



STILBENE GLYCOSIDES FROM GUIBOURTIA TESSMANNII

A. M. NYEMBA, T. NGANDO MPONDO, S. F. KIMBU and J. D. CONNOLLY*

Département de Chimie Organique, Université de Yaoundé I B.P. 812 Yaounde, Cameroon; *Department of Chemistry, The University of Glasgow, Glasgow, Glasgow, Glasgow, Glasgow, Glasgow, Glasgow, U.K.

(Received in revised form 27 October 1994)

Key Word Index—Guibourtia tessmannii; Cesalpiniaceae; stem bark; chemistry; stilbene glycoside.

Abstract—(E)-3,4'-Dimethoxy-5-rutinosyl stilbene, rhaponticin and piceid have been isolated from the stem bark of Guibourtia tessmannii.

INTRODUCTION

Guibourtia tessmannii (Harms Leonard), a plant which is used as a source of commercial timber (Bubinga), is also an important medicinal plant in the central and southern provinces of Cameroon. Its preparations are used for the treatment of hypertension, the prevention of abortion and as an anti-helmintic.

A preliminary pharmacological screening of an extract of this plant revealed the presence of an interesting antifungal activity [1].

Earlier chemical work on this plant afforded two flavanols, leucofisetinidin and guibourtacacidin, tannins and sugars [2, 3]. Studies on other *Guibourtia* species have resulted in the isolation of many stilbene glycosides [4-6]. (E)-3,4'-Dimethoxy-5-rutinosyl stilbene was identified in *G. coleosperma* and was characterized as its acetate [6]. This report describes the isolation and characterization of (E)-3,4'-dimethoxyl-5-rutinosyl stilbene and other stilbenes from *G. tesmanii*.

RESULTS AND DISCUSSION

From the acetone extract of the stem bark, three compounds, 1-3, were separated by a combination of different chromatographic techniques.

Compound 1, a glycoside, gave a positive Molish test. Hydrolysis of 1 afforded an aglycone 1a and two sugars characterized as D-glucose and L-rhamnose by chromatography.

The aglycone 1a was shown to be a stilbene from the following information. The IR spectrum showed absorptions at 3390 (OH), 1620 (C=C), 1600 and 1520 (benzene ring) cm⁻¹. Its ¹H NMR spectrum (90 MHz, CDCl₃) showed two *trans* olefinic protons appearing as an AB quartet at $\delta 6.83$ (1H, J=16 Hz) and 7.14 (1H, J=16 Hz). The occurrence of two separate aromatic rings was established by the presence of signals for three protons at $\delta 6.47$ (1H, t, J=2.3 Hz) and 6.76 (2H, d, J=8.7 Hz) indicating a 1,3,5-trisubstituted benzene ring

and an AA'BB' type system at $\delta 6.89$ (2H, J=8.7 Hz) and 7.43 (2H, J=8.7 Hz) for a 1,4-disubstituted benzene ring. The spectrum also contained two methoxy signals at $\delta 3.95$ (3H, s, OMe) and 3.98 (3H, s, OMe). From this evidence, the partial structure 1a was assumed to be a stilbene bearing two methoxy groups and a hydroxy function. The EI-mass spectrum (70 eV) showed positive ions at m/z 256 [M]⁺ (100) and 195 [M - 2 × OMe]⁺ (12). The position of each of these groups in the stilbene structure was assigned by comparison of the ¹³C NMR signals of 1a with those of a similar compound [7]. Thus the chemical shifts at C-5 ($\delta 161.1$) and C-3 ($\delta 156.9$) established that the methoxy and hydroxy groups were on the same benzene ring, and that the second methoxy group was on the other ring. Consequently, the aglycone

	\mathbf{R}^{1}	\mathbb{R}^2	\mathbb{R}^3	R^4
1	Me	rutinosyl	Me	H
1a	Me	Н	Me	Н
1b	Me	hexa-acetoxy rutinosyl	Me	Н
2	H	glucosyl	Me	OH
2a	Н	Н	Me	OH
3	Н	glucosyl	Н	Н
3a	Мe	glucosyl	Me	Н

1a was shown to be (E)-3,4'-dimethoxy-5-hydroxy stilbene. The EI-mass spectrum (70 eV) of 1 exhibited a molecular ion at m/z 564 [M]⁺. This was followed by two significant ions at m/z 418 [M – 148]⁺ (2.1) and 256 [M – 308]⁺ (100) due to successive elimination from the molecular ion of a rhamnosyl and a glucosyl moiety. The link between the rhamnosyl and glucosyl portions was deduced from an examination of the ¹³C NMR spectrum (25.15 MHz) which showed signal patterns similar to those of rutinose [8].

