



## TRITERPENOIDS FROM *WALSURA PISCIDIA*

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**Key Word Index**—*Walsura piscidia*; Meliaceae; triterpenoids; piscidinol F.

**Abstract**—Piscidinol F, a new apotirucallane, has been isolated from the leaves of *Walsura piscidia* in addition to the other piscidinols. The side chain configuration of piscidinol B has been established from  $^1\text{H}$  NMR data.

### INTRODUCTION

In the course of screening plants belonging to the family Meliaceae for insect control activity, it was observed that extracts from the leaves of *Walsura piscidia* Roxb. displayed significant antifeedant activity against some important insect pests. This plant has been investigated in some detail and shown to contain tirucallane and apotirucallane triterpenoids [1], but none of these compounds has been tested for biological activity. It was considered worthwhile to reinvestigate this plant and isolate the triterpenoidal compounds for testing as anti-feedants.

A number of compounds were obtained, piscidinols, B–E, but piscidinol A was not present in our extract. A new compound, piscidinol F, was isolated and its structure elucidated on the basis of spectral data. The configuration of the side chain of piscidinol B was established, as this was not done earlier.

### RESULTS AND DISCUSSION

Piscidinol F (1),  $\text{C}_{33}\text{H}_{48}\text{O}_9$ , contains seven tertiary methyls ( $\delta_{\text{H}}$  1.09, 1.11, 1.17, 1.22, 1.25, 1.28 and 1.44), one acetate ( $\delta_{\text{H}}$  2.17 s, 3H) and one methoxy ( $\delta_{\text{H}}$  3.23 s, 3H) group. Its spectroscopic properties indicate a close relationship to piscidinol D [1]. A ring-A enone system is shown by the presence of two well-separated doublets at  $\delta$  8.05 (d,  $J = 10.2$  Hz) and 5.80 (d,  $J = 10.2$  Hz). The signals at  $\delta$  6.37 (d,  $J = 2$  Hz) and 3.88 (s, 1H) are assigned to H-21 and H-24, respectively, as in other piscidinols. The signals at  $\delta$  4.04 (t,  $J = 3$  Hz) and 4.39 (m, 1H) are assigned to H-7 and H-11, respectively. The coupling constant value of 3 Hz for H-7 shows the  $\beta$ , equatorial orientation of H-7 and thus the  $\alpha$ , axial orientation of the OH-7 group. Irradiation of the H-11 signal at  $\delta$  4.39 collapsed the H-9 doublet at  $\delta$  2.38 ( $J = 8.8$  Hz) into a singlet. The coupling value of 8.8 Hz shows the axial orientation of H-11 and thus the OH-11 group is  $\alpha$ , equatorial. The signal at  $\delta$  3.92 is assigned to the proton on the carbon bearing the methoxy group.

The double bond proton at  $\delta$  5.93 which appears as a doublet ( $J = 2.9$  Hz) is assigned to H-15 showing that C-16 is substituted. Confirmation of the siting of the OMe at C-16 is obtained from the  $^{13}\text{C}$  NMR spectrum. The C-16 signal is shifted downfield ( $\Delta\delta$  8.2) and the C-17 signal shifted upfield ( $\Delta\delta$  2.1) [2], compared to piscidinol D which has a hydroxyl group at this position [1]. The signal for C-15 remains relatively unchanged.

On acetylation, piscidinol F formed a triacetate (1a). In the triacetate the H-7 and H-11 signals are shifted to  $\delta$  5.35 and 5.4 but there is no change in the positions of H-24 ( $\delta$  3.88) and H-16 ( $\delta$  3.92), hence there is no free hydroxyl at C-16 and the hydroxyl at C-24 is not acetylated. All these assignments were confirmed by irradiation experiments. The configuration at C-21 and C-24 is assigned from NOE studies. Irradiation of H-21 at  $\delta$  6.37 gave enhancements (NOE) of H-20 and H-22 $\alpha$  (6%) and of H-24 (2%). Since the normal tirucallane configuration is H-20 $\alpha$ , H-21 and H-24 can also be assigned  $\alpha$ -configurations, OAc-21 group and the OH-24 group are  $\beta$ . Thus piscidinol F is assigned the structure 1. That piscidinol F is not an artefact produced under chromatographic conditions was proved by demonstration of its presence in extracts obtained from leaves of *W. piscidia* with cold ethyl acetate and cold ethanol (TLC).

