



ISOCRYPTOLEPINE FROM *CRYPTOLEPIS SANGUINOLENTA*

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Key Word Index—*Cryptolepis sanguinolenta*; Asclepiadiaceae; indoloquinoline; alkaloid.

Abstract—A new alkaloid, 5-methyl, 5H-indolo-[3,2-c]-quinoline named isocryptolepine, has been isolated from the roots of *Cryptolepis sanguinolenta* and its structure determined from spectroscopic data, including IR, MS and NMR.

INTRODUCTION

Cryptolepis sanguinolenta (Lindl) Schlechter (Asclepiadiaceae), a shrub indigenous to tropical West Africa, has long been employed in the dyeing of textiles and leather [1]; it is also used in folk medicine as an antimalarial agent [2] and such bioactivities as antibacterial activity or action on vasodilation have been reported [3].

The first alkaloid isolated from *C. sanguinolenta* in 1951 by Gellert *et al.* [4] was cryptolepine (5-methyl, 5H-indolo-[3,2-b]-quinoline), previously extracted from *C. triangularis*. Later, the same alkaloid was re-isolated from *C. sanguinolenta* by Dwuma-Badu *et al.* [5], along with quindoline (norcryptolepine) and a non-characterized alkaloid (CSA-3). Recently, the spirononacyclic alkaloid cryptospirolepine was isolated from the same plant [3].

This paper reports the isolation of a new alkaloid **1** (5-methyl, 5H-indolo-[3,2-c]-quinoline) which is an isomer of cryptolepine **2**, and thus termed isocryptolepine.

RESULTS AND DISCUSSION

The EtOH extract of the defatted powdered roots of *C. sanguinolenta* was treated with aqueous HOAc, filtered, alkalized with NH_4OH , and extracted repeatedly with CH_2Cl_2 . The combined CH_2Cl_2 extracts were chromatographed over Al_2O_3 eluted with a CH_2Cl_2 -MeOH gradient. Elution with CH_2Cl_2 gave a mixture of quindoline and isocryptolepine, and elution with CH_2Cl_2 -MeOH (9:1) afforded cryptolepine. Quindoline and isocryptolepine (**1**) were separated by preparative silica gel thin-layer chromatography.

The EI mass spectrum of **1** showed the molecular ion at m/z 232 in agreement with the molecular formula $\text{C}_{16}\text{H}_{12}\text{N}_2$. The 16 carbon atoms were depicted in the ^{13}C NMR spectrum and distributed into 15 aromatic carbon atoms (9 CH and 6 quaternary sp^2 carbons) and one methyl group (Table 1). The ^1H NMR spectrum

displayed resonances for 12 protons forming two four-spin systems assigned to two *ortho*-disubstituted benzene rings and a singlet at δ 9.29. These spin systems were the same as those previously described for cryptolepine (**2**), but the chemical shifts were different, especially the aromatic singlet at δ 8.91 in the spectrum of cryptolepine was shifted to lower field.

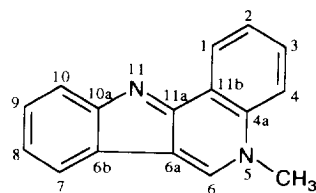
Analysis of the ^1H - ^1H COSY, ^{13}C - ^1H COSY and HMBC spectra allowed the assignment of one benzene ring in an indole substructure and the second in a quinoline substructure, as in the structure of cryptolepine. However, the assembling of the substructures was different for isocryptolepine, and was suggested to be as shown in structure **1**, due to the following long-range correlations observed in the HMBC. The quaternary carbon at δ 152.1 (11a) was 3J correlated to the proton at δ 8.77 (H-1); the singlet proton at δ 9.29 was 3J correlated to carbon atoms at δ 152.1 (C-11a), 137.5 (C-4a), 43.5 (*N*-Me) and 2J to that of δ 117.6 (C-6a), whereas the carbon atoms at 140.5 (C-6) and 137.5 (C-4a), gave strong correlations with the *N*-methyl protons. These data agreed with the position of the singlet methine α to the *N*-methyl group. The results led to structure **1** for isocryptolepine, which has thus an angular structure isomeric with cryptolepine for which the four rings are linear. Isocryptolepine could be the partially characterized alkaloid CSA-3 [5].

EXPERIMENTAL

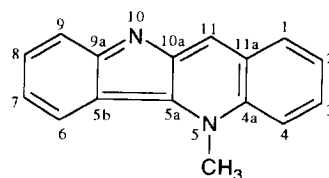
General. ^1H 300.13 MHz and ^{13}C 75.47 MHz NMR spectra were performed on an AC 300 Bruker spectrometer. The HMBC spectrum was optimized for $J_{\text{C-H}}$ 10 Hz. EIMS: obtained with a Nermag Sidar V 3.0 mass spectrometer.