Thus 1 was found to be (E)-3,4'-dimethoxy-5-rutinosyl stilbene. The configuration at the anomeric carbons was found to be β for the glucosyl and α for the rhamnosyl moieties from the ¹H NMR spectrum of 1.

Acetylation of 1 yielded a peracetate (1b) which was identical with the one reported in the literature [6].

Compound 2, $C_{21}H_{24}O_9$, gave a positive Molish test for a glycoside. Acid hydrolysis of 2 afforded an aglycone (2a) which was identified as rhapontigenin [7]. Comparison of the physical and spectral data of 2 with those described in the literature [7] showed that 2 is rhaponticin.

Compound 3, $C_{20}H_{22}O_8$, gave a positive Molish test for a glycoside. The ¹H NMR spectrum of 3 was similar to that of piceid [9]. An attempt to prepare the known derivative of piceid [4'-methoxy piceid (desoxyrhaponticin)] gave the dimethylated derivative 3,4'-dimethoxy piceid ($C_{22}H_{26}O_8$).

EXPERIMENTAL

Plant material. Stem bark of Guibourtia tessmannii was collected from Eseka (Centre Province of Cameroon). It was identified by the National Herbarium of Cameroon where a sample is deposited.

General. Mps: uncorr.; ¹H and ¹³C NMR: CD₃OD or CDCl₃ or DMSO with TMS as int. standard; flash chromatography: silica gel 60 H, spots detected by UV fluorescence and by spraying with H₂SO₄ and heating at 100° for 5–10 min, or by I₂ vapour; TLC: CHCl₃–MeOH (9:1 and 4:1).

Extraction and isolation. Pulverized stem bark (1 kg) of G. tessmannii was defatted with hexane and then extracted successively with Me₂CO and MeOH. The Me₂CO extract, after removal of solvent, was subjected to flash chromatography over silica gel using CHCl₃ and MeOH as solvents.

Two main frs were obtained. The first contained a viscous residue which was discarded and the second contained the mixt. of stilbenes (7.2 g). Crystallization of this mixt. in MeOH afforded a crystalline mixt. of 1-3. Three g of a mixt. of 1-3 was purified by prep. TLC (CHCl₃-MeOH, 4:1).

Compound 1. Crystals (EtOH) (360 mg) mp $142-143^{\circ}$. Found C 59.57, H 6.38 required for C₂₈H₃₆O₁₂: C 59.5, H 6.4% IR v_{max} (KBr) cm⁻¹: 3400 (OH), 3050, 1620 (C=C), 1600, 1590, 1510 (benzene ring); ¹H NMR (CD₃OD, 300 MHz): δ 1.2 (3H, d, J = 7.4 Hz, Me rhamnose), 3.76 (3H, s, OMe), 3.79 (3H, s, OMe), 3.29–3.41 (m,

CH sugars), 4.7 (1H, d, J = 1.6 Hz anomeric proton of α -L rhamnose), 4.9 (1H, d, J = 7.5 Hz anomeric proton of β -D-glucose), 6.58 (1H, t, J = 2.6 Hz, H-4), 6.75 (1H, t, J = 1.6 Hz, H-2), 6.82 (1H, t, J = 1.6 Hz, H-6), 6.87 and 7.04 (2H, ABq, J = 16 Hz, trans olefinic proton), 6.88 and 7.42 (4H, AA'BB', J = 8.7 Hz, H-2', H-3', H-5', H-6'); 13 C NMR: Table 1.

Acid hydrolysis of 1. Compound 1 (200 mg) was mixed with 50 ml 0.4 M HCl in MeOH and refluxed for 9 hr in a water bath. The solvent was reduced by evapn and upon cooling, crystals were obtained. Further crystallization from H₂O afforded the aglycone 1a (32 mg). Compound 1a: Found C 75, H 6.25, required for C₁₆H₁₆O₃: C 74.9, H 6.29%; crystals, mp 119–120°; IR v_{max} (KBr) cm⁻¹: 3390 (OH), 1620 (C=C), 1600, 1580, 1520 (benzene ring); ¹H NMR (90 MHz, CDCl₃): δ 3.95 (3H, s, OMe), 3.98 (3H, s, OMe), 6.47 (1H, t, J = 2.3 Hz, H-4), 6.76 (2H, d, J = 2.3 Hz, H-2, H-6), 6.83 and 7.14 (2H, ABq, J = 16 Hz, olefinic protons), 6.89 and 7.43 (4H, AA'BB', J = 8.0 Hz, H-2', H-3', H-5', H-6'); ¹³C NMR: Table 1.