Confirmatory evidence for the side chain configuration of piscidinol B comes from the  $^1\text{H}$  NMR spectral data. In piscidinol B there is no coupling between H-23 and H-24 but in triacetate (2a) a coupling of 1.5 Hz is observed as in other related tirucallanes, i.e. bourjotinolone [3], sapelin F and its tetraacetate [4, 5]. These assignments are confirmed by  $^1\text{H}$ – $^1\text{H}$  COSY and double irradiation experiments. Irradiation of the H-24 signal (at  $\delta$  4.88 d, 1.5 Hz) in the acetate spectrum results in the removal of the 1.5 Hz coupling from the H-23 signal. Similarly, irradiation of the H-23 signal at  $\delta$  5.43 simplified the H-24 signal to a singlet. These data indicate the similar configurations for the vicinal diol as in bourjotinolone and sapelin F. In the original paper [1], the configuration of the side chain in piscidinol B was not assigned, but from

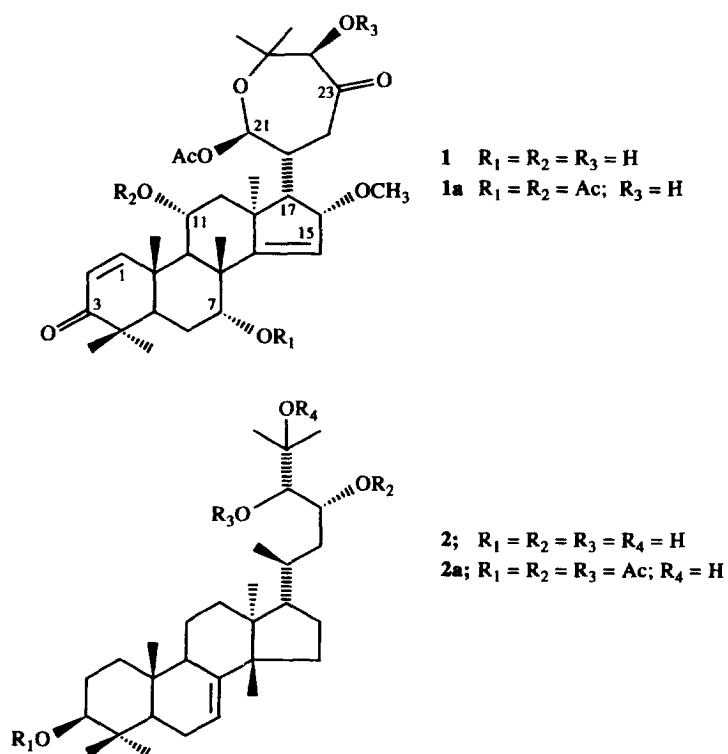


Table 1.  $^1H$ NMR (400 MHz) data for piscidinols and their derivatives ( $\delta$  in ppm, TMS as int. standard,  $J$  in Hz)

H	Piscidinol F (1) $CDCl_3$	Piscidinol B (2) (pyridine- $d_5$ )	Piscidinol B acetate (2a) $CDCl_3$
1	8.05 ( <i>d</i> (10.2)	—	—
2	5.80 ( <i>d</i> (10.2)	—	—
3	—	3.45 <i>m</i>	4.51 <i>dd</i> (4.4, 11.7)
5	2.50 <i>dd</i> (4.39, 11.71)	—	—
6	1.90 <i>m</i>	1.95 <i>m</i>	—
7	4.04 <i>t</i> (3)	5.29 <i>m</i>	5.22 <i>m</i>
9	2.38 <i>d</i> (8.8)	—	—
11	4.39 <i>m</i>	—	—
12 $\alpha$	1.75 <i>d</i> (13.3)	—	—
12 $\beta$	1.66 <i>dd</i> (5.5, 13.3)	—	—
15	5.93 <i>d</i> (2.9)	—	—
16	3.92 <i>dd</i> (3.4, 5.8)	—	—
17	1.55 <i>dd</i> (5.8, 11.23)	—	—
18-Me	1.17 <i>s</i>	0.79 <i>s</i>	0.80 <i>s</i>
19-Me	1.22 <i>s</i>	0.87 <i>s</i>	0.76 <i>s</i>
20	2.81 <i>m</i>	—	—
21	6.37 <i>d</i> (2.0)	1.62 <i>d</i> (8.3)	0.93 <i>d</i> (5.9)
22 $\alpha$	2.74 <i>dd</i> (5.8, 16.1)	2.35 <i>m</i>	—
22 $\beta$	2.45 <i>dd</i> (3.9, 16.1)	1.6 <i>m</i>	—
23	—	4.59 <i>m</i>	5.43 <i>ddd</i> (1.5, 5.0, 8.4)
24	3.88 <i>s</i>	3.67 <i>s</i>	4.88 <i>d</i> (1.5)
26-Me	1.25 <i>s</i>	1.61 <i>s</i>	1.19 <i>s</i>
27-Me	1.28 <i>s</i>	1.63 <i>s</i>	1.24 <i>s</i>
28-Me	1.09 <i>s</i>	1.15 <i>s</i>	0.97 <i>s</i>
29-Me	1.11 <i>s</i>	1.10 <i>s</i>	0.85 <i>s</i>
30-Me	1.44 <i>s</i>	0.99 <i>s</i>	0.94 <i>s</i>
OAc	2.17 <i>s</i>	—	2.05 <i>s</i> , 2.07 <i>s</i> , 2.19 <i>s</i>
OMe	3.23 <i>s</i>	—	—

