

A LUPENE-TYPE TRITERPENE FROM MIMUSOPS ELENGI

NUSRAT JAHAN, WASIM AHMED and ABDUL MALIK*

H.E.J. Research Institute of Chemistry, University of Karachi, Karachi 75 270, Pakistan

(Received 17 June 1994)

Key Word Index—*Mimusops elengi*; Sapotaceae; triterpene; 3β -hydroxy-lup-20(29)-ene-23,28-dioic acid.

Abstract—A new triterpene 3β -hydroxy-lup-20(29)-ene-23,28-dioic acid has been isolated from *Mimusops elengi*. Its structure was established through chemical and spectroscopic studies. The known triterpenes, β -amyrin, lupeol, α -taraxerol and ursolic acid were also isolated.

INTRODUCTION

Mimusops elengi grows wild in southern India, Burma and Pakistan. Various parts of the plant are used in the indigenous system of medicine as a febrifuge, astringent, purgative and stimulant [1]. Although the presence of saponins, steroids, terpenoids, and alkaloids have been reported from M. elengi [2–5], but very few individual constituents have so far been isolated or characterized [5a]. We now report on the isolation and characterization of a new pentacyclic triterpene (1) of the lupene series from this plant. The known triterpenes β -amyrin, lupeol, α -taraxerol and ursolic acid have also been isolated from this species for the first time.

RESULTS AND DISCUSSION

Compound 1 was assigned the molecular formula $C_{30}H_{46}O_5$ by HRMS ([M]⁺ at m/z 486.3307; calc. 486.3345). It gave positive Liebermann–Burchard and CeSO₄ tests for triterpenes and also a positive NaHCO₃ test for a carboxyl group(s). The IR spectrum showed the absorption for a hydroxy group (3400 cm⁻¹), a carboxyl group (2730 and 1700 cm⁻¹) and a disubstituted double bond (3075 and 1640 cm⁻¹). The formation of the dimethyl ester 1a and a monoacetyl derivative (1b) indicated the presence of two carboxylic functions and one hydroxyl. Oxidation of 1a with PCC in methylene chloride gave the keto diester 1c, confirming the secondary nature of the hydroxyl group.

The ¹H NMR spectrum of 1 showed two olefinic protons at $\delta 4.58$ and 4.71, a one-proton double proton at $\delta 3.9$ (J = 5.1, 10.7 Hz) and five methyl groups on quater-

1 $R^1 = \beta$ -OH; α -H; $R^2 = H$ 1a $R^1 = \beta$ -OH; α -H; $R^2 = Me$ 1b $R^1 = \beta$ -OAc; α -H; $R^2 = H$ 1c $R^1 = O$; $R^2 = Me$

nary carbons (δ : 0.79, 0.86, 0.98, 1.30 and 1.68). The ¹³C NMR spectrum (BB and DEPT) of 1 corroborated the presence of five methyls, nine methylenes, seven methines and five quaternary carbons, in addition to carboxylic groups (δ 178.7 and 177.3) and a 1,1-disubstituted double bond (δ 49.8 and δ 150.0).

In the HR-mass spectrum ions at m/z 250.1549 ($C_{15}H_{22}O_3$), 248.1771 ($C_{16}H_{24}O_2$), 231.1472 ($C_{14}H_{21}O_3$), 234.1602 ($C_{15}H_{22}O_2$) and 219.1373 ($C_{14}H_{19}O_2$) were characteristic of a lup-20(29)-ene skeleton with a carboxy group in ring D/E and the hydroxyl and remaining carboxylic function in ring A/B [6, 7]. The ready loss of one of the carboxyl groups from the molecular ion peak gave a fragment at m/z 441.3327 ($C_{29}H_{45}O_3$) allowing assignment to C-17. This was supported by the chemical shifts of all the carbon atoms of rings D and E in the ^{13}C NMR spectrum, which showed complete agreement to betulinic acid [6, 8].

^{*}Author to whom correspondence should be addressed.

Short Reports

In the ¹H-¹H one bond COSY spectrum of 1 the carbinol methine proton at $\delta 3.9$ showed a cross-peak with two other protons thus placing it in ring A or on C-7. It was assigned to C-3 on biogenetic grounds. This was supported by the characteristic chemical shifts of C-1 and C-2 in the ¹³C NMR spectrum. The large coupling constant allowed us to assign a β and equatorial configuration to the hydroxy group. This was also confirmed by NaBH₄ reduction of the keto diester 1c back to the dimethyl ester 1a. Evidence as to the position of the carboxylic group in ring A/B was provided by the mass spectrum of the keto diester 1c. This showed an intense peak at m/z 169.0847 ($C_9H_{13}O_3$) which is also found in the mass spectrum of 3-oxo-allobetulane [9] and thus placed the carbonyl group and the carboxylic function at positions C-3 and C-4, respectively.