Acetylation of 1. Compound 1 (100 mg) was left overnight in a mixt. of 5 ml pyridine and 5 ml Ac₂O and then worked up in the usual way. Purification by prep. TLC with CHCl₃ as solvent yielded a peracetate (1b) (88 mg).

Compound 1b was identified as (E)-3,4'-dimethoxy-5 [2,3,4-tri-O-acetyl-6-O-(2,3,4-tri-O-acetyl-o'-L-rhamnosyl)- β -D-glucopyranosyloxy]stilbene by comparison of its 1 H NMR, 13 C NMR and MS data with those reported in the lit. [6].

Compound 2. Rhaponticin (300 mg) crystals (EtOH) mp 241–242° (lit. [7] 246–248°). Found C 60, H 5.71 calc. for $C_{21}H_{24}O_9$: C 59.9, H 5.75%. IR v_{max} (KBr) cm⁻¹: 3400 (OH), 1620 (C=C-Ar), 1580, 1510 (benzene ring); ¹H NMR (DMSO- d_6 , 90 MHz): δ3.25–3.35 (6H, m, 4H and –CH₂), 3.76 (3H, s, OMe), 4.85 (1H, d, J = 6 Hz, anomeric proton), 5–5.3 (4H, br s, OH glucose), 6.34 (1H, br s, H-4), 6.55 (1H, br s, H-2), 6.75 (1H, br s, H-6), 6.8–7.5 (5H, m, H-2′, H-3′, H-5′, H-α and H-β), 8.9 (1H, s, OH phenolic), 9.35 (1H, s, OH phenolic).

Acid hydrolysis of **2**. Compound **2** (200 mg) was hydrolysed as previously described. The residue was purified over silica gel with CHCl₃–MeOH (19:1) and (9:1). Frs of 25 ml were collected. Frs 30–36 deposited, after evapn, a powder (40 mg) of rhapontigenin **2a** mp 196–197°. Found C 69.7, H 5.42 calc. for $C_{15}H_{14}O_4$: C 69.8, H 5.46%. ¹H NMR (300 MHz, CD₃OD): δ 3.75 (3H, s, OMe), 6.15 (1H, t, J = 2.1 Hz, H-4), 6.42 (2H, d, J = 2.1 Hz, H-2, H-6), 6.71 (1H, d, J = 8 Hz, H-5'), 6.83

	-

C	1	1a	1 b	1 b [6]	2	2a	3	3a
1	140.9 s	140.1	140.1	140.5	139.2	139.7	139.9 s	139.4 s
2	100.3 d	105.8	106.5	108.2	105	105.0	106.0 d	105.8 a
3	162.0 s	156.9	161.0	158.7	158.3	157.3	158.4 s	159.1
4	103.2 d	100.7	102.5	102.6	103	102.0	102.6 d	101.7
5	160.0 s	161.1	158.2	161.6	158.8	157.3	157.7 s	159.0
6	108.6 d	104.8	107.9	106.8	107.3	105.3	107.3 d	106.5
OMe	55.9 q	55.4	55.4	55.2	55.5	55.2		55.3
OMe	55.9 q	55.4	55.4	54.8		<u> </u>		55.2
α	127.1 d	126.3	126.1	126.6	126.1	126.8	125.2	126.1
β	129.8 d	129.0	129.4	129.7	128.6	126.5	128.7	128.8
C=O			170.3(1C) 170.1(1C)	169.9				
			169.8(2C)					
17	120.8	120.2	169.5(2C)	120.3	120.0	120.0	130.7	129.6
1'	130.8 s	130.2	129.9	130.2	129.9	130.9	128.7 127.6	129.0
2' 3'	126.1 d	127.9	128.0	128.3	112.3	112.2		114.3
	114.9 d	114.1	114.0	114.5	146.5	145.7	115.2	160.5
4'	160.9 s	158.4	159.0	160.0	147.7	147.0	156.5	
5' 6'	114.9 d	114.1	114.0	114.5 128.3	112.3	111.0	115.2	114.3
o Me C=0	126.1 d	127.8	128.0		118.6	119.0	127.6	128.0
			20.7	20.3				
Glucose	101.0.1		01.1	00.0	100 (100 (1	100.0
1	101.8 d		91.1	98.9	100.6		100.6 d	100.8
2 3	73.8 d		73.2	71.8	73.3		73.1 d	73.4 <i>a</i>
4	77.6 d		78.3	73.3	77.0		76.2 d 69.7 d	79.9
5	71.1 d		71.4	69.9	69.8		76.2 d	76.9
5 6	76.6 d		77.0 66.7	73.4	76.7 60.7		61.3 t	60.9
Rhamnose	67.5 t		00.7	67.1	OU. /		01.57	60.9
	102.2		000	98.4				
1	102.2 d		98.0					
2 3	72.1 d		72.8	69.9				
	71.8 d		71.0	69.8				
4 5	74.7 d		75.8	71.2				
	69.5 d		69.5	67.3				
6	17.8 q		17.3	17.7				