Table 2.  $^{13}\text{C}$  NMR (100 MHz) data for compounds 1 and 2

C	Piscidinol F (1) ( $\text{CDCl}_3$ )	Piscidinol B (2) (pyridine- $d_5$ )
1	161.1 <i>d</i>	36.7 <i>t</i>
2	123.6 <i>d</i>	27.7 <i>t</i>
3	204.7 <i>s</i>	77.4 <i>d</i>
4	40.7 <i>s</i>	38.6 <i>s</i>
5	44.1 <i>d</i>	50.2 <i>d</i>
6	24.3 <i>t</i>	23.5 <i>t</i>
7	71.6 <i>d</i>	117.5 <i>d</i>
8	44.2 <i>s</i>	145.1 <i>s</i>
9	45.2 <i>d</i>	48.4 <i>d</i>
10	44.4 <i>s</i>	34.3 <i>s</i>
11	66.3 <i>d</i>	17.5 <i>t</i>
12	45.6 <i>t</i>	33.5 <i>t</i>
13	46.5 <i>s</i>	42.9 <i>s</i>
14	168.8 <i>s</i>	50.5 <i>s</i>
15	119.6 <i>d</i>	33.3 <i>t</i>
16	82.0 <i>d</i>	27.9 <i>t</i>
17	56.7 <i>d</i>	53.4 <i>d</i>
18	24.6 <i>q</i>	12.5 <i>q</i>
19	20.0 <i>q</i>	21.2 <i>q</i>
20	36.0 <i>d</i>	33.5 <i>d</i>
21	91.0 <i>d</i>	18.6 <i>q</i>
22	38.6 <i>t</i>	41.4 <i>t</i>
23	208.6 <i>s</i>	68.5 <i>d</i>
24	80.7 <i>d</i>	75.8 <i>d</i>
25	72.2 <i>d</i>	72.9 <i>s</i>
26	24.3 <i>q</i>	26.3 <i>q</i>
27	26.2 <i>q</i>	26.9 <i>q</i>
28	21.5 <i>q</i>	27.4 <i>q</i>
29	26.8 <i>q</i>	14.6 <i>q</i>
30	29.7 <i>q</i>	26.5 <i>q</i>
COMe	169.8 <i>s</i>	—
COMe	20.9 <i>q</i>	—
OMe	56.2 <i>q</i>	—

Multiplicities determined by means of off-resonance decoupled spectra.

the above data it appears that piscidinol B is identical with hispidol B isolated from *Trichilia hispida* [6].

#### EXPERIMENTAL

Leaves of *Walsura piscidia* were collected from Guindy National Park. Their identity was confirmed by Dr Narasimhan, Dept of Botany, Madras Christian College and a voucher specimen is deposited in the Herbarium of Madras Christian College. The leaves were dried, powdered (1.325 kg) and extracted exhaustively with *n*-hexane and cold EtOH. The EtOH extract, following removal of the solvent, was suspended in  $\text{H}_2\text{O}$  and extracted with EtOAc. The EtOAc layer was concd and chromatographed on a column of silica gel using hexane,  $\text{CHCl}_3$  and  $\text{CHCl}_3$ -MeOH mixtures as eluents. All the piscidinols were obtained in the  $\text{CHCl}_3$ -MeOH (19:1) fraction. This fr. was again chromatographed using  $\text{CHCl}_3$ -MeOH (49:1).

**Isolation of piscidinol B (1).** Frs 1–8 ( $\text{CHCl}_3$ -MeOH, 19:1) on further chromatography yielded piscidinol B (60 mg), which was recrystallized from MeOH. Mp 248–250°;  $[\alpha]_D - 59.6$  (MeOH; *c* 0.5); IR  $\nu_{\text{max}} \text{ cm}^{-1}$ : 3362, 2951, 1457, 1364, 1160, 1033, 798  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR: Tables 1 and 2; EIMS *m/z* (rel. int.): 476  $[\text{M}]^+$  (6), 461 (2), 443 (11), 425 (18), 371 (50), 353 (34), 327 (15), 309 (11), 257 (7).

