



BENZYLISOQUINOLINE ALKALOIDS AND FLAVONOLS FROM OCOTEA VELLOSIANA

WALMIR S. GARCEZ,† MASSAYOSHI YOSHIDA and OTTO R. GOTTLIEB‡

Instituto de Química, Universidade de São Paulo, 05508-900 São Paulo, SP, Brazil

(Received 25 October 1994)

Key Word Index—Ocotea vellosiana; Lauraceae; benzylisoquinoline alkaloids; flavonol glycosides; acylated flavonol glycosides; thalictoside; asparagine.

Abstract—Unripe fruits of *Ocotea vellosiana* were found to contain 13 benzylisoquinoline alkaloids, four flavonol glycosides, thalictoside, *p*-hydroxybenzoyl-rutinoside and asparagine, besides three novel *p*-coumaroyl derivatives of afzelin.

INTRODUCTION

Species of Ocotea are found throughout tropical America. Chemically they appear to fall into two groups characterized by the predominance either of neolignans [2] or of benzylisoquinoline alkaloids [1]. Ocotea vellosiana (Meissn.) Mez, collected near Campo Grande, Mato Grosso do Sul, belongs to the latter group.

RESULTS AND DISCUSSION

(+)-Dicentrine (1) [3, D-00796] was found in wood, fruit and leaves. nor-Dicentrine (2) [3, D-00796], predicentrine (3) [3, P-01740], ocoteine (4) [3, O-00081], Omethylcassifoline (5) [3, C-00527], leucoxylonine (6) [3, L-00499], ocotominarine (7) [3, O-00082], and the benzylisoquinoline alkaloid (\pm)-reticuline (8) [3, R-00142] were additionally isolated from wood. In contrast, fruits were shown to contain in addition to dicentrine, glaucine (9) [3, G-00398], corydine (10) [3, C-01881], and (+)isocorydine (11) [3, I-00368], whereas in leaves ocopodine (12) [3, L-00497] and ocominarine (13) [3, O-00078] were additionally located. While all these compounds are known and were identified by detailed spectroscopic evidence, their distribution in the plant organs is of interest. Only 1,2,9,10-tetraoxygenated benzylisoquinoline alkaloids (represented by 1-3 and 9) seem to be widely distributed in the plant. However, 1,2,10,11-tetraoxygenated derivatives (10, 11) are limited to fruit;

1,2,8,9,10-pentaoxygenated derivatives (12, 13) are limited to leaves; and 1,2,3,9,10-pentaoxygenated as well as 1,2,3,8,9,10-hexaoxygenated derivatives (respectively 4, 5 and 6, 7) are limited to wood.

In addition to alkaloids, the amino acid asparagine (14) [3, A-02880] was isolated from fruit and the flavonol glycosides afzelin (15) [3, A-00540], astragalin (Kaempferol 3-glucoside) (16) [3, G-00470], quercitrin (quercetin 3-rhamnoside) (17) [3, Q-00019] and hirsutrin (myricetin 3-glucoside) (18) [3, H-00908] were isolated from leaves. Three further heterosides (19-21), upon methanolysis, yielded afzelin (kaempferol-3-α-L-rhamnoside) and the methyl ester of p-coumaric acid [3, H-02875]. The locations of the p-coumaroyl moieties were established by ¹H NMR spectral comparisons (Table 1). The rhamnose proton signals were easily assigned by their characteristic multiplicities. The H-4" triplets (J = ca10 Hz) in the spectra of 19-21 appeared at relatively low field with respect to the corresponding signals of 15. Hence all three compounds are esterified at this position.

K = 3-kaempferyl, C = p-coumaroyl

19 $R^1 = R^2 = H$

20 $R^1 = H$, $R^2 = C$

21 $R^2 = H$, $R^1 = C$

Part 105 in the series 'The Chemistry of Brazilian Lauraceae'. For Part 104 see ref. [1]. Based on the Doctorate thesis presented by W.S.G. to Universidade de São Paulo (1991).

[†]Present address: Departamento de Química, Universidade Federal do Mato Grosso do Sul, Campo Grande, MS Brazil.

[‡]Departmento de Fisiologia e Farmacodinâmica, Instituto Oswaldo Cruz, FIOCRUZ, 21045-900, Rio de Janeiro, RJ, Brazil.