(1H, dd, J = 2 Hz, J = 8 Hz, H-6'), 6.96 (1H, d, J = 2 Hz, H-2'), 6.70 and 6.82 (1H each, d, J = 16 Hz trans olefinic protons); ¹³C NMR: Table 1.

Compound 3. Piceid (446 mg): Found C 61.53, H 5.64 calc. for $\rm C_{20}H_{22}O_8$: C 61.53, H 5.68%. Pale yellow crystals, mp 220° (lit. 225° [9]). ¹H NMR (CD₃OD and CDCl₃, 200 MHz): δ 3.17–3.6 (6H, m, 4CH and CH₂ glucose), 4.7 (1H, d, J=5 Hz, anomeric proton), 6.27 (1H, dd, J=2 Hz, J=1.8 Hz, H-4), 6.44 (1H, dd, J=2 Hz, J=1.8 Hz, H-2), 6.5 (1H, dd, J=2 Hz, J=1.8 Hz, H-6), 6.54 and 6.64 (1H each, d, J=18 Hz, d1 trans olefinic protons), 6.58 and 7.13 (4H, AA'BB' type signal, H-2', H-3', H-5', H-6').

Methylation of 3. Compound 3 (70 mg) was dissolved in 50 ml dry Me₂CO. MeI (3 ml) was added together with 3 g K₂CO₃ then refluxed for 5 hr. The reaction mixt. was cooled and extracted with EtOAc. The residue (1 g) from the EtOAc extract after further purification by prep. TLC using CHCl₃-MeOH (4:1) as solvent gave 3a (48.5 mg) as yellow crystals, mp 174-175°. Found C 63.15, H 6.22 required for $C_{22}H_{26}O_8$: C 63.1, H 6.26%. ¹H NMR (200 MHz, DMSO- d_6): δ 3.1-3.6 (6H, m, 4H,

CH₂), 3.75 (3H, s, OMe), 3.76 (3H, s, OMe), 4.7 (1H, br s, OH), 4.9 (1H, d, J = 7 Hz anomeric proton), 5.15 (1H, br s, OH), 5.17 (1H, br s, OH), 5.45 (1H, br s, OH), 6.51 (1H, dd, J = 1.7 Hz, J = 2 Hz, H-4), 6.8 (1H, dd, J = 1.7 Hz, J = 2 Hz, H-6), 6.93 and 7.52 (4H, AA'BB' type signal, J = 8 Hz, H-2', H-3', H-5', H-6'), 7.0 and 7.21 (2H, ABq, J = 18 Hz, H_a and H_B); 13 C NMR: Table 1.

Acknowledgements—We are grateful to Mr B. Hagbe, CEPM, Manga Ondoa for the collection and identification of the plant, and to Mballa Madeleine, Ondoa Theophile and Azeme Emile of Eben village for information about its medicinal uses.

REFERENCES

- Ohigashi, H., Kaji, M., Jato, J., Hoshino, J. and Koshi, M. (1985) Depart. Food Sciences and Technology, Faculty of Agriculture Kyoto University.
- 2. Roux, D. G. (1959) Nature 183, 890.

- Drewes, S. G. and Roux, D. G. (1965) Biochem. J. 96, 689
- 4. Steynberg, J. P., Ferreira, D. and Roux, D. G. (1983) Tetrahedron Letters 24, 4147.
- 5. Steynberg, J. P., Ferreira, D. and Roux, D. G. (1987) J. Chem. Soc. Perkin Trans 1 1705.
- 6. Steynberg, J. P., Brant, E. V., Burger, J. F. W.,
- Bezuidenhoudt, B. C. B. and Ferreira, D. (1988) J. Chem. Soc. Perkin Trans 1 37.
- Kashimada, Y., Nonaka, G. I. and Nishioka, I. (1984) Chem. Pharm. Bull. 32, 3509.
- 8. Borel, C., Gupta, M. P. and Hostettmann, K. (1987) Phytochemistry 26, 2685.
- 9. Hillis, W. E. and Ishikura (1968) J. Chromat. 32, 323.