**Acetylation of piscidinol B.** A sample (15 mg) of 1 was treated with  $\text{Ac}_2\text{O}$  (1 ml) and pyridine (1 ml) and kept at room temp. for 24 hr. Usual work-up gave 12 mg of acetate (ex. MeOH). Mp 143°, IR  $\nu_{\text{max}} \text{ cm}^{-1}$ : 3600, 1728, 1380, 1229, 908;  $^1\text{H}$  NMR: Table 1.

**Isolation of piscidinol C.** Frs. 9–13, on further chromatography, gave piscidinol C (350 mg), which was recrystallized from EtOAc-hexane (2:3), mp 230°.

**Isolation of piscidinol F.** The mother liquors, following removal of piscidinol C, were subjected to prep. HPLC (Shimadzu ODS column 20 mm  $\times$  25 cm, 220 nm, 10 ml  $\text{min}^{-1}$ ) yielding piscidinol F (*R*<sub>f</sub> 33.4 min) (200 mg) which was recrystallized from EtOAc-hexane. Mp 220°; IR  $\nu_{\text{max}} \text{ cm}^{-1}$ : 3468, 1749, 1715, 1655, 1558, 1457, 1386, 1223, 1083 1016, 951, ;  $[\alpha]_D - 62.8$  ( $\text{CHCl}_3$ ; *c* 1); UV  $\lambda_{\text{max}}$  (EtOH) nm: 234 ( $\epsilon$  109.65), 213 ( $\epsilon$  134.75); Found: C, 66.89%; H, 8.04.  $\text{C}_{33}\text{H}_{48}\text{O}_9$  requires C. 67.3, and H, 8.16%. MS (FAB) *m/z*: 589  $[\text{M} + \text{H}]^+$  (100%).

**Acetylation of piscidinol F.** Under standard conditions piscidinol F (50 mg sample,  $\text{Ac}_2\text{O}$ -pyridine room temp. for 16 hr), gave on usual work-up of the reaction mixture a product which on CC gave two compounds; acetate 1 (17 mg) and acetate 2 (6 mg), which was not characterized further. Acetate 1 had mp 182–184° (ex. MeOH); IR  $\nu_{\text{max}} \text{ cm}^{-1}$  3618, 1730, 1668, 1379, 1223, 1084;  $^1\text{H}$  NMR of acetate 1 (400 MHz,  $\text{CDCl}_3$ );  $\delta$  1.06, 1.1, 1.21 (6H), 1.25, 1.26 1.4 (7-Me), 2.15, 2.13, 1.98 (3-OAc) 3.15 (s, 3H, OMe), 5.35 (t, H-7), 5.4 (m, H-11), 5.78 (d, *J* = 3 Hz), 5.82 (d, *J* = 11 Hz), 7.15 (d, *J* = 11 Hz), 3.88 (s, H-24)), 3.90 (m, H-16), 6.25 (d, *J* = 2 Hz).

**Isolation of piscidinol E.** Frs 14–16 on prep. TLC ( $\text{CHCl}_3$ -MeOH, 19:1) gave piscidinol E (200 mg) as a powder, mp 175–178°.

**Isolation of piscidinol D.** Frs. 17–20, on centrifugal chromatography (chromatotron) using  $\text{CHCl}_3$ -MeOH (97:3), gave piscidinol D (185 mg), mp 192°.

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#### REFERENCES

1. Purushothaman, K. K., Duraiswamy, K., Connolly, J. D. and Rycroft, D. S. (1985) *Phytochemistry* **24**, 2349.

2. Stothers (1972) *Carbon 13 NMR Spectroscopy* (Blomquist, A. T. and Wasserman, H. eds), Academic Press, New York.
3. Breen, G. J. W., Ritchie, E., Sidwell, W. T. L. and Taylor, W. C. (1966) *Australian J. Chem.* **19**, 455.
4. Chan, W. R., Taylor, D. R. and Yee, T. H. (1971) *J. Chem. Soc. C*, 2622.
5. Lyons, C. W. and Taylor, D. R. (1975) *J. Chem. Soc. Chem. Commun.* 517.
6. Jolad, S. D., Hoffmann, J. J., Schram, K. H. and Cole, J. R. (1981) *J. Org. Chem.* **46**, 4085.