The key to the configuration of the isopropenyl group at C-19 and that of the carboxylic group at C-4 was provided by the physical data of the keto diester 1c which showed complete agreement to those reported in the literature for the dimethyl ester of 3-oxo-lup-20(29)-ene-23,28-dioic acid [7]. Thus compound 1 is 3β -hydroxylup-20(29)-ene-23,28-dioic acid. In the ¹H-¹³C longrange COSY spectrum of 1 the carbonyl carbon of the carboxylic group at δ 178.70 showed a cross-peak to the protons of the methyl group at C-24, thus providing conclusive evidence for the assigned structure. Compound 1 is epimeric at C-3 with a triterpene isolated from Scheffera octophylla [7]. The diester obtained from the latter was reported to show two bands in the IR spectrum, one at 3630 cm⁻¹ for the free hydroxyl group and one at 3510 cm⁻¹ due to hydrogen bonding between OH and the ester group at C-23. No such hydrogen bonding is possible in 1a due to the trans disposition of these groups, hence only a single band for a free hydroxyl group was observed at 3620 cm⁻¹.

EXPERIMENTAL

General. Mps: uncorr.; IR: CHCl₃; ¹H NMR (400 MHz) and ¹³C NMR (100 MHz): CDCl₃ using TMS as int. standard; HRMS: 70 eV; TLC: silica gel, PF₂₅₄; CC: silica gel, 70-230 mesh. The DEPT, NOE and HMQC experiments were performed as reported earlier [10, 11].

Plant material. The plant material was collected from the Karachi region and was identified as Mimusops elengi by Prof. M. Qaiser, Department of Botany, University of Karachi. A voucher specimen is deposited in the herbarium of the Department of Botany, University of Karachi.

Isolation. The shade-dried plant material (70 kg) was extracted (\times 4) with MeOH at room temp. The residue from the methanolic extract was partitioned between hexane and H₂O. The hexane-soluble fr. was chromatographed over silica gel using various mixtures of hexane. CHCl₃ and MeOH.

3β-Hydroxy-lup-20(29)-ene-23,28-dioic acid (1). The fr. eluted in hexane-CHCl₃ (1:3) for CC contained only one major compounds on TIC. It was further purified by prep. TLC using hexane-EtOAc (3:2) and then crystal-

lized from Me₂CO-n-hexane (30 mg); mp: 262-264°; $[\alpha]_D^{25} + 13^\circ$ (CHCl₃; c 0.52); IR $\nu_{max}^{CHCl_3}$ cm⁻¹; 3400, 3075, 2730, 1700, 1640; MS m/z (rel. int.): 486.3307 [M]⁺ (18) $(C_{30}H_{46}O_5)$, 471.3088 (7) $(C_{29}H_{43}O_5)$, 468.3188 (22) $(C_{30}H_{44}O_4)$, 441.3327 (19) $(C_{29}H_{45}O_3)$, 250.1549 (53) $(C_{15}H_{22}O_3)$, 248.1771 (71) $(C_{16}H_{24}O_2)$, 237.1472 (44), $(C_{14}H_{21}O_3)$, 234.1602 (75), $(C_{15}H_{22}O_2)$, 219.1373 (78) $(C_{14}H_{19}O_2)$; ¹H NMR (CDCl₃): δ 0.79, 0.86, 0.98, 1.30, 1.68 (3H, each s, Me), 3.9 (dd, J = 5.1 and 10.7 Hz, H-3), 4.58 and 4.71 (1H each, m H₂-29); ¹³C NMR (CDCl₃, 100 MHz): δ39.0 (C-1), 29.4 (C-2), 84.4 (C-3), 43.0 (C-4), 44.2 (C-5), 18.3 (C-6), 33.9 (C-7), 41.5 (C-8), 49.6 (C-9), 37.0 (C-10), 23.4 (C-11), 25.3 (C-12), 38.5 (C-13), 42.8 (C-14), 30.5 (C-15), 32.2 (C-16), 56.1 (C.17), 46.8 (C-18), 49.8 (C-19), 150.0 (C-20), 29.7 (C-21), 37.0 (C-22), 178.7 (C-23), 18.1 (C-24), 18.4 (C-25), 16.1 (C-26), 14.4 (C-27), 177.3 (C-28), 109.4 (C-29), 19.0 (C-30). These assignments were made by comparison with published 13C NMR data of related compounds [6-8], and confirmed in each case by ¹H-¹³C correlated spectroscopy (HMQC).

Dimethyl ester 1a. Obtained from 1 by treatment with CH₂N₂ in MeOH. Amorphous; $[\alpha]_{D}^{25} + 14^{\circ}$ (CHCl₃, c 0.19); IR $v_{max}^{CHCl_3}$ cm⁻¹: 3450, 3070, 1730, 1640; MS m/z (rel. int.): 514 [M]⁺ (23), 499 (8), 496 (8), 496 (22), 454 (21), 264 (64), 262 (76), 251 (60), 248 (76), 233 (80).

