3.07 d (5.5)
Me-12	1.09 d (6.7)	1.09 d (6.5)
Me-13	0.88 s	$0.88 \ s$
Me-14	1.02 s	1.00 s
15	3.92 d (11.0)	3.87 s
15'	3.81 d (11.0)	3.87 s
	OAng (C-9)	OAng (C-9)
3	6.10 qq (7.5, 1.5)	6.10 qq (7.5, 1.5)
Me-4	2.00 dq (7.5, 1.5)	2.00 dq (7.5, 1.5)
Me-5	1.90 quin (1.5)	1.90 quin (1.5)
	OAng (C-15)	OTigl (C-15)
3	6.10 qq (7.5, 1.5)	6.85 qq (7.5, 1.5)
Me-4	2.00 dq (7.5, 1.5)	1.81 dq (7.5, 1.5)
Me-5	1.90 quin (1.5)	1.83 quin (1.5)

[†]The position of the ester group in 4 was revised. See reference [3].

^{1.80} indicate the presence of one tiglate and one angelate ester group in 2. The positional assignment of both esters can be done when the chemical shifts of the signals owing to the protons geminal to the oxygen atoms (H-9, H-15 and H-15') are compared with those of diangelate 1. While the signal for H-9 had a very similar chemical shift in both substances, the signals for H-15 and H-15' experienced an evident change (Table 1). Therefore, the angelate group was located at C-9 and the tiglate group at

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1: $R^1 = H$; $R^2 = OAng$ 2: $R^1 = H$; $R^2 = OTigl$

 $3: R^1 = R^2 = OAng$

5: $R = \alpha$ -Me **6**: $R = \beta$ -Me

Table 2. ¹³C NMR data of 1 and 2 (50.3 MHz, CDCl₃)

С	1	2
1	211.7	212.1
2	42.3	42.3
3	27.1	27.1
4	44.6	44.7
5	42.2	42.7
6	36.3	36.5
7	28.1	28.0
8	26.0	26.0
9	77.6	77.7
10	46.5	46.5
11	53.7	54.1
12	19.7	19.7
13	20.3	20.3
14	20.3	20.2
15	71.7	72.7
	OAng (C-9)*	OAng (C-9)*
1	167.3	167.5
2	127.9	128.1
3	138.4	138.7
4	15.9	16.0
5	20.7	20.7
	OAng (C-15)*	OTigl (C-15)*
1	167.7	168.2
2	127.5	128.5
3	138.7	137.8
4	15.9	14.4
5	20.7	12.1

^{*}The ester residue signals were assigned as deduced by HETCOR experiments and by analogy. See refs [8], [13].

C-15. This is also in agreement with the ¹³C NMR data (Table 2), which showed a 1 ppm chemical shift difference for C-15 on going from 1 to 2.

Although many longipinene esters functionalized at C-7 and C-15 have been isolated from several *Stevia* species [2, 6, 9, 12], the isolation of esters at C-9 and C-15 like 1 and 2 is rare. The only other known case is the unsaturated analogue 4, which has been isolated from *S. elatior* [10].

EXPERIMENTAL

¹H and ¹³C NMR spectra were measured in CDCl₃ with TMS as int. standard. The ¹³C signals were assigned from APT and HETCOR experiments and by comparison with published data [8, 13]. CC was performed on alumina (Merck) or silica gel (230–400 mesh). TLC was carried out on silica gel PF₂₅₄ (Merck) plates.

Plant material. Stevia viscida HBK was collected at km 283 of the México-Morelia federal road No. 15 in September 1992. A voucher specimen is deposited at the Herbarium of the Instituto de Ecología A. C., Pátzcuaro, Mich. México, where Prof. Jerzy Rzedowski identified the plant material.

Extraction and isolation. The air-dried roots (985 g) of S. viscida were extracted \times 3 with hexane under reflux for 4 hr. After removal of the solvent, the crude extract (2.2 g) was chromatographed over alumina (30 g), 50 ml frs being collected as follows: frs 1–16 (petrol), 17–24 (C_6H_6), 25–30 (CHCl₃), 31–44 (EtOAc). Fr. 31 was rechromatographed under the same conditions collecting 31 frs of 20 ml and monitoring by TLC (hexane–EtOAc, 7:3). Fr. 7 was subjected to prep. TLC (hexane–EtOAc, 4:1 three

developments) giving ca 40 mg of diangelate 1 and 10 mg of angelate-tiglate 2.

Longipinan-9α,15-diangeloyloxy-1-one (1). Oil. UV $\lambda_{\rm max}^{\rm EIOH}$ nm (log ε): 223, (3.95); [α] $_{589}$ – 26°; IR $\nu_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$: 1700, 1640. 1 H NMR see Table 1; 13 C NMR see Table 2.

Longipinan-9α-angeloyloxy-15-tigloyloxy-1-one (2). Oil. UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (log ε): 223, (3.88); [α]₅₈₉ – 33°; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1706, 1650; ¹H NMR see Table 1; ¹³C NMR see Table 2.

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