Isolation. The roots of *Cryptolepis sanguinolenta* (Lindl) Schlechter (60 g) were air-dried, and ground to provide a fine powder. The powder was defatted with hexane for 12 hr and then extracted by percolation with EtOH (500 ml). After evapn of the solvent, the residue was

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1



2

Table 1. ^{13}C and ^1H NMR data for isocryptolepine (1) (CD_3OD)

C	1 δ_{C}	1 δ_{H} m J (Hz)	HMBC C correlated to H
1	125.4	8.77 dd 8.1, 1.4	3
2	127.5	7.78 ddd 8.4, 8.1, 0.9	4
3	131.7	7.92 ddd 8.4, 8.1, 1.4	1
4	118.8	8.12 brd 8.1	2
4a	137.5	—	1, 3, 6, N-Me
6	140.5	9.29 s	N-Me
6a	117.6	—	6, 7
6b	125.7	—	8, 10
7	120.9	8.15 dd 8.1, 1.0	9
8	122.5	7.35 ddd 8.2, 8.2, 0.8	10
9	127.8	7.53 ddd 8.1, 8.2, 1.0	7
10	117.6	7.81 dd 8.1, 0.8	8
10a	152.0	—	7, 9
11a	152.1	—	1, 6
11b	121.6	—	2, 4
N-Me	43.5	4.36 s	6

mixed with aq. HOAc (10%, 200 ml), and allowed to stand overnight. The mixt. was filtered, alkalized with NH_4OH to pH 10, and extracted by CH_2Cl_2 (2×200 ml). The combined CH_2Cl_2 extracts were washed with H_2O , dried over Na_2SO_4 , filtered and evapd to afford a dark alkaloidal residue (450 mg). This residue was dissolved in MeOH (2 ml) and chromatographed over Al_2O_3 (30 g). Elution with hexane, and then CH_2Cl_2 afforded a mixt. of quindoline and isocryptolepine (40 mg). Elution with CH_2Cl_2 -MeOH (9:1) gave a violet residue of cryptolepine (400 mg). The mixt. of quindoline and isocryptolepine was further purified by prep. TLC (Kieselgel Merck 60) using toluene- Me_2CO - NH_4OH (50:50:2) as eluent yielding isocryptolepine (1, 5 mg, 0.08%, R_f 0.45) and quindoline (5 mg, R_f 0.80), which was identified by comparison with an authentic sample.

Isocryptolepine (1). $\text{C}_{16}\text{H}_{12}\text{N}_2$; (not crystallized); UV (EtOH) λ_{max} nm (log ϵ): 238 (4.18), 284 (4.26), 352 sh (3.64); IR (KBr) ν_{max} cm^{-1} : 1644, 1617, 1460, 1354, 1242, 1123, 762, 715, 617; EIMS (m/z) 232 [M] $^+$ (100%), 217 (16), 204 (4), 190 (10), 116 (12), 101 (5), 89 (6).

Cryptolepine (2). $\text{C}_{16}\text{H}_{12}\text{N}_2$; mp 167–168°, lit. 166–169° [4]; IR (KBr) ν_{max} cm^{-1} : 1644, 1615, 1466, 1354, 1308, 1242, 1031, 775, 715, 597; EIMS (m/z) 232 [M] $^+$ (100%), 217 (32), 190 (15), 116 (16), 102 (11), 94 (15), 89 (35). ^1H NMR (CD_3OD) δ ppm: 8.91 (s, H-11), 8.54 (d, 8.5, H-4), 8.50 (d, 9.1, H-6), 8.34 (dd, 7.8, 1.2, H-1), 8.05 (ddd, 8.5, 7.3, 1.2, H-3), 7.80 (dd, 7.8, 7.3, H-2), 7.74 (dd, 8.4, 1.3, H-9), 7.69 (dd, 8.4, 7.6, H-8), 7.34 (ddd, 9.1, 7.6, 1.3, H-7), 4.95 (s, N-Me).

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REFERENCES

1. Saxton, J. E. (1965) in *The Alkaloids, Chemistry and Physiology* (Manske, R. H. F. and Holmes, H. L., eds), Vol. 8, p. 19. Academic Press, New York.
2. Boye, G. L. and Ampofo, O. (1983) *Proceedings at the First International Symposium on Cryptolepine*, Kumasi, Ghana, University of Science and Technology.
3. Tackie, A. N., Boye, G. L., Sharaf, M. H. M., Schiff Jr, P. L., Crouch, R. C., Spitzer, T. D., Johnson, R. L., Dunn, J., Minick, D. and Martin, G. E. (1993) *J. Nat. Prod.* **56**, 653.
4. Gellert, E., Raymond-Hamet and Schlittler, E. (1951) *Helv. Chim. Acta* **34**, 642.
5. Dwuma-Badu, D., Ayim, J. S. K., Fiagbe, N. I. Y., Knapp, J. E., Schiff Jr, P. L. and Slatkin, D. J. (1978) *J. Pharm. Sci.* **67**, 433.