Acetate 1b. Obtained from 1 by treatment with Ac_2O -pyridine for 20 hr at 25° ; mp $209-212^\circ$ (Me_2CO -n-hexane); $[\alpha]_D^{25} + 20^\circ$ (CHCl₃; c 0.17); IR $v_{\max}^{\text{CHCl}_3}$ cm⁻¹: 3070, 1730, 1700, 1640, 1240; MS m/z (rel. int.): 528 [M]⁺ (10), 513 (6), 482 (11), 486 (16), 292 (55), 248 (69), 279 (46), 234 (76), 219 (80).

Oxidation of 1a to keto diester 1c. To PCC (67 mg) in dry CH_2Cl_2 (5 ml), compound 1a (58 mg) was added and the soln stirred at 20° for 4 hr. Standard work-up followed by CC over silica gel and elution with *n*-hexane-CHCl₃ (4:1) gave keto ester 1c (34 mg); mp $131-132^{\circ}$ (*n*-hexane); $[\alpha]_D^{25} + 6^{\circ}$ (CHCl₃; c 0.2). The MS, IR and ¹H NMR data were in good agreement with those reported in ref. [7].

Reduction of 1c to 1a. To a soln of 1c (10 mg) in 1 ml MeOH-THF (1:1) was added NaBH₄ (10 mg). The mixt. was stirred at 20° for 1 hr. Standard work-up followed by CC over silica gel (elution with n-hexane-CHCl₃, 7:3) provided an amorphous compound having the same R_f value and physical constants as 1a.

Compound 2. The fr. eluted in hexane–CHCl₃ (7:3) from CC showed one major spot and was crystallized from EtOH, mp: 197–198°; $[\alpha]_D^{25} + 99^\circ$ (CHCl₃); MS m/z (rel. int.): 426 [M]⁺ (15). The physical and spectral data identified it as β -amyrin [12].

Compound 3. The eluent obtained from hexane–CHCl₃ (13:7) showed one major spot and was further purified on silica gel CC using hexane–CHCl₃ (3:2) as solvent system and crystallized from MeOH, mp 214–14°, $[\alpha]_{\rm b}^{25} + 27^{\circ}$. MS m/z (rel. int.): 426 [M]⁺ (20). The physical and spectral data identified it as lupeol [8].

Compound 4. The fr. eluted with hexane-CHCl₃ (1:1) was subjected to prep. TLC using as hexane-CHCl₃ (3:2) solvent system. It crystallized from benzene as needles, mp $271-272^{\circ}$; $\lceil \alpha \rceil_D^{25} - 9.9^{\circ}$. MS m/z (rel. int.): 426 [M]⁺

Short Reports 257

(18). It was identified as α -taraxerol on the basis of physical and spectral data [13].

Compound 5. The material in the hexane–CHCl₃ (3:7) eluate was further subjected to prep. TLC using the solvent system CHCl₃–MeOH (49:1) and the major spot crystallized from EtOH, mp: $283-285^{\circ}$; $[\alpha]_D^{21} + 62.5-68^{\circ}$ (MeOH). The physical and spectral data identified it as ursolic acid [14].

REFERENCES

- 1. Nadkarni, K. M. (1976) in *Indian Materia Medica*, Vol. 1, p. 801. Popular Prakashan Pvt. Ltd, Bombay.
- 2. Misra, G. and Mitra, C. R. (1967) Phytochemistry 6, 1309
- Gupta, G. K., Dhar, K. L. and Atal. C. K. (1976) Indian J. Chem, 14B, 818.
- 4. Misra, G. and Mitra, C. R. (1968) *Phytochemistry* 7, 501.
- Misra, G. and Mitra, C. R. (1967) Phytochemistry 6, 453.

5a. Misra, G., Nigam, S. K. and Mitra, C. R. (1974) *Planta Med.* **24**, 15.

- Lischewski, M., Ty, Ph.D., Schmidt, J., Preiss, A., Phiet, H. V. and Adam, J. (1984) *Phytochemistry* 23, 1695.
- Adam, G., Lischewski, M., Phiet, H. V., Preiss, A., Schmidt, J. and Sung, T. V. (1982) *Phytochemistry* 21, 1385.
- 8. Sholichin, M., Yamasaki, K., Kasai, R. and Tanaka, O. (1980) Chem. Pharm. Bull. 28, 1006.
- 9. Schmidt, J. and Huneck, S. (1979) Org. Mass Spectrom. 14, 646.
- Rasool, N., Khan, A. Q. and Malik, A. (1989) *Phytochemistry* 28, 193.
- Ahmed, W., Khan, A. Q. and Malik, A. (1992) J. Nat. Prod. 55, 1764.
- 12. Knight, S. A. (1974) Org. Magn. Reason. 6, 603.
- Hui, W. H. and Sung, M. L. (1968) Aust. J. Chem. 21, 2137.
- 14. Seo, S., Tomita, Y. and Tori, K. (1975) J. Chem. Soc. Chem. Commun 270.