Table 1. 1 H NMR data [δ , mult. (J in Hz)] for rhamnose units of heterosides (Me₂CO- d_6 , 200 MHz)

H	15	19	20	21	
 1"	5.47	5.64	5.74	5.82	
	d(1.4)	br s	br s	br s	
?"	4.21	4.28	4.53	5.62	
	m	m	br s	br s	
3"		3.92	5.42	4.20	
		dd (3.4, 9.7)	br d (10.5)	m	
"	3.2 - 3.8	4.97	5.33 t	5.00	
		t (9.7)	t (10.5)	t (9.7)	
"		3.2-3.6	3.51	3.96	
		m	m	m	
<i>"</i>	0.86	0.79	0.88	0.87	
	d(6.0)	d (6.2)	d (6.0)	d(6.0)	

Table 2. ¹³C data (δ) , for glucose (Glc) and rhamnose (Rha) units of *p*-hydroxybenzoyl- β -D-rutinoside (23) and kaempferol-3-rutinoside (24)

Glc	C	23*	24 [4]†	Rha	C	23*	24 [4]†
	1	101.4	100.6		2	74.5	74.2
	2	70.8	70.3		3	76.8	76.5
	3	71.2	70.7		4	70.0	70.1
	4	72.6	72.0		5	75.6	75.8
	5	68.9	68.1		6	67.6	66.9
	6	17.4	17.4				

^{*} $Me_2CO-d_6-D_2O$, 50 MHz.

Compounds 20 and 21 showed additional low field signals for H-3" (br s, J = 10.5 Hz) and H-2" (br s), respectively. Thus 19, 20 and 21 are 4"-p-coumaroy!, 3", 4"-di-p-coumaroy! and 2", 4"-di-p-coumaroy! afzelins, three seemingly new compounds.

Finally, leaves afforded two further heterosides, thalictoside (22) [3, N-00491] and the novel p-hydroxybenzoyl- β -D-rutinoside (23). Methanolysis of 23 gave the methyl ester of p-hydroxybenzoic acid [3, H-01256] and rutinose [3, R-00495]. The 13 C NMR spectra of 23 and kaempferol-3-rutinoside [4] were practically superimposable with respect to the glycosidic signals of the molecules (Table 2). Hence the aglycones of both com-

pounds must be located at the same carbon, i.e. C-1 of the glucose moieties.

EXPERIMENTAL

Unripe fruit (without calyxes), leaves and wood (from a branch measuring 10 cm in diameter) were collected from a tree (height ca 6 m) growing on sandy soil in a deforested region near Campo Grande, MS, Brasil. The fruits were crushed and percolated with EtOH. The extracts were washed with hexane and partitioned between CHCl₃ and EtOH-H₂O. The latter solution, after addition of Me₂CO and cooling, pptd crystalline 11. After filtration it was extracted with dilute (2%) HOAc. Separation of alkaloids from the aqueous phase by the usual process and fractionation of the crude mixture (CC, alumina, hexane-AcOEt gradient) gave 11, 1, 9 and 10.

Wood, treated in the same way, gave 4, 1, 2, 6, 7, 3, 5 and 8.

Leaves were percolated with EtOH. An EtOH-H₂O solution of the extract was washed successively with hexane, CHCl₃ and EtOAc. The CHCl₃ soln was evapd. Fractionation of the residue (CC, silica, CH₂Cl₂-EtOH, 19:1 and 9:1) gave two fractions. The first one (by CC, alumina, C₆H₆-EtOAc, 9:1) gave 12, 13 and 1. The second fraction (by CC and TLC, silica H, MeOH-H₂O-HCO₂H gradient) gave 19, 20, 21, 15, 16 and 1. The EtOAc soln was evapd. Fractionation of the residue (CC, Sephadex LH-20, MeOH) gave 22 and 23 besides a mixture. This was separated (CC and TLC, silica, EtOAc-H₂O-MeOH gradient) into 15-18.

Acknowledgements—Fellowships and financial support were provided by CAPES (to W.S.G.), CNPq (to M.Y. and O.R.G.), PADCT and FAPESP.

REFERENCES

- Chavez, J. P., Gottlieb, O. R. and Yoshida, M. (1995) Phytochemistry 39, 849.
- 2 Botea, C., Pagliosa, F. M., Bolzani, V. da S., Yoshida, M. and Gottlieb, O. R. (1993) Phytochemistry 32, 1331.
- Buckingham, J., Macdonald, F. M. and Bradley, H. M. (eds) (1994) Dictionary of Natural Products. Chapman & Hall, London.
- Markham, K. R., Chari, V. M. and Mabry, T. J. (1982) in *The Flavonoids: Advances in Research* (Harborne, J. B. and Mabry, T. J., eds), p. 40. Chapman & Hall, London.

[†]DMSO-d₆, 20 MHz.